

SUPPORTING INFORMATION

Discovery of 5-nitro-6-thiocyanatopyrimidines as inhibitors of *Cryptococcus neoformans* and *Cryptococcus gattii*

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Short running title: 5-Nitro-6-thiocyanatopyrimidine antifungals

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Experimental Procedures

Reagents and physicochemical analysis of the synthesized compounds

All reagents and solvents were purchased from commercial suppliers and used without further purification. ¹H spectra were measured on Bruker AC-300 (300 MHz). Chemical shifts were measured in DMSO-d₆, using tetramethylsilane as an internal standard, and reported as units (ppm) values. Mass spectra were recorded on Finnigan MAT INCO 50 mass spectrometer (EI, 70 eV) with direct injection. The purity of the final compounds were analyzed on an Agilent 1290 Infinity II HPLC system coupled to Agilent 6460 triple-quadrupole mass spectrometer equipped with an electrospray ionization source. All final compounds are > 95 % pure. Elemental analysis (% C, H, N) was carried out by an elemental analyzer EURO EA. Melting points were determined on Electrothermal 9001 (10 °C per min) and are uncorrected. Merck KGaA silica gel 60 F₂₅₄ plates were used for analytical thin-layer chromatography. Yields refer to purified products and are not optimized.

2-Substituted 4,6-dihydroxy-5-nitropyrimidines **1-6** and 4,6-dichloro-5-nitropyrimidines **7-12** were synthesized with minimal changes according to the procedures described in Latli *et al.*¹. Physicochemical data for intermediates **13-62** are reported in the literature²⁻⁵ and are not shown in the present file. General synthetic procedures and physicochemical characteristics of the final 4-thiocyano-5-nitropyrimidines **63-112** are presented as follows.

General procedure for the synthesis of 2-substituted 4-amino-6-chloro-5-nitropyrimidines

To a solution of 2-substituted 4,6-dichloro-5-nitropyrimidines (1 eqv.) in dioxane (40 mL), freshly prepared corresponding amine acetate (2.5 eqv.) was added at $T < 15$ °C. The mixture was kept at room temperature for 3 h and then diluted with water (200 mL). The precipitate was collected, washed with cold water and used without purification.

General procedure for the synthesis of 2-substituted 4-alkoxy-6-chloro-5-nitropyrimidines

To a solution of 2-substituted 4,6-dichloro-5-nitropyrimidines (1 eqv.) in the corresponding anhydrous alcohol (40 mL), the corresponding freshly prepared sodium alkoxide (1 eqv., 20 mL) was added at $T < 0$ °C, and the mixture was kept at room temperature from 2 h to 4 h. After completion of the reaction, the mixture was diluted with water (200 mL), and the precipitate of the corresponding products was collected, washed with cold water and recrystallized from benzene.

General procedure for the synthesis of 2,4-disubstituted 5-nitro-6-thiocyanatopyrimidines

A mixture of 2,4-substituted 6-chloro-5-nitropyrimidines (1 eqv.) in the corresponding alcohol (30 mL) and potassium thiocyanate (1.1 eqv.) was refluxed for 4 h. After completion of the reaction, activated carbon (0.1 g) was added, the hot mixture was stirred for 1 h and filtered off. The filtrate was cooled and then was diluted with water (100 mL). The precipitate of the final products was collected, washed with cold water and recrystallized from the corresponding solvent.

4-Methoxy-5-nitro-6-thiocyanatopyrimidine 63

Yield 75 %, 131-133 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.05 (3H, s, CH₃), 8.24 (1H, s, CH_{ar}). MS (EI): m/z 212 (M⁺). Anal. calcd for C₆H₄N₄O₃S: C, 33.96; H, 1.90; N, 26.41. Found: C, 33.89; H, 1.96; N, 26.37.

4-Propoxy-5-nitro-6-thiocyanatopyrimidine 64

Yield 76 %, 107-109 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.21 (3H, t, CH₃), 1.86 (2H, m, CH₂), 4.46 (2H, t, OCH₂), 8.29 (1H, s, CH_{ar}). MS (EI): m/z 240 (M⁺). Anal. calcd for C₈H₈N₄O₃S: C, 40.00; H, 3.36; N, 23.32. Found: C, 40.06; H, 3.30; N, 23.29.

4-Isopropoxy-5-nitro-6-thiocyanatopyrimidine 65

Yield 79 %, 96-99 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.24 (6H, s, 2CH₃), 4.72 (1H, qui, CH), 8.27 (1H, s, CH_{ar}). MS (EI): m/z 240 (M⁺). Anal. calcd for C₈H₈N₄O₃S: C, 40.00; H, 3.36; N, 23.32. Found: C, 40.08; H, 3.39; N, 23.38.

5-Nitro-6-thiocyanatopyrimidin-4-amine 66

Yield 84 %, 215 °C (decomp.) (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 8.34 (1H, s, CH_{ar}), 8.53 (2H, brs, NH₂). MS (EI): m/z 197 (M⁺). Anal. calcd for C₅H₃N₅O₂S: C, 30.46; H, 1.53; N, 35.52. Found: C, 30.42; H, 1.57; N, 35.47.

N-Isopropyl-5-nitro-6-thiocyanatopyrimidin-4-amine 67

Yield 81 %, 151-153 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.37 (6H, s, 2CH₃), 4.86 (1H, m, CH), 8.16 (1H, s, CH_{ar}), 9.46 (1H, brs, NH). MS (EI): m/z 239 (M⁺). Anal. calcd for C₈H₉N₅O₂S: C, 40.16; H, 3.79; N, 29.27. Found: C, 40.03; H, 3.88; N, 29.11.

N-(3-Methylbutan-2-yl)-5-nitro-6-thiocyanatopyrimidin-4-amine 68

Yield 78 %, 132-134 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.08 (6H, s, 2CH₃), 1.17 (3H, s, CH₃), 1.55 (1H, m, CH), 2.53 (1H, qui, CH), 8.18 (1H, s, CH_{ar}), 9.39 (1H, brs, NH).

MS (EI): m/z 267 (M⁺). Anal. calcd for C₁₀H₁₃N₅O₂S: C, 44.93; H, 4.90; N, 26.20. Found: C, 44.88; H, 4.97; N, 26.25.

N-(Pentan-3-yl)-5-nitro-6-thiocyanatopyrimidin-4-amine 69

Yield 72 %, 125-126 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.13 (6H, s, 2CH₃), 1.51 (4H, brm, 2CH₂), 2.43 (1H, qui, CH), 8.23 (1H, s, CH_{ar}), 9.41 (1H, brs, NH). MS (EI): m/z 267 (M⁺). Anal. calcd for C₁₀H₁₃N₅O₂S: C, 44.93; H, 4.90; N, 26.20. Found: C, 44.97; H, 4.86; N, 26.25.

