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Emission spectra of BODIPY₁-Pep₄ in mixture solvent of buffer and DMSO

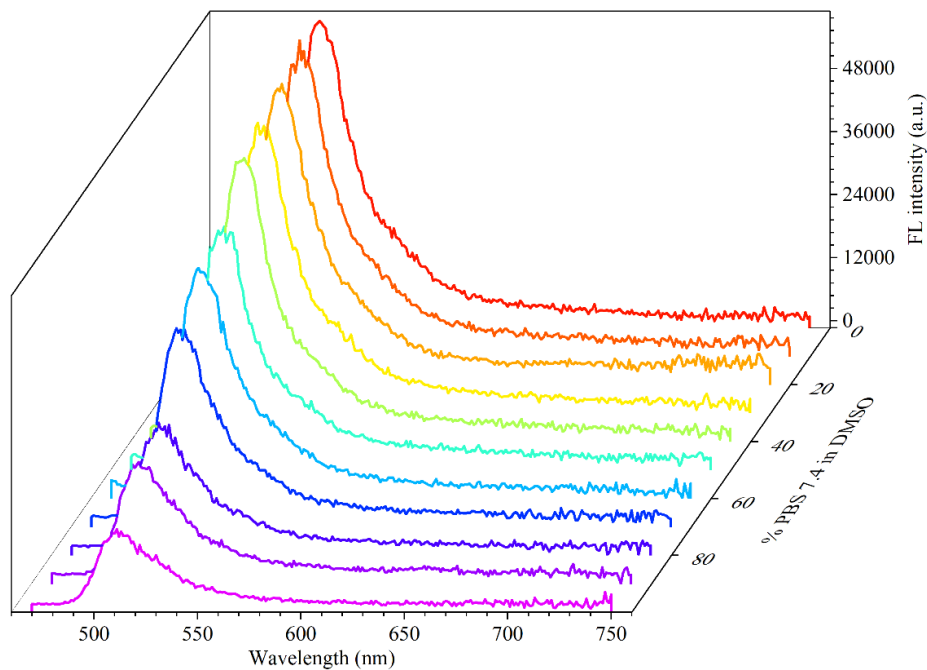


Fig. S1. Comparison of emission spectrum of BODIPY₁-Pep₄ (1.2 μM) in solvents with different ratios of PBS7.4 and DMSO.

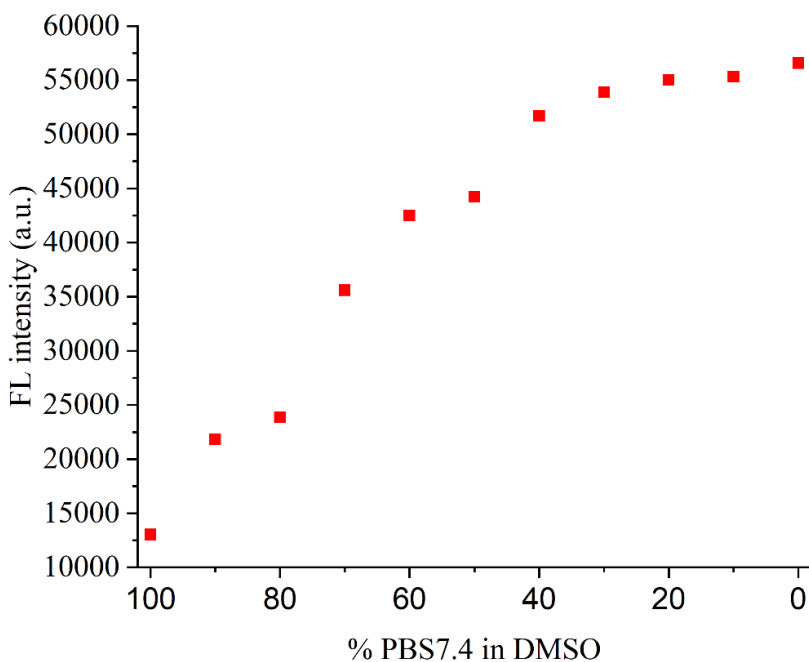


Fig. S2. Comparison of fluorescent intensity of BODIPY₁-Pep₄ (1.2 μM) at 512 nm in solvents with different ratios of PBS7.4 and DMSO.

Photophysical properties of BODIPY-peptide conjugates

Table. S1. Photophysical property of BODIPY_n-Pep₁.^a

	λ absorption (nm)	ϵ ($\times 10^4 \text{ M}^{-1} \text{ cm}^{-1}$)	λ excitation (nm)	λ emission (nm)	τ (ns)	Φ (%) ^b
BODIPY₁-Pep₁	503	5.81	501	515	3.18	50.1
BODIPY₃-Pep₁	526	5.10	526	542	5.23	34.0
BODIPY₉-Pep₁	560	4.07	557	599	0.991	5.58

^a. all data are recorded in DMSO

^b. the quantum yields are measured by comparative method with reference rhodamine 6G in water ($\Phi = 95\%$).

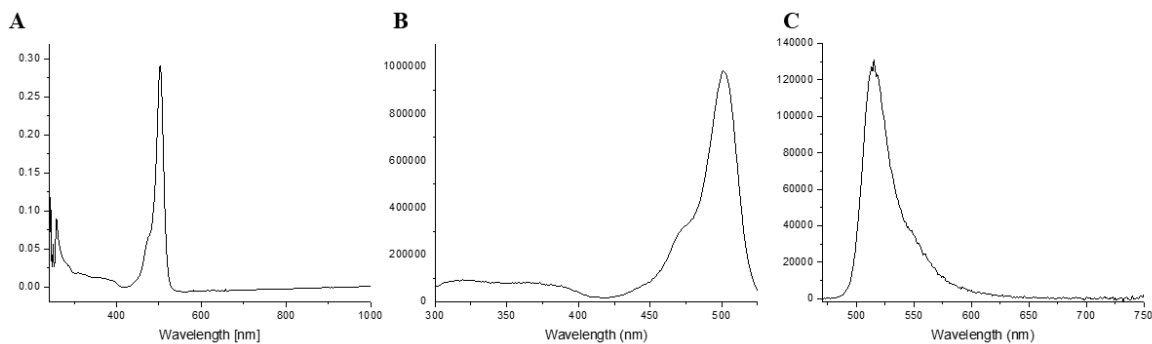


Fig. S3. Absorption (A), excitation (B) and emission (C) spectra of **BODIPY₁-Pep₁**.

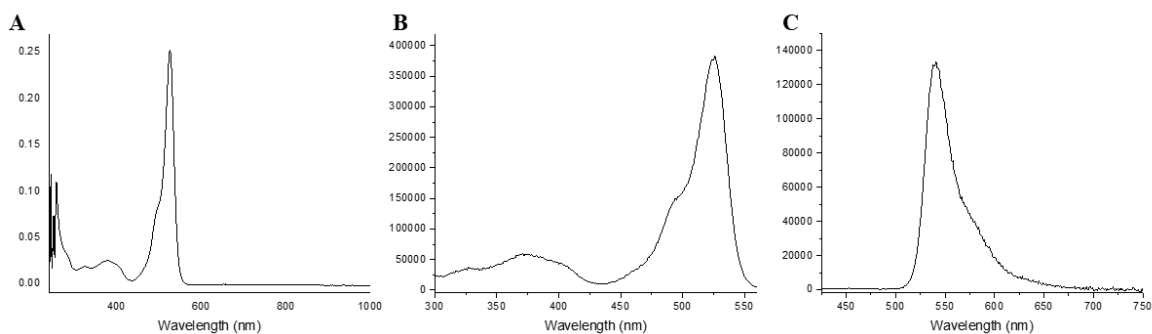


Fig. S4. Absorption (A), excitation (B) and emission (C) spectra of **BODIPY₃-Pep₁**.

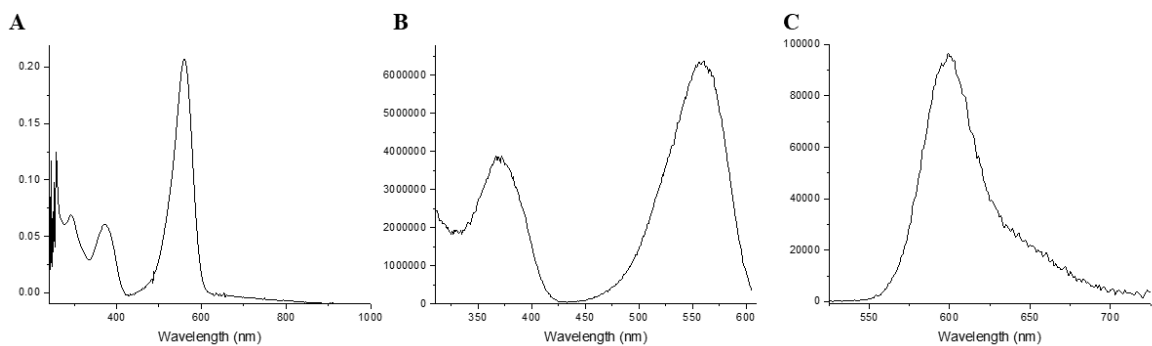


Fig. S5. Absorption (A), excitation (B) and emission (C) spectra of **BODIPY₉-Pep₁**.

