Electronic Supporting Information

Conformational Control of Pd₂L₄ Assemblies with Unsymmetrical Ligands

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General Experimental

Synthesis: Unless otherwise stated, all reagents, including anhydrous solvents, were purchased from commercial sources and used without further purification. $CDCl_3$ was stored over 4 Å molecular sieves prior to use. All reactions were carried out under an atmosphere of N₂ using degassed, anhydrous solvents unless otherwise stated. Petrol refers to the fraction of petroleum ether boiling in the range 40-60 °C. Analytical TLC was performed on pre-coated silica gel plates (0.25 mm thick, 60F254, Merck, Germany) and observed under UV light. EDTA solution refers to a 0.1 M solution of EDTA-Na₂ in 3% $NH_{3(aq)}$.

Analysis: NMR spectra were recorded on Bruker AV400 or AV500 instrument, at a constant temperature of 298 K. Chemical shifts are reported in parts per million from low to high field and referenced to residual solvent. Standard abbreviations indicating multiplicity were used as follows: m = multiplet, quint = quintet, q = quartet, t = triplet, d = doublet, s = singlet, app. = apparent, br. = broad. Signal assignment was carried out using 2D NMR methods (HSQC, HMBC, COSY, NOESY) where necessary. In the case of some signals absolute assignment was not possible. Here indicative either/or assignments (e.g. H_A/H_B for H_A or H_B) are provided. All melting points were determined using a hot stage apparatus and are uncorrected. Mass spectroscopy Service using Waters LCT Premier for HR-ESI-MS and Thermo Scientific Q-Exactive for tandem MS.

The following compounds were synthesised according to literature procedures: 5-Iodoisoquinoline.¹

Synthetic Procedures



S1



3-Ethynylpyridine (0.209 g, 3.00 mmol, 1 eq.), 1,3-diiodobenzene (1.48 g, 4.50 mmol, 1.5 eq.), Pd(PPh₃)₂Cl₂ (0.042 g, 0.060 mmol, 2 mol%) and CuI (0.023 g, 0.12 mmol, 4 mol%) were stirred at rt in ⁱPr₂NH (15 mL) for 16 h. EDTA solution (25 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:4 EtOAc/petrol) the product was obtained as a yellow oil (0.657 g, 72%) that solidified on standing. M.p. 75-77 °C. ¹H NMR (400 MHz, CDCl₃) δ : 8.75 (dd, *J* = 2.1, 0.8 Hz, 1H, H_a), 8.56 (dd, *J* = 4.9, 1.7 Hz, 1H, H_b), 7.91 (app. t, *J* = 1.6 Hz, 1H, H_h), 7.79 (app. dt, *J* = 7.9, 1.9 Hz, 1H, H_d), 7.70 (ddd, *J* = 8.0, 1.8, 1.1 Hz, 1H, H_g), 7.50 (ddd, *J* = 7.7, 1.5, 1.0 Hz, 1H, H_e), 7.29 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 1H, H_c), 7.10 (app. t, *J* = 7.9 Hz, 1H, H_f). ¹³C NMR (101 MHz, CDCl₃) δ : 152.4, 149.0, 140.4, 138.6, 137.9, 130.9, 130.1, 124.7, 123.2, 120.1, 93.9, 91.0, 87.3. HR-ESI-MS *m/z* = 305.9777 [M+H]⁺ calc. 305.9780.











Figure S4 HSQC NMR (CDCl₃) of S1.







5-Bromo-2-methylpyridine (0.344 g, 2.0 mmol, 1 eq.), trimethylsilylacetylene (0.42 mL, 3.0 mmol, 1.5 eq.), Pd(PPh₃)₂Cl₂ (0.070 g, 010 mmol, 5 mol%) and CuI ((0.038 g, 0.20 mmol, 10 mol%) were stirred at 80 °C in ^{*i*}Pr₂NH (10 mL) in a sealed vial for 21 h. EDTA solution (20 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:19 EtOAc/petrol) the product was obtained as a brown oil (0.350 g, 92%). ¹H NMR (400 MHz, CDCl₃) δ : 8.57 (dd, *J* = 2.2, 0.9 Hz, 1H, H_a), 7.62 (dd, *J* = 8.1, 2.2 Hz, 1H, H_c), 7.09 (d, *J* = 8.0 Hz, 1H, H_b), 2.55 (s, 3H, H_d), 0.25 (s, 9H, H_e). ¹³C NMR (101 MHz, CDCl₃) δ : 158.1, 152.1, 139.2, 122.7, 117.3, 101.9, 97.3, 24.7, 0.0. HR-ESI-MS *m/z* = 190.1063 [M+H]⁺ calc. 190.1052.



Figure S6 ¹H NMR (CDCl₃, 400 MHz) of S2.



Figure S7 13 C NMR (CDCl₃, 101 MHz) of S2.



8-Bromoisoquinoline (0.624 g, 3.00 mmol, 1 eq.), trimethylsilylacetylene (0.50 mL, 3.6 mmol, 1.2 eq.), Pd(PPh₃)₂Cl₂ (0.053 g, 0.075 mmol, 2.5 mol%) and CuI (0.029 g, 0.15 mmol, 5 mol%) were stirred at 75 °C in ⁱPr₂NH (15 mL) in a sealed vial for 18 h. EDTA solution (25 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 25 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:9 EtOAc/pentane) the product was obtained as a yellow oil (0.601 g, 89%). ¹H NMR (400 MHz, CDCl₃) δ : 9.72 (s, 1H, H_a), 8.58 (d, *J* = 5.7 Hz, 1H, H_b), 7.79-7.74 (m, 2H, H_d, H_f), 7.64-7.59 (m, 2H, H_c, H_e), 0.34 (s, 9H, H_g). ¹³C NMR (101 MHz, CDCl₃) δ : 151.5, 143.9, 135.8, 131.9, 129.8, 128.3, 127.3, 121.6, 120.5, 101.7, 101.0, 0.1. HR-ESI-MS *m/z* = 226.1047 [M+H]⁺ calc. 226.1052.











