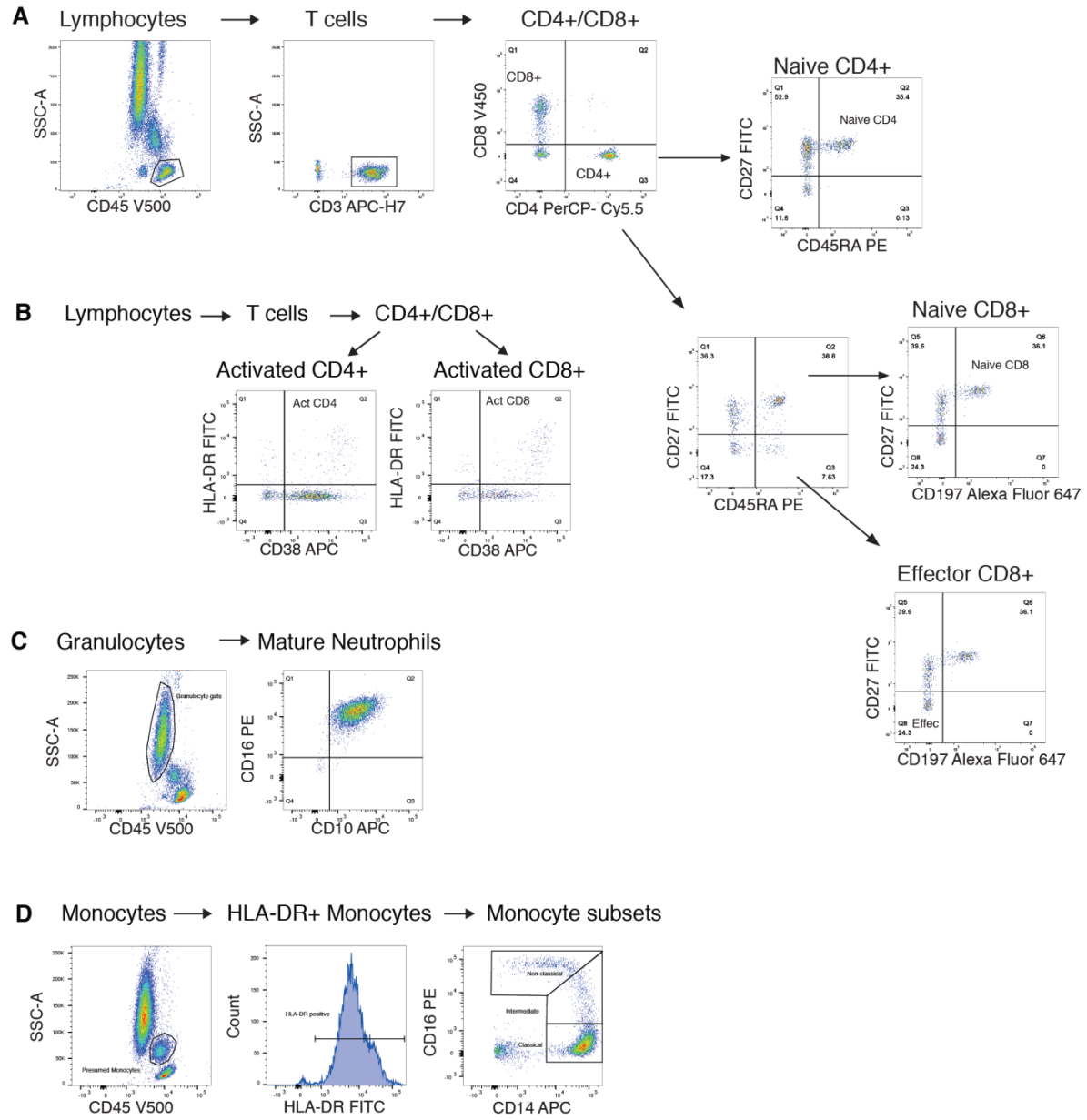