N-Cyclopentyl-5-nitro-6-thiocyanatopyrimidin-4-amine 70

Yield 77 %, 146-149 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.50-2.02 (8H, m, 4CH₂), 2.64 (1H, t, CH), 8.21 (1H, s, CH_{ar}), 9.39 (1H, brs, NH). MS (EI): m/z 265 (M⁺). Anal. calcd for C₁₀H₁₁N₅O₂S: C, 45.27; H, 4.18; N, 26.40. Found: C, 45.31; H, 4.13; N, 26.49.

N-Cyclohexyl-5-nitro-6-thiocyanatopyrimidin-4-amine 71

Yield 74 %, 141-142 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.11-1.21 (4H, m, 2CH₂), 1.48-1.57 (6H, m, 3CH₂), 2.59 (1H, qui, CH), 8.23 (1H, s, CH_{ar}), 9.38 (1H, brs, NH). MS (EI): m/z 279 (M⁺). Anal. calcd for C₁₁H₁₃N₅O₂S: C, 47.30; H, 4.69; N, 25.07. Found: C, 47.37; H, 4.64; N, 25.01.

4-(Piperidin-1-yl)-5-nitro-6-thiocyanatopyrimidine 72

Yield 58 %, 152-154 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.51-1.60 (6H, m, 3CH₂), 3.69-3.73 (4H, m, 2CH₂), 8.25 (1H, s, CH_{ar}). MS (EI): m/z 265 (M⁺). Anal. calcd for C₁₀H₁₁N₅O₂S: C, 45.27; H, 4.18; N, 26.40. Found: C, 45.35; H, 4.21; N, 26.36.

4-(2-Methylpiperidin-1-yl)-5-nitro-6-thiocyanatopyrimidine 73

Yield 54 %, 158-160 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.33 (3H, s, CH₃), 1.70 and 3.65 (9H, two brm, CH(CH₂)₄), 8.21 (1H, s, CH_{ar}). MS (EI): m/z 279 (M⁺). Anal. calcd for C₁₁H₁₃N₅O₂S: C, 47.30; H, 4.69; N, 25.07. Found: C, 47.11; H, 4.54; N, 25.12.

4-(2,6-Dimethylpiperidin-1-yl)-5-nitro-6-thiocyanatopyrimidine 74

Yield 61 %, 115-116 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.18 (6H, s, 2CH₃), 1.30-1.56 (6H, 3CH₂), 3.19 (2H, m, 2CH), 8.28 (1H, s, CH_{ar}). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.17; H, 5.19; N, 23.80.

4-(3,5-Dimethylpiperidin-1-yl)-5-nitro-6-thiocyanatopyrimidine 75

Yield 67 %, 123-125 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.19 (6H, s, 2CH₃), 1.21 (2H, m, CH₂), 1.97 (2H, m, 2CH), 3.02 and 3.97 (4H, 2 brm, N(CH₂)₂), 8.34 (1H, s, CH_{ar}). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.20; H, 5.12; N, 23.83.

1-(5-Nitro-6-thiocyanatopyrimidin-4-yl)azepane 76

Yield 71 %, 112 °C (decomp.) (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.51-1.77 (8H, brm, 4CH₂), 3.69 (4H, brm, N(CH₂)₂), 8.26 (1H, s, CH_{ar}). MS (EI): m/z 279 (M⁺). Anal. calcd for C₁₁H₁₃N₅O₂S: C, 47.30; H, 4.69; N, 25.07. Found: C, 47.35; H, 4.64; N, 25.01.

1-(5-Nitro-6-thiocyanatopyrimidin-4-yl)azocane 77

Yield 74 %, 93-96 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.32-1.50 (10H, brm, 5CH₂), 3.86 (4H, t, N(CH₂)₂), 8.30 (1H, s, CH_{ar}). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.08; H, 5.14; N, 23.80.

N-Benzyl-5-nitro-6-thiocyanatopyrimidin-4-amine 78

Yield 43 %, 144-146 °C (iPrOH). ¹H NMR (DMSO-d₆; δ, ppm): 4.41 (2H, s, CH₂), 7.38-7.25 (5H, m, Ph), 8.38 (1H, s, CH_{ar}), 9.40 (1H, brs, NH). MS (EI): m/z 287 (M⁺). Anal. calcd for C₁₂H₉N₅O₂S: C, 50.17; H, 3.16; N, 24.38. Found: C, 50.29; H, 3.24; N, 24.29.

5-Nitro-6-thiocyanato-N-(3-(trifluoromethyl)benzyl)pyrimidin-4-amine 79

Yield 51 %, 114-116 °C (iPrOH). ¹H NMR (DMSO-d₆; δ, ppm): 4.99 (2H, d, CH₂), 7.48-7.70 (4H, m, Ph), 8.59 (1H, s, CH_{ar}), 9.20 (1H, brs, NH). MS (EI): m/z 355 (M⁺). Anal. calcd for C₁₃H₈F₃N₅O₂S: C, 43.95; H, 2.27; N, 19.71. Found: C, 43.99; H, 2.21; N, 19.78.

N-(4-Chlorobenzyl)-5-nitro-6-thiocyanatopyrimidin-4-amine 80

Yield 45 %, 126-128 °C (iPrOH). ¹H NMR (DMSO-d₆; δ, ppm): 4.81 (2H, d, CH₂), 7.25-7.45 (4H, m, Ph), 8.59 (1H, s, CH_{ar}), 9.18 (1H, brs, NH). MS (EI): m/z 321 (M⁺). Anal. calcd for C₁₂H₈ClN₅O₂S: C, 44.80; H, 2.51; N, 21.77. Found: C, 44.85; H, 2.47; N, 21.72.

5-Nitro-N-phenethyl-6-thiocyanatopyrimidin-4-amine 81

Yield 44 %, 126-128 °C (iPrOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.97 (2H, t, CH₂), 3.81 (2H, m, CH₂), 7.25 (5H, m, Ph), 8.60 (1H, s, CH_{ar}), 9.42 (1H, brs, NH). MS (EI): m/z 301 (M⁺). Anal. calcd for C₁₃H₁₁N₅O₂S: C, 51.82; H, 3.68; N, 23.24. Found: C, 51.76; H, 3.73; N, 23.29.

N,N-Dimethyl-5-nitro-6-thiocyanatopyrimidin-4-amine 82

Yield 78 %, 142-144 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 3.48 (6H, s, N(CH₃)₂), 8.37 (1H, s, CH_{ar}). MS (EI): m/z 225 (M⁺). Anal. calcd for C₇H₇N₅O₂S: C, 37.33; H, 3.13; N, 31.10. Found: C, 37.23; H, 3.27; N, 29.97.

N,N-Diethyl-5-nitro-6-thiocyanatopyrimidin-4-amine 83

Yield 74 %, 114-116 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.01 (6H, t, 2CH₃), 3.46 (4H, m, 2CH₂), 8.31 (1H, s, CH_{ar}). MS (EI): m/z 253 (M⁺). Anal. calcd for C₉H₁₁N₅O₂S: C, 42.68; H, 4.38; N, 27.65. Found: C, 42.60; H, 4.33; N, 27.74.