Distribution of signal of BODIPY₁-Pep₄ and nuclear blue in the cell lines

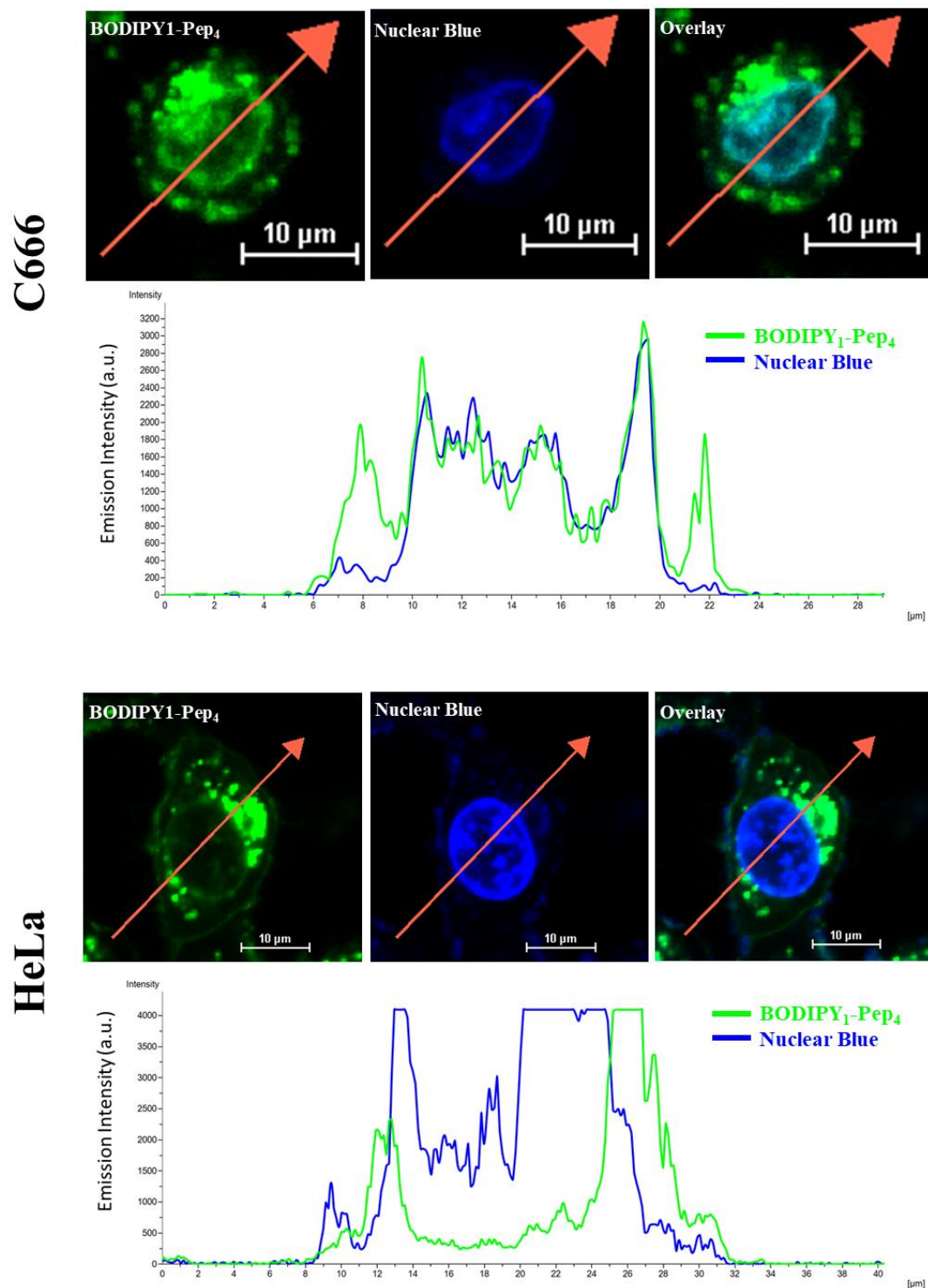


Fig. S6. Profiles of the emission intensity of the BODIPY₁-Pep₄ and nuclear blue were plotted along the red arrow A. C666, B. HeLa. The signal of BODIPY₁-Pep₄ and nuclear blue overlapped well in C666 cell line (EBNA+) but not in HeLa cell line (EBNA-).

Chemical Experiments

Reagents: All amino acid building blocks for Fmoc-strategy SPPS were purchased from Bidepharm. For synthesizing **Pep₁~Pep₁₀**, Fmoc-Ala-OH, Fmoc-Val-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Met-OH, Fmoc-Phe-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Tyr(PO₃(MDPSE)₂)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Thr(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Gly-OH, Fmoc-Pro-OH, Fmoc-Arg(Pdf)-OH, Fmoc-His(Boc)-OH, Fmoc-Lys(Boc)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Glu(tBu)-OH, Fmoc-6-Ahx-OH, Fmoc-2-Nal-OH, Fmoc-4-Cpa-OH, Fmoc-3-Pal-OH, Fmoc-Cit-OH. For **Pep₁₁**, Fmoc-Lys(Alloc)-OH was used to take place Fmoc-Lys(Boc)-OH as building block of lysine. The Rink Amide resin (100-200 mesh) and Wang resin (100-200 mesh) were purchased from Sigma-Aldrich for synthesizing peptides with amide and acid C-terminal respectively. All aldehyde and pyrrole building blocks are commercially available except the pyrrole for synthesizing **DP₈** and **DP₁₁** are synthesized according literature.^[1] Solvents and other reagent were purchased and used without further purification.

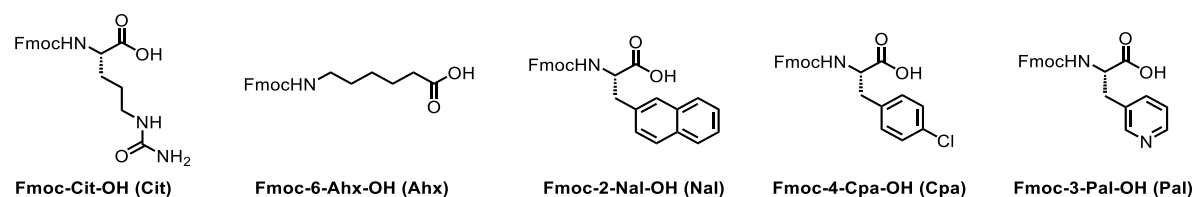


Fig. S7. Structure and abbreviate of unnatural amino acids.

Analytical HPLC: Analytical HPLC was performed on an Agilent 1100 series HPLC system (Agilent Technologies, Stockport, UK) equipped with a diode-array detection (DAD) detector and Agilent C18 column (250 mm x 4.6 mm) at the following gradients:

Gradient A:

Time (min)	A % (H ₂ O + 0.1 % TFA)	B % (MeCN + 0.1 % TFA)	Flow (mL/min)
0	80	20	0.5
60	0	100	0.5
70	0	100	0.5

Gradient B:

Time (min)	A % (H ₂ O + 0.1 % TFA)	B % (MeCN + 0.1 % TFA)	Flow (mL/min)
0	80	20	0.5
40	20	80	0.5
41	0	100	0.5
55	0	100	0.5

Preparative HPLC: The purifications of products were carried out on Waters semi-preparative system with Waters 2707 Autosampler, Water 1525 Binary HPLC Pump, Waters 2998 Photodiode Array Detector and Waters Fraction Collector III and Atlantis® T3 Prep OBDTM column (C18, 5 μm, 19×250 mm). The Gradient usually refer to the analytical HPLC but flow rate is 5 mL/min. The fractions were collected and verified by ESI-MS.

Mass spectrometry: High-resolution mass spectra, reported as m/z , were obtained from Bruker Autoflex MALDI-TOF mass spectrometer. Low-resolution mass spectra were conducted by SCIEX 3200Q ESI mass spectrometer was also used for monitoring reaction and determining correct fraction during purification of product.

Nuclear magnetic resonance: NMR spectra were recorded on a Bruker Ultrashield 400 Plus NMR spectrometer (^1H NMR on 400 MHz, ^{13}C NMR on 101 MHz and ^{19}F NMR on 376 MHz. The ^1H NMR chemical shifts were referenced to corresponding solvent peak (2.50 for DMSO- d_6 and 3.31 for methanol- d_4). The following abbreviations were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, dd = doublet of doublets, m = multiplet, br = broad.

General procedure of solid phase peptide synthesis: Standard Fmoc-based SPPS procedure was carried out manually. The 5 mL SPE filtration tube with Frits were used. The Rink Amide resin and Wang resin were used for synthesizing peptides with amide and acid C-terminal respectively. To load first amino acid onto Rink Amide resin, Fmoc-deprotected resin was shaken with amino acid building block (3 eq.), PyBOP (3 eq.) and DIPEA (6 eq.) in DMF (4 mL/ 0.1 mmol) overnight. To load first amino acid onto Wang resin, resin was shaken with amino acid building block (3 eq.), PyBOP (3 eq.) and DMAP (1 eq.) in DCM/DMF, v/v, 8/2 (4 mL/ 0.1 mmol) overnight. After loading first amino acid, the resin was washed with DMF (4 mL \times 3) and DCM (4 mL \times 3). Then, the shaken with Ac_2O /Pyridine, v/v, 3/2 (4 mL/ 0.1 mmol) for 30 min to cap all remaining reacting site on resin. During peptide elongation, coupling was carried out with amino acid building block (3 eq.), PyBOP (3 eq.) and DIPEA (6 eq.) in DMF (4 mL/ 0.1 mmol) for 2~12 h, and Fmoc deprotection was carried out with 4 mL 20% piperidine in DMF (4 mL/ 0.1 mmol) for 25 min. After all amino acid building blocks were coupled, the substitute value of resin bound peptide was calculated by the method from literature^[2]. Global cleavage was carried out with different cocktail (4 mL/ 0.1 mmol) and reaction time according different amino acid composition of each peptide. Cocktail A: TFA/TIPS/ H_2O , v/v/v, 95/2.5/2.5; Cocktail B: TFA/EDT/p-Cresol/ H_2O , v/v/v/v, 90/5/2.5/2.5.