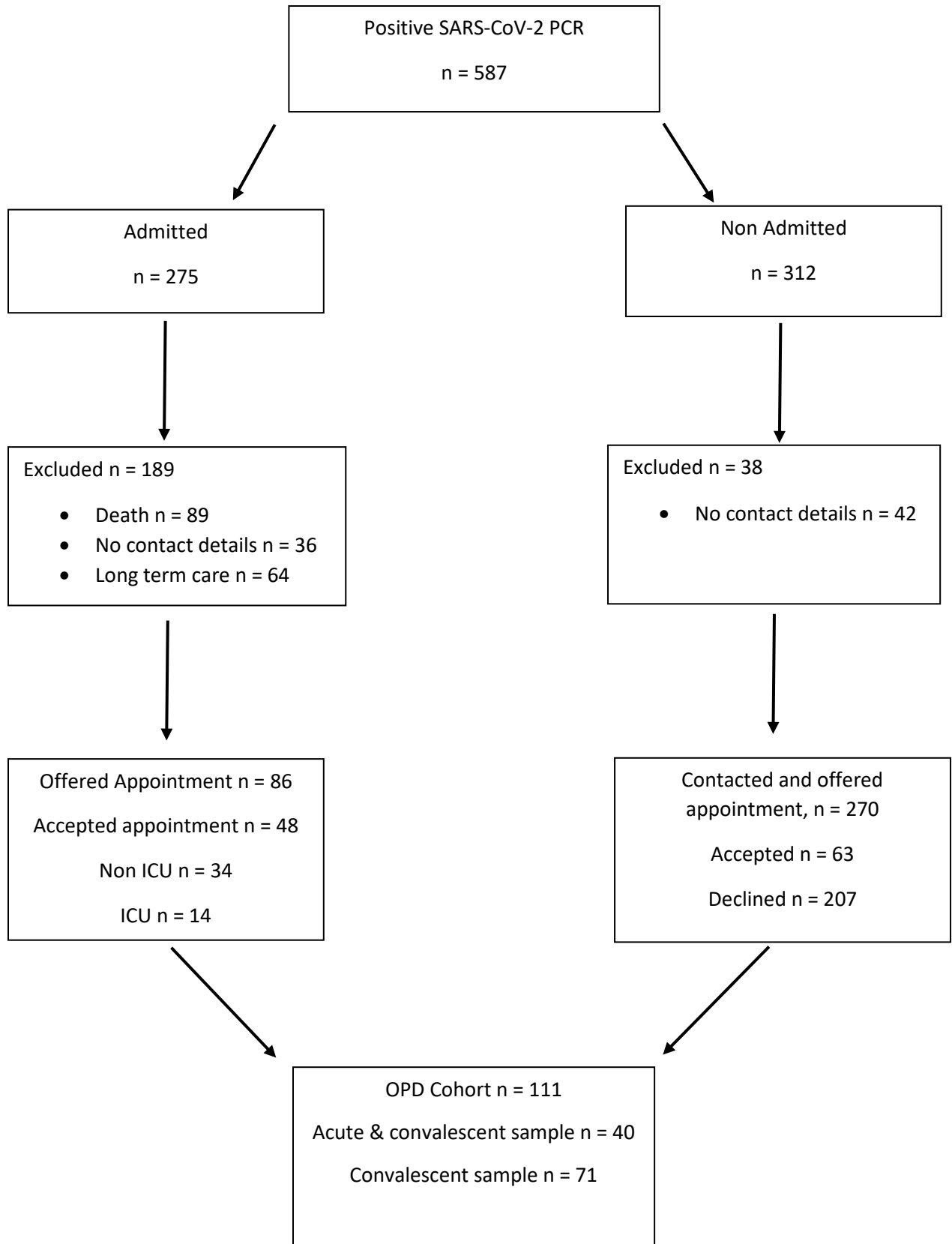


