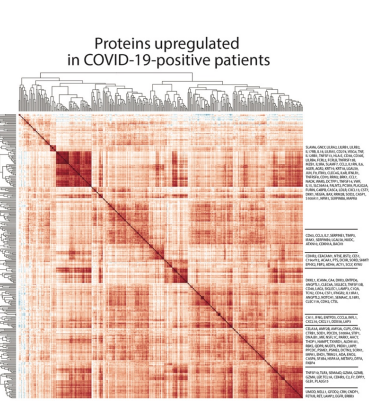
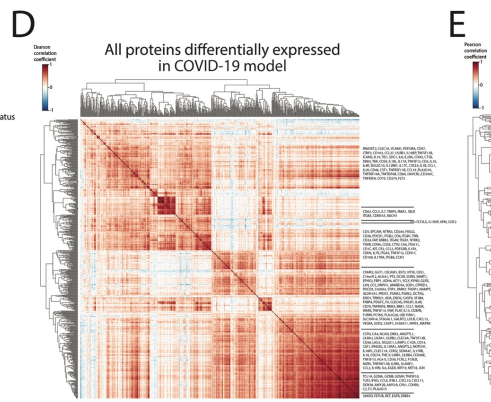
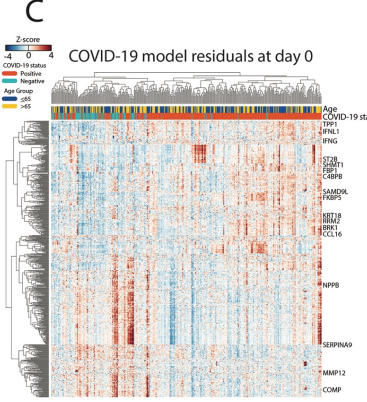
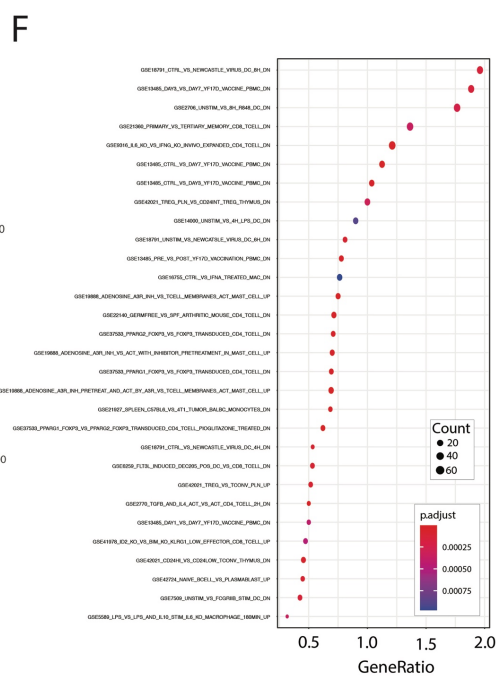
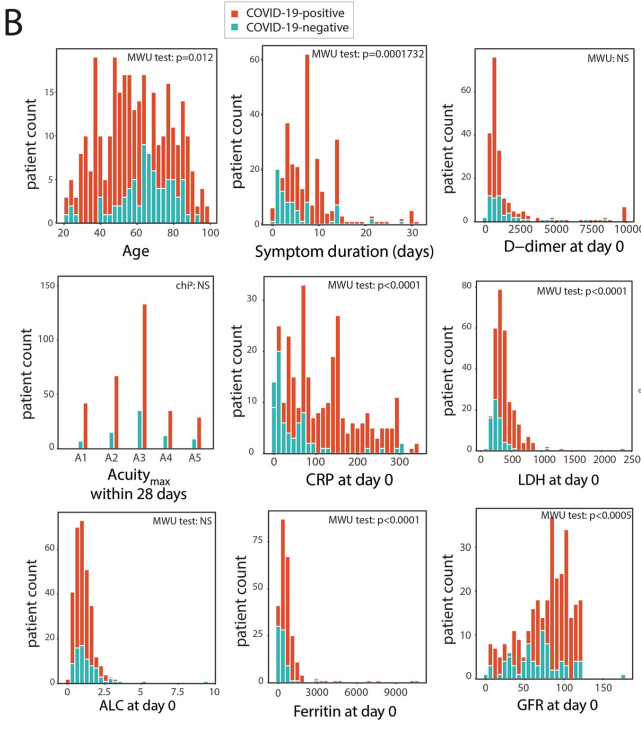
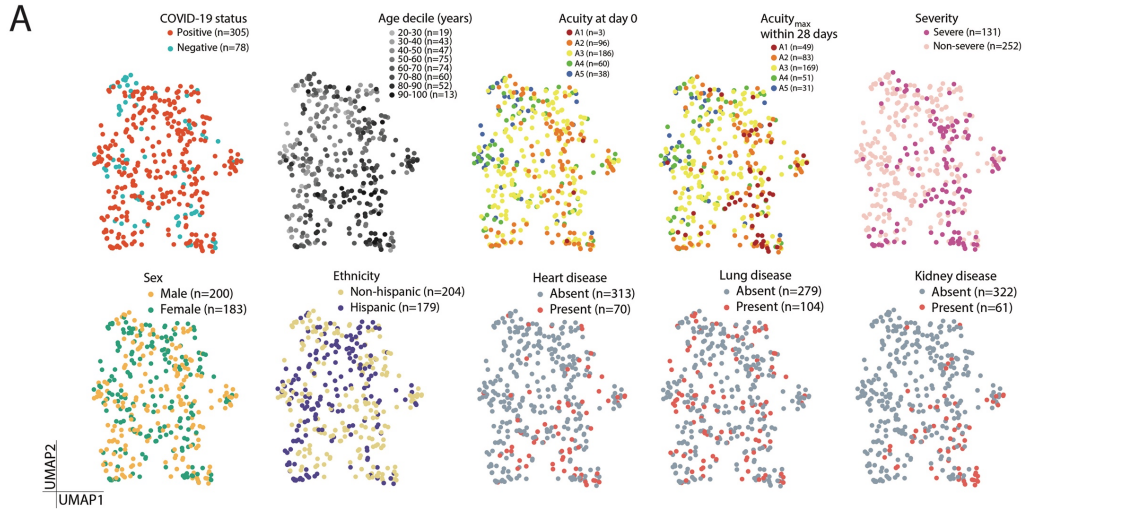