To **S3** (0.225 g, 1.0 mmol, 1 eq.), 1,3-diiodobenzene (0.495 g, 1.5 mmol, 1.5 eq.), Pd(PPh₃)₂Cl₂ (0.018 g, 0.025 mmol, 2.5 mol%) and CuI (0.010 g, 0.050 mmol, 5 mol%) in CH₃CN (5 mL) was added DBU (2.7 mL, 18 mmol, 18 eq.) and the reaction stirred at rt for 18 h. EDTA solution (25 mL) was added, and the aqueous extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:9 EtOAc/petrol followed by 1:3 EtOAc/petrol) the product was obtained as a yellow oil that solidified on standing (0.243 g, 68%). M.p. 91-93 °C. ¹H NMR (400 MHz, CDCl₃) δ : 9.78 (s, 1H, H_a), 8.61 (d, *J* = 5.7 Hz, 1H, H_b), 8.01 (app. t, *J* = 1.6 Hz, 1H, H_j), 7.83-7.80 (m, 2H, 2 of H_d, H_e, H_f), 7.73 (ddd, *J* = 8.0, 1.8, 1.1 Hz, 1H, H_g/H_i), 7.69-7.65 (m, 2H, H_c, 1 of H_d, H_e, H_f), 7.61 (app. dt, *J* = 7.7, 1.3 Hz, 1H, H_g/H_i), 7.14 (app. t, *J* = 7.8 Hz, 1H, H_h). ¹³C NMR (101 MHz, CDCl₃) δ : 151.3, 144.0, 140.4, 138.0, 135.9, 131.7, 131.0, 130.2, 129.9, 128.0, 127.5, 124.9, 121.3, 120.6, 94.1, 94.0, 87.0. HR-ESI-MS *m/z* = 355.9941 [M+H]⁺ calc. 355.9936.











To 3-pyridinylboronic acid (0.369 g, 3.0 mmol, 1 eq.), 1,3-dibromobenzene (0.708 g, 3.0 mmol, 1 eq.) and Pd(PPh₃)₂Cl₂ (0.105 g, 0.15 mmol, 5 mol%) in dioxane (10 mL) was added a solution of K₂CO₃ (1.04 g, 7.5 mmol, 2.5 eq.) in H₂O (5 mL) whilst stirring vigorously. The reaction mixture was stirred at 100 °C for 15 h before EDTA solution (25 mL) was added. The aqueous phase was extracted with CH₂Cl₂ (3 × 25 mL), the combined organic phases dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:9 acetone/CH₂Cl₂) the product was obtained as a yellow oil (0.277 g, 39%). ¹H NMR (400 MHz, CDCl₃) δ : 8.82 (d, *J* = 2.3 Hz, 1H, H_a), 8.62 (dd, *J* = 4.8, 1.5 Hz, 1H, H_b), 7.85 (app. dt, *J* = 7.9, 1.7 Hz, 1H, H_d), 7.73 (app. t, *J* = 1.7 Hz, 1H, H_h), 7.55-7.50 (m, 2H, 2 of H_e, H_f, H_g), 7.39-7.34 (m, 2H, H_c, H_e/H_f/H_g). ¹³C NMR (101 MHz, CDCl₃) δ : δ 149.2, 148.4, 140.1, 135.4, 134.5, 131.2, 130.7, 130.3, 125.9, 123.7, 123.3. HR-ESI-MS *m/z* = 233.9926 [M+H]⁺ calc. 233.9918.







Figure S20 COSY NMR (CDCl₃) of S5.



Figure S22 HMBC NMR (CDCl₃) of S5.



3-(Pyridin-3-yl)bromobenzene (0.277 g, 1.2 mmol, 1 eq.), trimethylsilylacetylene (0.25 mL, 1.8 mmol, 1.5 eq.), Pd(PPh₃)₂Cl₂ (0.021 g, 0.030 mmol, 2.5 mol%) and Cul (0.011 g, 0.059 mmol, 5 mol%) in ^{*i*}Pr₂NH (10 mL) were stirred at 80 °C in a sealed vial for 18 h. EDTA solution (25 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 25 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:9 acetone/CH₂Cl₂) the product was obtained as a brown oil (0.281 g, 95%). ¹H NMR (400 MHz, CDCl₃) δ : 8.83 (d, *J* = 1.9 Hz, 1H, H_a), 8.61 (dd, *J* = 4.8, 1.4 Hz, 1H, H_b), 7.86 (ddd, *J* = 7.9, 2.4, 1.6 Hz, 1H, H_d), 7.69 (app. t, *J* = 1.5 Hz, 1H, H_h), 7.54-7.49 (2H, H_e, H_g), 7.41 (app. t, *J* = 7.7 Hz, 1H, H_f), 7.36 (ddd, *J* = 7.9, 4.8, 1.07 Hz, 1H, H_c), 0.27 (s, 9H, H_i). ¹³C NMR (101 MHz, CDCl₃) δ : 149.0, 148.4, 138.1, 136.0, 134.5, 131.6, 130.8, 129.1, 127.4, 124.2, 123.7, 104.7, 95.1, 0.1. HR-ESI-MS *m*/*z* = 252.1214 [M+H]⁺ calc. 252.1209.



Figure S23 ¹H NMR (CDCl₃, 400 MHz) of S6.



Figure S25 COSY NMR (CDCl₃) of S6.



Figure S27 HMBC NMR (CDCl₃) of S6.



5-lodoisoquinoline (0.510 g, 2.0 mmol, 1 eq.), trimethylsilylacetylene (0.33 mL, 2.4 mmol, 1.2 eq.), Pd(PPh_3)₂Cl₂ (0.028 g, 0.040 mmol, 2 mol%) and CuI (0.015 g, 0.080 mmol, 4 mol%) were stirred at rt in ⁱPr₂NH (15 mL) for 36 h. EDTA solution (25 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:4 EtOAc/petrol) the product was obtained as a brown oil (0.445 g, 99%). ¹H NMR (400 MHz, CDCl₃) δ : 9.25 (d, *J* = 1.0 Hz, 1H, H_c), 8.62 (d, *J* = 5.8 Hz, 1H, H_b), 8.07 (d, *J* = 5.9 Hz, 1H, H_a), 7.94 (d, *J* = 8.3 Hz, 1H, H_d), 7.88 (dd, *J* = 7.2, 1.1, 1H, H_f), 7.55 (dd, *J* = 8.2, 7.2 Hz, 1H, H_e), 0.34 (s, 9H, H_g). ¹³C NMR (101 MHz, CDCl₃) δ : 152.9, 144.2, 136.3, 134.6, 128.4, 128.4, 126.7, 120.3, 119.0, 101.4, 101.2, 0.1. HR-ESI-MS *m*/*z* = 226.1055 [M+H]⁺ calc. 226.1052.



Figure S28 ¹H NMR (CDCl₃, 400 MHz) of **S7**.







Figure S32 HMBC NMR (CDCl₃) of S7.



