

Supplementary Information

Hanley et al, 2021

Supplementary Table 1. Subset demographics.

Supplementary Table 2. List of reagents used.

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Supplementary Table 1. Subset demographics.

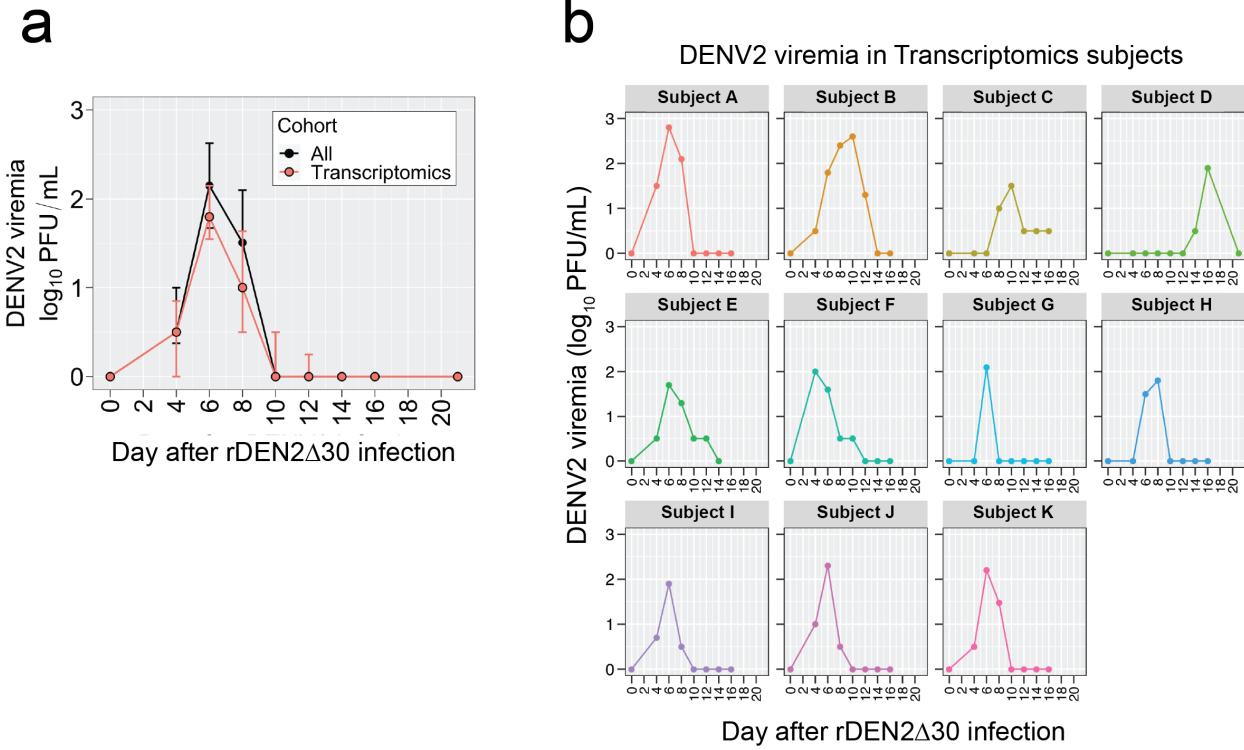
| | <i>Parent study</i> (n = 20) | <i>Gene expression subset</i> (n = 11) |
|------------------|---------------------------------|--|
| <i>Female</i> | 6 (30%) | 2 (18%) |
| <i>Male</i> | 14 (70%) | 9 (82%) |
| <i>Black</i> | 9 (45%) | 5 (45%) |
| <i>White</i> | 11 (55%) | 6 (55%) |
| <i>Baltimore</i> | 11 (55%) | 6 (55%) |
| <i>Vermont</i> | 9 (45%) | 5 (45%) |

Supplementary Table 2. List of reagents used.