**Supplemental information**

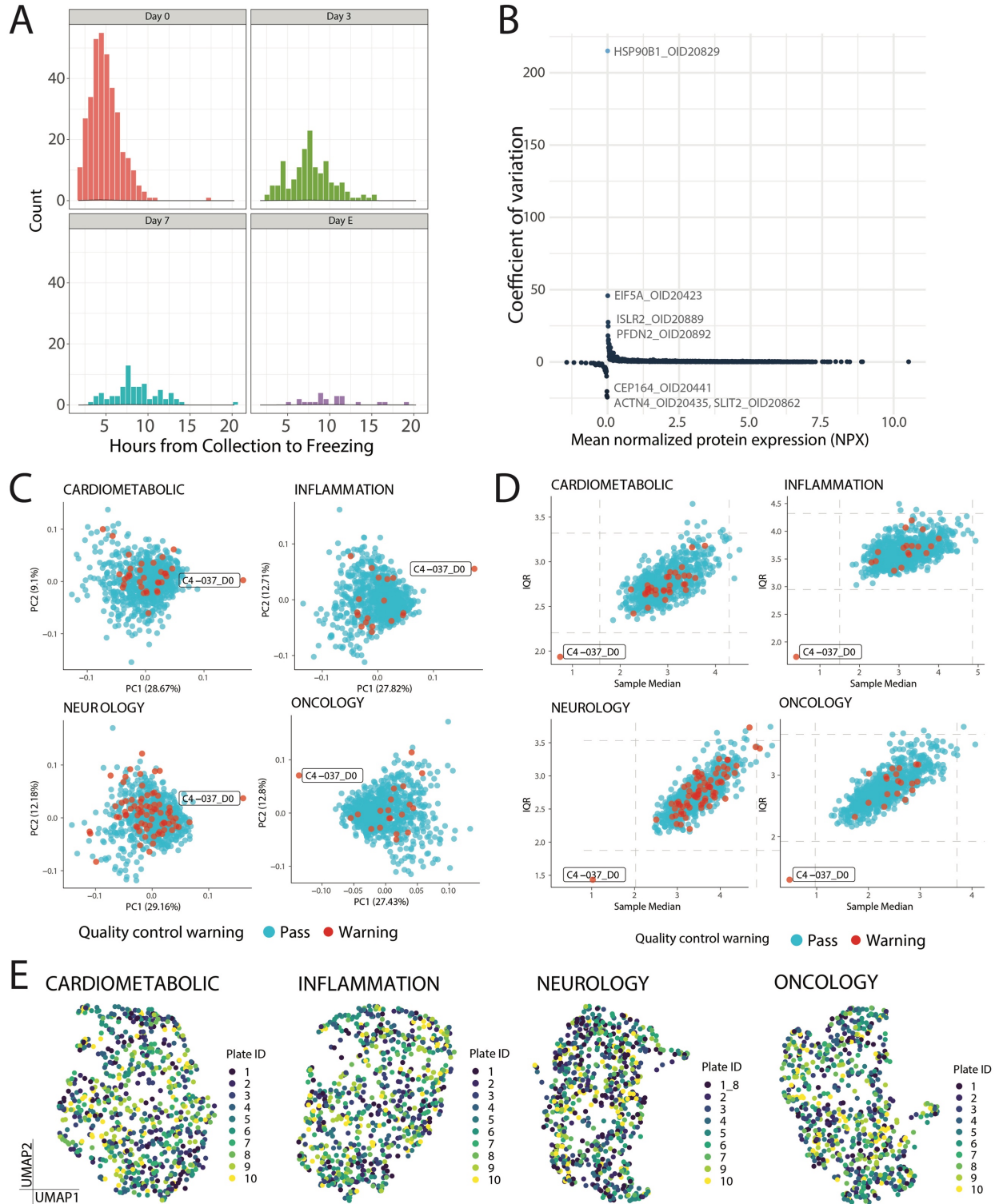
**Longitudinal proteomic analysis of severe COVID-19  
reveals survival-associated signatures, tissue-specific  
cell death, and cell-cell interactions**

**Michael R. Filbin, Arnav Mehta, Alexis M. Schneider, Kyle R. Kays, Jamey R. Guess, Matteo Gentili, Bánk G. Fenyves, Nicole C. Charland, Anna L.K. Gonye, Irena Gushterova, Hargun K. Khanna, Thomas J. LaSalle, Kendall M. Lavin-Parsons, Brendan M. Lilley, Carl L. Lodenstein, Kasidet Manakongtreecheep, Justin D. Margolin, Brenna N. McKaig, Maricarmen Rojas-Lopez, Brian C. Russo, Nihaarika Sharma, Jessica Tantivit, Molly F. Thomas, Robert E. Gerszten, Graham S. Heimberg, Paul J. Hoover, David J. Lieb, Brian Lin, Debby Ngo, Karin Pelka, Miguel Reyes, Christopher S. Smillie, Avinash Waghay, Thomas E. Wood, Amanda S. Zajac, Lori L. Jennings, Ida Grundberg, Roby P. Bhattacharyya, Blair Alden Parry, Alexandra-Chloé Villani, Moshe Sade-Feldman, Nir Hacohen, and Marcia B. Goldberg**