N-Ethyl-N-isopropyl-5-nitro-6-thiocyanatopyrimidin-4-amine 84

Yield 79 %, 103-105 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.17 (9H, m, 3CH₃), 2.83 (1H, m, NCH), 3.12 (2H, q, NCH₂), 8.23 (1H, s, CH_{ar}). MS (EI): m/z 267 (M⁺). Anal. calcd for C₁₀H₁₃N₅O₂S: C, 44.93; H, 4.90; N, 26.20. Found: C, 44.87; H, 4.95; N, 26.13.

N-Cyclohexyl-N-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 85

Yield 34 %, 112-114 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.0-2.0 (10H, brm, 5CH₂), 3.12 (3H, s, CH₃), 3.64 (1H, m, NCH), 8.32 (1H, s, CH_{ar}). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.17; H, 5.08; N, 23.84.

N-Benzyl-N-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 86

Yield 46 %, 127-129 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 3.15 (3H, s, CH₃), 4.34 (2H, s, CH₂), 7.20-7.40 (5H, m, Ph), 8.31 (1H, s, CH_{ar}). MS (EI): m/z 301 (M⁺). Anal. calcd for C₁₃H₁₁N₅O₂S: C, 51.82; H, 3.68; N, 23.24. Found: C, 51.76; H, 3.57; N, 23.19.

N-Benzyl-N-isopropyl-5-nitro-6-thiocyanatopyrimidin-4-amine 87

Yield 51 %, 110-112 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.18 (6H, s, 2CH₃), 2.85 (1H, m, NCH), 4.71 (2H, m, NCH₂), 7.25-7.35 (5H, m, Ph), 8.29 (1H, s, CH_{ar}). MS (EI): m/z 329 (M⁺). Anal. calcd for C₁₅H₁₅N₅O₂S: C, 54.70; H, 4.59; N, 21.26. Found: C, 54.71; H, 4.52; N, 21.29.

4-Ethoxy-2-methyl-5-nitro-6-thiocyanatopyrimidine 88

Yield 40 %, 60-62 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.43 (3H, t, CH₃), 2.66 (3H, s, CH₃), 4.64 (2H, q, OCH₂). MS (EI): m/z 240 (M⁺). Anal. calcd for C₈H₈N₄O₃S: C, 40.00; H, 3.36; N, 23.32. Found: C, 39.93; H, 3.46; N, 23.31.

2-Methyl-5-nitro-4-propoxy-6-thiocyanatopyrimidine 89

Yield 26 %, 50-52 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.17 (3H, t, CH₃), 1.77 (2H, m, CH₂), 2.58 (3H, s, CH₃), 4.32 (2H, t, OCH₂). MS (EI): m/z 254 (M⁺). Anal. calcd for C₉H₁₀N₄O₃S: C, 42.51; H, 3.96; N, 22.04. Found: C, 42.45; H, 3.89; N, 22.09.

4-Isopropoxy-2-methyl-5-nitro-6-thiocyanatopyrimidine 90

Yield 33 %, 54-56 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.18 (6H, s, 2CH₃), 2.39 (3H, s, CH₃), 4.65 (1H, m, OCH). MS (EI): m/z 254 (M⁺). Anal. calcd for C₉H₁₀N₄O₃S: C, 42.51; H, 3.96; N, 22.04. Found: C, 42.45; H, 3.92; N, 21.97.

2-Methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 91

Yield 39 %, 209-212 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.43 (3H, s, CH₃), 8.19 (2H, brs, NH₂). MS (EI): m/z 211 (M⁺). Anal. calcd for C₆H₅N₅O₂S: C, 34.12; H, 2.39; N 33.16. Found: C, 34.25; H, 2.47; N, 33.04.

N,2-Dimethyl-5-nitro-6-thiocyanatopyrimidin-4-amine 92

Yield 76 %, 200-202 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.35 (3H, s, CH₃), 3.12 (3H, s, CH₃), 9.29 (1H, brs, NH). MS (EI): m/z 225 (M⁺). Anal. calcd for C₇H₇N₅O₂S: C, 37.33; H, 3.13; N, 31.10. Found: C, 37.21; H, 3.27; N, 31.22.

2-Methyl-5-nitro-N-(pentan-3-yl)-6-thiocyanatopyrimidin-4-amine 93

Yield 79 %, 95-97 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.07 (6H, t, 2CH₃), 1.46 (4H, brm, 2CH₂), 2.40 (4H, m, CH, CH₃), 9.32 (1H, brs, NH). MS (EI): m/z 281 (M⁺). Anal. calcd for C₁₁H₁₅N₅O₂S: C, 46.96; H, 5.37; N, 24.89. Found: C, 47.02; H, 5.32; N, 24.81.

N-Cyclopropyl-2-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 94

Yield 67 %, 104-107 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 0.83-1.12 (4H, brm, 2CH₂), 2.25 (1H, brm, CH), 2.39 (3H, s, CH₃), 9.41 (1H, brs, NH). MS (EI): m/z 251 (M⁺). Anal. calcd for C₉H₉N₅O₂S: C, 43.02; H, 3.61; N, 27.87. Found: C, 43.06; H, 3.68; N, 27.80.

N-Cyclopentyl-2-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 95

Yield 70 %, 124-125 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.57-1.82 (8H, m, 4CH₂), 2.39 (3H, s, CH₃), 2.64 (1H, t, CH), 9.35 (1H, brs, NH). MS (EI): m/z 279 (M⁺). Anal. calcd for C₁₁H₁₃N₅O₂S: C, 47.30; H, 4.69; N, 25.07. Found: C, 47.37; H, 4.75; N, 25.10.

N-Cyclohexyl-2-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 96

Yield 69 %, 95-96 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.01-1.13 (4H, m, 2CH₂), 1.45-1.63 (6H, m, 3CH₂), 2.39 (3H, s, CH₃), 2.57 (1H, qui, CH), 9.38 (1H, brs, NH). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.18; H, 5.21; N, 23.81.

2-Methyl-5-nitro-4-(pyrrolidin-1-yl)-6-thiocyanatopyrimidine 97

Yield 68 %, 142-144 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.91 (4H, m, CH₂CH₂), 2.41 (3H, s, CH₃), 3.50 (4H, m, N(CH₂)₂). MS (EI): m/z 265 (M⁺). Anal. calcd for C₁₀H₁₁N₅O₂S: C, 45.27; H, 4.18; N, 26.40. Found: C, 45.29; H, 4.34; N, 26.43.

2-Methyl-4-(2-methylpiperidin-1-yl)-5-nitro-6-thiocyanatopyrimidine 98

Yield 76 %, 75-78 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.17 (3H, s, CH₃), 1.45-1.55 (6H, m, 2CH₂), 2.37 (3H, s, CH₃), 3.04-3.19 (3H, m, CH₂, NCH). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.17; H, 5.19; N, 23.81.