To **S7** (0.225 g, 1.0 mmol, 1 eq.), 1,3-diiodobenzene (0.660 g, 2.0 mmol, 2 eq.), Pd(PPh₃)₂Cl₂ (0.014 g, 0.020 mmol, 2 mol%) and CuI (0.008 g, 0.04 mmol, 4 mol%) in CH₃CN (5 mL) was added DBU (1.8 mL, 12 mmol, 12 eq.) via syringe and the reaction mixture stirred at rt for 22 h. EDTA solution (20 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (pentane followed by 1:1 EtOAc/pentane) the product was obtained as a yellow oil that solidified on standing (0.226 g, 64%). ¹H NMR (400 MHz, CDCl₃) δ : 9.28 (d, *J* = 1.0 Hz, 1H, H_c), 8.65 (d, *J* = 5.8 Hz, 1H, H_b), 8.13 (app. dt, *J* = 5.7, 1.0 Hz, 1H, H_a), 8.00-7.97 (m, 2H, H_d/H_f, H_j), 7.93 (dd, *J* = 7.2, 1.2 Hz, 1H, H_d/H_f), 7.73 (ddd, *J* = 8.0, 1.7, 1.0 Hz, 1H, H_g/H_i), 7.62-7.58 (m, 2H, H_e, H_g/H_i), 7.14 (app. t, *J* = 7.9 Hz, 1H, H_h). ¹³C NMR (101 MHz, CDCl₃) δ : 153.0, 144.3, 140.3, 137.9, 136.1, 134.4, 131.0, 130.1, 128.5, 128.5, 126.9, 125.0, 120.0, 118.9, 94.0, 93.7, 87.3. HR-ESI-MS *m/z* = 355.9942 [M+H]⁺ calc. 355.9936.











Figure S37 HMBC NMR (CDCl₃) of S8.



To a stirring solution of 2-methylpyridine-5-boronic acid pinacol ester (0.657 g, 3.0 mmol, 1 eq.), 1,3dibromobenzene (0.708 g, 3.0 mmol, 1 eq.) and Pd(PPh₃)₂Cl₂ (0.105 g, 0.15 mmol, 5 mol%) in dioxane (10 mL) was added a solution of K₂CO₃ (1.04 g, 7.5 mmol, 2.5 eq.) in H₂O (5 mL). The reaction mixture was stirred at 100 °C for 19 h. EDTA solution (20 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (CH₂Cl₂ followed by 1:9 acetone/CH₂Cl₂), the product was obtained as a colourless oil that solidified on standing (0.400 g, 54%). M.p. 54-56 °C. ¹H NMR (400 MHz, CDCl₃) δ : 8.69 (d, *J* = 2.1 Hz, 1H, H_g), 7.74 (dd, *J* = 8.0, 2.4 Hz, 1H, H_e), 7.70 (app. t, *J* = 1.8 Hz, 1H, H_d), 7.52-7.47 (m, 2H, H_a, H_c), 7.33 (app. t, *J* = 7.9 Hz, 1H, H_b), 7.23 (d, *J* = 8.0 Hz, 1H, H_f). ¹³C NMR (101 MHz, CDCl₃) δ : 158.1, 147.6, 140.2, 134.8, 132.5, 130.9, 130.7, 130.1, 125.7, 123.4, 123.3, 24.3. HR-ESI-MS *m*/*z* = 248.0067 [M+H]⁺ calc. 248.0075.



Figure S38 ¹H NMR (CDCl₃, 400 MHz) of S9.







To **S1** (0.153 mg, 0.50 mmol, 1 eq.), **S2** (0.142 g, 0.75 mmol, 1.5 eq.), Pd(PPh₃)₂Cl₂ (0.018 g, 0.025 mmol, 5 mol%) and CuI (0.010 g, 0.050 mmol, 10 mol%) in CH₃CN (2.5 mL) was added DBU (0.90 mL, 6.0 mmol, 12 eq.) via syringe and the reaction stirred at rt for 18 h. EDTA solution (25 mL) was added and the aqueous phase was extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (1:1 pentane/Et₂O followed by neat Et₂O) the product was obtained as a yellow solid (0.132 g, 90%). M.p. 92-94 °C. ¹H NMR (400 MHz, CDCl₃) δ : 8.77 (dd, *J* = 2.1, 0.9 Hz, 1H, H_a), 8.66 (d, *J* = 1.8 Hz, 1H, H_k), 8.57 (dd, *J* = 4.9, 1.7 Hz, 1H, H_b), 7.81 (app. dt, *J* = 7.9, 1.9 Hz, 1H, H_d), 7.74 (t, *J* = 1.4 Hz, 1H, H_h), 7.70 (dd, *J* = 8.0, 2.2 Hz, 1H, H_i/H_j), 7.54-7.51 (m, 2H, H_e, H_g), 7.37 (app. t, *J* = 7.8 Hz, 1H, H_f), 7.30 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 1H, H_c), 7.16 (d, *J* = 8.0 Hz, 1H, H_i/H_j), 2.59 (s, 3H, H_i). ¹³C NMR (101 MHz, CDCl₃) δ : 158.2, 152.5, 151.8, 149.0, 138.8, 138.6, 134.8, 131.9, 131.8, 128.8, 123.4, 123.2, 123.1, 122.9, 120.3, 117.1, 91.8, 90.9, 87.2, 86.8, 24.7. HR-ESI-MS *m/z* = 295.1244 [M+H]⁺ calc. 295.1235.



Figure S43 ¹H NMR (CDCl₃, 400 MHz) of **1**.









To a vigorously stirring solution of **S4** (0.178 g, 0.50 mmol, 1 eq.), 4-pyridinylboronic acid (0.074 g, 0.60 mmol, 1.2 eq.) and Pd(PPh₃)₂Cl₂ (0.018 g, 0.025 mmol, 5 mol%) in dioxane (2.0 mL) was added K₂CO₃ (0.173 g, 1.25 mmol, 2.5 eq.) in H₂O (1.0 mL) via syringe. The reaction mixture was stirred at 80 °C for 16 h. EDTA solution (20 mL) was added, and the aqueous phase extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (2:3 MeCN/CH₂Cl₂ followed by 2:3 acetone/CH₂Cl₂) the product was obtained as a colourless oil that solidified on standing (0.031 g, 20%). M.p. 112-114 °C. ¹H NMR (400 MHz, CDCl₃) δ : 9.85 (s, 1H, H_a), 8.71 (m, 2H, H_i), 8.62 (d, *J* = 5.7 Hz, 1H, H_b), 7.93 (t, *J* = 1.8 Hz, 1H, H_j), 7.86-7.82 (m, 2H, 2 of H_d, H_e, H_f, H_g, H_h), 7.74-7.65 (4H, H_c, H_i, 2 of H_d, H_e, H_f, H_g, H_h), 7.56-7.52 (m, 3H, H_k, 1 of H_d, H_e, H_f, H_g, H_h). ¹³C NMR (101 MHz, CDCl₃) δ : 151.3, 150.6, 147.5, 144.0, 138.8, 135.9, 132.3, 131.7, 130.4, 129.9, 129.5, 128.1, 127.6, 127.4, 123.9, 121.8, 121.5, 120.7, 95.2, 86.5. HR-ESI-MS *m*/*z* = 307.1240 [M+H]⁺ calc. 307.1235.



Figure S48 ¹H NMR (CDCl₃, 400 MHz) of **2**.