**Supplemental Figure 1: Flow cytometry gating strategy.**

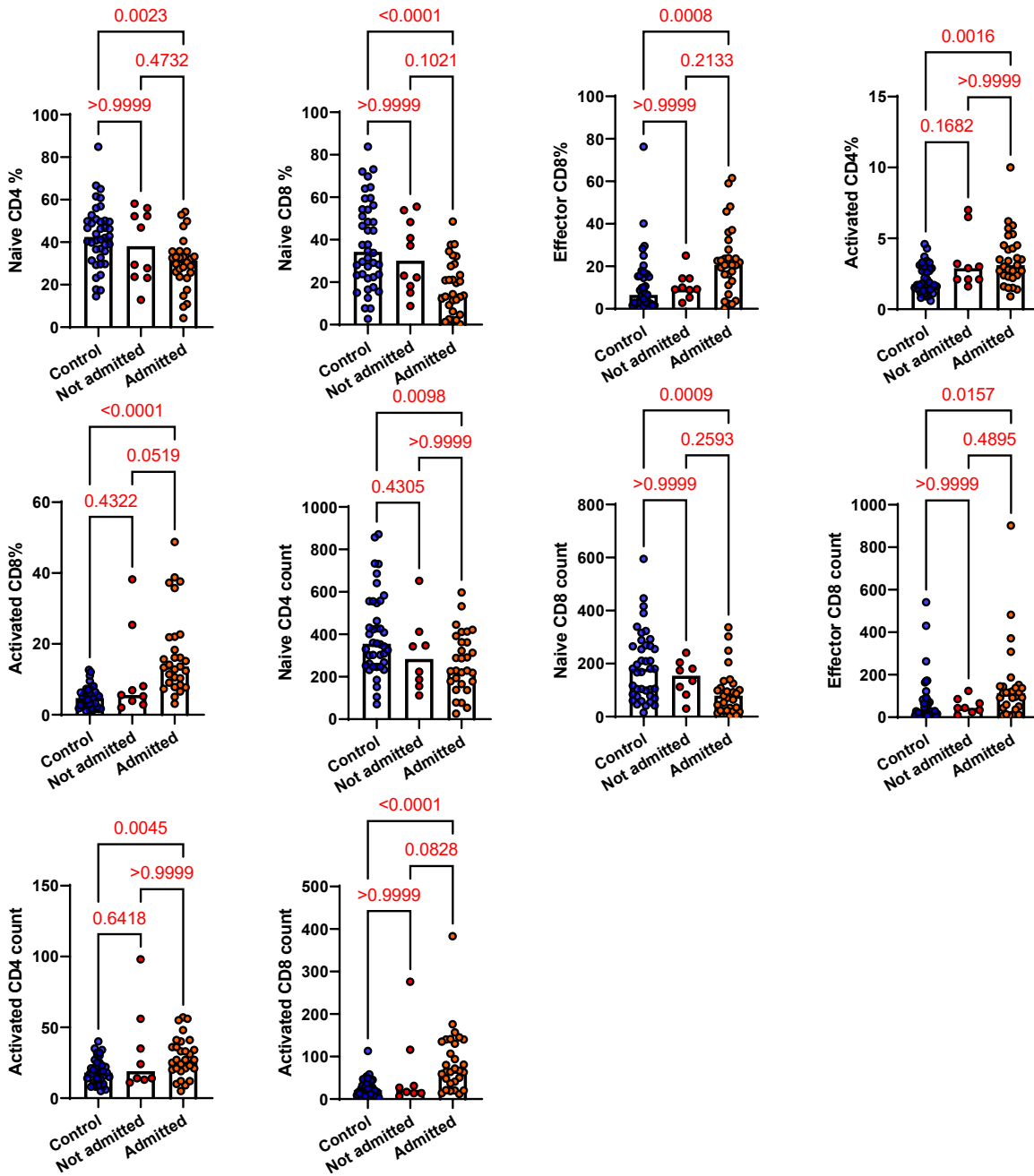


**Supplemental Figure 1: Gating strategy for flow cytometry shown (A) naïve and effector T lymphocytes (B) activated T lymphocytes (C) neutrophils (D) monocytes**

**Supplemental Figure 2: Patient enrolment diagram**

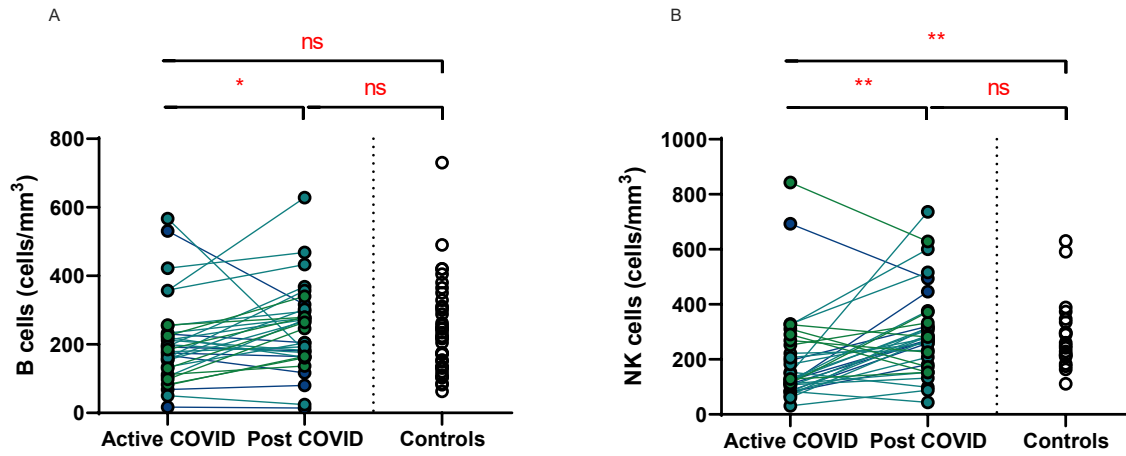


**Supplemental Figure 3: Lymphoid subsets in the matched cohort at 68 days post-infection versus controls, stratified by need for admission during acute infection**