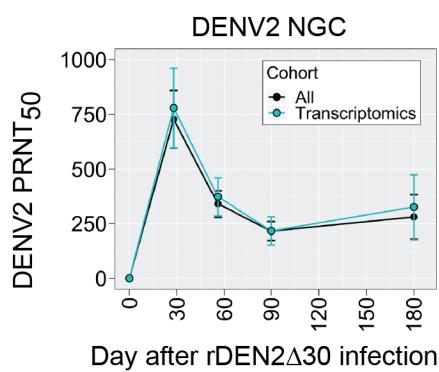
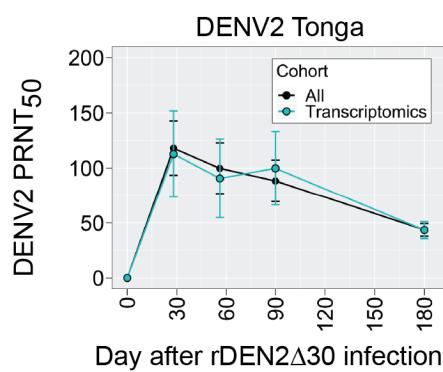
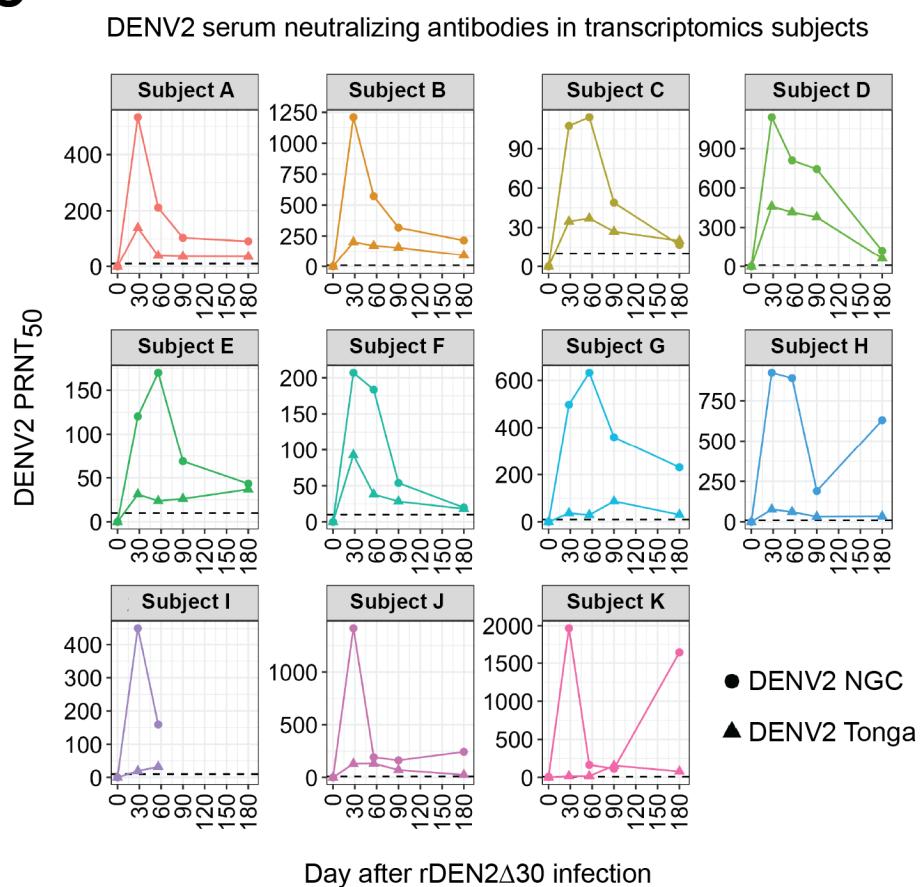
| REAGENT or RESOURCE | SOURCE | IDENTIFIER |
|---|---------------------------|------------------------------|
| Antibodies | | |
| Anti-human CD3 (UCHT1) FITC-conjugated | 0.25 µL/test Biolegend | Cat. #300406 Lot #B279208 |
| Anti-human CD4 (OKT4) BV510-conjugated | 1 µL/test Biolegend | Cat. #317444 Lot #B248141 |
| Anti-human CD8 (RPA-T8) BV650-conjugated | 0.5 µL/test Biolegend | Cat. #301041 Lot #B275821 |
| Anti-human CD14 (M5E2) BV711-conjugated | 1 µL/test Biolegend | Cat. #301837 Lot #B275829 |