To **S6** (0.126 g, 0.50 mmol, 1 eq.), 5-iodoisoquinoline (0.128 g, 0.50 mmol, 1 eq.), Pd(PPh₃)₂Cl₂ (0.0088 g, 0.013 mmol, 2.5 mol%) and CuI (0.0048 g, 0.025 mmol, 5 mol%) in MeCN (5 mL) was added via syringe DBU (0.90 mL, 6.0 mmol, 12 eq.). N₂ was bubbled through the reaction solution for 5 minutes before stirring at rt for 21 h. The solvent was removed *in vacuo* and the residue was dissolved in CH₂Cl₂ (50 mL) before washing with EDTA solution (20 mL), H₂O (2 × 20 mL) and brine (20 mL), dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (CH₂Cl₂ with a step gradient of 5% increments up to 30% acetone) the product was obtained as a light yellow solid (0.098 g, 64%). M.p. 116-118 °C. ¹H NMR (400 MHz, CDCl₃) δ : 9.28 (s, 1H, H_c), 8.89 (d, *J* = 1.9 Hz, 1H, H_n), 8.66-8.62 (m, 2H, H_b, H_m), 8.18 (dt, *J* = 5.8, 1.0 Hz, 1H, H_a), 7.98-7.94 (m, 2H, H_f, H_d/H_e), 7.91 (ddd, *J* = 7.9, 2.4, 1.6 Hz, 1H, H_k), 7.85 (t, *J* = 1.5 Hz, 1H, H_i), 7.67 (dt, *J* = 7.6, 1.4 Hz, 1H, H_g), 7.61-7.58 (m, 2H, H_i, H_d/H_e), 7.52 (app. td, *J* = 7.7, 0.6 Hz, 1H, H_h), 7.39 (ddd, *J* = 7.9, 4.8, 0.9 Hz, 1H, H_l). ¹³C NMR (101 MHz, CDCl₃) δ : 152.9, 149.1, 148.4, 144.2, 138.5, 136.1, 135.8, 134.5, 134.3, 131.4, 130.5, 129.4, 128.5, 128.3, 127.6, 126.9, 123.9, 123.7, 120.2, 118.9, 95.0, 86.6. HR-ESI-MS *m/z* = 307.1241 [M+H]⁺ calc. 307.1235.



Figure S53 ¹H NMR (CDCl₃, 400 MHz) of **3**.





Figure S57 HMBC NMR (CDCl₃) of 3.



S8 (0.080 g, 0.23 mmol, 1 eq.), 3-ethynylpyridine (0.028 g, 0.27 mmol, 1.2 eq.), Pd(PPh₃)₂Cl₂ (0.0079 g, 0.011 mmol, 5 mol%) and CuI (0.0043 g, 0.023 mmol, 10 mol%) were stirred at rt in ^{*i*}Pr₂NH (5 mL) for 22 h. EDTA solution (10 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 10 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (gradient of 1:9 to 1:3 acetone/CH₂Cl₂) the product was obtained as a yellow solid (0.069 g, quant.). M.p. 136-138 °C. ¹H NMR (400 MHz, CDCl₃) δ :9.29 (d, *J* = 1.0 Hz, 1H, H_c), 8.79 (dd, *J* = 2.1, 0.9 Hz, 1H, H_n), 8.66 (d, *J* = 5.8 Hz, 1H, H_b), 8.57 (dd, *J* = 4.9, 1.7 Hz, 1H, H_m), 8.17 (d, *J* = 5.9 Hz, 1H, H_a), 7.98 (d, *J* = 8.2 Hz, 1H, H_d), 7.95 (dd, *J* = 7.2, 1.1 Hz, 1H, H_f), 7.84-7.82 (m, 2H, H_j, H_k), 7.65-7.55 (m, 3H, H_e, H_g, H_i), 7.42 (app. td, *J* = 7.8, 0.6 Hz, 1H, H_h), 7.31 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 1H, H_i). ¹³C NMR (101 MHz, CDCl₃) δ : 153.0, 152.5, 149.0, 144.3, 138.6, 136.2, 134.9, 134.4, 132.0, 132.0, 128.9, 128.5, 128.4, 126.9, 123.5, 123.3, 123.2, 120.3, 120.1, 118.9, 94.5, 91.7, 86.9, 86.8. HR-ESI-MS *m*/*z* = 331.1241 [M+H]⁺ calc. 331.1235.



Figure S58 ¹H NMR (CDCl₃, 400 MHz) of **4**.



Figure S60 COSY NMR (CDCl₃) of 4.



Figure S62 HMBC NMR (CDCl₃) of 4.



S5 (0.117 g, 0.50 mmol, 1 eq.), 3-ethynylpyridine (0.062 g, 0.60 mmol, 1.2 eq), Pd(PPh₃)₂Cl₂ (0.0088 g, 0.013 mmol, 2.5 mol%) and CuI (0.0048 g, 0.025 mmol, 5 mol%) were stirred at 80 °C in ^{*i*}Pr₂NH (5 mL) in a sealed vial for 17 h. EDTA solution (20 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20 mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (CH₂Cl₂ with a step gradient of 5% increments up to 25% acetone) the product was obtained as a light yellow oil (0.083 g, 65%). ¹H NMR (400 MHz, CDCl₃) δ : 8.86 (dd, *J* = 2.4, 0.9 Hz, 1H, H_i), 8.79 (dd, *J* = 2.1, 0.9 Hz, 1H, H_a), 8.62 (dd, *J* = 4.8, 1.6 Hz, 1H, H_k), 8.56 (dd, *J* = 4.9, 1.7 Hz, 1H, H_b), 7.89 (ddd, *J* = 7.9, 2.4, 1.7 Hz, 1H, H_i), 7.83 (app. dt, *J* = 7.9, 1.9 Hz, 1H, H_d), 7.77 (t, *J* = 1.5 Hz, 1H, H_b), 7.59-7.57 (m, 2H, H_e, H_g), 7.51-7.47 (m, 1H, H_f), 7.38 (ddd, *J* = 7.9, 4.8, 0.9 Hz, 1H, H_j), 7.30 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 1H, H_c). ¹³C NMR (101 MHz, CDCl₃) δ : 152.4, 149.1, 148.9, 148.4, 138.6, 138.4, 135.8, 134.5, 131.3, 130.5, 129.4, 127.7, 123.7, 123.6, 123.2, 120.3, 92.3, 86.7. HR-ESI-MS *m/z* = 257.1070 [M+H]⁺ calc. 257.1079.











Figure S67 HMBC NMR (CDCl₃) of 5.