4-(3,5-Dimethylpiperidin-1-yl)-2-methyl-5-nitro-6-thiocyanatopyrimidine 99

Yield 74 %, 122-125 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.15 (6H, s, 2CH₃), 1.25-1.60 (4H, m, HCCH₂CH), 2.39 (3H, s, CH₃), 2.90 and 3.15 (4H, 2 brm, N(CH₂)₂). MS (EI): m/z 307 (M⁺). Anal. calcd for C₁₃H₁₇N₅O₂S: C, 50.80; H, 5.58; N, 22.79. Found: C, 50.89; H, 5.63; N, 22.73.

1-(2-Methyl-5-nitro-6-thiocyanatopyrimidin-4-yl)azepane 100

Yield 68 %, 71-73 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.32-1.48 (8H, brm, 4CH₂), 2.39 (3H, s, CH₃), 3.64 (4H, brm, N(CH₂)₂). MS (EI): m/z 293 (M⁺). Anal. calcd for C₁₂H₁₅N₅O₂S: C, 49.13; H, 5.15; N, 23.87. Found: C, 49.21; H, 5.18; N, 23.92.

1-(2-Methyl-5-nitro-6-thiocyanatopyrimidin-4-yl)azocane 101

Yield 76 %, 95-98 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.18-1.82 (10H, brm, 5CH₂), 2.56 (3H, s, CH₃), 3.91 (4H, brs, N(CH₂)₂). MS (EI): m/z 307 (M⁺). Anal. calcd for C₁₃H₁₇N₅O₂S: C, 50.80; H, 5.58; N, 22.79. Found: C, 50.89; H, 5.63; N, 22.72.

N-Benzyl-2-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 102

Yield 58 %, 175-177 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.69 (3H, s, CH₃), 4.82 (2H, d, CH₂), 7.45 (5H, m, Ph), 9.04 (1H, brs, NH). MS (EI): m/z 301 (M⁺). Anal. calcd for C₁₃H₁₁N₅O₂S: C, 51.82; H, 3.68; N, 23.24. Found: C, 51.90; H, 3.63; N, 23.26.

2-Methyl-N-(4-methylbenzyl)-5-nitro-6-thiocyanatopyrimidin-4-amine 103

Yield 67 %, 160-162 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.27 (3H, s, CH₃), 2.71 (3H, s, CH₃), 4.79 (2H, d, CH₂), 7.18 and 7.26 (2H, m, Ph), 9.70 (1H, brs, NH). MS (EI): m/z 315 (M⁺). Anal. calcd for C₁₄H₁₃N₅O₂S: C, 53.32; H, 4.16; N, 22.21. Found: C, 53.39; H, 4.18; N, 22.27.

2-Methyl-5-nitro-N-phenethyl-6-thiocyanatopyrimidin-4-amine 104

Yield 52 %, 151-153 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.37 (3H, s, CH₃), 2.90 (2H, t, CH₂), 3.81 (2H, m, CH₂), 7.25 (5H, brm, Ph), 9.29 (1H, brs, NH). MS (EI): m/z 315 (M⁺). Anal. calcd for C₁₄H₁₃N₅O₂S: C, 53.32; H, 4.16; N, 22.21. Found: C, 53.41; H, 4.11; N, 22.16.

N,N,2-Trimethyl-5-nitro-6-thiocyanatopyrimidin-4-amine 105

Yield 86 %, 159-161 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.36 (3H, s, CH₃), 3.48 (6H, s, 2CH₃). MS (EI): m/z 239 (M⁺). Anal. calcd for C₈H₉N₅O₂S: C, 40.16; H, 3.79; N, 29.27. Found: C, 40.26; H, 3.88; N, 29.34.

N,N-Diethyl-2-methyl-5-nitro-6-thiocyanatopyrimidin-4-amine 106

Yield 53 %, 69-70 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.10 (6H, m, 2CH₃), 2.56 (3H, s, CH₃), 3.44 (4H, m, N(CH₂)₂). MS (EI): m/z 267 (M⁺). Anal. calcd for C₁₀H₁₃N₅O₂S: C, 44.93; H, 4.90; N, 26.20. Found: C, 45.08; H, 4.93; N, 26.32.

N-Benzyl-2-(methylthio)-5-nitro-6-thiocyanatopyrimidin-4-amine 107

Yield 69 %, 157-159 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.15 (3H, s, CH₃S), 4.77 (2H, d, CH₂), 7.27 (5H, brm, Ph), 9.81 (1H, brs, NH). MS (EI): m/z 333 (M⁺). Anal. calcd for C₁₃H₁₁N₅O₂S₂: C, 46.84; H, 3.33; N, 21.01. Found: C, 46.89; H, 3.40; N, 20.97.

N-(4-Methylbenzyl)-2-(methylthio)-5-nitro-6-thiocyanatopyrimidin-4-amine 108

Yield 80 %, 115-117 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.25 (3H, s, CH₃), 2.61 (3H, s, CH₃S), 4.75 (2H, d, CH₂), 7.07-7.27 (4H, m, Ph), 9.79 (1H, brs, NH). MS (EI): m/z 347 (M⁺). Anal. calcd for C₁₄H₁₃N₅O₂S₂: C, 48.40; H, 3.77; N, 20.16. Found: C, 48.49; H, 3.71; N, 20.21.

2-(Methylthio)-5-nitro-N-phenethyl-6-thiocyanatopyrimidin-4-amine 109

Yield 71 %, 93-95 °C (MeOH). ¹H NMR (DMSO-d₆; δ, ppm): 2.59 (3H, s, CH₃S), 2.87 (2H, t, CH₂), 3.78 (2H, t, CH₂), 7.25 (5H, brm, Ph), 9.46 (1H, brs, NH). MS (EI): m/z 347 (M⁺). Anal. calcd for C₁₄H₁₃N₅O₂S₂: C, 48.40; H, 3.77; N, 20.16. Found: C, 48.51; H, 3.86; N, 20.13.

N-Isopropyl-5-nitro-6-thiocyanato-2-(trifluoromethyl)pyrimidin-4-amine 110

Yield 75 %, 74-76 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.24 (6H, s, 2CH₃), 3.97 (1H, m, CH), 9.59 (1H, brs, NH). MS (EI): m/z 307 (M⁺). Anal. calcd for C₉H₈F₃N₅O₂S: C, 35.18; H, 2.62; N, 22.79. Found: C, 35.25; H, 2.67; N, 22.72.

N-Isopropyl-5-nitro-2-phenyl-6-thiocyanatopyrimidin-4-amine 111

Yield 76 %, 156-158 °C (EtOH). ¹H NMR (DMSO-d₆; δ, ppm): 1.18 (6H, s, 2CH₃), 3.85 (1H, m, CH), 7.50 (3H, m, CH_{ar}), 8.36 (2H, m, CH_{ar}). 9.47 (1H, brs, NH). MS (EI): m/z 315 (M⁺). Anal. calcd for C₁₄H₁₃N₅O₂S: C, 53.32; H, 4.16; N, 22.21. Found: C, 53.39; H, 4.23; N, 22.16.