## Supplemental Figures

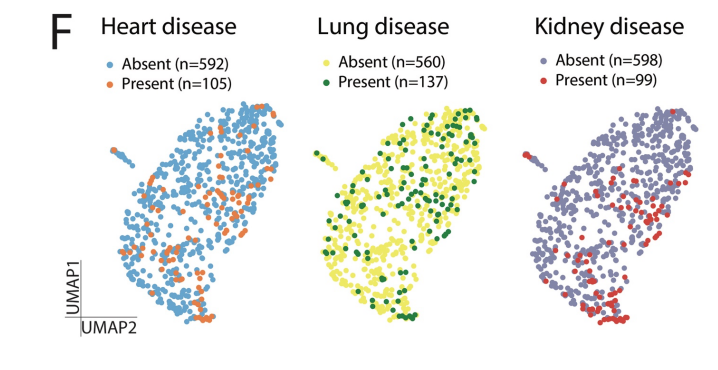
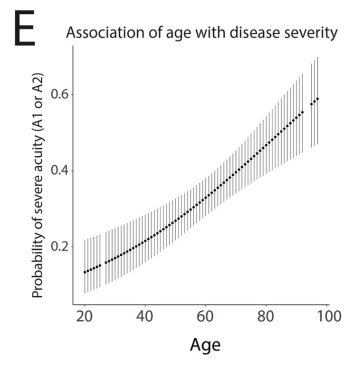
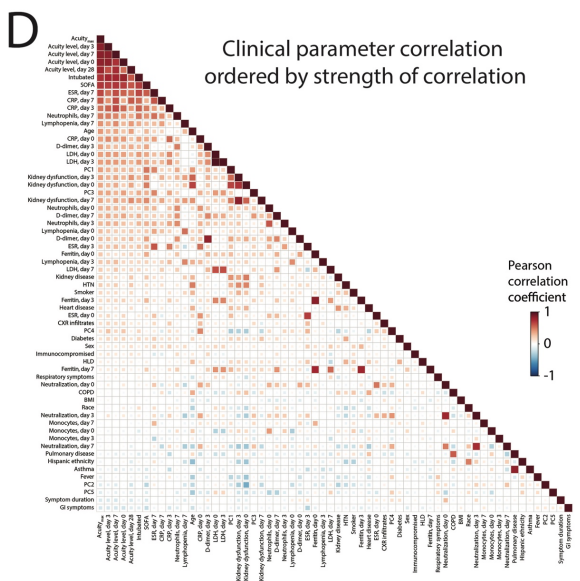
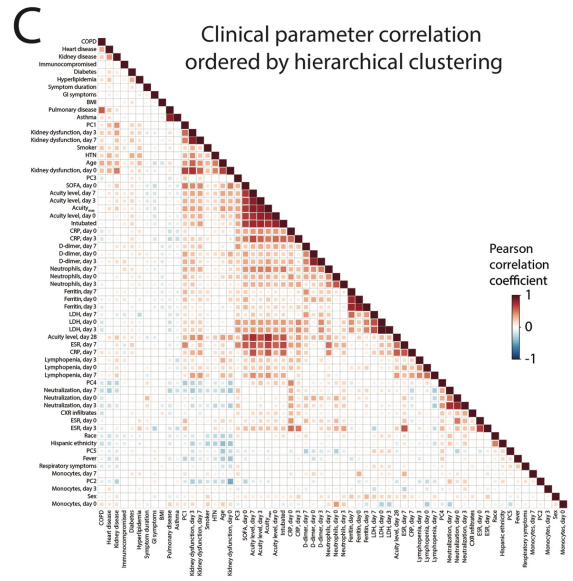
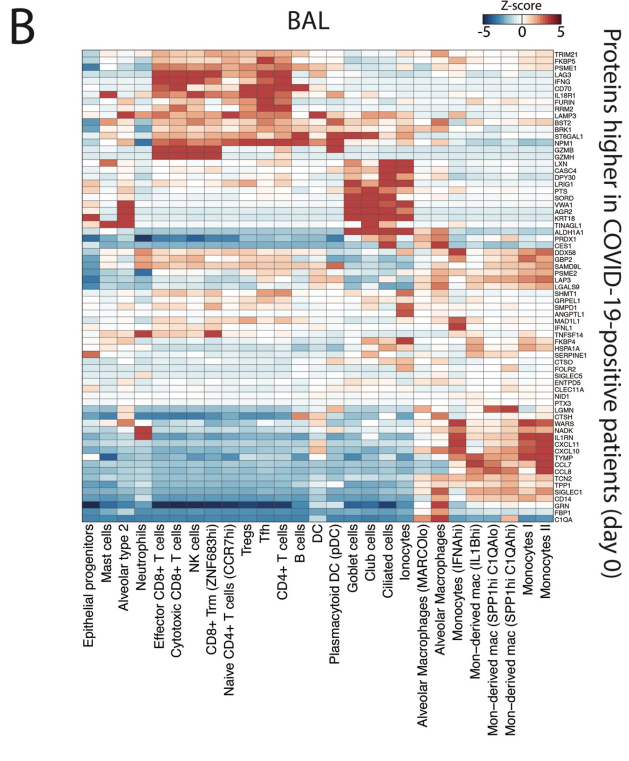
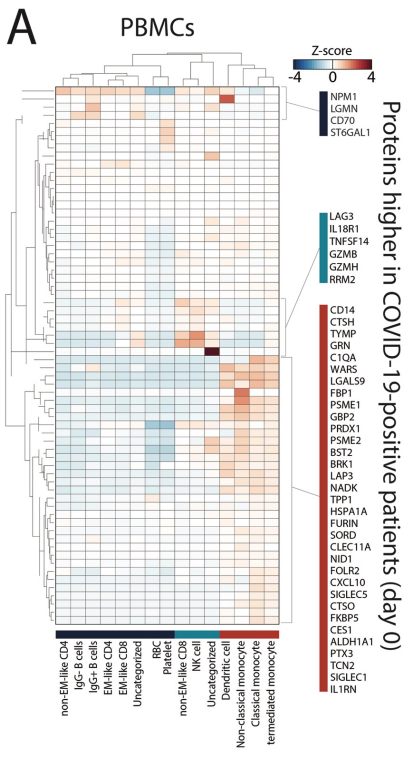