$\text{H}_2\text{N-YFMVF-CONH}_2$ (Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 60 %. White powder. Analytic HPLC: Gradient A, retention time: 23.9 min, purity: 96.6 %. HRMS(MALDI-TOF): calc. for $\text{C}_{37}\text{H}_{49}\text{N}_6\text{O}_6\text{S}^+ [\text{M}+\text{H}]^+$ 705.3429, found 705.3489; calc. for $\text{C}_{37}\text{H}_{48}\text{N}_6\text{NaO}_6\text{S}^+ [\text{M}+\text{Na}]^+$ 727.3248, found 727.3251; calc. for $\text{C}_{37}\text{H}_{48}\text{KN}_6\text{O}_6\text{S}^+ [\text{M}+\text{K}]^+$ 743.2988, found 743.3027. ^1H NMR (400 MHz, Methanol- d_4) δ 7.26 – 7.14 (m, 10H), 7.11 – 7.04 (m, 2H), 6.81 – 6.72 (m, 2H), 4.89 (s, 1H), 4.70 (tt, J = 8.3, 5.9 Hz, 2H), 4.40 – 4.31 (m, 1H), 4.07 (dd, J = 8.6, 4.8 Hz, 1H), 3.20 – 3.05 (m, 3H), 2.98 – 2.79 (m, 3H), 2.56 – 2.38 (m, 2H), 2.01 (s, 6H), 0.90 (dd, J = 6.8, 5.1 Hz, 6H); ^{13}C NMR (101 MHz, Methanol- d_4) δ 129.58, 128.28, 128.22, 127.43, 127.34, 125.74, 114.80, 53.39, 47.52, 47.31, 47.10, 46.89, 46.67, 46.46, 46.25, 37.23, 37.05, 35.71, 31.38, 30.27, 28.97, 17.65, 16.99, 13.27; ^1H NMR (400 MHz, DMSO- d_6) δ 9.38 (s, 1H), 8.77 (d, J = 8.1 Hz, 1H), 8.44 (d, J = 7.9 Hz, 1H), 8.02 (d, J = 8.2 Hz, 1H), 7.99 – 7.92 (m, 2H), 7.83 (d, J = 8.8 Hz, 1H), 7.38 (d, J = 2.2 Hz, 1H), 7.30 – 7.15 (m, 10H), 7.06 (s, 1H), 7.04 (d, J = 8.5 Hz, 2H), 6.72 – 6.67 (m, 2H), 4.65 (td, J = 8.7, 4.2 Hz, 1H), 4.50 – 4.42 (m, 2H), 4.18 – 4.13 (m, 1H), 3.89 (s, 1H), 3.08 – 2.93 (m, 3H), 2.82 (ddd, J = 14.0, 9.3, 6.6 Hz, 3H), 2.42 (ddp, J = 13.1, 9.3, 6.9, 6.2 Hz, 2H), 2.03 (s, 3H), 1.97 – 1.87 (m, 2H), 1.79 (dtd, J = 14.2, 9.4, 6.1 Hz, 1H), 0.79 (dd, J = 6.8, 2.2 Hz, 6H); ^{13}C NMR (101 MHz, Methanol- d_4) δ 129.58, 128.28, 128.22, 127.43, 127.34, 125.74, 114.80, 53.39, 47.52, 47.31, 47.10, 46.89, 46.67, 46.46, 46.25, 37.23, 37.05, 35.71, 31.38, 30.27, 28.97, 17.65, 16.99, 13.27; ^{13}C NMR (101 MHz, DMSO- d_6) δ 172.61, 170.66, 170.58, 170.40, 156.52, 137.70, 137.37, 130.53, 129.19, 129.00,

128.06, 127.91, 126.32, 126.14, 124.42, 115.25, 57.57, 53.96, 53.45, 53.23, 51.93, 37.57, 37.39, 36.08, 31.92, 30.59, 29.47, 19.08, 17.90, 14.58.

H₂N-C-Ahx-RrRKGGYFMVF-COOH (Pep₂). Global cleavage condition: cocktail B, 4 h. Yield: 54 %. White powder. Analytic HPLC: Gradient B, retention time: 16.8 min, purity: 98.4 %. HRMS(MALDI-TOF): calc. for C₇₁H₁₁₀N₁₉O₁₄⁺ [M+H]⁺ 1452.8474, found 1452.8474; calc. for C₇₁H₁₀₉N₁₉NaO₁₄⁺ [M+Na]⁺ 1474.8294, found 1474.8294.

H₂N-C-Ahx-YFMVFGGRrRK-COOH (Pep₃). Global cleavage condition: cocktail B, 4 h. Yield: 60 %. White powder. Analytic HPLC: Gradient B, retention time: 18.9 min, purity: 97.1 %. HRMS(MALDI-TOF): calc. for C₇₄H₁₁₈N₂₃O₁₅S₂⁺ [M+H]⁺ 1632.8614, found 1632.8623; calc. for C₇₄H₁₁₇N₂₃NaO₁₅S₂⁺ [M+Na]⁺ 1654.8433, found 1654.8431.

H₂N-C-Ahx-YFMVFGGRrRK-COOH (Pep₄). Global cleavage condition: cocktail B, 4 h. Yield: 53 %. White powder. Analytic HPLC: Gradient B, retention time: 17.6 min, purity: 95.9 %. HRMS(MALDI-TOF): calc. for C₇₄H₁₁₈N₂₃O₁₅S₂⁺ [M+H]⁺ 1632.8614, found 1632.8604; calc. for C₇₄H₁₁₇N₂₃NaO₁₅S₂⁺ [M+Na]⁺ 1654.8433, found 1654.8440.

H₂N-NWYFIVF-COOH (Pep₅). Global cleavage condition: cocktail A, 2 h. Yield: 57 %. White powder. Analytic HPLC: Gradient A, retention time: 28.7 min, purity: 94.9 %. HRMS(MALDI-TOF): calc. for C₅₈H₇₃N₁₁NaO₁₂⁺ [M+Na]⁺ 1138.5332, found 1138.5323.

H₂N-DEHYFIVF-COOH (Pep₆). Global cleavage condition: cocktail A, 2 h. Yield: 60 %. White powder. Analytic HPLC: Gradient A, retention time: 22.8 min, purity: 96.8 %. HRMS(MALDI-TOF): calc. for C₅₃H₆₉N₁₀O₁₄⁺ [M+H]⁺ 1069.4989, found 1069.4988; calc. for C₅₃H₆₈KN₁₀O₁₄⁺ [M+K]⁺ 1107.4548, found 1107.4424.

H₂N-Ahx-P-(pTyr)-LKTK-COOH (Pep₇). Global cleavage condition: cocktail B, 3 h. Yield: 59 %. White powder. Analytic HPLC: Gradient B, retention time: 8.4 min, purity: 93.7 %. HRMS(MALDI-TOF): calc. for C₄₂H₇₃N₉O₁₃P⁺ [M+H]⁺ 942.5060, found 942.5063; calc. for C₄₂H₇₂N₉NaO₁₃P⁺ [M+Na]⁺ 964.4879, found 964.4882; calc. for C₄₂H₇₂KN₉O₁₃P⁺ [M+K]⁺ 980.4619, found 980.4627.

H₂N-Ahx-P-(pTyr)-LKTKRrRK-COOH (Pep₈). Global cleavage condition: cocktail B, 3 h. Yield: 62 %. White powder. Analytic HPLC: Gradient B, retention time: 33.5 min, purity: 98.4 %. HRMS(MALDI-TOF): calc. for C₆₆H₁₂₁N₂₃O₁₇P⁺ [M+H]⁺ 1538.9043, found 1538.9043; calc. for C₆₆H₁₂₀N₂₃NaO₁₇P⁺ [M+Na]⁺ 1560.8862, found 1560.8860.

H₂N-Nal-Cpa-Pal-SY-Cit-LRPA-CONH₂ (Pep₉). Global cleavage condition: cocktail A, 3 h. Yield: 64 %. White powder. Analytic HPLC: Gradient B, retention time: 16.3 min, purity: 99.0 %. HRMS(MALDI-TOF): calc. for C₆₈ClH₉₁N₁₇O₁₃⁺ [M+H]⁺ 1388.6665, found 1388.6555; calc. for C₆₈ClH₉₀N₁₇NaO₁₃⁺ [M+Na]⁺ 1410.6485, found 1410.6362.

H₂N-DRVYIHPF-CONH₂ (Pep₁₀). Global cleavage condition: cocktail A, 2 h. Yield: 58 %. White powder. Analytic HPLC: Gradient B, retention time: 13.3 min, purity: 96.2 %. HRMS(MALDI-TOF): calc. for C₅₀H₇₃N₁₄O₁₁⁺ [M+H]⁺ 1045.5578, found 1045.5859; calc. for C₅₀H₇₂N₁₄NaO₁₁⁺ [M+Na]⁺ 1067.5397, found 1067.5725.

H₂N- GHK-CONH₂ (Pep₁₁). Before global cleavage, Alloc deprotection was required. Soak resin in 4 mL DCM, the Pd(PPh₃)₄ (0.5 eq) and PhSiH₃ (10 eq.) was added. The SPE tube was sealed tightly, and shaken overnight. After reaction, little resin was taken for cleaving, and resulting solution was verified by ESI-MS to check if Alloc was deprotected completely. Global cleavage condition: cocktail A, 2 h. Yield: 70 %. White powder. Analytic HPLC: Gradient B, retention time: 6.0 min, purity: 92.8 %. HRMS(MALDI-TOF): calc. for C₁₄H₂₆N₇O₃⁺ [M+H]⁺ 340.2092, found 340.2039; calc. for C₁₄H₂₅N₇NaO₃⁺ [M+Na]⁺ 362.1911, found 362.1856. ¹H NMR (400 MHz, DMSO-*d*₆) δ 14.51 (s, 2H), 8.97

(t, $J = 1.1$ Hz, 1H), 8.68 (d, $J = 8.0$ Hz, 1H), 8.27 (d, $J = 7.5$ Hz, 1H), 8.07 (t, $J = 6.0$ Hz, 3H), 7.82 (s, 3H), 7.55 (s, 1H), 7.37 (s, 1H), 7.19 (s, 1H), 4.71 (q, $J = 6.8$ Hz, 1H), 4.14 (td, $J = 8.3, 5.0$ Hz, 1H), 3.59 (d, $J = 5.6$ Hz, 2H), 3.04 (qd, $J = 15.3, 6.2$ Hz, 2H), 2.75 (q, $J = 6.5$ Hz, 2H), 1.74 – 1.41 (m, 4H), 1.30 (q, $J = 9.5, 8.1$ Hz, 2H); ^{13}C NMR (101 MHz, DMSO- d_6) δ 173.65, 169.17, 165.91, 133.76, 128.64, 117.10, 52.47, 51.48, 39.97 (read from HSQC), 38.53, 31.09, 27.44, 26.57, 22.30.