N-Methyl-5-nitro-2-styryl-6-thiocyanatopyrimidin-4-amine 112

Yield 79 %, 215-217 °C (EtOH/DMF). ¹H NMR (DMSO-d₆; δ, ppm): 2.73 (3H, s, CH₃), 6.95-6.99 (2H, m, HC=CH), 7.36-7.49 (3H, brm, CH_{ar}), 7.60 (2H, m, CH_{ar}), 9.57 (1H, brs, NH). MS (EI): m/z 313 (M⁺). Anal. calcd for C₁₄H₁₁N₅O₂S: C, 53.67; H, 3.54; N, 22.35. Found: C, 53.74; H, 3.49; N, 22.30.

Strains and media

KN99α, a strain of *C. neoformans* serotype A ⁶, was used as the wild-type strain and was obtained from Dr. Jennifer Lodge, Washington University. The *C. neoformans*

fluconazole-resistant DUMC-158.03, and *C. gattii*, RSA-3615, clinical strains were obtained from Dr. John Perfect, Duke University ⁷. *Candida albicans* (Robin) Berkhout strain was purchased from the ATCC (ATCC #90028).

Strains were grown on YPD (1% yeast extract, 2% bacto-peptone, and 2% dextrose). Solid media contained 2% bacto-agar. YNB-02 (0.67% yeast nitrogen base, 0.2% dextrose, pH 7.0 with 50 mM MOPs) was used for all limiting dilution inhibition assays unless otherwise noted. The media is prepared in deionized water passed through ion exchange Milli Q filter system (Sigma-Millipore, USA).

Determination of minimal inhibitory concentration

Cells were grown overnight in 4 mL cultures of YPD at 30°C with shaking and then diluted to with an optical density (650 nm) of 0.001 (~5 x 10⁵ cells/mL) in YNB-02 + 1% DMSO for the limiting dilution assay. The compound dilution series and cells were prepared in round-bottom clear 96-well plates (Costar #3488), incubated without shaking for 48 hours at 35°C and the optical density measured. The minimal inhibitory concentration (MIC) was determined using compound concentrations from 0.19 to 50 mM, unless otherwise noted, of the compound in YNB-02 + 1% DMSO. Each assay was done in triplicate and all values are the average of two or more independent assays. The data are presented as the average cell density as a percent of DMSO-only treated cells. MICs are reported as the minimal concentration needed to inhibit 80% of *C. neoformans* or *C. gatti* growth relative to vehicle-treated controls.

Drug combinations

Drug combinations were assessed using the checkerboard assay^{8,9}. The MIC of the selected compounds were measured using compound concentrations from 0.19 to 12.5 μM and MICs for fluconazole (FLC) and amphotericin B (AMB) were measured using concentrations from 0.08 to 12.5 μM . Each assay was performed in triplicate and all values are the average of two or more independent assays. The fractional inhibitory concentration index (FICI) model is expressed as $\Sigma\text{FIC} = \text{FIC}_A + \text{FIC}_B = \text{MIC}_{A'}/\text{MIC}_A + \text{MIC}_{B'}/\text{MIC}_B$, where MIC_A , and MIC_B are the MIC values of agents A and B used alone and $\text{MIC}_{A'}$ and $\text{MIC}_{B'}$ are the MICs of agents A and B used in combination. The interaction between FLC or AMB and the test compounds was interpreted as synergistic when FICI was ≤ 0.5 , as indifferent when FICI was between > 0.5 and 4 and as antagonistic when FICI was > 4 ¹⁰.

Cytotoxicity in hepatoma cells

HepDES19 cells (1.0×10^4 cells per well) were seeded in 96-well plates and incubated in DMEM with 10% fetal bovine serum (FBS) plus 1% penicillin and streptomycin, 1% nonessential amino acids, and 1% glutamine. The compounds were diluted in the medium at concentrations ranging from 0.78 to 100 μM plus 1% DMSO and added to the cells 48 hours after plating, with each concentration tested in triplicate. Cells were incubated with the compound for 72 hours and cytotoxicity was measured using a mitochondrial metabolic assay with MTS (Promega). The data were transformed to $\log[\text{inhibitor}]$ and fit to a 4-variable slope curve using GraphPad Prism (v8, www.graphpad.com). The concentration at which 50% of cells were inhibited relative to vehicle-treated control is reported as the CC_{50} value.

Assay Central

Assay Central is a proprietary software developed from open-source descriptors and algorithms that have been previously described ¹¹⁻¹⁷ combined with additional proprietary scripts. Structure-activity datasets for the molecules previously described were collated in Molecular Notebook (Molecular Materials Informatics, Inc. in Montreal, Canada) and are thoroughly curated to generate a Bayesian machine learning model with multiple scripts. We employed a series of rules to detect any problematic data, corrections were implemented by a combination of automated and human re-curation for structure standardization. This approach produces a high-quality dataset and a Bayesian model to predict activities for proposed compounds. These Bayesian models utilize extended-connectivity fingerprints of maximum diameter 6 (ECFP6) descriptors generated from the Chemistry Development Kit library ¹⁸. These descriptors have widely been previously noted for their ability to map structure-activity relationships ¹⁹. From all the training set molecules, the Assay Central software enumerates all possible fingerprints from the training set and determines a given fingerprint's contribution to a binary activity classification from the ratio of its presence in active and inactive molecules. Assay Central also uses the summation of these contributions for a given molecule to produce a probability-like score. Metrics such as Receiver Operator Characteristic (ROC), Recall, Precision, F1 Score, Cohen's Kappa and Matthew's Correlation Coefficient are generated from internal five-fold cross-validation of the model. To maximize these internal performance statistics, the software can select a reasonable activity threshold, and generate predictions as well as applicability scores for any desired compound. Higher

prediction scores are desirable as scores higher than 0.5 are assigned to active compounds (inhibitors). Higher applicability scores are also desirable as it ensures the representation of the drug in the training set ¹⁹. Assay Central has been used in various drug discovery projects and the applicability of the model statistics have also been previously described ^{11-13, 16, 20-25}.

NIAID ChemDB Curation of Training Data

ChemDB was initially received as a transfer from NIAID as a series of comma-separated value files (.csv) ²⁶. Each measurement was assigned to a molecule identifier (AIDSNo) and depicted in a structure file (.sdf) with additional molecular information. We utilized workflows to combine the structures with measurements into new datasheet structures files (.ds) with the AIDSNo, restricted to *Cryptococcus neoformans* data.

Supplemental References

1. Latli, B.; Jones, P.-J.; Krishnanmurthy, D.; Senanayake, C. H., Synthesis of [14C]-, [13C4]-, and [13C4, 15N2]- 5-amino-4-iodopyrimidine. . *Journal of Labelled Compounds and Radiopharmaceuticals* **2008**, *51*, 54-58.
2. Demina, G. R.; Makarov, V. A.; Nikitushkin, V. D.; Ryabova, O. B.; Vostroknutova, G. N.; Salina, E. G.; Shleeva, M. O.; Goncharenko, A. V.; Kaprelyants, A. S., Finding of the low molecular weight inhibitors of resuscitation promoting factor enzymatic and resuscitation activity. *PLoS One* **2009**, *4* (12), e8174.
3. Brown, D. J.; Jacobsen, N. W., 690. Pyrimidine reactions. Part X. The methylation of triaminopyrimidines; conversion of the resulting imines into pteridines. *J Chem Soc* **1965**, 3770-3778.
4. Rousseaux, O.; Blondeau, D.; Sliwa, H., Synthesis of 8-nitro and 7-methoxy-8-nitro-1,2,4-triazolo[1,5-c]pyrimidines. *Heterocycles* **1990**, *31*, 288-281.

