

Figure S1. Dose dependent effects of *cmn^{spod/+}* mutation on vertebral patterning. Related to Figure 1. Alizarin red stained skeletal preparations imaged on a Fluoview FV3000 (Olympus) confocal microscope. Wild-type and *cmn^{spod/+}* fish were fixed and stained at 6 weeks old. Homozygous mutants were 4 months of age. Homozygous *spondo* mutants display highly irregular vertebral morphology.

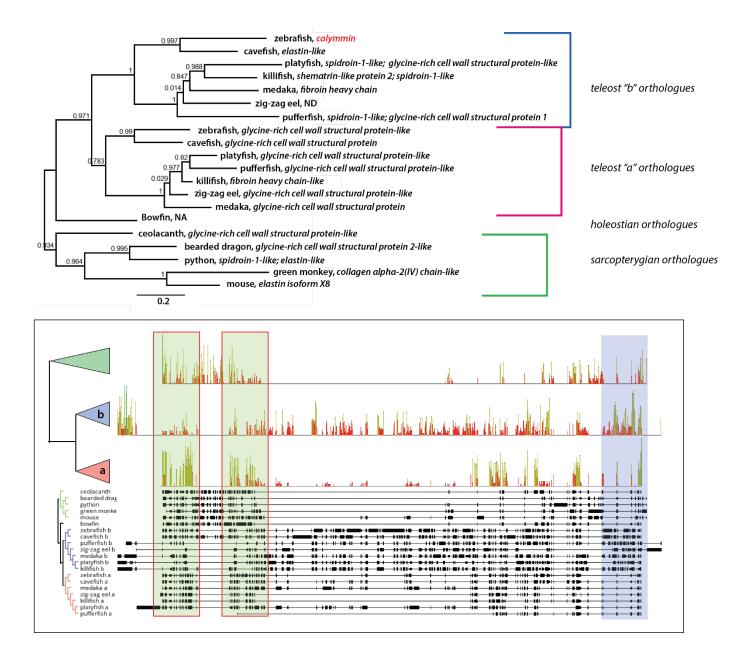


Figure S2. Phylogeny, alignment, and conservation of *calymmin* related genes. Related to Figure 3. **Top**) Maximum likelihood phylogeny of *cmn* coding regions and identified orthologues across vertebrates. Designation of 'a' and 'b' paralogous groups within teleosts based on prior notation of genes syntenic to *cmn*. No orthologues were found for gar or any bird species. Numbers indicate bootstrap support. **Bottom**) Multiple sequence alignment and percent identity of *cmn* coding regions and orthologues across vertebrates. These genes are rapidly evolving, but conservation within orthologous groups reveals both shared and unique critical regions (highlighted), suggesting neofunctionalization. Bar height and color indicate percent identity: green = 100%, yellow > 30%, red < 30%.

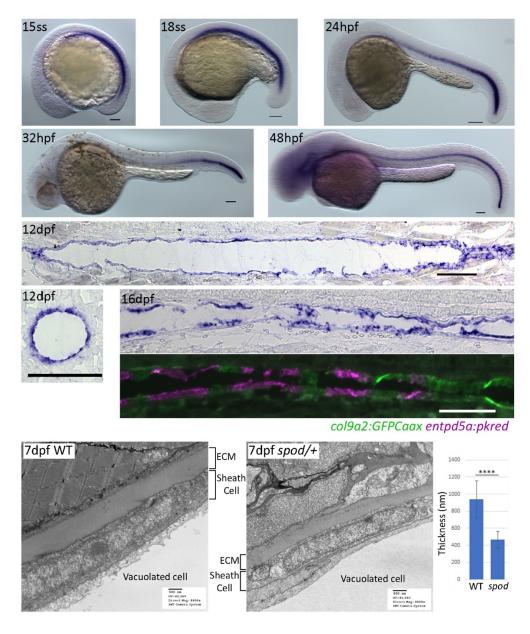
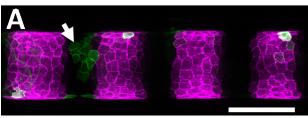
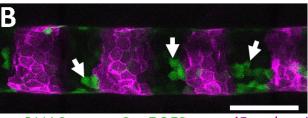