Synthetic procedure of symmetric dipyrin peptide conjugates: Resin-bound peptide (0.1 mmol) from SPPS was placed in SPE tube, then the aldehyde building block was coupled as routine coupling procedure of SPPS. Upon completion, the liquid phase was removed, and the resin was washed with DMF (4 mL \times 3) and DCM (4 mL \times 3), little part of resin could be taken and be cleaved as a reference. The resin was soared in 4 mL DMF, then pyrrole building block (1.0 mmol) and $\text{BF}_3 \cdot \text{OEt}_2$ (0.1 mmol) was added into SPE tube directly by pipette. The SPE tube was shaken for 3-12 h (usually overnight) in dark. After that, the liquid phase was removed by filtration and resin was washed with DMF (4 mL \times 3) and DCM (4 mL \times 3). Resin was soared in DCM, and DDQ (0.5 mmol) solid was added directly and shaking for 1 h. The resin was washed thoroughly with DMF and DCM until liquid phase was colorless. Finally, 4 mL cleaving cocktail was use for global cleavage. The crude product was purified by preparative HPLC, dry crude product may used for boron complexation directly.

DP₁-HN-YFMVF-CONH₂ (DP₁-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 51 %. Red powder. Analytic HPLC: Gradient A, retention time: 33.3 min, purity: 96.3 %. HRMS(MALDI-TOF): calc. for $\text{C}_{57}\text{H}_{67}\text{N}_8\text{O}_7\text{S}^+$ $[\text{M}+\text{H}]^+$ 1007.4848, found 1007.4843.

DP₂-HN-YFMVF-CONH₂ (DP₂-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 41 %. Orange powder. Analytic HPLC: Gradient A, retention time: 31.3 min, purity: 94.0 %. HRMS(MALDI-TOF): calc. for $\text{C}_{55}\text{H}_{63}\text{N}_8\text{O}_7\text{S}^+$ $[\text{M}+\text{H}]^+$ 979.4535, found 979.4522; calc. for $\text{C}_{55}\text{H}_{62}\text{N}_8\text{NaO}_7\text{S}^+$ $[\text{M}+\text{Na}]^+$ 1001.4354, found 1001.4316.

DP₃-HN-YFMVF-CONH₂ (DP₃-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 53 %. Red powder. Analytic HPLC: Gradient A, retention time: 39.3 min, purity: 96.6 %. HRMS(MALDI-TOF): calc. for $\text{C}_{61}\text{H}_{75}\text{N}_8\text{O}_7\text{S}^+$ $[\text{M}+\text{H}]^+$ 1063.5474, found 1063.5477; calc. for $\text{C}_{61}\text{H}_{74}\text{N}_8\text{NaO}_7\text{S}^+$ $[\text{M}+\text{Na}]^+$ 1085.5293, found 1085.5297.

DP₄-HN-YFMVF-CONH₂ (DP₄-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 10 %. Yellow powder. Analytic HPLC: Gradient A, retention time: 28.9 min, purity: 96.0 %. HRMS(MALDI-TOF): calc. for $\text{C}_{53}\text{H}_{59}\text{N}_8\text{O}_7\text{S}^+$ $[\text{M}+\text{H}]^+$ 951.4222, found 951.4234; calc. for $\text{C}_{53}\text{H}_{58}\text{N}_8\text{NaO}_7\text{S}^+$ $[\text{M}+\text{Na}]^+$ 973.4041, found 973.4025.

DP₅-HN-YFMVF-CONH₂ (DP₅-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 51 %. Red powder. Analytic HPLC: Gradient A, retention time: 32.6 min, purity: 97.6 %. HRMS(MALDI-TOF): calc. for $\text{C}_{61}\text{H}_{63}\text{N}_8\text{O}_7\text{S}^+$ $[\text{M}+\text{H}]^+$ 1051.4535, found 1051.4539; calc. for $\text{C}_{61}\text{H}_{62}\text{N}_8\text{NaO}_7\text{S}^+$ $[\text{M}+\text{Na}]^+$ 1073.4354, found 1073.4337.

DP₆-HN-YFMVF-CONH₂ (DP₆-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 50 %. Red powder. Analytic HPLC: Gradient A, retention time: 33.2 min, purity: 94.8 %. HRMS(MALDI-TOF): calc. for $\text{C}_{63}\text{H}_{67}\text{N}_8\text{O}_9\text{S}^+$ $[\text{M}+\text{H}]^+$ 1111.4746, found 1111.4750.

DP₉-HN-YFMVF-CONH₂ (DP₉-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 35 %. Purple powder. Analytic HPLC: Gradient A, retention time: 40.2 min, purity: 95.2 %. HRMS(MALDI-TOF): calc. for $\text{C}_{65}\text{H}_{67}\text{N}_8\text{O}_7\text{S}^+$ $[\text{M}+\text{H}]^+$ 1103.4848, found 1103.5282.

DP₁₀-HN-YFMVF-CONH₂ (DP₁₀-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 47 %. Red powder. Analytic HPLC: Gradient A, retention time: 32.4 min, purity: 95.2 %. HRMS(MALDI-TOF): calc. for C₅₇H₆₇N₈O₇S⁺ [M+H]⁺ 1007.4848, found 1007.4864; calc. for C₅₇H₆₆N₈NaO₇S⁺ [M+Na]⁺ 1029.4667, found 1029.4450.

DP₁₂-HN-YFMVF-CONH₂ (DP₁₂-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 40 %. Red powder. Analytic HPLC: Gradient A, retention time: 33.5 min, purity: 96.0 %. HRMS(MALDI-TOF): calc. for C₅₈H₆₉N₈O₈S⁺ [M+H]⁺ 1037.4954, found 1037.4929.

DP₁₃-HN-YFMVF-CONH₂ (DP₁₃-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 40 %. Red powder. Analytic HPLC: Gradient A, retention time: 32.8 min, purity: 97.6 %. HRMS(MALDI-TOF): calc. for C₅₈H₆₉N₈O₈S⁺ [M+H]⁺ 1037.4954, found 1037.4922; calc. for C₅₈H₆₈N₈NaO₈S⁺ [M+Na]⁺ 1059.4773, found 1059.4777.

DP₁₄-HN-YFMVF-CONH₂ (DP₁₄-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 34 %. Orange powder. Analytic HPLC: Gradient A, retention time: 33.1 min, purity: 96.4 %. HRMS(MALDI-TOF): calc. for C₅₇H₇₀N₉O₇S⁺ [M+H]⁺ 1024.5113, found 1024.5127.

DP₁-HN-C-Ahx-RrRKGGYFMVF-COOH (DP₁-Pep₂). Global cleavage condition: cocktail B, 4 h. Yield: 44 %. Red powder. Analytic HPLC: Gradient B, retention time: 23.1 min, purity: 95.8 %. HRMS(MALDI-TOF): calc. for C₉₁H₁₂₈N₂₁O₁₅⁺ [M+H]⁺ 1755.9927, found 1755.9933.

DP₁-HN-C-Ahx-YFMVFGGRrRK-COOH (DP₁-Pep₃). Global cleavage condition: cocktail B, 4 h. Yield: 49 %. Red powder. Analytic HPLC: Gradient B, retention time: 21.8 min, purity: 97.0 %. HRMS(MALDI-TOF): calc. for C₉₄H₁₃₆N₂₅O₁₆S₂⁺ [M+H]⁺ 1936.0066, found 1936.0118.

DP₁-HN-C-Ahx-YFMVFGGRrRK-COOH (DP₁-Pep₄). Global cleavage condition: cocktail B, 4 h. Yield: 42 %. Red powder. Analytic HPLC: Gradient B, retention time: 22.2 min, purity: 96.6 %. HRMS(MALDI-TOF): calc. for C₉₄H₁₃₆N₂₅O₁₆S₂⁺ [M+H]⁺ 1936.0066, found 1936.0017.

DP₁-HN-NWYFIVF-COOH (DP₁-Pep₅). Global cleavage condition: cocktail A, 2 h. Yield: 47 %. Red powder. Analytic HPLC: Gradient A, retention time: 34.2 min, purity: 99.0 %. HRMS(MALDI-TOF): calc. for C₇₈H₉₂N₁₃O₁₃⁺ [M+H]⁺ 1418.6932, found 1418.6955.

DP₁-HN-DEHYFIVF-COOH (DP₁-Pep₆). Global cleavage condition: cocktail A, 2 h. Yield: 50 %. Red powder. Analytic HPLC: Gradient A, retention time: 27.3 min, purity: 96.7 %. HRMS(MALDI-TOF): calc. for C₇₃H₈₇N₁₂O₁₅⁺ [M+H]⁺ 1371.6408, found 1371.6353; calc. for C₇₃H₈₆N₁₂NaO₁₅⁺ [M+Na]⁺ 1393.6228, found 1393.6185.

DP₁-HN-Ahx-P-(pTyr)-LKTK-COOH (DP₁-Pep₇). Global cleavage condition: cocktail B, 3 h. Yield: 47 %. Red powder. Analytic HPLC: Gradient B, retention time: 17.6 min, purity: 98.2 %. HRMS(MALDI-TOF): calc. for C₆₂H₉₁N₁₁O₁₄P⁺ [M+H]⁺ 1244.6479, found 1244.6422.

DP₁-HN-Ahx-P-(pTyr)-LKTKRrRK-COOH (DP₁-Pep₈). Global cleavage condition: cocktail B, 3 h. Yield: 48 %. Red powder. Analytic HPLC: Gradient B, retention time: 25.0 min, purity: 96.8 %. HRMS(MALDI-TOF): calc. for C₈₆H₁₃₉N₂₅O₁₈P⁺ [M+H]⁺ 1842.0496, found 1842.0535.

DP₁-HN-DRVYIHPF-CONH₂ (DP₁-Pep₁₀). Global cleavage condition: cocktail A, 2 h. Yield: 53 %. Red powder. Analytic HPLC: Gradient B, retention time: 15.8 min, purity: 98.9 %. HRMS(MALDI-TOF): calc. for C₇₀H₉₁N₁₆O₁₂⁺ [M+H]⁺ 1347.6997, found 1347.7072.