| | | | |
|---|--|--------------|--------------------------------|
| Anti-human CD16 (3G8) APC-Cy7-conjugated | 0.25 µL/test | Biolegend | Cat. #302017 Lot #B295391 |
| Anti-human CD19 (HIB19) PE-Dazzle594-conjugated | 0.5 µL/test | Biolegend | Cat. #302252 Lot #B277039 |
| Anti-human CD25 (M-A251) BV421-conjugated | 0.5 µL/test | Biolegend | Cat. #356113 Lot #B301467 |
| Anti-human CD27 (M-T271) PE-Cy7-conjugated | 0.125 µL/test | Biolegend | Cat. #356412 Lot #B279971 |
| Anti-human CD38 (HIT2) Alexa Fluor 647-conjugated | 0.125 µL/test | Biolegend | Cat. #303514 Lot #B233813 |
| Anti-human CD45RA (HI100) BUV395-conjugated | 0.25 µL/test | BD OptiBuild | Cat. #740298 Lot #0293615 |
| Anti-human CD56 (NCAM16.2) BUV563-conjugated | 0.125 µL/test | BD Horizon | Cat. #612929 Lot #0044064 |
| Anti-human CD57 (QA17A04) BV605-conjugated | 0.5 µL/test | Biolegend | Cat. #393303 Lot #270939 |
| Anti-human CD127 (HIL-7R-M21) BUV805-conjugated | 0.25 µL/test | BD OptiBuild | Cat. #748486 Lot #0294340 |
| Anti-human CD134 (ACT35) BUV737-conjugated | 0.125 µL/test | BD OptiBuild | Cat. #749286 Lot #0294339 |
| Anti-human CD154 (24-31) BV785-conjugated | 0.5 µL/test | Biolegend | Cat. #310841 Lot #B264809 |
| Anti-human HLA-DR (L243) BV570-conjugated | 2.5 µL/test | Biolegend | Cat. #307637 Lot #B314475 |
| Anti-human IgM (MHM-88) PerCP-Cy5.5-conjugated | 0.5 µL/test | Biolegend | Cat. #314512 Lot #B231968 |
| Anti-human CCR7 (2-L1-A) APC-R700-conjugated | 0.5 µL/test | BD Horizon | Cat. #566767 Lot #0283646 |
| Anti-human CD279 (EH12.2H7) PE-conjugated | 0.5 µL/test | Biolegend | Cat. #329905 Lot #B252642 |
| Human Trustain FcX blocker | 5 µL/test | Biolegend | Cat. #422302 Lot #B313422 |
| True-Stain Monocyte blocker | 5 µL/test | Biolegend | Cat. #426102 Lot #B311012 |
| Live/Dead Blue Viability Dye | 0.5 µL/test | Invitrogen | Cat. #L23105 A Lot #2214471 |
| Brilliant stain buffer | 50 µL/test | BD Horizon | Cat. #566349 Lot #9192749 |
| Bacterial and Virus Strains | | | |
| Dengue virus serotype 1 (West Pacific 74) WHO reference strain | Stephen Whitehead, National Institute of Allergy and Infectious Disease (NIAID) | | Genbank AY145121 |
| Dengue virus serotype 2 (New Guinea C) | Stephen Whitehead, NIAID | | Genbank AF038403.1 |
| Dengue virus serotype 2 (strain S-16803) WHO reference strain | Aravinda De Silva, University of North Carolina-Chapel Hill (UNC) | | Genbank GU289914 |
| Dengue virus serotype 3 (Sleman/78) | Stephen Whitehead, NIAID | | Genbank AY656169 |