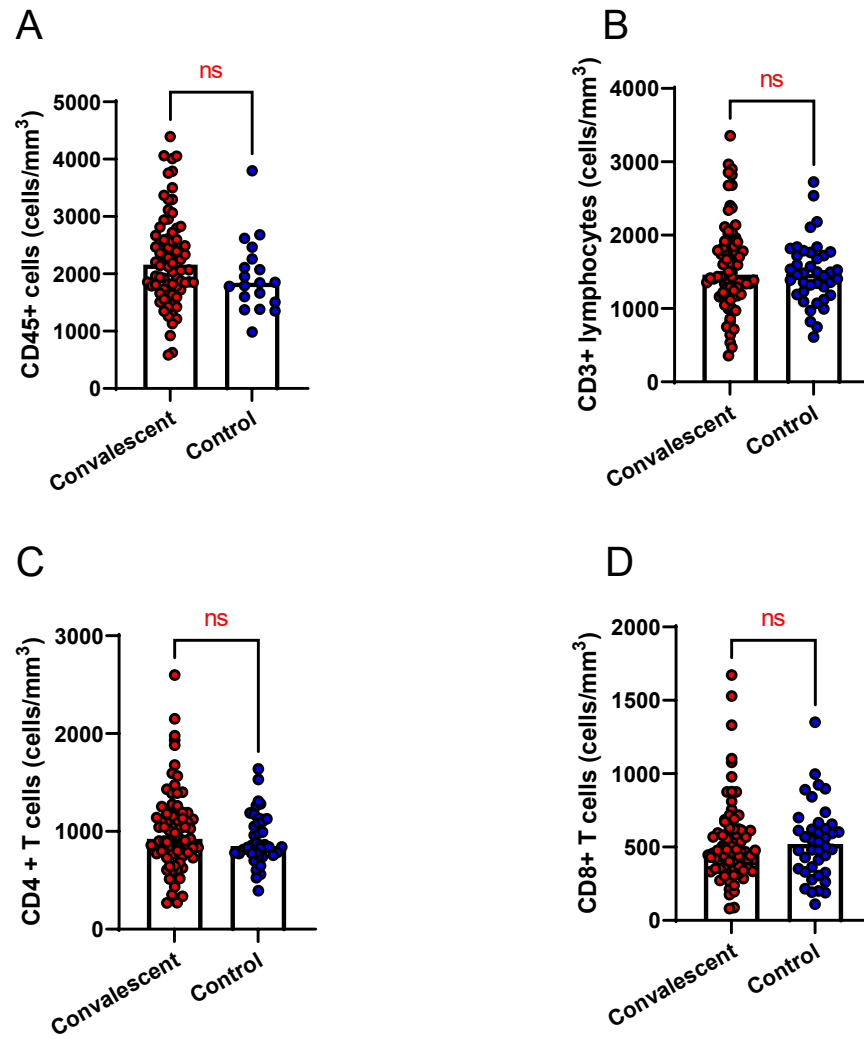


**Supplemental Figure 3:** Lymphocyte subpopulations in n=40 convalescent COVID patients, divided by need for admission during acute infection, in comparison to n=40 controls. Proportions and absolute counts of cell populations shown. Kruskal-Wallis test with Dunn’s multiple comparison post-hoc test used to determine between-group differences.