DP₁-HN-GHK-CONH₂ (DP₁-Pep₁₁). Before global cleavage, Alloc deprotection was required (the procedure described on Pep₁₁). Global cleavage condition: cocktail A, 2 h. Yield: 59 %. Red powder. Analytic HPLC: Gradient B, retention time: 12.1 min, purity: 95.9 %. HRMS(MALDI-TOF): calc. for C₃₄H₄₄N₉O₄⁺ [M+H]⁺ 642.3511, found 642.3905; calc. for C₃₄H₄₃N₉NaO₄⁺ [M+Na]⁺ 664.3330, found

664.3734. ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 14.44 (s, 2H), 12.20 (s, 2H), 9.10 (t, $J = 5.8$ Hz, 1H), 9.00 (d, $J = 1.5$ Hz, 1H), 8.38 (d, $J = 8.0$ Hz, 1H), 8.13 (d, $J = 7.5$ Hz, 1H), 8.11 – 8.02 (m, 2H), 7.81 (s, 3H), 7.53 (d, $J = 8.1$ Hz, 2H), 7.50 (d, $J = 2.0$ Hz, 1H), 7.41 (d, $J = 1.3$ Hz, 1H), 7.22 – 7.15 (m, 1H), 6.53 (s, 2H), 4.65 (td, $J = 7.8, 5.4$ Hz, 1H), 4.16 (td, $J = 8.5, 4.9$ Hz, 1H), 3.94 (d, $J = 5.7$ Hz, 2H), 3.20 – 2.98 (m, 2H), 2.78 (h, $J = 6.0$ Hz, 2H), 2.43 (s, 6H), 1.83 – 1.38 (m, 10H), 1.33 (q, $J = 9.2, 8.3$ Hz, 2H); ^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ 173.67, 169.67, 168.94, 165.75, 152.41, 145.34, 142.32, 138.40, 136.91, 133.69, 129.09, 127.93, 120.48, 116.98, 52.48, 51.49, 42.70, 38.59, 31.07, 27.07, 26.60, 22.31, 13.79, 13.73.

DP₁-HN-GHK-CONH₂ (DP₁-Pep₁₁)*. Fmoc and Alloc dual-protected peptide on resin was used. Alloc deprotection was conducted as procedure described on **Pep₁₁**. Then, the dipyrin was constructed as general procedure above. Fmoc was deprotected before global cleavage. Global cleavage condition: cocktail A, 2 h. Yield: 51 %. Red powder. Analytic HPLC: Gradient B, retention time: 13.1 min, purity: 98.9 %. HRMS(MALDI-TOF): calc. for $\text{C}_{34}\text{H}_{44}\text{N}_9\text{O}_4^+ [\text{M}+\text{H}]^+$ 642.3511, found 642.3593; calc. for $\text{C}_{34}\text{H}_{43}\text{N}_9\text{NaO}_4^+ [\text{M}+\text{Na}]^+$ 664.3330, found 664.3391. ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 14.38 (s, 2H), 12.16 (s, 2H), 8.99 (s, 1H), 8.78 (t, $J = 5.7$ Hz, 1H), 8.68 (d, $J = 8.0$ Hz, 1H), 8.29 (d, $J = 7.2$ Hz, 1H), 8.04 (s, 3H), 8.02 (s, 2H), 7.58 (s, 1H), 7.50 (d, $J = 8.0$ Hz, 2H), 7.40 (s, 1H), 7.21 (s, 1H), 6.53 (s, 2H), 4.74 (q, $J = 6.7$ Hz, 1H), 4.19 – 4.15 (m, 1H), 3.63 (s, 2H), 3.27 (t, $J = 6.8$ Hz, 2H), 3.09 – 3.01 (m, 2H), 2.43 (s, 6H), 1.60 (ddp, $J = 37.7, 15.0, 7.7, 6.9$ Hz, 10H), 1.39 – 1.30 (m, 2H).

Synthetic procedure of asymmetric dipyrin peptide conjugates: Resin-bound peptide was loaded in SPE tube, then the 5-formyl-2,4-dimethyl-3-pyrrolicarboxylic acid was coupled as general coupling procedure of SPPS. Upon completion, the liquid phase was removed by filtration, and the resin was washed with DMF (4 mL \times 3) and DCM (4 mL \times 3). The resin was soaked in DCM, and the pyrrole building block (0.5 mmol) and POCl_3 (0.5 mmol) was added by pipette into SPE tube directly. The mixture was shaken overnight in dark. Then, the resin was washed thoroughly with DMF and DCM until liquid phase was colorless, suitable cleavage cocktail was used for global cleavage. The crude product was purified by preparative HPLC.

DP₁₅-HN-YFMVF-CONH₂ (DP₁₅-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 45 %. Yellow powder. Analytic HPLC: Gradient A, retention time: 30.3 min, purity: 95.2 %. HRMS(MALDI-TOF): calc. for $\text{C}_{51}\text{H}_{62}\text{N}_8\text{NaO}_7\text{S}^+ [\text{M}+\text{Na}]^+$ 953.4354, found 953.4309; calc. for $\text{C}_{51}\text{H}_{62}\text{KN}_8\text{O}_7\text{S}^+ [\text{M}+\text{K}]^+$ 969.4094, found 969.4078.

DP₁₆-HN-YFMVF-CONH₂ (DP₁₆-Pep₁). Global cleavage condition: cocktail A, 2 h. Yield: 42 %. Red powder. Analytic HPLC: Gradient A, retention time: 31.4 min, purity: 95.1 %. HRMS(MALDI-TOF): calc. for $\text{C}_{54}\text{H}_{63}\text{N}_8\text{O}_8\text{S}^+ [\text{M}+\text{H}]^+$ 983.4484, found 983.5748; calc. for $\text{C}_{54}\text{H}_{62}\text{N}_8\text{NaO}_8\text{S}^+ [\text{M}+\text{Na}]^+$ 1005.4304, found 1005.5553. ^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 13.78 (s, 1H), 12.55 (s, 1H), 9.03 (s, 1H), 8.48 (s, 1H), 8.28 (d, $J = 7.8$ Hz, 2H), 8.21 (dd, $J = 8.3, 3.3$ Hz, 1H), 8.00 (d, $J = 8.3$ Hz, 1H), 7.87 (d, $J = 2.4$ Hz, 1H), 7.82 (d, $J = 8.7$ Hz, 1H), 7.54 (d, $J = 8.8$ Hz, 1H), 7.36 (d, $J = 2.3$ Hz, 1H), 7.25 – 7.18 (m, 10H), 7.08 (d, $J = 8.4$ Hz, 2H), 7.05 (s, 1H), 7.01 (dd, $J = 8.8, 2.4$ Hz, 1H), 6.66 (d, $J = 8.4$ Hz, 2H), 4.67 – 4.61 (m, 2H), 4.48 (m, 1H), 4.41 (m, 1H), 4.17 – 4.14 (m, 1H), 3.88 (s, 3H), 3.00 (td, $J = 8.5, 7.9, 3.7$ Hz, 3H), 2.81 (d, $J = 9.1$ Hz, 3H), 2.41 (td, $J = 7.3, 6.7, 3.1$ Hz, 2H), 2.35 (s, 3H), 2.30 (s, 3H), 2.02 (s, 3H), 1.94 – 1.88 (m, 2H), 1.80 – 1.76 (m, 1H), 0.79 (dd, $J = 6.8, 4.0$ Hz, 6H).

General procedure of boron complexation of dipyrin conjugates: To a suspension of dipyrin peptide conjugate (0.02 mmol) in acetonitrile (1.0 mL), DIPEA (170 μL) was added by pipette. The mixture was shaken or sonicated for 5 min. $\text{BF}_3\cdot\text{OEt}_2$ (280 μL) was added by pipette in one portion. The mixture was

further shaken for 10 min. The crude product was concentrated under vacuum and was purified by preparative HPLC.

BODIPY₁-HN-YFMVF-CONH₂ (BODIPY₁-Pep₁). Yield: 73 %. Red powder. Analytic HPLC: Gradient A, retention time: 47.5 min, purity: 97.0 %. HRMS(MALDI-TOF): calc. for BC₅₇H₆₈N₈O₉S⁺ [M-2F+2OH+H]⁺ 1051.4987, found 1051.4934; calc. for BC₅₇F₂H₆₅KN₈O₇S⁺ [M+K]⁺ 1093.4460, found 1093.4703. ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.20 (s, 1H), 8.74 (d, *J* = 8.2 Hz, 1H), 8.27 (dd, *J* = 28.2, 7.9 Hz, 2H), 8.05 (s, 1H), 8.02 (d, *J* = 8.4 Hz, 2H), 7.85 (d, *J* = 8.7 Hz, 1H), 7.55 (d, *J* = 8.1 Hz, 2H), 7.47 – 7.37 (m, 1H), 7.35 – 7.19 (m, 10H), 7.18 – 7.15 (m, 2H), 7.14 – 7.05 (m, 1H), 6.71 – 6.64 (m, 2H), 6.25 (s, 2H), 4.64 (dtd, *J* = 24.6, 8.6, 4.1 Hz, 2H), 4.54 (td, *J* = 8.6, 5.2 Hz, 1H), 4.47 (td, *J* = 8.3, 4.9 Hz, 1H), 4.20 (dd, *J* = 8.7, 6.7 Hz, 1H), 3.18 – 2.97 (m, 3H), 2.95 – 2.85 (m, 3H), 2.52 (s, 6H), 2.46 (tt, *J* = 6.6, 2.9 Hz, 2H), 2.07 (s, 3H), 2.01 – 1.92 (m, 2H), 1.84 (dt, *J* = 13.3, 4.7 Hz, 1H), 1.39 (s, 6H), 0.84 (dd, *J* = 6.8, 2.6 Hz, 6H); ¹³C NMR (101 MHz, DMSO-*d*₆) δ 172.65, 171.48, 170.88, 170.75, 170.45, 165.61, 155.68, 155.13, 142.63, 141.07, 137.76, 136.91, 134.52, 130.39, 130.06, 129.26, 129.06, 128.37, 127.98, 127.91, 126.20, 121.51, 114.83, 57.66, 55.29, 53.90, 53.49, 51.91, 37.61, 37.23, 35.98, 32.01, 30.60, 29.49, 29.01, 19.13, 17.99, 14.65, 14.22, 14.15; ¹⁹F NMR (376 MHz, DMSO-*d*₆) δ -145.48 (dd, *J* = 65.6, 30.6 Hz).