**Supplemental Figure 1.** Distributions of clinical characteristics and differentially-expressed plasma proteins between COVID-19-positive and COVID-19-negative patients in the study cohort. Related to Figure 1 and Tables S1, S2, and S3. (A) Unsupervised clustering (by UMAP), generated from plasma proteins of all patients on day 0, color-coded (left to right) by COVID-19 status, age decile, acuity level at day 0, Acuity<sub>max</sub> within 28 days, severity, gender, ethnicity, previously known heart disease, previously known lung disease, and previously known kidney disease. (B) Age, symptom duration prior to ED arrival, D-dimer, Acuity<sub>max</sub>, CRP, LDH, absolute lymphocyte count (ALC), ferritin, and glomerular filtration rate (GFR). All measurements, except Acuity<sub>max</sub> and symptom duration are at day 0. Significance is by the Mann-Whitney U-test (MWU) and chi-squared test ( $\chi^2$ ); p-values as indicated. (C)-(F) Differentially-expressed proteins by COVID-19 status. (C) Identification of differentially-expressed proteins by COVID-19 status using linear model of Olink proteins with putative confounders as covariates (see **STAR methods**). Heatmap of residuals from this model for each protein found to be significantly differentially-expressed between COVID-19-positive and negative patients, using the model described in **Figure 1C-1E**. (D) Correlation heatmap of plasma protein levels of all proteins significantly differentially-expressed between COVID-19-positive and negative patients. (E) Correlation heatmap of plasma protein levels of all proteins significantly higher in COVID-19-positive patients than in COVID-19-negative patients. (F) Gene-set enrichment analysis for pathways enriched among plasma proteins differentially-expressed between COVID-19-positive and negative patients. Linear model fitting each Olink protein, with COVID-19 status as a main effect and putative confounders as covariates (see **STAR methods**). p-values were adjusted to control the false discovery rate (FDR) at  $<0.05$ , Benjamini-Hochberg method.



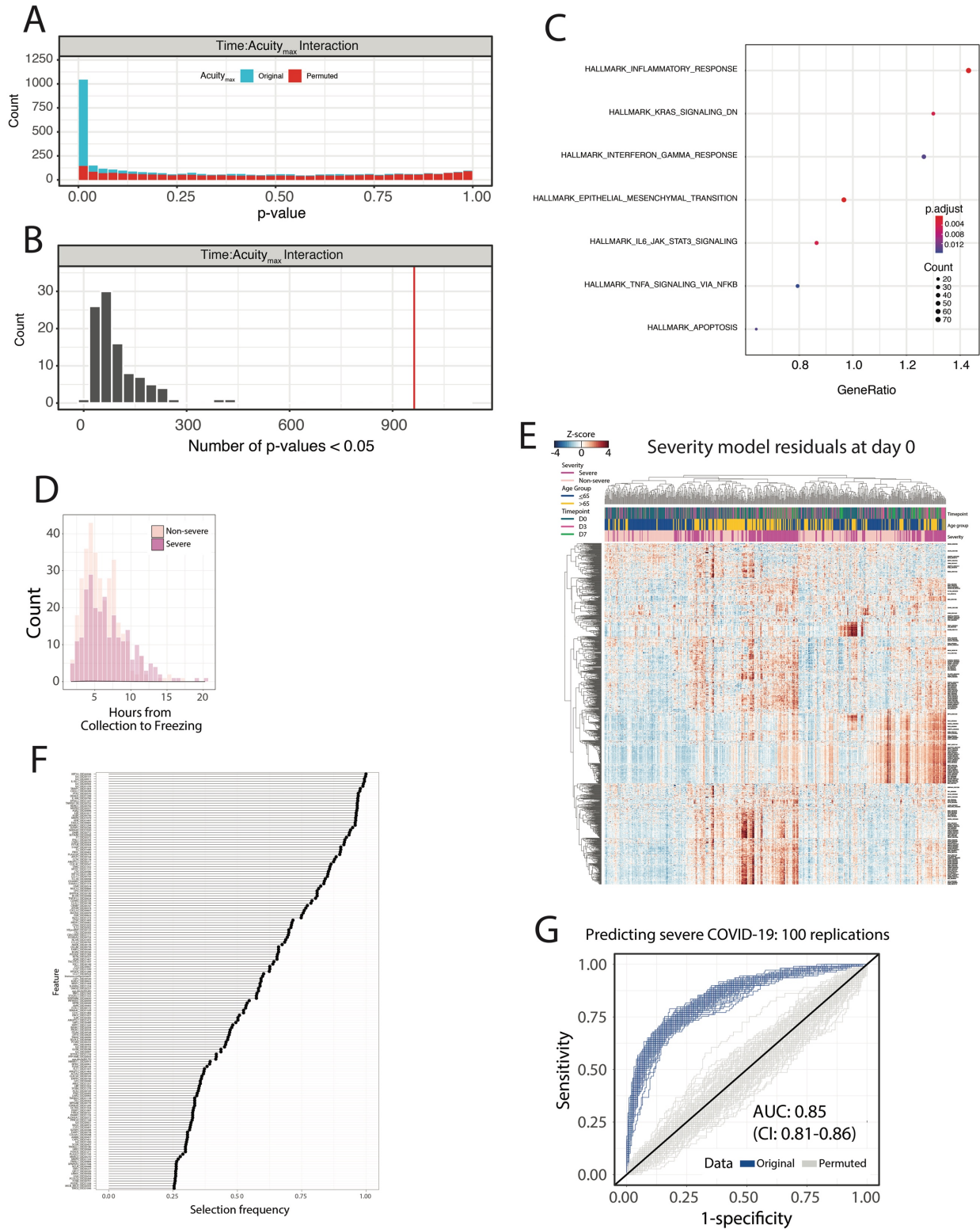
**Supplemental Figure 2.** Quality control for Olink plasma proteomics data. Related to Figures 1 and 2, and Table S2. (A) Histogram showing the time from collection to freezing for each sample, separated by time point of collection. E, event-driven samples (see **STAR methods**). (B) Histogram showing the