## Supplemental Figure 4: Recovery of B and NK cells

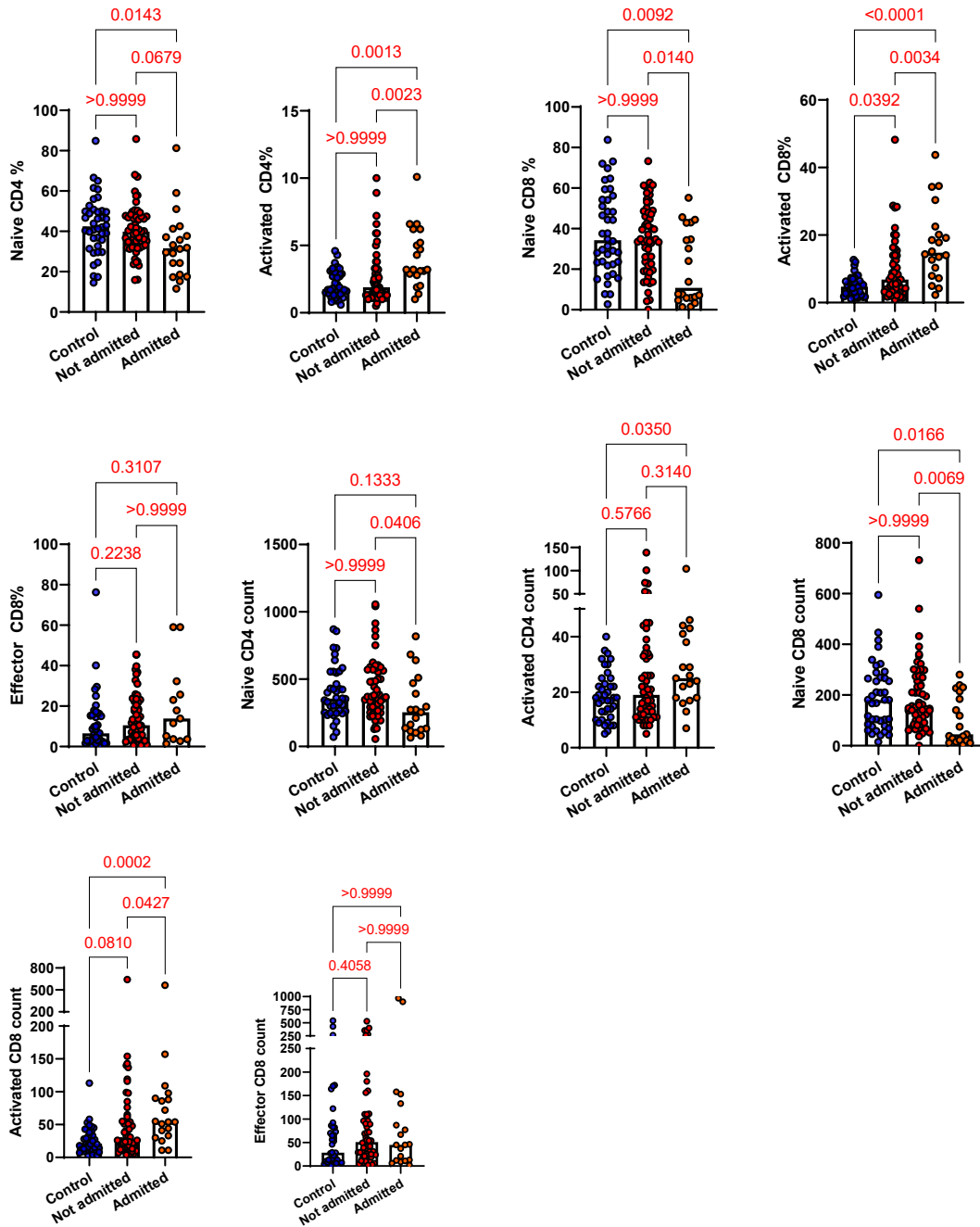


**Supplemental Figure 5: Major lymphoid populations in convalescent COVID-19.**



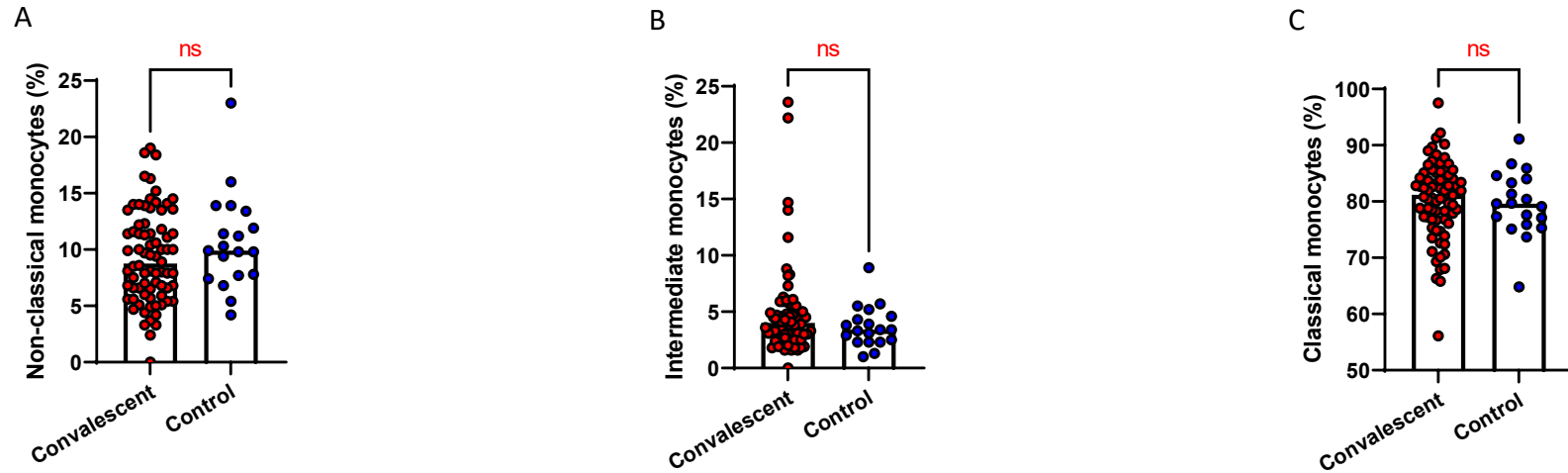
**Supplemental Figure 5:** Major lymphoid populations in n=71 convalescent COVID patients in comparison to n=40 controls **(A)** CD45+ cells **(B)** CD3+ lymphocytes **(C)** CD4+ T cells **(D)** CD8+ T cells. Wilcoxon rank-sum test ns=not significant