BODIPY₁-HN-YFMVF-CONH₂ (BODIPY₁-Pep₁). Yield: 73 % (from purified **DP₁-Pep₁**) or 45 % (from dry crude product of **DP₁-Pep₁** from SPPS) Red powder. Analytic HPLC: Gradient A, retention time: 47.5 min, purity: 97.0 %. HRMS(MALDI-TOF): calc. for C₅₇H₆₈N₈O₉S⁺ [M-2F+2OH+H]⁺ 1051.4987, found 1051.4934; calc. for C₅₇H₆₅BF₂KN₈O₇S⁺ [M+K]⁺ 1093.4460, found 1093.4703.

BODIPY₂-HN-YFMVF-CONH₂ (BODIPY₂-Pep₁). Yield: 76 %. Orange powder. Analytic HPLC: Gradient A, retention time: 46.2 min, purity: 95.0 %. HRMS(MALDI-TOF): calc. for BC₅₅FH₆₁N₈O₇S⁺ [M-F]⁺ 1007.4525, found 1007.4451; calc. for C₅₅H₆₁BF₂N₈NaO₇S⁺ [M+Na]⁺ 1049.4407, found 1049.3821; calc. for C₅₅H₆₁BF₂KN₈O₇S⁺ [M+K]⁺ 1065.4147, found 1065.4182.

BODIPY₃-HN-YFMVF-CONH₂ (BODIPY₃-Pep₁). Yield: 70 %. Red powder. Analytic HPLC: Gradient A, retention time: 54.7 min, purity: 96.0 %. HRMS(MALDI-TOF): calc. for C₆₁H₇₅N₈O₇S⁺ [M-BF₂+2H]⁺ 1063.5474, found 1063.5389; calc. for C₆₁H₇₃BF₂N₈NaO₇S⁺ [M+Na]⁺ 1133.5346, found 1133.5359; calc. for C₆₁H₇₃BF₂KN₈O₇S⁺ [M+K]⁺ 1149.5086, found 1149.5101.

BODIPY₉-HN-YFMVF-CONH₂ (BODIPY₉-Pep₁). Yield: 55 %. Purple powder. Analytic HPLC: Gradient A, retention time: 53.0 min, purity: 94.6 %. HRMS(MALDI-TOF): calc. for BC₆₅H₆₈N₈O₉S⁺ [M-2F+2OH+H]⁺ 1147.4987, found 1147.5021; calc. for C₆₅H₆₅BF₂N₈NaO₇S⁺ [M+Na]⁺ 1173.4720, found 1173.4887; calc. for C₆₅H₆₅BF₂KN₈O₇S⁺ [M+K]⁺ 1189.4460, found 1189.4789.

BODIPY₁₅-HN-YFMVF-CONH₂ (BODIPY₁₅-Pep₁). Yield: 71 %. Orange powder. Analytic HPLC: Gradient A, retention time: 42.8 min, purity: 94.1 %. HRMS(MALDI-TOF): calc. for BC₅₁FH₆₁N₈O₇S⁺ [M-F]⁺ 959.4525, found 959.5180; calc. for C₅₁H₆₁BF₂N₈NaO₇S⁺ [M+Na]⁺ 1001.4407, found 1001.5133; calc. for BC₅₁F₂H₆₁KN₈O₇S⁺ [M+K]⁺ 1017.4147, found 1017.5093.

BODIPY₁-HN-C-Ahx-YFMVFGGRrRK-COOH (BODIPY₁-Pep₄). Yield: 49 %. EDT was used as reductant during reaction to prevent the oxidation of Cys. Red powder. Analytic HPLC: Gradient B, retention time: 29.5 min, purity: 98.7 %. HRMS(MALDI-TOF): calc. for C₉₄H₁₃₅BF₂N₂₅O₁₆S₂⁺ [M+H]⁺ 1984.0119, found 1983.9865.

BODIPY₁-HN-NWYFIVE-COOH (BODIPY₁-Pep₅). Yield: 63 %. Red powder. Analytic HPLC: Gradient A, retention time: 46.4 min, purity: 97.7 %. HRMS(MALDI-TOF): calc. for C₇₈H₉₀BF₂N₁₃NaO₁₃⁺ [M+Na]⁺ 1488.6804, found 1488.6771; calc. for C₇₈H₉₀BF₂KN₁₃O₁₃⁺ [M+K]⁺ 1504.6544, found 1504.6543.

BODIPY₁-HN-DEHYFIVF-COOH (BODIPY₁-Pep₆). Yield: 31 %. Red powder. Analytic HPLC: Gradient A, retention time: 35.8 min, purity: 95.2 %. HRMS(MALDI-TOF): calc. for C₇₃H₈₅BF₂N₁₂NaO₁₅⁺ [M+Na]⁺ 1441.6281, found 1441.5949; calc. for C₇₃H₈₅BF₂KN₁₂O₁₅⁺ [M+K]⁺ 1457.6020, found 1457.5786.

DP₁-HN-Ahx-P-(pTyr)-LKTK-COOH (BODIPY₁-Pep₇). Yield: 50 %. Red powder. Analytic HPLC: Gradient B, retention time: 19.3 min, purity: 95.9 %. HRMS(MALDI-TOF): calc. for C₆₂H₈₉BFN₁₁O₁₄P⁺ [M-F]⁺ 1272.6470, found 1272.7580.

BODIPY₁-HN-DRVYIHPF-CONH₂ (BODIPY₁-Pep₁₀). Yield: 37 %. Red powder. Analytic HPLC: Gradient B, retention time: 19.3 min, purity: 95.9 %. HRMS(MALDI-TOF): calc. for C₇₀H₉₁BF₂N₁₆O₁₂⁺ [M+H]⁺ 1396.7128, found 1396.6222.

Biological experiments

Cell Culture Conditions. Nasopharyngeal carcinoma (NPC) cell line C666 and Cervical carcinoma cell line HeLa were used in this study. C666 and HeLa were cultivated in Roswell Park Memorial Institute Medium (RPMI 1640) and Dulbecco's Modified Eagle Medium (DMEM) containing 10% v/v Fetal Bovine Serum (FBS) and 1% v/v Penicillin Streptomycin respectively. Both C666 and HeLa were maintained at 37 °C and 5% CO₂.

Dark Cytotoxicity Assay. The dark cytotoxicity and were assessed by MTT viability assay. Cells (6 x 10³ per well) were seeded onto 96-well plates and then incubated at 37 °C with 5% CO₂ in dark for 24 h prior to the addition of samples. The cells were then incubated with samples for another 24 h. Medium were then removed, and the cell monolayers were washed with 1X PBS and then incubated with 100 mL medium with 5% v/v MTT solution (5 mg/mL) at 37 °C for 2.5 h. 80 mL of solution were removed and 100 mL Dimethyl sulfoxide (DMSO) were added to dissolve the formazan crystals. The absorbance of the formazan crystal was measured at 540 nm and 650 nm by dual-wavelength Azure microplate reader after 1 h of shaking.

Light Cytotoxicity Assay. The light cytotoxicity of samples was assessed with MTT viability assay. The procedures of the MTT viability assay were previously described. Cells were irradiated with optical dose of 5J/cm² after 24 h incubation with sample and the medium were replaced with fresh medium prior to irradiation.

In vitro imaging and co-staining. Cell were incubated with **BODIPY₁-Pep₄** (10mM) for 24h and then co-stained with Hoechst 33342 nuclear dye for 15 minutes. Imaging was performed by a Nikon Eclipse Ti2 confocal laser-scanning microscope.

Expression and purification of EBNA1 (a.a. 468-607). The gene construct for N-terminally hexahistidine tagged SMT3-EBNA1 DNA binding domain (residues 468-607) fusion protein was chemically synthesized and cloned into pET28a (+) vector (Genscript). The plasmid was transformed into *Escherichia coli* BL21 (DE3) competent cells and grown in 2YT media supplemented with 50 µg/ml kanamycin at 37 °C. When the OD₆₀₀ of the cells reached 0.8, expression was induced with 0.5 mM isopropyl β-D-1-thiogalactopyranoside (IPTG) and the cells were grown at 25 °C overnight. The cells were harvested by centrifugation at 5000 rpm (6,238 x g) for 10 mins at 4 °C. The following protein purification protocol was adapted from a previously published report^[3]. The cell pellet was resuspended in lysis buffer (20 mM Tris, pH 8.0, 500 mM NaCl, 10 mM imidazole, 5% (v/v) glycerol, 5 mM β-mercaptoethanol (BME), 17.4 µg/ml phenylmethylsulfonyl fluoride (PMSF) and 1 mM MgCl₂). The resuspended cells were lysed by sonication and the lysate was separated from the insoluble fractions by centrifugation at 24,000 rpm (69,673 x g) for 30 min at 4 °C. The protein was purified by nickel affinity using a His-trap column (GE Healthcare) equilibrated with lysis buffer. The lysate was first loaded onto the His-trap column, then washed with 10 column volumes of wash buffer (20 mM Tris pH 8.0, 500 mM NaCl, 30 mM imidazole, 5% glycerol and 5 mM BME) and the protein was eluted with 3 - 4 column volumes of elution buffer (20 mM Tris pH 8.0, 500 mM NaCl, 300 mM imidazole, 5% glycerol and 5 mM BME). The eluted protein was treated with ULP1 to cleave the hexahistidine-SMT3 protein tag. Any residual uncleaved fusion protein and the histidine-tagged ULP1 protease were removed by nickel affinity while the tag-free EBNA1 (468-607) protein was recovered by flowing wash buffer through the His-trap column (GE Healthcare). The fractions were pooled, concentrated and further purified by size exclusion chromatography using a Superdex 200 Increase 10/300 column (GE Healthcare), equilibrated with a buffer containing 1 mM HEPES, pH 7.2, 500

mM NaCl and 10 mM dithiothreitol (DTT). The relevant protein fractions were analysed using SDS-PAGE, pooled, concentrated to 6 mg/ml and stored at -80 °C.