| | | |
|---|---|---|
| Dengue virus serotype 3 (CH53489) WHO reference strain | Aravinda De Silva, UNC | Genbank DQ863638 |
| Dengue virus serotype 4 Dominica/81 | Stephen Whitehead, NIAID | Genbank AY648301 |
| Dengue virus serotype 4 (TVP-376) WHO reference strain | Aravinda De Silva, UNC | Genbank KC963424 |
| Biological Samples | | |
| Human Serum | University of Vermont Vaccine Testing Center and Johns Hopkins Center for Immunization Research | Clinicaltrials.gov identifiers: NCT01072786, NCT02021968 |
| Peripheral blood mononuclear cells (PBMC) | University of Vermont Vaccine Testing Center and Johns Hopkins Center for Immunization Research | Clinicaltrials.gov identifiers: NCT01072786, NCT02021968 |
| Experimental Models: Cell Lines | | |
| African green monkey kidney cells (Vero-81) | Stephen Whitehead, NIAID | RRID:CVC_0059 |



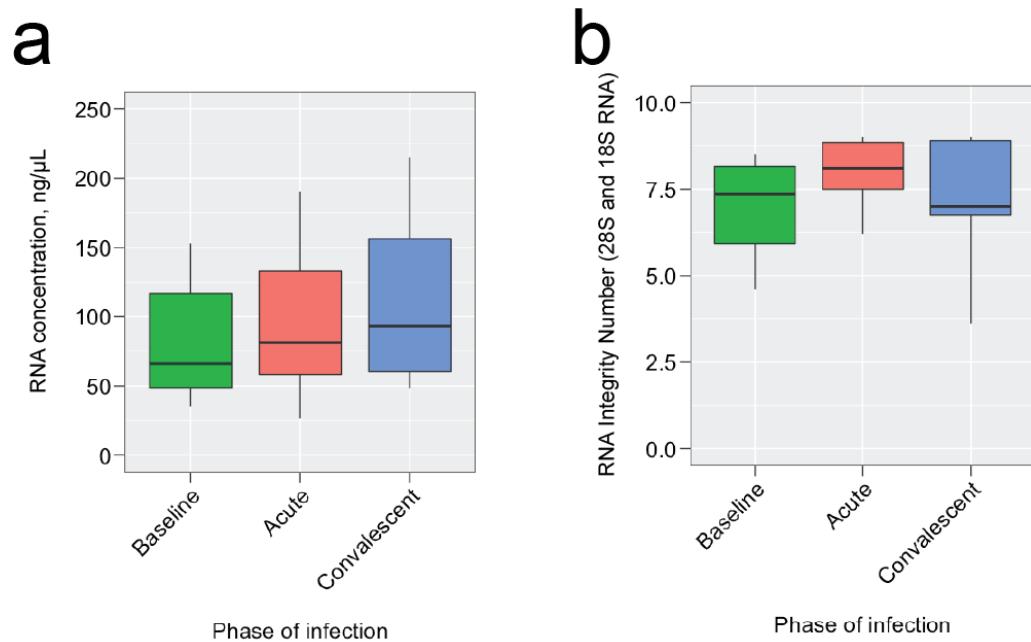
Supplementary Figure 1 | DENV2 viremia after rDEN2Δ30 infection. **(a)** Comparison of DENV2 serum viremia as assessed by plaque assay (expressed as \log_{10} plaque-forming units/mL of serum) in the parent cohort (All, $n = 20$) from which a subset was selected for transcriptomics analysis ($n = 11$). Median \pm standard deviation is plotted. **(b)** Individual viremia profiles for each of the subjects in the transcriptomics analysis.

a**b****c**

Supplementary Figure 2 | DENV2 serum neutralizing antibodies after rDEN2 Δ 30 infection.