S9 (0.248 g, 1.0 mmol, 1 eq.), 3-ethynylpyridine (0.124 g, 1.2 mmol, 1.2 eq.), Pd(PPh₃)₂Cl₂ (0.018 g, 0.025 mmol, 2.5 mol%) and CuI (0.010 g, 0.050 mmol, 5 mol%) were stirred at 80 °C in i Pr₂NH (10 mL) in a sealed vial for 24 h. EDTA solution (20 mL) was added and the aqueous phase extracted with CH₂Cl₂ (3 × 20mL). The combined organic phases were dried (MgSO₄) and the solvent removed *in vacuo*. After purification by column chromatography on silica (CH₂Cl₂ with a step gradient of 5% increments up to 25% acetone) the product was obtained as a light brown solid (0.170 g, 63%). M.p. 90-92 °C. ¹H NMR (400 MHz, CDCl₃) δ : 8.79 (dd, *J* = 2.1, 0.8 Hz, 1H, H_a), 8.74 (dd, *J* = 2.4, 0.9 Hz, 1H, H_k), 8.56 (dd, *J* = 4.9, 1.7 Hz, 1H, H_b), 7.83 (dt, *J* = 7.9, 1.9 Hz, 1H, H_d), 7.79 (dd, *J* = 8.0, 2.4 Hz, 1H, H_i), 7.74 (td, *J* = 1.8, 0.6 Hz, 1H, H_b), 7.58-7.55 (m, 2H, H_e, H_g), 7.47 (ddd, *J* = 8.0, 7.3, 0.6 Hz, 1H, H_f), 7.30 (ddd, *J* = 7.9, 4.9, 0.9 Hz, 1H, H_a), 8.154. (101 MHz, CDCl₃) δ : 157.9, 152.5, 148.9, 147.6, 138.6, 138.5, 134.8, 132.9, 131.1, 130.3, 129.3, 127.5, 123.5, 123.4, 123.2, 120.4, 92.4, 86.6, 24.3. HR-ESI-MS *m/z* = 271.1230 [M+H]⁺ calc. 271.1235.



Figure S68 ¹H NMR (CDCl₃, 400 MHz) of **6**.





[Pd₂(1)₄](BF₄)₄



1 (8.8 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in d_6 -DMSO (0.75 mL) until all solids were dissolved. After standing at rt for 24 h, the formation of two major species could be observed by ¹H NMR. Diffusion coefficient (500 MHz, d_6 -DMSO) D: 1.12 × 10⁻¹⁰ m²s⁻¹. ESI-MS m/z = 781.13 {[Pd₂(C₂₁H₁₄N₂)₄](BF₄)₂}²⁺ calc. 781.14.



Figure S73 ¹H NMR (*d*₆-DMSO, 500 MHz) of [Pd₂(**1**)₄](BF₄)₄.



Figure S74 ROESY NMR (*d*₆-DMSO, 500 MHz) of [Pd₂(1)₄](BF₄)₄.



Figure S75 Partial 2D TOCSY NMR (*d*₆-DMSO, 500 MHz) of [Pd₂(**1**)₄](BF₄)₄.



Figure S77 Calculated (top) and experimental (bottom) isotopic patterns for $\{[Pd_2(1)_4](BF_4)_2\}^{2+}$.



Figure S78 ESI-MS of $[Pd_2(1)_4](BF_4)_4$ formed in d_6 -DMSO.

1 (8.8 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in CD₃CN (0.75 mL) until all solids were dissolved. After standing at 60 °C for 24 h, the formation of a single major species could be observed by ¹H NMR. ¹H NMR (400 MHz, CD₃CN) δ : 9.45 (d, *J* = 1.6 Hz, 4H, H_k), 9.35 (d, *J* = 1.7 Hz, 4H, H_a), 8.66 (m, 4H, H_b), 8.12 (ddd, *J* = 8.0, 1.8, 1.3 Hz, 4H, H_d), 8.02 (dd, *J* = 8.2, 1.9 Hz, 4H, H_i), 7.89 (app. t, *J* = 1.3 Hz, 4H, H_h), 7.70-7.65 (m, 8H, H_e, H_g), 7.58-7.49 (m, 12H, H_c, H_j, H_f), 3.60 (s, 12H, H_i). Diffusion coefficient (500 MHz, CD₃CN) *D*: 8.10 × 10⁻¹⁰ m²s⁻¹. ¹³C NMR (101 MHz, CD₃CN) δ : 161.2, 153.7, 153.1, 152.0, 144.3, 144.0, 134.6, 134.5, 134.1, 130.7, 129.1, 128.4, 125.1, 123.3, 123.0, 122.2, 95.2, 94.4, 85.4, 85.1, 26.7. ESI-MS *m/z* = 781.06 {[Pd₂(C₂₁H₁₄N₂)₄](BF₄)₂}²⁺ calc. 781.14.



Figure S79 ¹H NMR (CD₃CN, 400 MHz) of [Pd₂(1)₄](BF₄)₄.



Figure S81 COSY NMR (CD₃CN) of $[Pd_2(1)_4](BF_4)_4$.



Figure S83 HMBC NMR (CD₃CN) of [Pd₂(1)₄](BF₄)₄.



Figure S85 DOSY NMR (CD₃CN, 500 MHz) of [Pd₂(1)₄](BF₄)₄.



Figure S86 Calculated (top) and experimental (bottom) isotopic patterns for ${[Pd_2(1)_4](BF_4)_2}^{2+}$.



Figure 87 ESI-MS of $[Pd_2(1)_4](BF_4)_4$ formed in CD₃CN.



Figure S88 ¹H NMR (CD₃CN, 400 MHz) of 2:1 mixture of **1** and [Pd(CH₃CN)₄](BF₄)₂ after a) 0 h, b) 2 h, c) 6 h and d) 22 h at room temperature, and e) after heating at 60 °C.





[Pd₂(2)₄](BF₄)₄



2 (9.2 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in d_6 -DMSO (0.75 mL) until all solids were dissolved. After standing at rt for 2 h, quantitative conversion to $[Pd_2(2)_4](BF_4)_4$ was observed by ¹H NMR. ¹H NMR (400 MHz, d_6 -DMSO) δ : 10.02 (s, 4H, H_a), 9.67 (d, J = 6.6 Hz, 4H, H_b), 9.59 (d, J = 6.6 Hz, 8H, H_l), 8.48 (d, J = 6.5 Hz, 4H, H_c), 8.37 (d, J = 6.9 Hz, 8H, H_k), 8.30 (s, 4H, H_j), 8.25 (d, J = 8.3 Hz, 4H, H_d), 8.13 (dd, J = 7.2, 1.0 Hz, 4H, H_f), 8.07-8.02 (m, 8H, H_e, H_l), 7.87 (dt, J = 7.6, 1.2 Hz, 4H, H_g), 7.70 (app. t, J = 7.8 Hz, 4H, H_h). Diffusion coefficient (500 MHz, d_6 -DMSO) *D*: 1.15 × 10⁻¹⁰ m²s⁻¹. ¹³C NMR (101 MHz, d_6 -DMSO) δ : 153.0, 151.0, 149.8, 142.9, 136.4, 134.7, 134.3, 134.1, 133.8, 131.8, 130.4, 128.2, 128.0, 127.7, 125.1, 124.3, 122.3, 120.4, 96.2, 86.0. ESI-MS $m/z = 805.15 \{[Pd_2(C_{22}H_{14}N_2)_4](BF_4)_2\}^{2+}$ calc. 805.14.