5. Clark, J.; Pendergast, W., Synthesis and covalent hydration of 4-trifluoromethylpteridine and some methyl derivatives. *J Chem Soc* **1969**, 1751-1754.
6. Nielsen, K.; Cox, G. M.; Wang, P.; Toffaletti, D. L.; Perfect, J. R.; Heitman, J., Sexual cycle of *Cryptococcus neoformans* var. *grubii* and virulence of congenic α and α isolates. *Infect Immun* **2003**, *71* (9), 4831-41.
7. Shaw, K. J.; Schell, W. A.; Covell, J.; Duboc, G.; Giamberardino, C.; Kapoor, M.; Moloney, M.; Soltow, Q. A.; Tenor, J. L.; Toffaletti, D. L.; Trzoss, M.; Webb, P.; Perfect, J. R., In Vitro and In Vivo Evaluation of APX001A/APX001 and Other Gwt1 Inhibitors against *Cryptococcus*. *Antimicrob Agents Chemother* **2018**, *62* (8).
8. White, R. L.; Burgess, D. S.; Manduru, M.; Bosso, J. A., Comparison of three different in vitro methods of detecting synergy: time-kill, checkerboard, and E test. *Antimicrob Agents Chemother* **1996**, *40* (8), 1914-8.
9. Banerjee, D.; Burkard, L.; Panepinto, J. C., Inhibition of nucleotide biosynthesis potentiates the antifungal activity of amphotericin B. *PLoS One* **2014**, *9* (1), e87246.
10. Odds, F. C., Synergy, antagonism, and what the checkerboard puts between them. *J Antimicrob Chemother* **2003**, *52* (1), 1.
11. Anantpadma, M.; Lane, T.; Zorn, K. M.; Lingerfelt, M. A.; Clark, A. M.; Freundlich, J. S.; Davey, R. A.; Madrid, P. B.; Ekins, S., Ebola Virus Bayesian Machine Learning Models Enable New in Vitro Leads. *ACS Omega* **2019**, *4* (1), 2353-2361.
12. Dalecki, A. G.; Zorn, K. M.; Clark, A. M.; Ekins, S.; Narmore, W. T.; Tower, N.; Rasmussen, L.; Bostwick, R.; Kutsch, O.; Wolschendorf, F., High-throughput screening and Bayesian machine learning for copper-dependent inhibitors of *Staphylococcus aureus*. *Metallomics* **2019**, *11* (3), 696-706.
13. Hernandez, H. W.; Soeung, M.; Zorn, K. M.; Ashoura, N.; Mottin, M.; Andrade, C. H.; Caffrey, C. R.; de Siqueira-Neto, J. L.; Ekins, S., High Throughput and Computational Repurposing for Neglected Diseases. *Pharm Res* **2018**, *36* (2), 27.
14. Chiarelli, L. R.; Mori, G.; Orena, B. S.; Esposito, M.; Lane, T.; de Jesus Lopes Ribeiro, A. L.; Degiacomi, G.; Zemanova, J.; Szadocka, S.; Huszar, S.; Palcekova, Z.; Manfredi, M.; Gosetti, F.; Lelievre, J.; Ballell, L.; Kazakova, E.; Makarov, V.; Marengo, E.; Mikusova, K.; Cole, S. T.; Riccardi, G.; Ekins, S.; Pasca, M. R., A multitarget approach to drug discovery inhibiting *Mycobacterium tuberculosis* PyrG and PanK. *Sci Rep* **2018**, *8* (1), 3187.
15. Kumar, P.; Capodagli, G. C.; Awasthi, D.; Shrestha, R.; Maharaja, K.; Sukheja, P.; Li, S. G.; Inoyama, D.; Zimmerman, M.; Ho Liang, H. P.; Sarathy, J.; Mina, M.; Rasic, G.; Russo, R.; Perryman, A. L.; Richmann, T.; Gupta, A.; Singleton, E.; Verma, S.; Husain, S.; Soteropoulos, P.; Wang, Z.; Morris, R.; Porter, G.; Agnihotri, G.; Salgame, P.; Ekins, S.; Rhee, K. Y.; Connell, N.; Dartois, V.; Neiditch, M. B.;

Freundlich, J. S.; Alland, D., Synergistic Lethality of a Binary Inhibitor of Mycobacterium tuberculosis KasA. *mBio* **2018**, *9* (6).

16. Sandoval, P. J.; Zorn, K. M.; Clark, A. M.; Ekins, S.; Wright, S. H., Assessment of Substrate-Dependent Ligand Interactions at the Organic Cation Transporter OCT2 Using Six Model Substrates. *Mol Pharmacol* **2018**, *94* (3), 1057-1068.

17. He, S.; Ye, T.; Wang, R.; Zhang, C.; Zhang, X.; Sun, G.; Sun, X., An In Silico Model for Predicting Drug-Induced Hepatotoxicity. *Int J Mol Sci* **2019**, *20* (8).

18. Willighagen, E. L.; Mayfield, J. W.; Alvarsson, J.; Berg, A.; Carlsson, L.; Jeliazkova, N.; Kuhn, S.; Pluskal, T.; Rojas-Cherto, M.; Spjuth, O.; Torrance, G.; Evelo, C. T.; Guha, R.; Steinbeck, C., The Chemistry Development Kit (CDK) v2.0: atom typing, depiction, molecular formulas, and substructure searching. *J Cheminform* **2017**, *9* (1), 33.

19. Clark, A. M.; Dole, K.; Coulon-Spektor, A.; McNutt, A.; Grass, G.; Freundlich, J. S.; Reynolds, R. C.; Ekins, S., Open Source Bayesian Models. 1. Application to ADME/Tox and Drug Discovery Datasets. *J Chem Inf Model* **2015**, *55* (6), 1231-45.

20. Ekins, S.; Gerlach, J.; Zorn, K. M.; Antonio, B. M.; Lin, Z.; Gerlach, A., Repurposing Approved Drugs as Inhibitors of Kv7.1 and Nav1.8 to Treat Pitt Hopkins Syndrome. *Pharm Res* **2019**, *36* (9), 137.

21. Anderson, E.; Havener, T. M.; Zorn, K. M.; Foil, D. H.; Lane, T. R.; Capuzzi, S. J.; Morris, D.; Hickey, A. J.; Drewry, D. H.; Ekins, S., Synergistic drug combinations and machine learning for drug repurposing in chordoma. *Sci Rep* **2020**, *10* (1), 12982.

22. Ekins, S.; Puhl, A. C.; Zorn, K. M.; Lane, T. R.; Russo, D. P.; Klein, J. J.; Hickey, A. J.; Clark, A. M., Exploiting machine learning for end-to-end drug discovery and development. *Nat Mater* **2019**, *18* (5), 435-441.