coefficient of variation versus mean normalized protein expression (NPX) for each of the 1356 assays performed by Olink. (C) Principal component plots using Olink assays in four predefined Olink panels (cardiometabolic, inflammation, neurology, and oncology), each with >300 protein assays. Shown are first two principal components (PC1 vs. PC2) for all patient samples. (D) Scatterplot showing interquartile range (IQR) versus sample median for each sample. In (C) and (D), sample C4-037\_D0 was noted to be an outlier and was therefore excluded from downstream analyses. (E) Unsupervised clustering by UMAP of all samples by assays in each predefined panel as in (C-D), color-coded by plate label, confirming there were no plate-related biases.



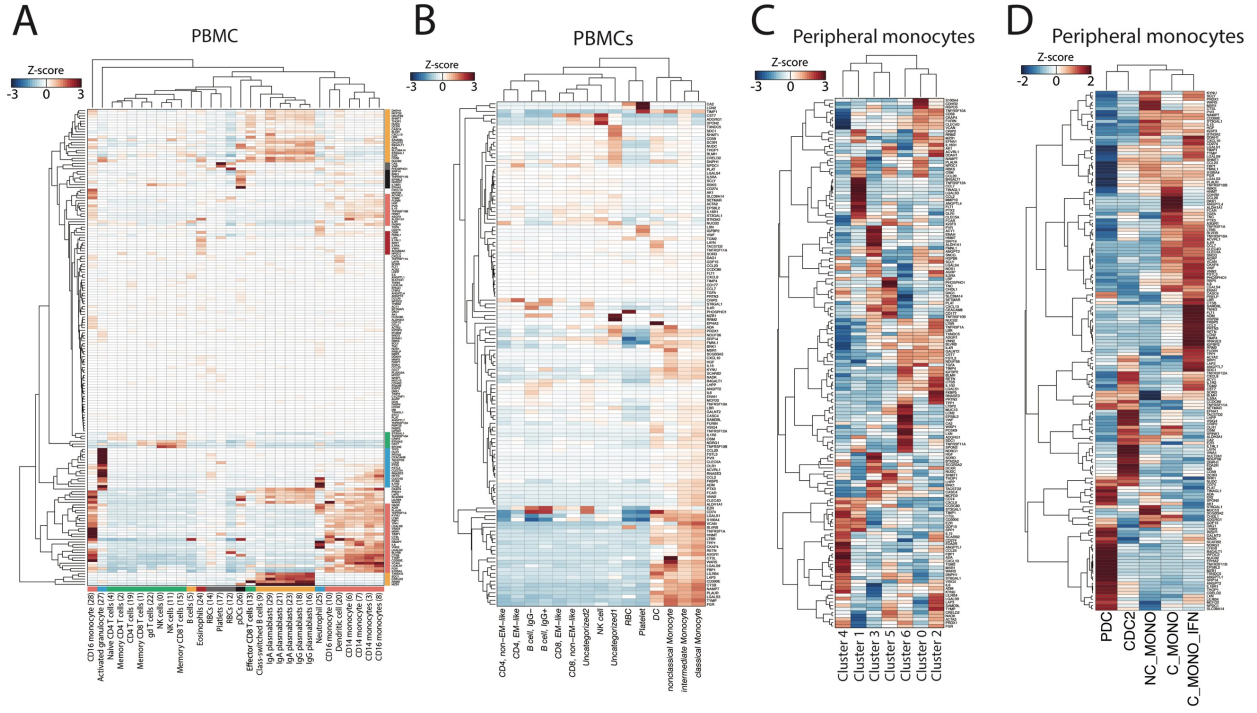
**Supplemental Figure 3.** Inference of cell of origin and clinical correlates derived by mapping gene expression of differentially-expressed plasma proteins (elevated in COVID-19-positive versus COVID-19-negative patients) onto scRNAseq datasets. Related to Figures 1 and 2, and Table S2. (A)-(B) Heatmaps of mean expression of COVID-19-related proteins (y-axis) in immune cell subtypes (x-axis) from peripheral blood cell scRNAseq dataset in Lee *et al.*<sup>1</sup> (A) and BAL COVID-19 scRNAseq dataset in Bost *et al.*<sup>2</sup> (B). non-EM-like, non-effector memory-like; EM-like, effector memory-like; Trm, resident memory CD8<sup>+</sup> T cells; DC, dendritic cells; Mon-derived mac, monocyte-derived macrophages. (C)-(D) Correlation heatmap of all clinically associated variables collected in this study ordered by hierarchical clustering (C) or correlation to Acuity<sub>max</sub> (D). For the purpose of displaying positive correlation with increasing disease severity, inverse numbering was assigned to A1-A5 for these calculations, and kidney function was measured as the inverse of the glomerular filtration rate (GFR). PC1 to PC5 are the loadings of the first five principal components for the PCA plot of all samples using all proteins. SOFA, sequential organ failure assessment score; monocytes, monocyte count; neutrophils, absolute neutrophil count. (E) Logistic regression model predicting probability of severe acuity (Acuity<sub>max</sub> of A1 or A2) using patient age. (F) Unsupervised clustering by UMAP using plasma proteins of all COVID-19-positive patients at days 0, 3 and 7, color-coded (left to right) by previously known heart disease, previously known lung disease, and previously known kidney disease.