Figure S91 ¹³C NMR (*d*₆-DMSO, 101 MHz) of [Pd₂(**2**)₄](BF₄)₄.



Figure S93 HSQC NMR (d_6 -DMSO) of $[Pd_2(2)_4](BF_4)_4$.





Figure S96 Experimental (top) and calculated (bottom) isotopic patterns for $\{[Pd_2(2)_4](BF_4)_2\}^{2+}$.



Figure S97 ESI-MS of [Pd₂(**2**)₄](BF₄)₄.

[Pd₂(3)₄](BF₄)₄



3 (9.2 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in d_6 -DMSO (0.75 mL) until all solids were dissolved. After standing at rt for 2 h, quantitative conversion to $[Pd_2(\mathbf{3})_4](BF_4)_4$ was observed by ¹H NMR. ¹H NMR (400 MHz, d_6 -DMSO) δ : 10.00-9.98 (m, 8H, H_b, H_c), 9.81 (d, J = 1.6 Hz, 4H, H_n), 9.59 (dd, J = 5.7, 1.3 Hz, 4H, H_m), 8.94 (d, J = 6.6 Hz, 4H, H_a), 8.58 (d, J = 7.8 Hz, 4H, H_k), 8.40 (s, 4H, H_j), 8.29 (d, J = 8.5 Hz, 4H, H_a), 8.17 (dd, J = 7.2, 1.1 Hz, 4H, H_f), 7.99 (dd, J = 8.1, 5.7 Hz, 4H, H_i), 7.88 (m, 4H, H_i), 7.84 (dd, J = 8.3, 7.3 Hz, 4H, H_e), 7.67 (app. dt, J = 7.7, 1.3 Hz, 4H, H_g), 7.56 (app. t, J = 7.8 Hz, 4H, H_h). Diffusion coefficient (500 MHz, d_6 -DMSO) *D*: 1.16 × 10⁻¹⁰ m²s⁻¹. ¹³C NMR (101 MHz, d_6 -DMSO) δ : 155.7, 150.2, 149.2, 143.2, 138.3, 136.8, 136.6, 135.8, 134.4, 132.2, 130.1, 129.6, 129.4, 129.3, 128.2, 128.0, 127.8, 124.0, 122.7, 119.4, 95.7, 85.4. ESI-MS m/z = 805.1 { $[Pd_2(C_{22}H_{14}N_2)_4](BF_4)_2$ ⁺ calc. 805.1; 1697.3 { $[Pd_2(C_{22}H_{14}N_2)_4](BF_4)_3$ ⁺ calc. 1697.3.











Figure S102 HMBC NMR (d_6 -DMSO) of $[Pd_2(3)_4](BF_4)_4$.



Figure S104 Calculated (top) and experimental (bottom) isotopic patterns for ${[Pd_2(3)_4](BF_4)_2}^{2+}$.



Figure S105 Calculated (top) and experimental (bottom) isotopic patterns for $\{[Pd_2(3)_4](BF_4)_3\}^+$.



Figure S106 ESI-MS of [Pd₂(**3**)₄](BF₄)₄.

[Pd₂(4)₄](BF₄)₄



4 (9.9 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in d_6 -DMSO (0.75 mL) until all solids were dissolved. After standing at rt for 18 h, quantitative conversion to $[Pd_2(\mathbf{4})_4](BF_4)_4$ was observed by ¹H NMR. ¹H NMR (400 MHz, d_6 -DMSO) δ : 10.10 (s, 4H, H_c), 9.62 (d, J = 1.9 Hz, 4H, H_n), 9.54 (dd, J = 5.8, 1.1 Hz, 4H, H_m), 9.49 (d, J = 6.7 Hz, 4H, H_b), 8.58 (d, J = 6.7 Hz, 4H, H_a), 8.34 (d, J = 8.5 Hz, 4H, H_d), 8.30 (app. dt, J = 8.0, 1.5 Hz, 4H, H_k), 8.25 (dd, J = 7.3, 1.0 Hz, 4H, H_f), 7.98 (app. td, J = 1.7, 1.0 Hz, 4H, H_g), 7.94-7.89 (m, 8H, H_e, H_l), 7.74 (app. dt, J = 7.7, 1.3 Hz, 4H, H_g/H_l), 7.69 (app. dt, J = 7.8, 1.3 Hz, 4H, H_g/H_l), 7.59-7.55 (m, 4H, H_h). Diffusion coefficient (500 MHz, d_6 -DMSO) D: 9.78 × 10⁻¹¹ m²s⁻¹. ¹³C NMR (101 MHz, d_6 -DMSO) δ : 156.8, 153.0, 150.7, 142.7, 141.9, 137.6, 135.4, 134.3, 132.7 (×2), 130.2, 130.1, 129.7, 128.4, 127.6, 122.8, 122.2, 121.7, 121.5, 118.9, 95.0, 93.3, 85.6, 84.8. ESI-MS $m/z = 853.13 \{[Pd_2(C_{24}H_{14}N_2)_4](BF_4)_2\}^{2+}$ calc. 853.14; 539.75 $\{[Pd_2(C_{24}H_{14}N_2)_4](BF_4)\}^{3+}$ calc. 539.76.



Figure S108 ¹³C NMR (*d*₆-DMSO, 101 MHz) of [Pd₂(4)₄](BF₄)₄.



Figure S110 HSQC NMR (d_6 -DMSO) of [Pd₂(4)₄](BF₄)₄.



Figure S112 DOSY NMR (*d*₆-DMSO, 500 MHz) of [Pd₂(**4**)₄](BF₄)₄.



Figure S113 Calculated (top) and experimental (bottom) isotopic patterns for ${[Pd_2(4)_4](BF_4)_2\}^{2+}}$.



Figure S114 Calculated (top) and experimental (bottom) isotopic patterns for {[Pd₂(4)₄](BF₄)}³⁺.




[Pd₂(5)₄](BF₄)₄



5 (7.7 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in d_6 -DMSO (0.75 mL) until all solids were dissolved. After standing at rt for 4 d, a number of species were observed by ¹H NMR. Heating at 60 °C for several days had no effect on the speciation observed by ¹H NMR. Diffusion coefficient (500 MHz, d_6 -DMSO) *D*: 1.16 × 10⁻¹⁰ m²s⁻¹. ESI-MS m/z = 1497.23 { $[Pd_2(C_{18}H_{12}N_2)_4](BF_4)_3$ ⁺ calc. 1497.23; 705.11 { $[Pd_2(C_{18}H_{12}N_2)_4](BF_4)_2$ ²⁺ calc. 705.11.