23. Lane, T.; Russo, D. P.; Zorn, K. M.; Clark, A. M.; Korotcov, A.; Tkachenko, V.; Reynolds, R. C.; Perryman, A. L.; Freundlich, J. S.; Ekins, S., Comparing and Validating Machine Learning Models for Mycobacterium tuberculosis Drug Discovery. *Mol Pharm* **2018**, *15* (10), 4346-4360.

24. Russo, D. P.; Zorn, K. M.; Clark, A. M.; Zhu, H.; Ekins, S., Comparing Multiple Machine Learning Algorithms and Metrics for Estrogen Receptor Binding Prediction. *Mol Pharm* **2018**, *15* (10), 4361-4370.

25. Wang, P. F.; Neiner, A.; Lane, T. R.; Zorn, K. M.; Ekins, S.; Kharasch, E. D., Halogen Substitution Influences Ketamine Metabolism by Cytochrome P450 2B6: In Vitro and Computational Approaches. *Mol Pharm* **2019**, *16* (2), 898-906.

26. Zorn, K. M.; Lane, T. R.; Russo, D. P.; Clark, A. M.; Makarov, V.; Ekins, S., Multiple Machine Learning Comparisons of HIV Cell-based and Reverse Transcriptase Data Sets. *Mol Pharm* **2019**, *16* (4), 1620-1632.

Figure S1. “Subvalidations” of the compounds from Table 1 using the literature cryptococcus model derived from the NIAID ChemDB database at thresholds of 10 μ M (A) and 3.2 μ M (B).

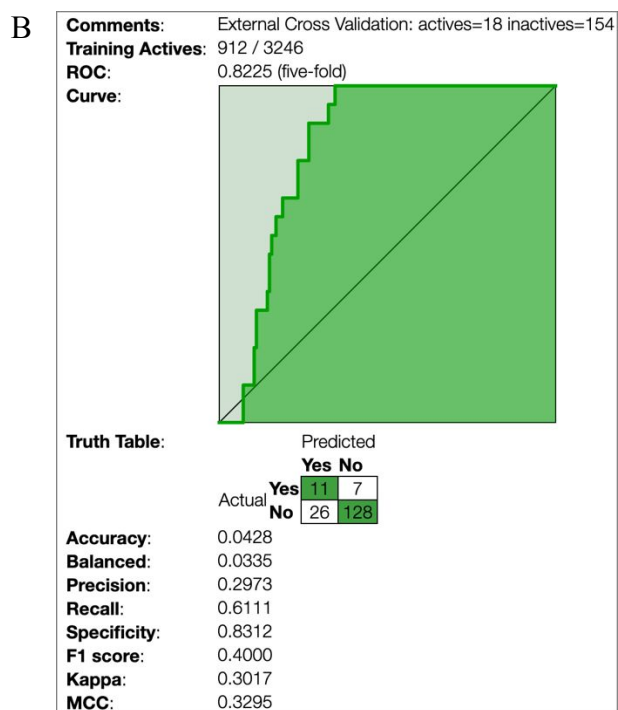
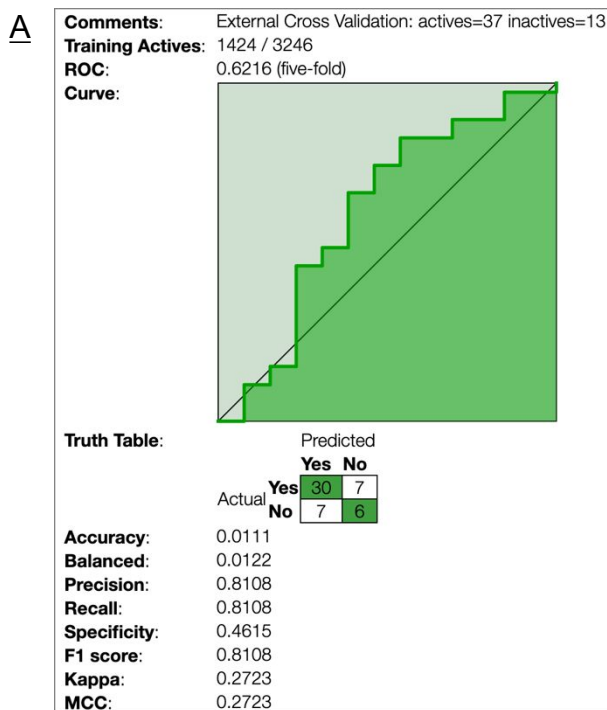


Figure S2. Bayesian Model 5-fold cross validation statistics for *C. gatti* model with an MIC₈₀ threshold for activity threshold of MIC 1.52-3.12 μM.

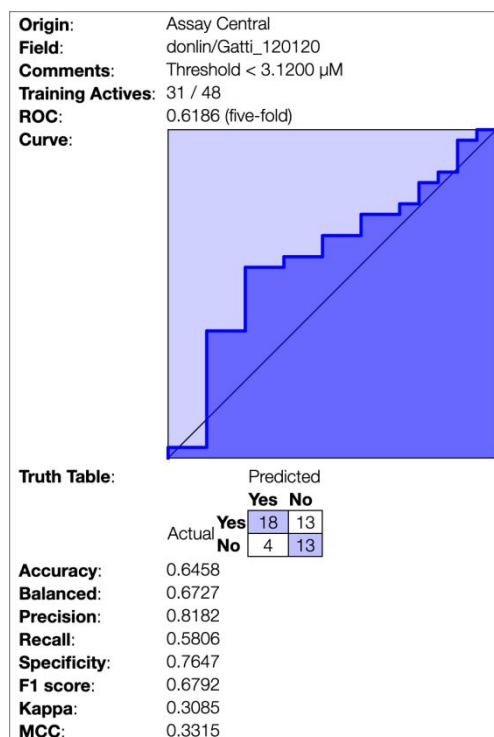


Table S1. Model 5-fold cross validation or subvalidation statistics for models described in each figure.

Model	ROC	truePos	trueNeg	falsePos	falseNeg	accuracy	balanced	precision	recall	specificity	F1	kappa	MCC
Fig 3A	0.70	8	103	4	6	0.92	0.77	0.67	0.57	0.96	0.62	0.57	0.57
Fig 3B	0.51	13	32	4	2	0.33	0.49	0.29	0.87	0.11	0.43	-0.01	-0.03
Fig 3C	0.90	18	120	34	0	0.80	0.89	0.35	1.00	0.78	0.51	0.42	0.52
Fig 3D	0.87	1202	1347	475	222	0.79	0.79	0.72	0.84	0.74	0.78	0.57	0.58
Fig 4	0.82	36	31	96	9	0.77	0.78	0.54	0.80	0.76	0.64	0.48	0.50
Fig S1A	0.62	30	7	6	7	0.72	0.64	0.81	0.81	0.46	0.81	0.27	0.27
Fig S1B	0.64	14	21	13	2	0.54	0.63	0.40	0.88	0.38	0.55	0.20	0.26
Fig S2	0.62	18	13	4	13	0.65	0.67	0.82	0.58	0.76	0.68	0.31	0.33

Table S2. Prediction validation for the 50 highlighted compounds using the NIAID *C. neoformans* training set.