Fluorescent titration. The EBNA1 protein was prepared as the procedure above, while Bovine serum albumin (BSA) protein was purchased from Thermo Fisher Scientific. Both BSA and EBNA1 were further prepared as solution (120 µM) in buffer (1 mM HEPES, pH 7.2, 500 mM NaCl and 10 mM dithiothreitol). The protein solution was gradually added into **BODIPY₁-Pep₄** (2000 µL 1.2 µM in same buffer) by pipette. Each time after addition of protein, the resulting solution was shaken slowly for around 2 min, then the fluorescent spectrum was recorded. The experiment was stopped when the influence on fluorescence was saturated. The increased of volume is less than 4% in total.

Reference:

- [1] M. Kondo, S. Furukawa, K. Hirai, S. Kitagawa, *Angew. Chemie - Int. Ed.* **2010**, *49*, 5327–5330.
- [2] S. Eissler, M. Kley, D. Bächle, G. Loidl, T. Meier, D. Samson, *J. Pept. Sci.* **2017**, *23*, 757–762.
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NMR spectra of products

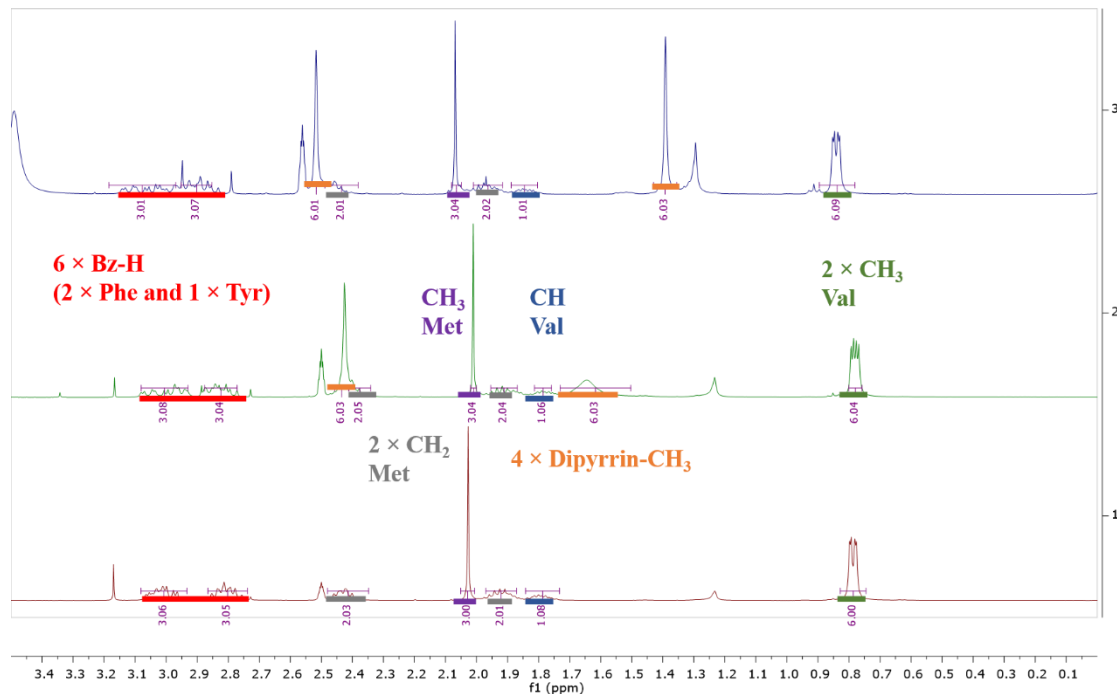


Fig. S8. Comparison and assignment of $^1\text{H-NMR}$ for **Pep₁** (bottom), **DP₁-Pep₁** (middle) and **BODIPY₁-Pep₁** (top) in range 3.5~0.0 ppm. Signal of side chain $\text{sp}^3\text{C-H}$ and methyl groups on dipyrrin/BODIPY.

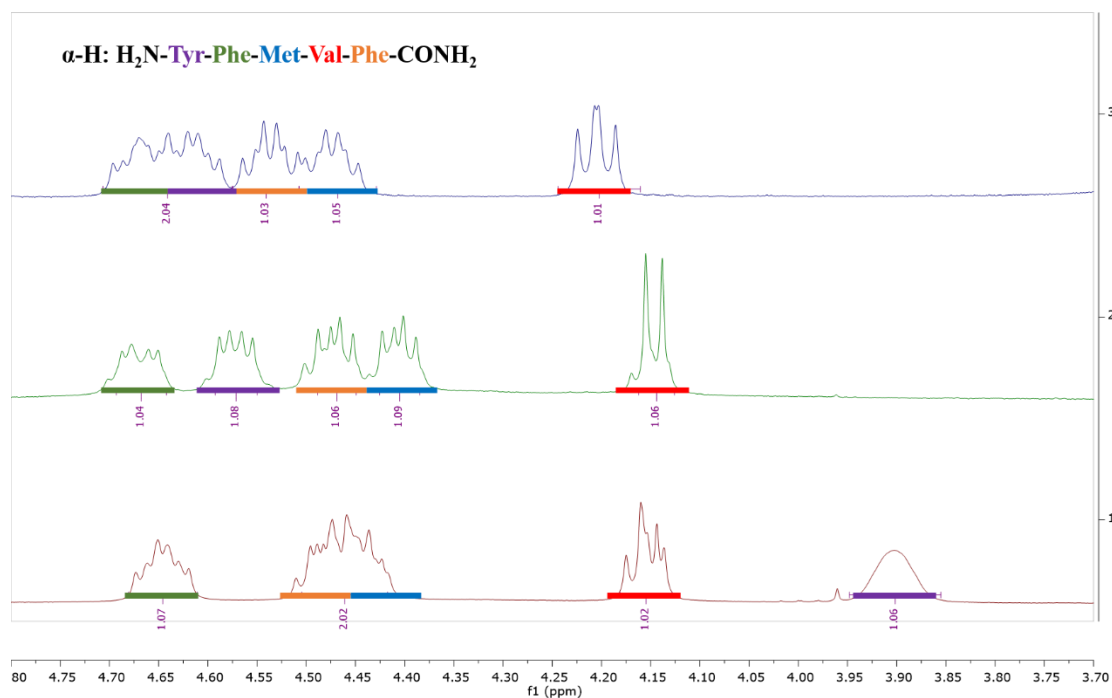


Fig. S9. Comparison and assignment of $^1\text{H-NMR}$ for **Pep₁** (bottom), **DP₁-Pep₁** (middle) and **BODIPY₁-Pep₁** (top) in range 4.8~3.7 ppm. Signal of $\alpha\text{-H}$ on the backbone of peptide.

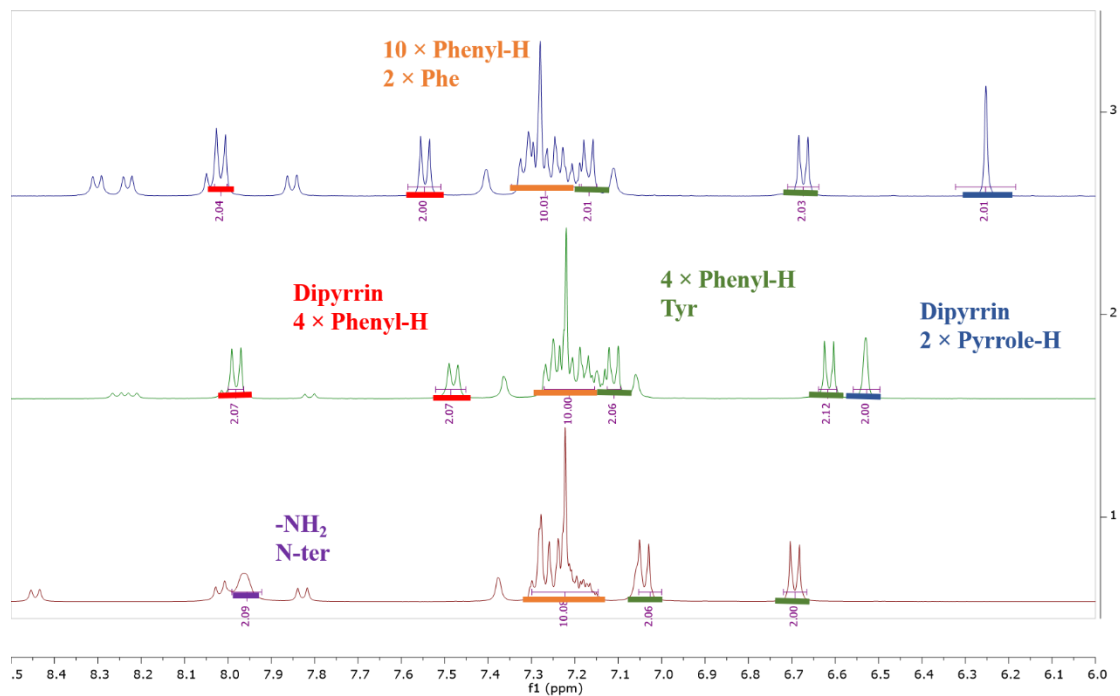


Fig. S10. Comparison and assignment of ^1H -NMR for **Pep₁** (bottom), **DP₁-Pep₁** (middle) and **BODIPY₁-Pep₁** (top) in range 9.0~6.0 ppm. Signal of aryl proton from Phe, Tyr and dipyrrin/BODIPY, as well as amide proton from backbone of peptide.