Figure S116 ¹H NMR (*d*₆-DMSO, 500 MHz) of [Pd₂(**5**)₄](BF₄)₄.







Figure S119 Calculated (top) and experimental (bottom) isotopic patterns for ${[Pd_2(5)_4](BF_4)_2}^{2+}$.



Figure S120 ESI-MS of $[Pd_2(5)_4](BF_4)_4$.

5 (7.7 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in CD₃CN (0.75 mL) until all solids were dissolved. After standing at 60 °C for 1 d, a number of species were observed by ¹H NMR. Continued heating at 60 °C for several days had no effect on the speciation observed by ¹H NMR. Diffusion coefficient (500 MHz, CD₃CN) *D*: 8.50 × 10⁻¹⁰ m²s⁻¹.



[Pd₂(6)₄](BF₄)₄



6 (8.1 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in d_6 -DMSO (0.75 mL) until all solids were dissolved. After standing at rt for 24 h, conversion to $[Pd_2(6)_4](BF_4)_4$ was observed by ¹H NMR. ¹H NMR (400 MHz, d_6 -DMSO) δ : 9.29 (br. s, 4H, H_a), 9.15 (m, 4H, H_b), 9.09 (d, J = 1.8 Hz, 4H, H_k), 8.50 (dd, J = 8.4, 2.0 Hz, 4H, H_i), 8.28 (dt, J = 8.0, 1.4 Hz, 4H, H_d), 8.24 (br. s, 4H, H_h), 7.86 (dt, J = 7.8, 1.7 Hz, 4H, H_g), 7.78-7.73 (m, 8H, H_c, H_j), 7.64-7.56 (m, 8H, H_e, H_f), 3.30 (s, 12H, H_i). Diffusion coefficient (500 MHz, d_6 -DMSO) D: 1.15 × 10⁻¹⁰ m²s⁻¹. ¹³C NMR (101 MHz, d_6 -DMSO) δ : 160.5, 152.4, 151.8, 146.9, 143.2, 139.1, 134.7, 134.3, 132.4, 130.3, 128.8 (×2), 128.5, 127.1, 122.9, 121.9, 94.5, 84.2, 25.9. ESI-MS $m/z = 459.76 \{[Pd_2(C_{19}H_{14}N_2)_4](BF_4)\}^{3+}$ calc. 459.76; 733.14 $\{[Pd_2(C_{19}H_{14}N_2)_4](BF_4)\}^{4+}$ calc. 1553.28.



Figure S124 ¹³C NMR (*d*₆-DMSO, 101 MHz) of [Pd₂(6)₄](BF₄)₄.







Figure S127 HMBC NMR (d_6 -DMSO) of [Pd₂(6)₄](BF₄)₄.



Figure S128 ROESY NMR (d_6 -DMSO) of [Pd₂(6)₄](BF₄)₄.



Figure S130 Calculated (top) and experimental (bottom) isotopic patterns for ${[Pd_2(6)_4](BF_4)}^{3+}$.



Figure S131 Calculated (top) and experimental (bottom) isotopic patterns for {[Pd₂(6)₄](BF₄)₂}²⁺.







Figure S133 ESI-MS of [Pd₂(6)₄](BF₄)₄.

X-ray Crystallographic Data

X-ray crystal structure of cis-[Pd₂(2)₄](BF₄)₄

2 (12.3 mg, 0.040 mmol) and $[Pd(CH_3CN)_4](BF_4)_2$ (8.9 mg, 0.020 mmol) were stirred in DMF (3 mL) at rt for 2 h. X-ray quality crystals were grown by vapour diffusion of Et₂O into the solution over a period of several days.

Crystal data for cis-[Pd₂(**2**)₄](BF₄)₄: [C₈₈H₅₆N₈Pd₂](BF₄)₄·4(C₄H₁₀O), *M* = 2081.92, monoclinic, *I*2/*a* (no. 15), *a* = 28.3600(4), *b* = 12.35285(17), *c* = 27.6660(4) Å, β = 94.6541(14)°, *V* = 9660.2(2) Å³, *Z* = 4 [*C_i* symmetry], *D_c* = 1.431 g cm⁻³, μ (Cu-Kα) = 3.749 mm⁻¹, *T* = 173 K, colourless platy needles, Agilent Xcalibur PX Ultra A diffractometer; 9259 independent measured reflections (*R*_{int} = 0.0268), *F*² refinement,^{2,3} *R*₁(obs) = 0.0419, *wR*₂(all) = 0.1190, 7879 independent observed absorption-corrected reflections [|*F*₀| > 4σ(|*F*₀|), completeness to θ_{full}(67.7°) = 98.7%], 555 parameters. CCDC 1946272.

The N22- and N52-based pyridyl rings, and the B60-based tetrafluoroborate anion, in the structure of cis-[Pd₂(**2**)₄](BF₄)₄ were all found to be disordered. In each case two orientations were identified, of ca. 67:33, 68:32, and 69:31% occupancy respectively, their geometries were optimised, the thermal parameters of adjacent atoms were restrained to be similar, and only the non-hydrogen atoms of the major occupancy orientations were refined anisotropically (those of the minor occupancy orientations were refined isotropically).

The included solvent was found to be highly disordered, and the best approach to handling this diffuse electron density was found to be the SQUEEZE routine of PLATON.⁴ This suggested a total of 662 electrons per unit cell, equivalent to 165.5 electrons per complex. Before the use of SQUEEZE the solvent clearly resembled diethyl ether (C₄H₁₀O, 42 electrons), and 4 dichloromethane molecules corresponds to 168 electrons, so this was used as the solvent present. As a result, the atom list for the asymmetric unit is low by $2(C_4H_{10}O) = C_8H_{20}O_2$ (and that for the unit cell low by $C_{64}H_{160}O_{16}$) compared to what is actually presumed to be present.



Figure S134 The structure of the C_i-symmetric cation present in the crystal of cis-[Pd₂(2)₄](BF₄)₄.

X-ray crystal structure of cis-[Pd₂(4)₄](BF₄)₄

4 (9.9 mg, 0.030 mmol, 2 eq.) and $[Pd(CH_3CN)_4](BF_4)_2$ (6.7 mg, 0.015 mmol, 1 eq.) were sonicated in DMF (3 mL) until all solids were dissolved. After stirring at rt overnight, X-ray quality crystals were grown by vapour diffusion of Et_2O into the solution over a period of several days.