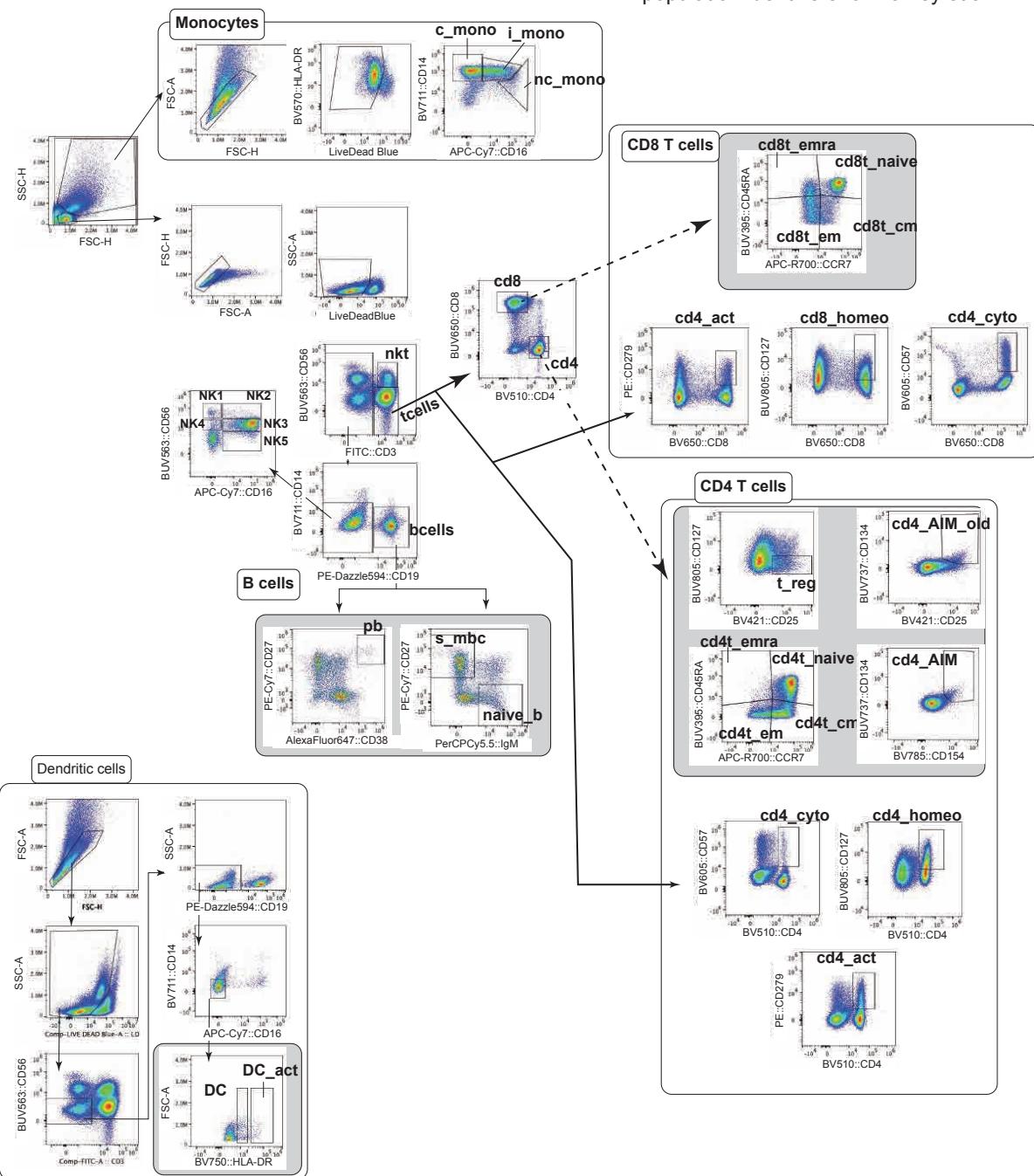
(a) Comparison of DENV2 serum neutralizing antibodies as assessed by plaque reduction neutralization titer 50% (PRNT $_{50}$) to (a) DEN2 NGC (b) DEN2 Tonga in the parent cohort (All,