Figure S3. Expression dynamics of *calymmin* during larval development and effects of the *spondo* **mutation on ECM organization and thickness. Related to Figure 3.** Top: *In situ* hybridization of *cmn* expression at different stages of zebrafish development (15ss-16dpf). *15ss-48hpf*) At each stage, *cmn* expression is found throughout the notochord. During initial chordablast differentiation, expression is enriched in the posterior tail region. *12dpf*) At later larval stages, *calymmin* expression is found throughout the notocryte-like sheath cells. *16dpf*) During notochord segmentation, *cmn* expression is enriched in chondrocyte-like sheath cells as well as cells that are transitioning into mineralizing cells and express both *col9a2* and *entpd5a*. Following sheath cell differentiation, *calymmin* expression is downregulated in cells that form the chordacentra. Image at 16dpf is an uncropped version of Figure 2G and G'. 7dpf WT, 7dpf *spod/+*). **Bottom,** transmission electron microscopy was performed on 7dpf wild-type and *cmn^{spod/+}* mutants. The sheath extracellular matrix in *spondo* mutants contains more loosely packed fibrils and is thinner than wild-type siblings. Quantification of matrix thickness is shown to the right. For quantification, 55 measurements per genotype were obtained (n=2 fish per genotype). Two-tailed p value = 1.67E-23



QUAS:GFPCaax entpd5a:pkred



QUAS:cmn-p2a-EGFP entpd5a:pkred

Figure S4. Overexpression of wild-type *cmn* **does not lead to ectopic** *entpd5a* **activation in** *col9a2+* **domains. Related to Figure 3**. **A,B**) Overexpression of either a *QUAS:GFPCaax* or *QUAS:cmn-p2a-EGFP* DNA construct in fish containing a *col9a2:QF2* transgene. Neither cells overexpressing the GFPCaax control nor the wild-type *cmn* construct began to ectopically express *entpd5a* win the prospective IVD domains. The number of GFP+/entpd5a+ cells were counted within each prospective IVD domain. An average of 4.1% of cells were double positive for GFPCaax and *entpd5a* in the control group. Of the cells within the IVD domains overexpressing wild-type *cmn*, 9.3% were also *entpd5a*+ (two-tailed *p* value of 0.09). These data indicate that overexpression of wild-type *cmn* is not sufficient to induce the differentiation of notochord sheath cells into mineralizing cells. Arrows point to cells overexpressing the DNA constructs that have not activated *entpd5a*

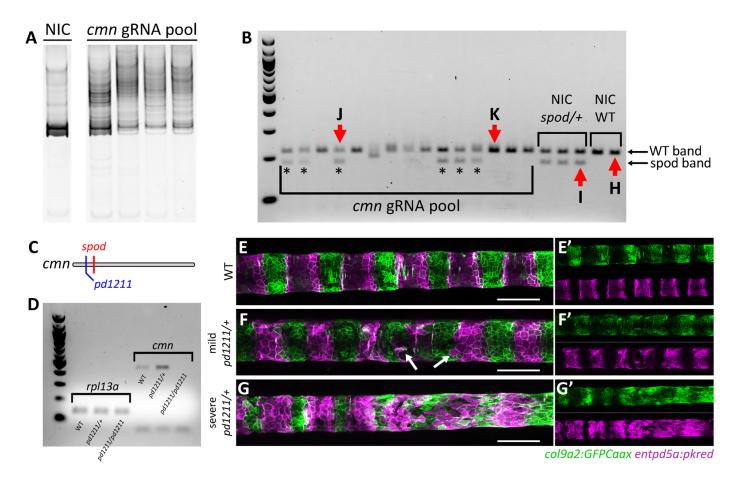


Figure S5. Loss of *cmn* function leads to irregular boundaries between notochord segments. Related to Figure 3. A) Heteroduplex mobility shift assay run on an acrylamide gel to visualize indels generated by injection of a gRNA pool targeting the *cmn* locus. NIC = non-injected control. **B**) Genotyping of injected and non-injected fish. Upper band is wild type *cmn* and the lower band is only present in spondo mutants after a restriction digest with BseRI (NEB). Arrows indicate fish depicted in Figure 3 H-K. Asterisks indicate injected spondo mutants. C) A stable mutant line (cmn^{pd1211}) containing a 2bp deletion upstream of the spondo mutation was generated using CRISPR/Cas9 genome editing. D) RT-PCR performed on 5dpf embryos revealed that the *cmn* transcript is degraded in *cmn*^{pd1211} homozygous mutants. E-G') Loss of cmn function leads to irregular segment boundaries. E) Wild-type sibling displaying normal notochord segmentation. **F,G**) *cmn^{pd1211}* heterozygotes had a notochord segmentation phenotype that was highly variable, ranging from mild to severe. In mild cases (n=8/25 mutant fish), jagged boundaries between the col9a2 and entpd5a domains were apparent similar to the pooled gRNA experiment. In more severe expressivity in heterozygous fish, the two segment domains are indistinguishable and blend into one another (n=3/25 mutant fish). Homozygous cmn^{pd1211} were not recovered at this stage (14-16dpf). E'-G') Isolated green and red channels for each image depicted in E-G.