Crystal data for cis-[Pd₂(**4**)₄](BF₄)₄: [C₉₆H₅₆N₈Pd₂](BF₄)₄·9(C₃H₇NO), *M* = 2539.38, monoclinic, *C2/c* (no. 15), *a* = 23.2193(9), *b* = 15.8896(5), *c* = 38.7298(15) Å, β = 112.281(5)°, *V* = 13222.3(9) Å³, *Z* = 4 [*C_i* symmetry], *D_c* = 1.276 g cm⁻³, μ(Cu-Kα) = 2.884 mm⁻¹, *T* = 173 K, colourless tablets, Agilent Xcalibur PX Ultra A diffractometer; 14084 independent measured reflections (*R*_{int} = 0.0507), *F*² refinement,^{2,3} *R*₁(obs) = 0.0994, *wR*₂(all) = 0.2743, 12479 independent observed absorption-corrected reflections [|*F*_o| > 4σ(|*F*_o|), completeness to θ_{full}(67.7°) = 98.7%], 479 parameters. CCDC 1946273.

The crystal of *cis*-[Pd₂(**4**)₄](BF₄)₄ that was studied was found to be a two component twin in a *ca*. 62:38 ratio, with the two lattices related by the approximate twin law [1.00 0.00 0.00 0.00 -1.00 0.00 -0.74 -0.01 -1.00].

Both the presumed tetrafluoroborate anions and the included solvent were found to be highly disordered, and the best approach to handling this diffuse electron density was found to be the SQUEEZE routine of PLATON.⁴ This suggested a total of 2090 electrons per unit cell, equivalent to 522.5 electrons per complex. Before the use of SQUEEZE, one tetrafluoroborate anion (BF₄, 41 electrons) and one dimethylformamide solvent molecule (C₃H₇NO, 40 electrons) were identified amongst the diffuse electron density. Removing the electron density of the presumed four tetrafluoroborate anions per complex leaves 522.5 – (4 x 41) = 358.5 electrons for the solvent; 9 dimethylformamide molecules corresponds to 360 electrons, so this was used as the solvent present. As a result, the atom list for the asymmetric unit is low by $2(BF4) + 4.5(C_3H_7NO) = C_{13.5}H_{31.5}B_2F_8N_{4.5}O_{4.5}$ (and that for the unit cell low by $C_{108}H_{252}B_{16}F_{64}N_{36}O_{36}$) compared to what is actually presumed to be present.



Figure S135 The structure of the C_i-symmetric cation present in the crystal of cis-[Pd₂(4)₄](BF₄)₄.

Density Functional Theory Calculations

All models were constructed using Avogadro⁵ starting from the crystal structures reported herein. The semi-empirical method $GFN2-xTB^6$ was used to geometry optimise all structures in implicit solvent before optimisation using density functional theory (DFT). The xTB family of functionals have been shown to reproduce the geometries of metal-containing supramolecular architectures.⁷ DFT optimisations were carried out using Gaussian 16.⁸ All optimisations were carried out in dimethyl sulfoxide (DMSO) using the polarizable continuum model (PCM), unless stated otherwise. All geometries were optimised at the B3LYP/6-31G(d) level of theory with the D3 empirical dispersion correction.⁹ The Stuttgart-Dresden (SDD) effective core potentials were used for the Pd²⁺ centres. Each cage has a charge of 4 and a multiplicity of 1. A similar approach has been used by Clever and co-workers on similar cage systems.¹⁰ The total electronic energies are reported in all cases (Tables S1 and S2). Numerical frequency calculations were performed on all optimised structures to confirm that the optimised structure was a minimum. However, there was at most one small negative frequency in some cases. We note that the accuracy of the Hessian calculations was not sufficient in all cases to reliably use the results of the thermochemistry calculations. The geometry of *cis*-[Pd₂(**4**)₄] optimised with DFT matches well with the solid-state structure (Fig. S127).

Input files and output files for DFT calculations can be accessed at: <u>https://github.com/andrewtarzia/citable_data/tree/master/lewis_2019</u>





Table S1 Relative energy (the minimum of each *cis-trans* pair is set to 0 kJ mol⁻¹) and the magnitude of the dipole moment of *cis* (**C**) and *trans* (**D**) isomers of the cages formed from ligand **1** using two implicit solvent models.

Ligand	Isomer	Solvent	Relative energy (kJ mol ⁻¹)	Dipole moment (Debye)	
1	С	Acetonitrile	6.1	0.40	
1	D	Acetonitrile	0.0	0.00	
1	С	DMSO	6.1	0.39	
1	D	DMSO	0.0	0.00	

Ligand	Isomer	Relative energy (kJ mol ⁻¹)	(kJ mol ⁻¹) Dipole moment (Debye)	
4	Α	26.8	12.60	
4	В	4.4	6.20	
4	С	0.0	0.96	
4	D	8.9	0.06	
5	Α	18.0	2.94	
5	В	0.1	1.12	
5	С	0.0	1.09	
5	D	12.0	2.85	
6	C	0.0	1.34	
6	D	12.2	2.20	

Table S2 Relative energy (the minimum of each set of isomers is set to 0 kJ mol⁻¹) and the magnitude of the dipole moment of optimised cage structures formed from ligands **4**, **5** and **6**.

Ligand Distortion Measurements

Ligand distortion measures (Fig. S128) used in this work were adapted from related work reported by Lusby, Duarte and co-workers.¹¹

The optimised pore diameters were calculated using pyWindow¹² (Python scripts are made available). All guests, ions and solvent molecules were removed from the cages extracted from SCXRD structures for pyWindow analysis.



Figure S137 Alkyne twist angles (ϕ) were measured as the torsion angle defined by the carbon atoms *a*, *b*, *c* and *d* for both a) ligands **L** and **1**, and b) **5** and **6**. c) Ligand twist angles (θ) were measured as the torsion angle defined by the two Pd-N bonds of an individual ligand.

Table S3 Ligand distortion and intrinsic porosity parameters obtained from SCXRD and calculated structures. (a) SCXRD data reported by Hooley and co-workers;¹³ (b) SCXRD data reported by Lusby and co-workers.¹⁴

Ligand	Isomer	Lissend twist angles (0)	Alkyne twist angles (φ)	Pd…Pd	Pore
Ligano		Liganu twist angles (0)		distance	diameter
L ^(a)	_		3.01°	11 9F2 Å	c cr Å
		1.00°	3.12°		
		0.78°	0.78° 6.99°		0.05 A
			7.21°		
L ^(b)	_		6.42°	12.215 Å	6.35 Å
		0.56°	8.47°		
		0.86°	12.53°		
			22.40°		
1	D	41.83°	17.24°	11.866 Å	6.26 Å
			29.65°		
5	с	25.82°	0.55°	9.697Å	4.36 Å
		27.25°	2.41°		
		27.53°	4.21°		
		27.54°	47.70°		
6	с	39.48°	11.16°		
		40.01°	18.05°	9.904 Å	л л <u>ө</u> Å
		42.05°	18.32°		4.40 A
		42.81°	57.06°		

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