$n = 20$) and the subset selected for transcriptomics analysis ($n = 11$). Mean \pm standard deviation values are shown. (c) Individual PRNT₅₀ profiles for DEN2-NGC (circles) and -Tonga (triangles) for each of the subjects in the transcriptomics analysis.



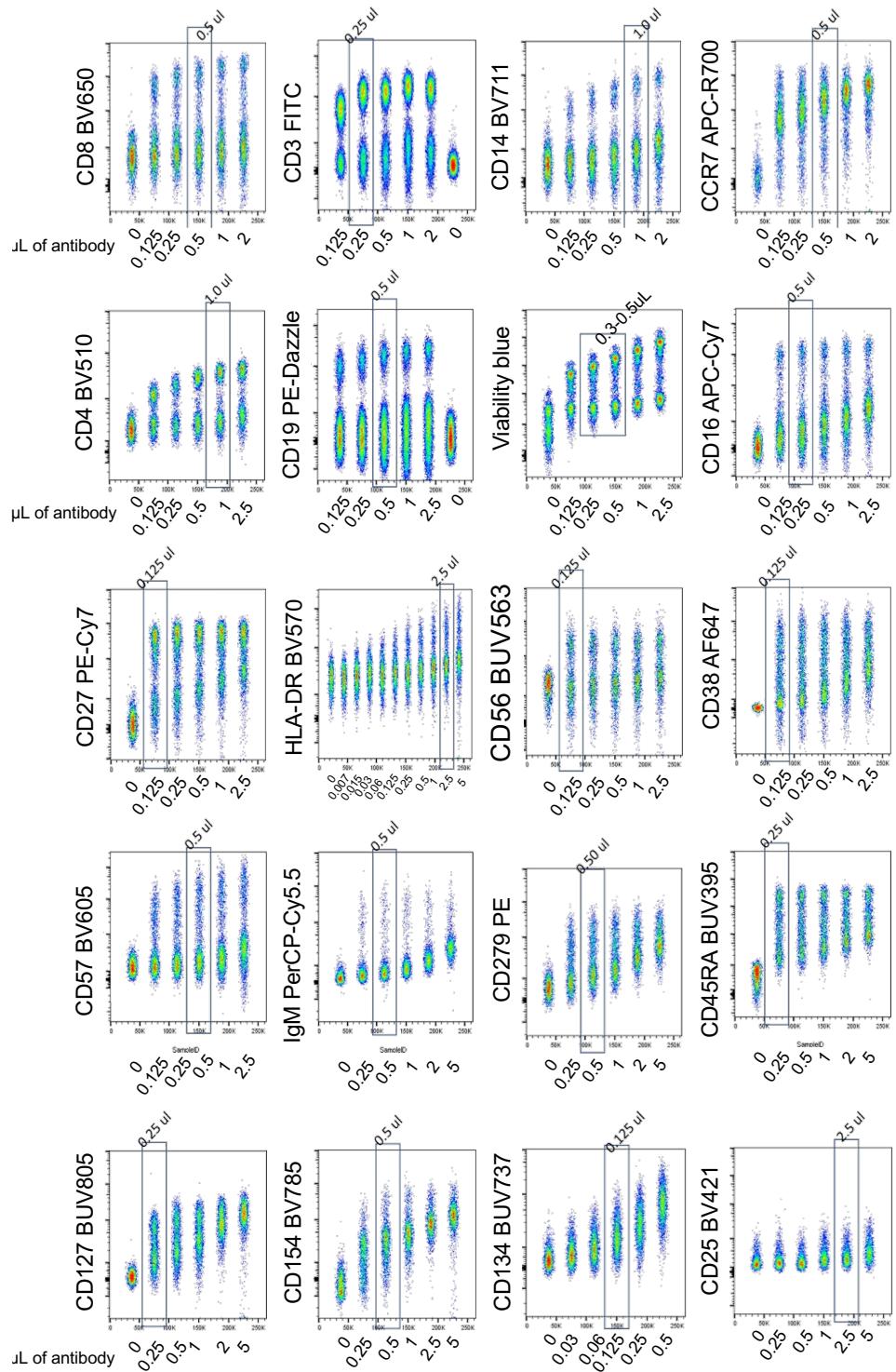
Supplementary Figure 3 | Quantity and quality of whole blood mRNA during rDEN2Δ30 infection. (a) RNA quantity (a) and quality (b) in samples isolated at Day 0 (baseline), day 8 (acute) and day 28 (convalescent) after primary rDEN2Δ30 infection (see methods). Boxplots show the median (horizontal line), 95% confidence level (box bounds), and minimum and maximum values (whiskers).