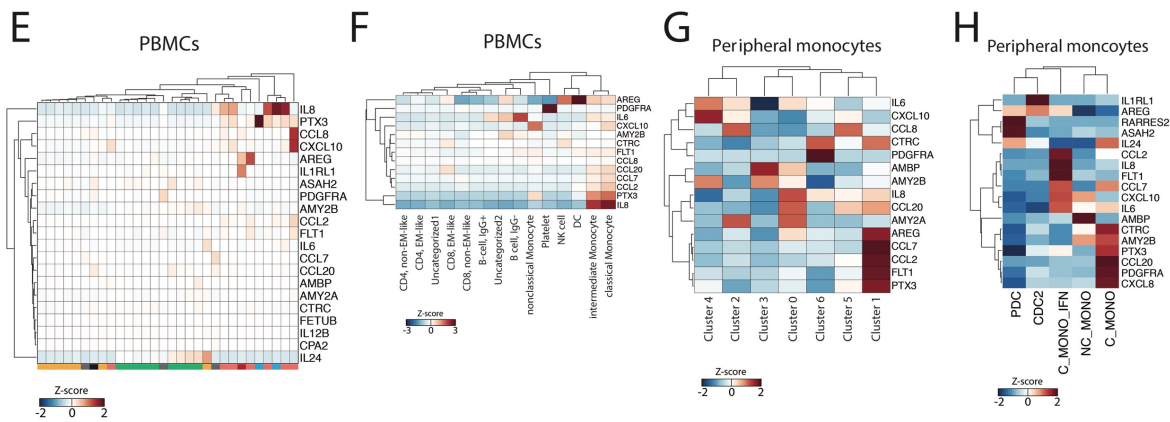


**Supplemental Figure 4.** Permutation analysis for linear mixed model on Olink data, with Acuity<sub>max</sub> and time as main effects. Related to Figure 2, and Tables S2 and S3. (A) Linear mixed model fitting each

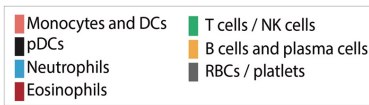
Olink protein, with Acuity<sub>max</sub>, timepoint, and the interaction of the two terms as main effects and putative confounders as covariates (see **STAR methods**). Significance of the three model terms was determined with an F-test using Satterthwaite degrees of freedom and type III sum of squares. p-values for the three model terms of interest were adjusted to control the FDR at < 0.05, Benjamini-Hochberg method. Group differences were calculated for each protein passing the 0.05 FDR threshold with p-values adjusted using the Tukey method. Shown are the distribution of p-values obtained from this linear mixed model and the distribution of p-values obtained from permutations of random Acuity<sub>max</sub> assignments to all patients. This result shows that the obtained distribution of p-values does not happen by chance. (B) Distribution of the number of p-values < 0.05 obtained from 100 permutations of random Acuity<sub>max</sub> assignments for the linear mixed model described in (A). The red line indicates the number of significant assays obtained from our true model with correct patient Acuity<sub>max</sub> assignments. (C)-(G) Protein associations with disease severity. (C) Gene-set enrichment analysis for pathways enriched among plasma proteins differentially-expressed between severe and non-severe COVID-19-positive patients. (D) Time from sample collection to freezing for all samples, color-coded by COVID-19 severity, indicating no clear difference between samples from severe and samples from non-severe patients. (E) Linear model fitting each Olink protein, with putative confounders as covariates (see **STAR methods**). Heatmap of residuals from this model for each protein that was found to be significant for interaction term in the model described in **Fig. 2C**. (F) Frequency of protein feature selection over the 100 iterations of the predictor in (G). (G) Receiver operating characteristic (ROC) curve showing predictive performance of an elastic net logistic regression classifier of disease severity using Olink plasma proteins for each patient at day 0. Shown are curves for each of the 100 repeats of 5-fold cross validation. Measures of performance are reported as medians and 95% confidence intervals. Shown in the figure are the ROC curves for the original data with the true severity labels and the curves for our data with random (permuted) severity assignments.