Molecule Number	Prediction Score	Domain	Predicted Active	Active (10 μM Threshold)	<i>C. neoformans</i> MIC₈₀ μM (μg/ml)	MIC80 (μM)	Predicted Correctly	Category
63	0.508	0.585	yes	yes	9.37 \pm 3.12 (1.98 \pm 0.66)	9.3 \pm 3.1	yes	TP
64	0.507	0.625	yes	yes	3.9 \pm 0 (0.93 \pm 0)	3.8 \pm 0	yes	TP
65	0.513	0.556	yes	yes	1.9 \pm 0 (0.45 \pm 0)	1.9 \pm 0	yes	TP
66	0.516	0.553	yes	yes	4.68 \pm 1.56 (0.92 \pm 0.3)	4.6 \pm 1.5	yes	TP
67	0.517	0.578	yes	yes	2.34 \pm 0.78 (0.55 \pm 0.18)	2.3 \pm 0.7	yes	TP
68	0.524	0.521	yes	yes	3 \pm 0 (0.8 \pm 0)	3 \pm 0	yes	TP

69	0.520	0.571	yes	no	18.5 ± 0 (4.94 ± 0)	18.5 ± 6.5	no	FP
70	0.526	0.571	yes	yes	4.9 ± 0 (1.29 ± 0)	2.2 ± 0.7	yes	TP
71	0.531	0.585	yes	yes	6 ± 0 (1.67 ± 0)	4.5 ± 1.5	yes	TP
72	0.512	0.571	yes	yes	2.34 ± 0.78 (0.61 ± 0.2)	2.3 ± 0.7	yes	TP
73	0.505	0.614	yes	yes	9.37 ± 3.12 (2.61 ± 0.86)	9.3 ± 3.1	yes	TP
74	0.494	0.566	no	yes	9.37 ± 3.12 (2.74 ± 0.9)	9.3 ± 3.1	no	FN
75	0.509	0.528	yes	yes	2.34 ± 0.78 (0.68 ± 0.22)	2.3 ± 0.7	yes	TP
76	0.517	0.571	yes	yes	2.34 ± 0.78 (0.65 ± 0.22)	2.3 ± 0.7	yes	TP

77	0.519	0.580	yes	yes	4.68 ± 1.56 (1.37 ± 0.46)	4.6 ± 1.5	yes	TP
78	0.510	0.636	yes	yes	2.34 ± 0.78 (0.66 ± 0.22)	2.3 ± 1.5	yes	TP
79	0.552	0.667	yes	no	9.37 ± 12.5 (3.32 ± 4.44)	18.5 ± 0	no	FP
80	0.505	0.632	yes	yes	5.85 ± 7.8 (1.88 ± 2.5)	9 ± 0	yes	TP
81	0.492	0.603	no	yes	3.51 ± 4.68 (1.05 ± 1.4)	4.5 ± 0	no	FN
82	0.517	0.512	yes	yes	4.68 ± 1.56 (1.05 ± 0.34)	4.6 ± 1.5	yes	TP
83	0.517	0.511	yes	yes	9.37 ± 3.12 (2.37 ± 0.78)	9.3 ± 3.1	yes	TP
84	0.529	0.490	yes	yes	4.55 ± 4.24 (1.4 ± 1.12)	0.9 ± 0.3	yes	TP

85	0.538	0.509	yes	yes	6 ± 0 (1.76 ± 0)	9 ± 3	yes	TP
86	0.527	0.596	yes	yes	2.34 ± 0.78 (0.7 ± 0.22)	2.3 ± 1.5	yes	TP
87	0.526	0.550	yes	no	>50 (>16.46)	50 ± 0	no	FP
88	0.499	0.533	no	yes	1.17 ± 0.38 (0.27 ± 0.08)	1.1 ± 0.3	no	FN
89	0.494	0.563	no	yes	0.6 ± 0 (0.15 ± 0)	0.6 ± 0	no	FN
90	0.500	0.489	yes	yes	1 ± 0 (0.25 ± 0)	1.2 ± 0	yes	TP
91	0.505	0.487	yes	yes	2.34 ± 0.78 (0.48 ± 0.16)	2.3 ± 0.7	yes	TP
92	0.513	0.463	yes	yes	2.34 ± 0.78 (0.52 ± 0.16)	2.3 ± 0.7	yes	TP

93	0.507	0.510	yes	yes	9 ± 0 (2.53 ± 0)	9 ± 3	yes	TP
94	0.507	0.500	yes	yes	0.6 ± 0 (0.15 ± 0)	0.6 ± 0	yes	TP
95	0.513	0.520	yes	yes	9 ± 0 (2.51 ± 0)	9 ± 3	yes	TP
96	0.472	0.583	no	no	>50 (>24.62)	50 ± 0	yes	TN
97	0.501	0.522	yes	yes	4.68 ± 1.56 (1.23 ± 0.4)	4.6 ± 1.5	yes	TP
98	0.493	0.561	no	no	18.75 ± 6.24 (5.49 ± 1.82)	18.7 ± 6.2	yes	TN
99	0.498	0.472	no	no	18.75 ± 6.24 (5.76 ± 1.92)	18.7 ± 6.2	yes	TN
100	0.506	0.520	yes	no	18.75 ± 6.24 (5.49 ± 1.82)	18.7 ± 6.2	no	FP

101	0.508	0.529	yes	no	18.75 ± 6.24 (5.76 ± 1.92)	18.7 ± 6.2	no	FP
102	0.497	0.596	no	no	50 ± 0 (15.06 ± 0)	50 ± 0	yes	TN
103	0.504	0.561	yes	no	50 ± 0 (15.76 ± 0)	50 ± 0	no	FP
104	0.479	0.567	no	no	50 ± 0 (15.76 ± 0)	50 ± 0	yes	TN
105	0.508	0.463	yes	yes	4.68 ± 1.56 (1.11 ± 0.36)	4.6 ± 1.5	yes	TP
106	0.508	0.467	yes	yes	9.37 ± 3.12 (2.5 ± 0.82)	9.3 ± 3.1	yes	TP
107	0.482	0.661	no	yes	4.68 ± 4.68 (1.56 ± 1.56)	4.5 ± 0	no	FN
108	0.489	0.633	no	yes	9.37 ± 3.12 (3.25 ± 1.08)	9 ± 0	no	FN

109	0.465	0.629	no	yes	9.37 ± 9.36 (3.25 ± 3.24)	9 ± 0	no	FP
110	0.518	0.531	yes	yes	12.25 ± 6.5 (4.4 ± 2)	4.5 ± 1.5	yes	TP
111	0.493	0.571	no	no	50 ± 0 (15.76 ± 0)	50 ± 0	yes	TN
112	0.511	0.552	yes	no	>50 (>15.66)	50 ± 0	no	FP