Flow cytometric gating strategy and cell population identifiers for Hanley et al.



Supplementary Figure 4 | Flow cytometric gating strategy. Labels in bold on plots correspond to graph headings in Figure 4. Lineage-negative ($CD3^-CD19^-CD56^-CD14^-CD16^-$) HLA-DR+ Dendritic cells (**DC**), HLA-DR^{hi} activated DC (**DC_act**); monocytes: classical (**c_mono**), intermediate (**i_mono**), and non-classical (**nc_mono**); natural killer T cells (**nkt**), natural killer cell (**NK1-5**) populations (defined by CD56 and CD16 staining; $CD3^+$ T cells (**tcells**), $CD4^+$ T

cells (**cd4**), and CD8⁺ T cells (**cd8**); CD8⁺ T cell populations: Activated CD279⁺ (**cd8_act**), cytotoxic CD57⁺ (**cd8_cyto**), CCR7⁻CD45RA⁺ T effector re-expressing CD45RA (**cd8t_emra**), CCR7⁺CD45RA⁺ naïve (**cd8t_naive**), CCR7⁻CD45RA⁻ effector memory (**cd8t_em**), CCR7⁺CD45RA⁻ central memory (**cd8t_cm**), and homeostatic CD127⁺ (**cd8_homeo**); CD4⁺ T cell populations: activated CD279⁺ (**cd4_act**), cytotoxic CD57⁺ (**cd4_cyto**), activation-induced marker (AIM)-positive CD154⁺CD134⁺ (**cd4_AIM**), CD25⁺CD134⁺ (**cd4_AIM_old**), regulatory T cells CD127⁻CD25⁺ (**t_regs**), homeostatic CD127⁺ (**cd4_homeo**), CCR7⁻CD45RA⁺ T effector re-expressing CD45RA (**cd4t_emra**), CCR7⁺CD45RA⁺ naïve (**cd4t_naive**), CCR7⁻CD45RA⁻ effector memory (**cd4t_em**), CCR7⁺CD45RA⁻ central memory (**cd4t_cm**); CD19⁺ B cells (**bcells**), plasmablasts CD19⁺CD38^{hi}CD27^{hi} (**pb**), IgM⁺CD27⁻ naïve B cells (**naïve_b**), and IgM⁻CD27⁺ switched memory B cells (**s_mbc**).



Supplementary Figure 5 | Titration of flow cytometry staining antibodies for multi-color staining panel. A range of dilutions of each single antibody was used to stain 10^6 PBMCs in 50

μ L of staining buffer. The fluorescence profiles of each individual antibody dilution (μ L of antibody used on x-axis) was plotted on a single plot with fluorescent intensity on y-axis). Based on the brightness and separation of signal (positive or negative or range of staining), the final staining amount for each antibody required in the multi-color panel are indicated by the box and label.