Proteins significantly increased at day 0 in severe (A1, A2) vs. non-severe (A3-5) patient

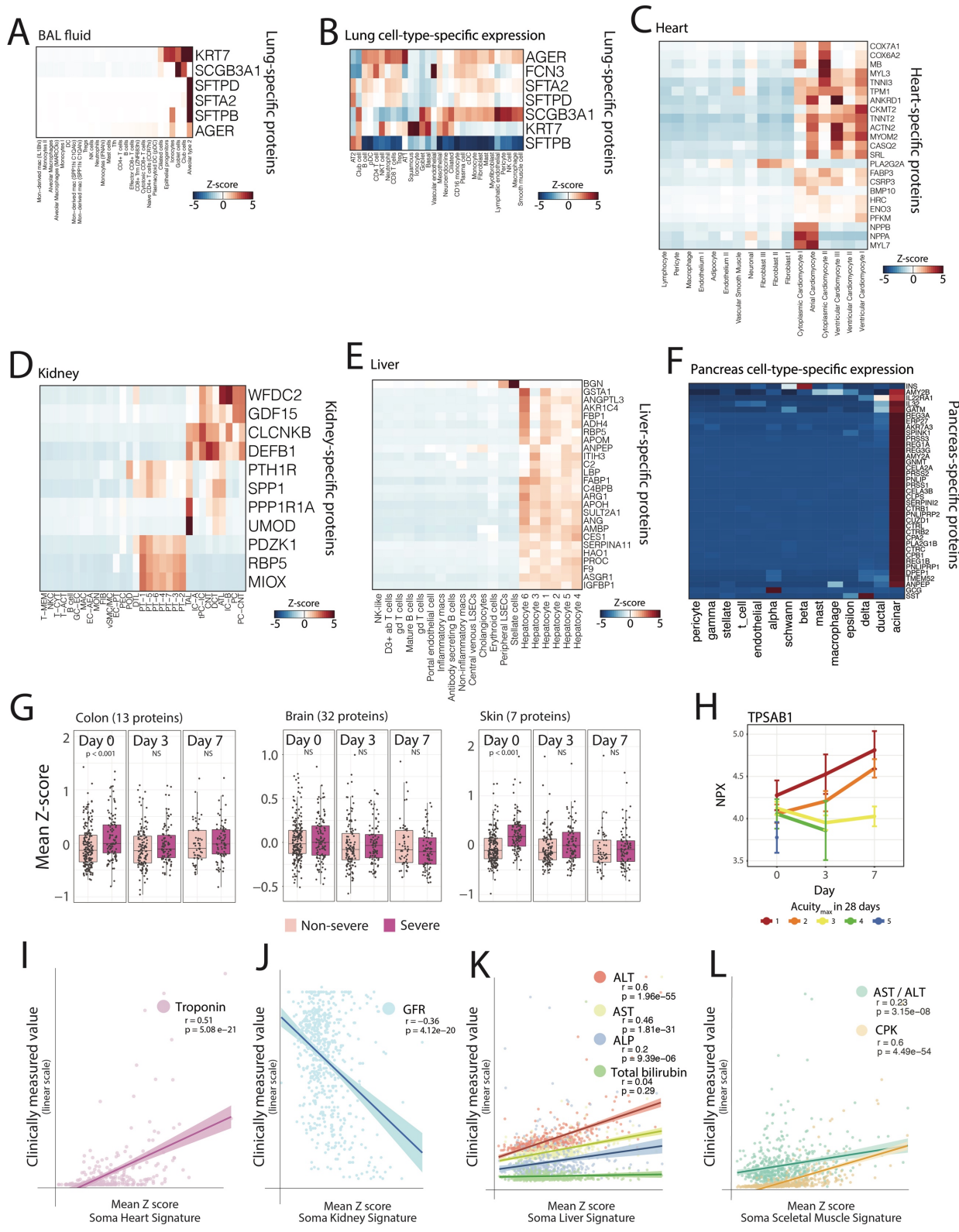


Differentially expressed proteins at day 7 between Acuity<sub>max</sub> A1 vs. A2



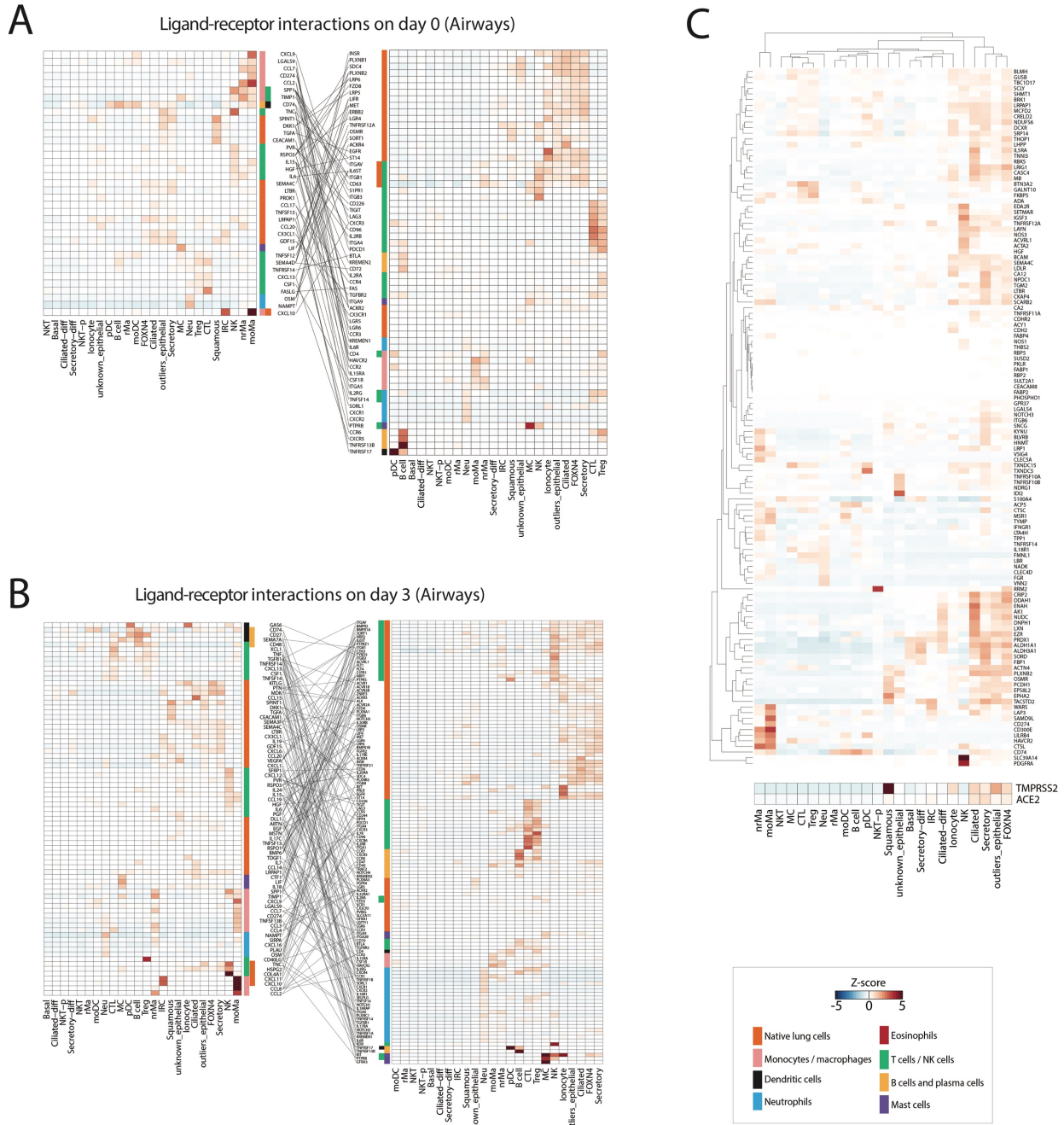
**Supplemental Figure 5.** Expression of severity-associated plasma proteins in PBMCs from COVID-19 patients. Related to Figures 1 and 5, and Tables S2 and S4. (A) Gene expression of Olink plasma proteins expressed more highly in severe COVID-19 patients shown within PBMCs using scRNAseq data.<sup>3</sup> (B)-