**Supplemental Figure 6: Lymphoid subsets at convalescence, stratified by need for admission during acute infection**



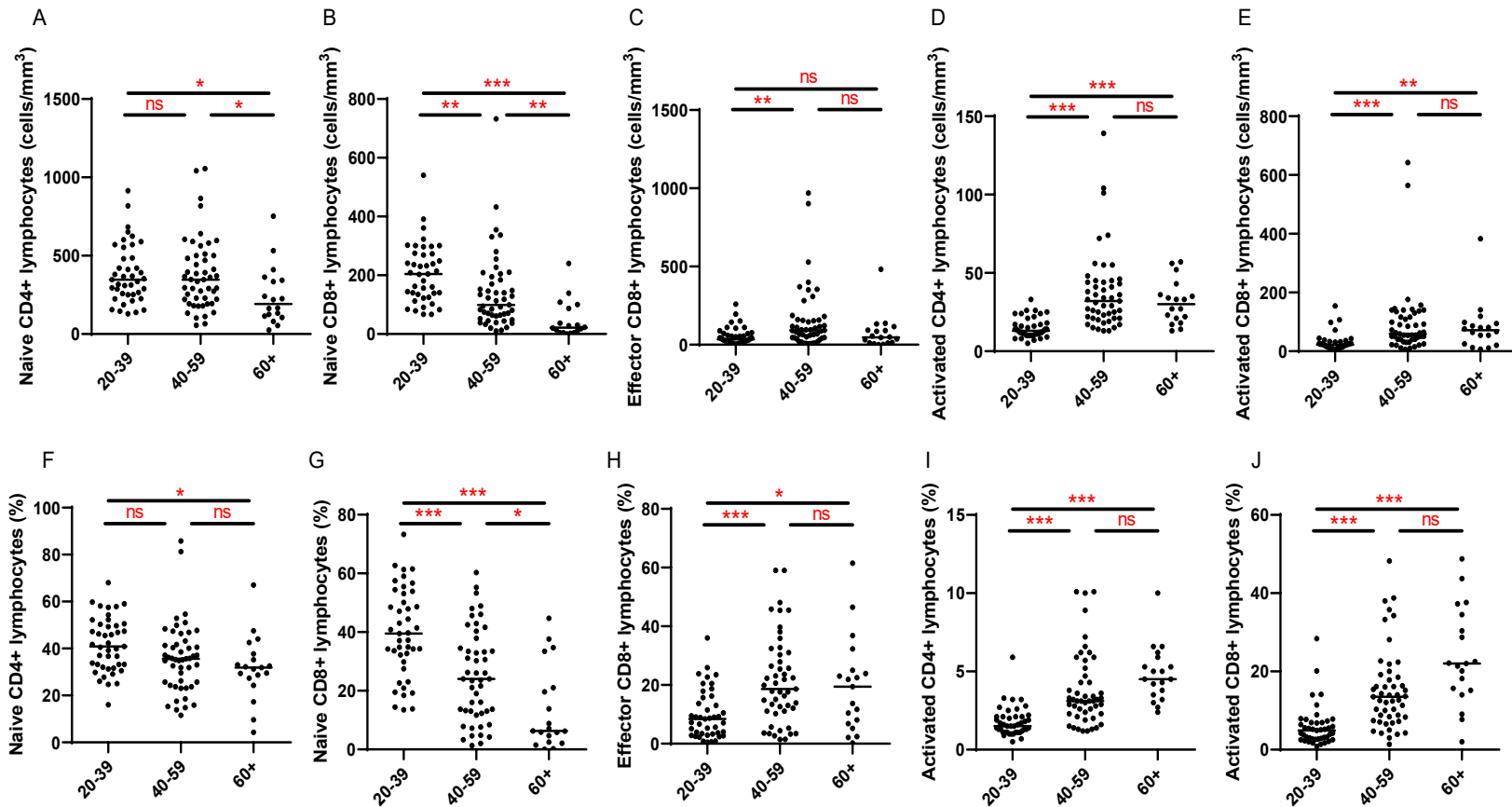
**Supplemental Figure 6:** Lymphocyte subpopulations in n=71 convalescent COVID patients, divided by need for admission during acute infection, in comparison to n=40 controls. Proportions and absolute counts of cell populations shown. Kruskal-Wallis test with Dunn's multiple comparison post-hoc test used to determine between-group differences.