Homologue	Species	Source*	Annotated Name
			glycine-rich cell wall structural
Calymmin	Pogona vitticeps	https://www.ncbi.nlm.nih.gov/gene/110088379	protein 2-like
	Chlorocebus		collagen alpha-2(IV) chain-like
Calymmin	sabaeus	https://www.ncbi.nlm.nih.gov/gene/103227069	
Calymmin	Mus musculus	https://www.ncbi.nlm.nih.gov/gene/320309	elastin isoform X8
		https://www.ncbi.nlm.nih.gov/gene/103054235	spidroin-1-like
Calymmin	Python bivittatus	https://www.ncbi.nlm.nih.gov/gene/103054480	elastin-like
	Latinmeria		glycine-rich cell wall structural
Calymmin	chalumnae	https://www.ncbi.nlm.nih.gov/gene/102353231	protein-like
Calymmin	Amia calva	Ingo Braasch, personal communication	
calymmin b	Danio rerio	https://www.ncbi.nlm.nih.gov/gene/30208	calymmin
calymmin b	Oryzias latipes	https://www.ncbi.nlm.nih.gov/gene/101174706	fibroin heavy chain
,		http://useast.ensembl.org/Mastacembelus_armatus/Transcrip	GENSCAN0000024764.1
	Mastacembelus	t/Summary?db=core;pt=GENSCAN00000024764;r=OOHQ0100	
calymmin b	armatus	0019.1:10867594-10914660	
	Astyanax		elastin-like
calymmin b	mexicanus	https://www.ncbi.nlm.nih.gov/gene/103038914	
	Marken have		spidroin-1-like
	Xiphophorus	https://www.ncbi.nlm.nih.gov/gene/114151854	glycine-rich cell wall structural
calymmin b	couchianus	https://www.ncbi.nlm.nih.gov/gene/114161560	protein 1.8-like
	Kryptolebias	https://www.ncbi.nlm.nih.gov/gene/108247781	shematrin-like protein 2 spidroin-1-like
calymmin b	marmoratus	https://www.ncbi.nlm.nih.gov/gene/108247328	
	mannoratas	<u></u>	spidroin-1-like
		https://www.ncbi.nlm.nih.gov/gene/105416331	glycine-rich cell wall structural
calymmin b	Takifugu rubripes	https://www.ncbi.nlm.nih.gov/gene/105416667	protein 1.8-like
			glycine-rich cell wall structural protein-like
calymmin a	Danio rerio	https://www.ncbi.nlm.nih.gov/gene/100001051	protein-like
			glycine-rich cell wall structural
calymmin a	Oryzias latipes	https://www.ncbi.nlm.nih.gov/gene/101167291	protein 1.0
	Xiphophorus		glycine-rich cell wall structural
calymmin a	couchianus	https://www.ncbi.nlm.nih.gov/gene/114160123	protein-like
, .	Kryptolebias		fibroin heavy chain-like
calymmin a	marmoratus	https://www.ncbi.nlm.nih.gov/gene/108237682	glucing rich call wall structure!
calummin a	Mastacembelus	https://www.pchi.plm.pih.gov/gopg/112140701	glycine-rich cell wall structural
calymmin a	armatus	https://www.ncbi.nlm.nih.gov/gene/113140701	protein 1.0-like glycine-rich cell wall structural
calymmin a	Takifugu rubripes	https://www.ncbi.nlm.nih.gov/gene/105416492	protein-like
	Astyanax	10093.// WWW.1001.1111.1111.gov/gelle/103410432	glycine-rich cell wall structural
	. istyanan	https://www.ncbi.nlm.nih.gov/gene/103039514	protein

Table S1. Data sources for *calymmin* **orthologues. Related to Figure 3.** Annotations used to compile multiple species alignment of calymmin across vertebrates. *spliced with adjacent annotation where notated.