(D) Expression of Olink plasma proteins expressed more highly in severe COVID-19 patients within PBMCs show within another independent dataset<sup>1</sup> (B), or circulating monocytes, from two independent datasets<sup>4,5</sup> (C)-(D). (E)-(H) Expression of differentially-expressed Olink proteins at day 7 between patients whose maximum classification was A1 (death) versus A2 (ARDS but survived) within day 28, for two independent PBMC datasets<sup>1,3</sup> (E)-(F) and two independent peripheral monocyte datasets<sup>4,5</sup> (G)-(H). non-EM-like, non-effector memory-like; EM-like, effector memory-like; DC, dendritic cells; pDC, plasmacytoid DCs; CDC2, C\_MONO, NC\_MONO, dendritic and monocyte clusters as defined in <sup>4,5</sup>.



**Supplemental Figure 6.** Organ-specific cellular death signatures (from SomaScan data). (A)-(B)

Expression of derived lung specific plasma proteins (secreted, membrane and intracellular) within BAL fluid from COVID-19 patients.<sup>2</sup> Related to Figures 5 and 6. (A) and an integrated analysis of several lung scRNAseq datasets (B). (C) Heatmap showing cell-type expression of severity-associated intracellular proteins mapped to cell types in scRNAseq data of normal cardiac tissue.<sup>6</sup> Kidney, pancreatic and liver-specific cellular death signatures. (D) Heatmap showing cell-type expression of severity-associated intracellular proteins mapped to normal kidney tissue.<sup>7</sup> (E) Heatmap showing cell-type expression of severity-associated intracellular proteins mapped to normal liver tissue.<sup>8</sup> (F) Heatmap showing cell-type expression of severity-associated intracellular proteins mapped to normal pancreatic tissue expression matrix obtained from the Itai Yanai lab.<sup>9</sup> (G) Expression of tissue-specific plasma protein signatures in severe versus non-severe patients at each timepoint in select tissues. (H) Expression of the mast cell marker tryptase (TPSAB1 assay by Olink) over time, by Acuity<sub>max</sub> level. (I)-(L) Organ damage plasma protein signature correlations with indicated clinical laboratory measurements at day 0. GFR, glomerular filtration rate; ALT, alanine aminotransferase; AST, aspartate aminotransferase; ALP, alkaline phosphatase; CPK, creatinine phosphokinase.



**Supplemental Figure 7.** Cellular communication and intracellular severity-associated proteins in the airway. Related to Figures 5 and 6, and Table S2. (A)-(B) Ligand-receptor relationships of severity-associated plasma ligands in an independent dataset of nasopharyngeal and bronchial samples from COVID-19 patients<sup>10, 11</sup> for ligands significant on day 0 (A) or day 3 (B). (C) Heatmap showing mean gene expression per cell type of severity-associated intracellular plasma proteins at D0 derived from

Olink data that map to scRNAseq of the nasopharyngeal and bronchial airway,<sup>10, 11</sup> with TMPRSS2 and ACE2 expression indicated. NKT, natural killer (NK) T cells; Ciliated-diff, differentiating secretory cells; Secretory-diff, differentiating secretory cells; NKT-p, proliferating NK T cells; pDC, plasmacytoid dendritic cells; rMa, resident macrophages; moDC, monocyte-derived dendritic cells; MC, mast cells; Neu, neutrophils; Treg, regulatory T cells; CTL, cytotoxic T cells; IRC, IFN $\gamma$  responsive cells; nrMa, non-resident macrophages; moMa, monocyte-derived macrophages; rMa, resident macrophages.



## Supplemental References

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