**Supplemental Figure 7: Monocyte subpopulations in convalescent COVID-19.**



**Supplemental Figure 7:** Monocyte subpopulations in n=71 convalescent COVID patients in comparison to n=20 controls **(A)** non-classical monocytes **(B)** intermediate monocytes **(C)** classical monocytes. Wilcoxon rank-sum test. ns=not significant

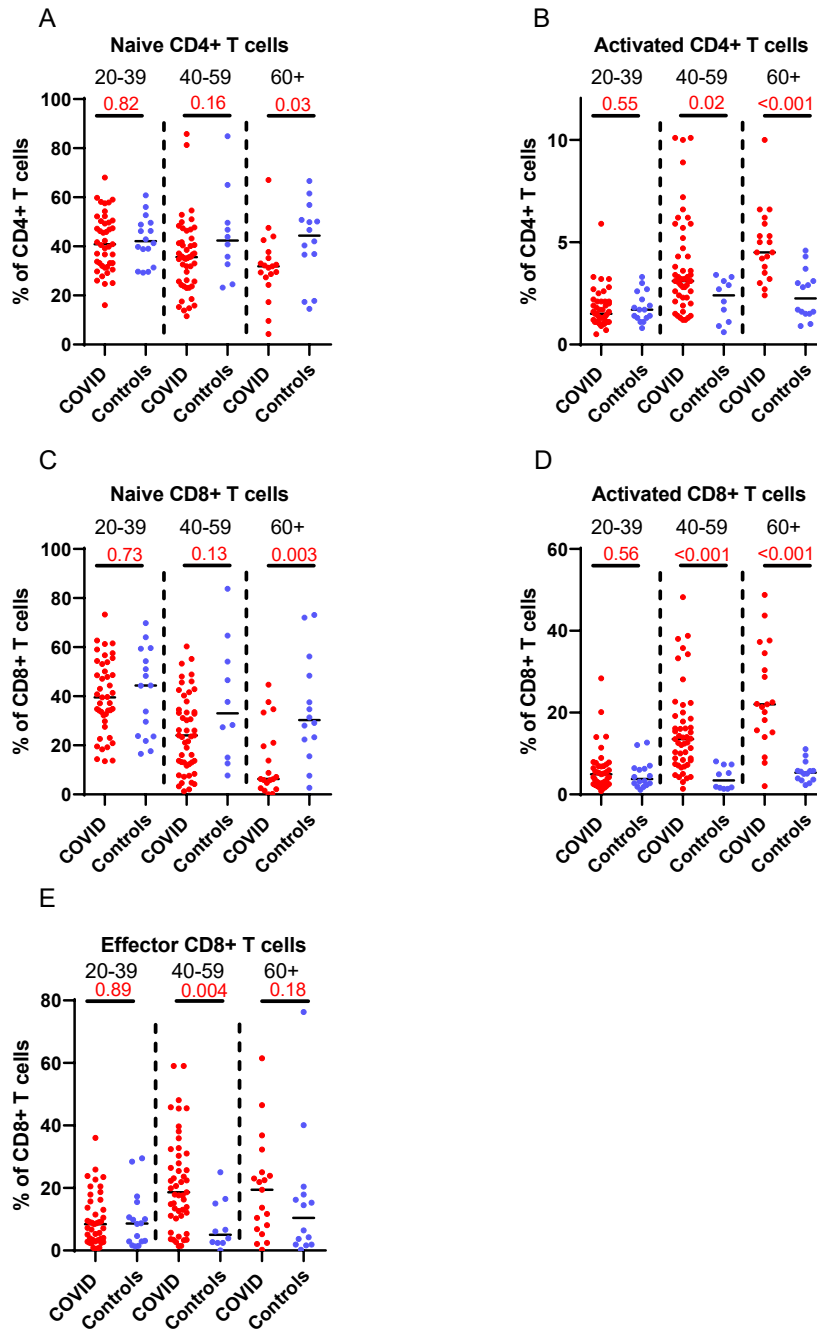
**Supplemental Figure 8: Age-associated changes in convalescent lymphocyte subsets**



**Supplemental Figure 8:** Lymphocyte immunophenotyping of convalescent COVID patients (n=111) broken down by age. **(A)** naïve CD4+ T cell count **(B)** naïve CD8+ T cell count **(C)** effector CD8+ T cell count **(D)** activated CD4+ T cell count **(E)** activated CD8+ T cell count **(F)** naïve CD4+ T cell proportion **(G)** naïve CD8+ T cell proportion **(H)** effector CD8+ T cell proportion **(I)** activated CD4+ T cell proportion **(J)** activated CD8+ T cell proportion. Kruskal-Wallis test with post-hoc Dunn test. \* p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001, ns = Not Significant



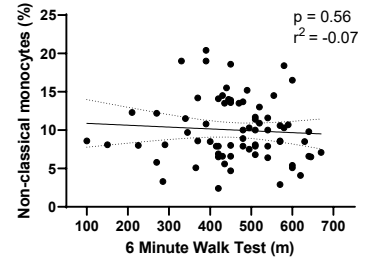
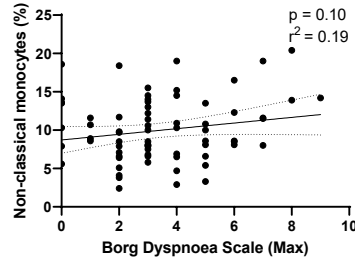
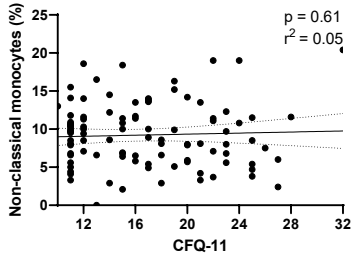
**Supplemental Figure 9: Age-associated changes in convalescent lymphocyte subset proportions versus age-matched controls**



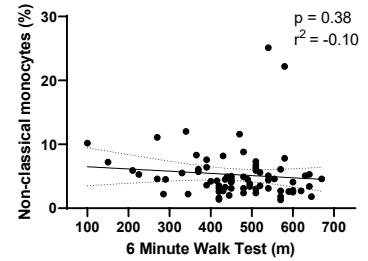
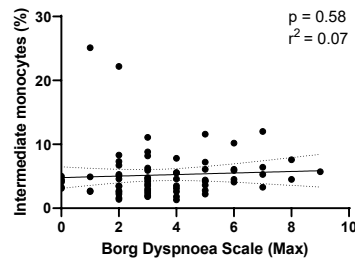
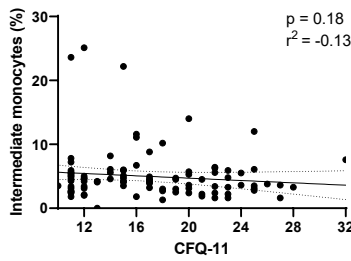
**Supplemental Figure 9:** Lymphocyte immunophenotyping of convalescent COVID patients (n=111) broken down by age with age-matched controls, showing proportions of naïve (A) and activated (B) CD4+ T cells, and naïve (C), activated (D) and effector (E) CD8+ T cells. Differences assessed by Wilcoxon rank-sum test. Bonferroni correction, significance  $p < 0.02$

## Supplemental Figure 10: Relationships of monocyte subsets with physical health measures

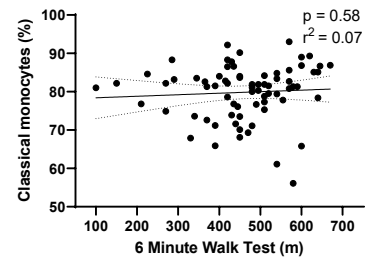
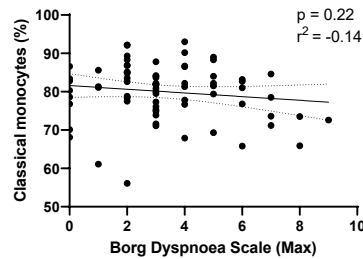
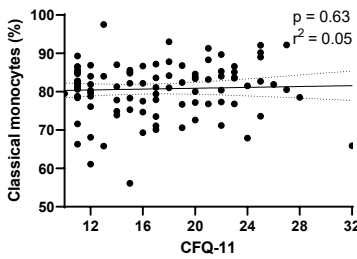
### A. Non-classical monocytes



### B. Intermediate monocytes



### C. Classical monocytes



**Supplemental Figure 10:** Relationship with fatigue, perceived exertion and 6MWT distance in n=101 convalescent COVID patients with (A) non-classical monocytes (B) intermediate monocytes (C) classical monocytes. Correlation assessed with Pearson's chi-squared test. CFQ-11 = Chalder Fatigue Questionnaire-11

**Supplemental Table 1: Flow cytometry antibodies**

Antibody	Fluorophore	Clone	Company	Catalogue #
CD45	V500-C	2D1	BD Biosciences	655873
CD8	V450	RPA-T8	BD Biosciences	560347
CD3	APC-H7	SK7	BD Biosciences	641415
CD4	PerCP/CY5.5	Sk3	BD Biosciences	332772
CD45RA	PE	-	BD Biosciences	556627
CD27	FITC	-	BD Biosciences	555440
CD197	Alexa Fluor 647	150503	BD Biosciences	560816
HLA DR	FITC	L243	BD Biosciences	347400
CD38	APC	HB-7	BD Biosciences	345807
CD14	APC	MΦP9	BD Biosciences	345787
CD16	PE	B73.1	BD Biosciences	332779
CD10	APC	HI10a	BD Biosciences	332777