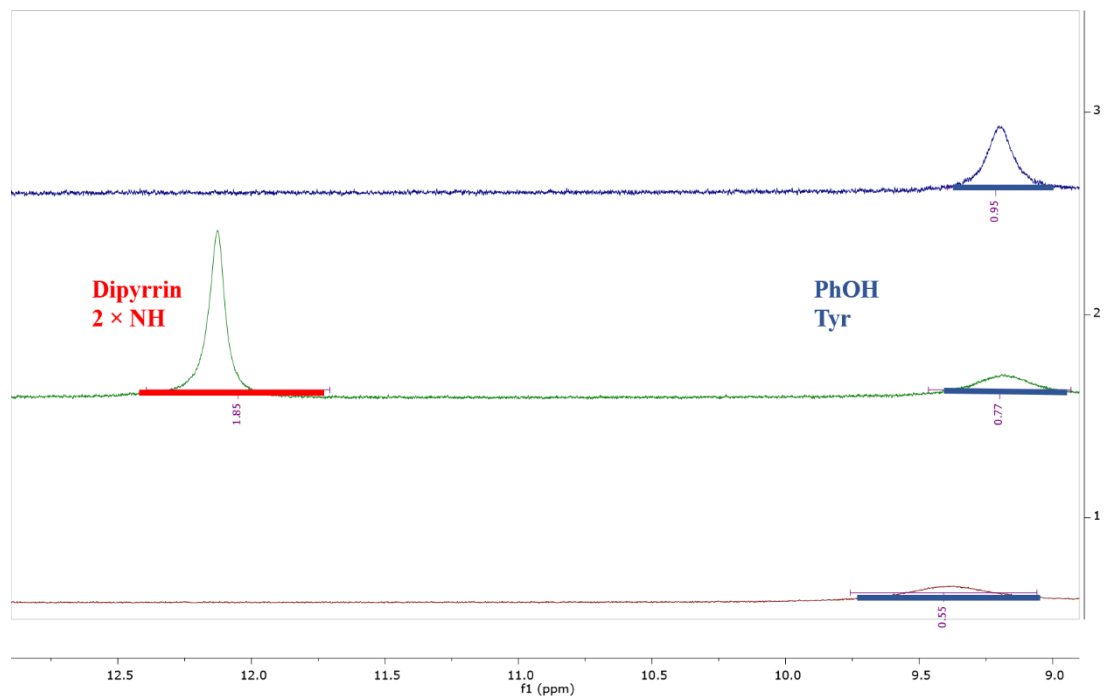
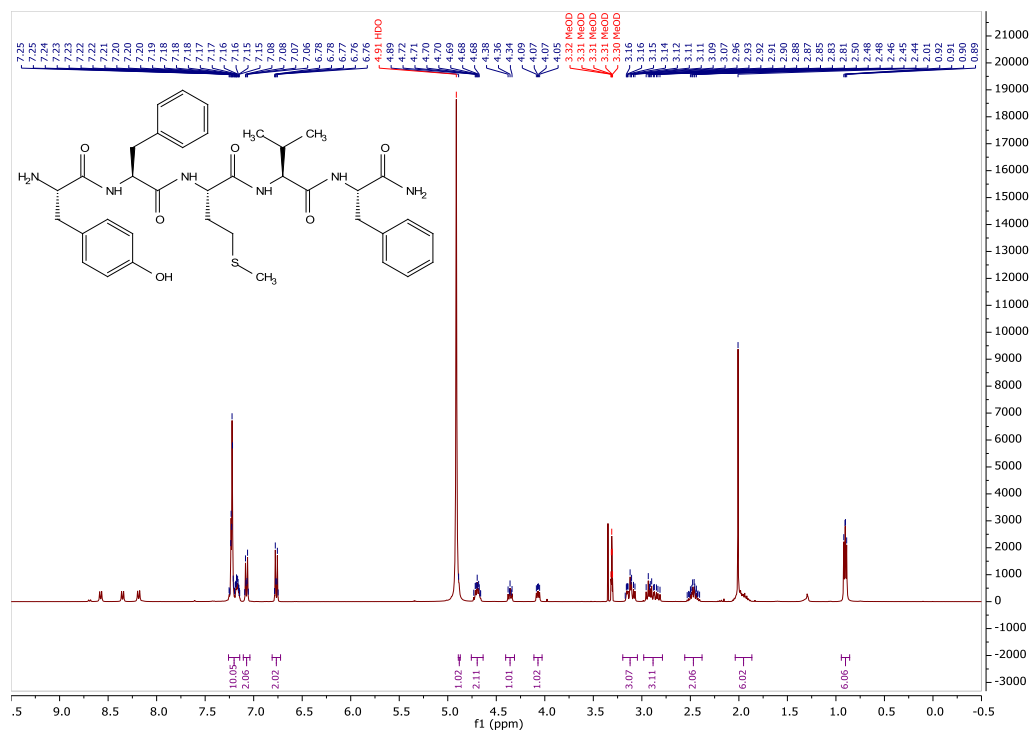
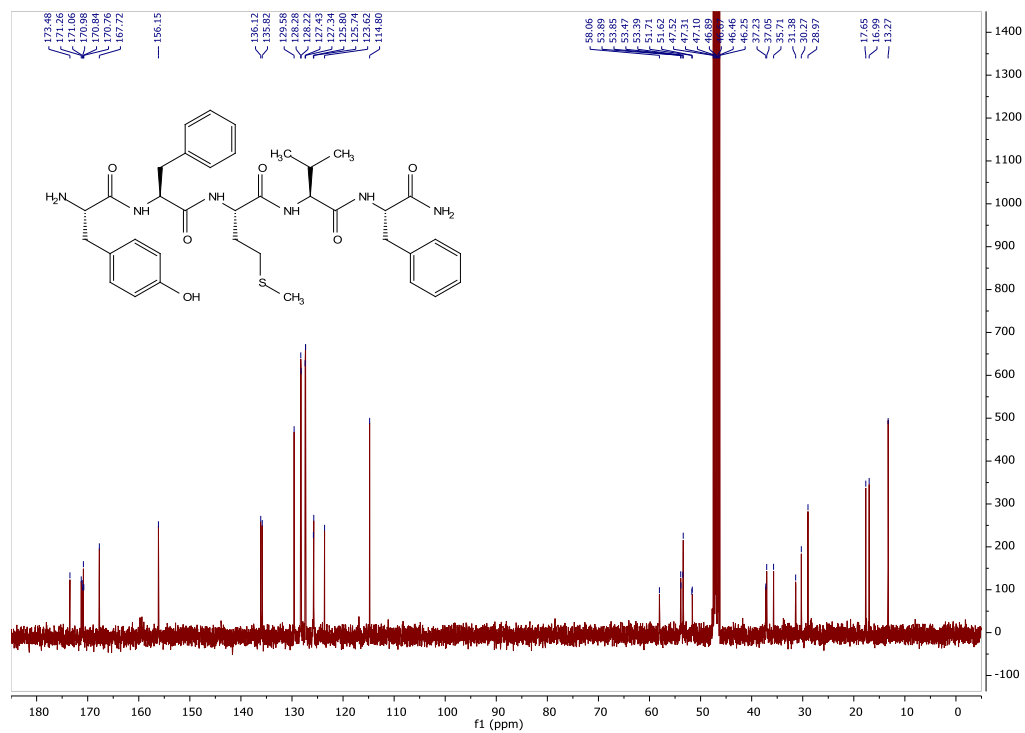


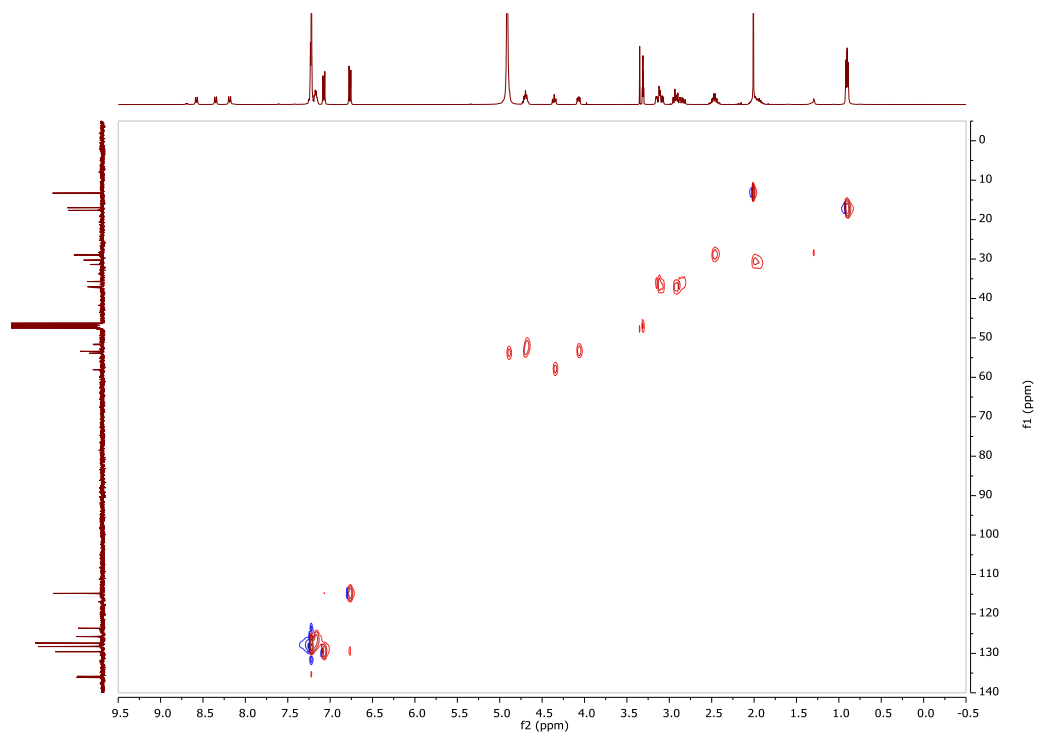
Fig. S11. Comparison and assignment of ^1H -NMR for **Pep₁** (bottom), **DP₁-Pep₁** (middle) and **BODIPY₁-Pep₁** (top) in range 12.9~8.9 ppm. Signal of acidic proton from Tyr (phenol-H) and dipyrrin (NH).



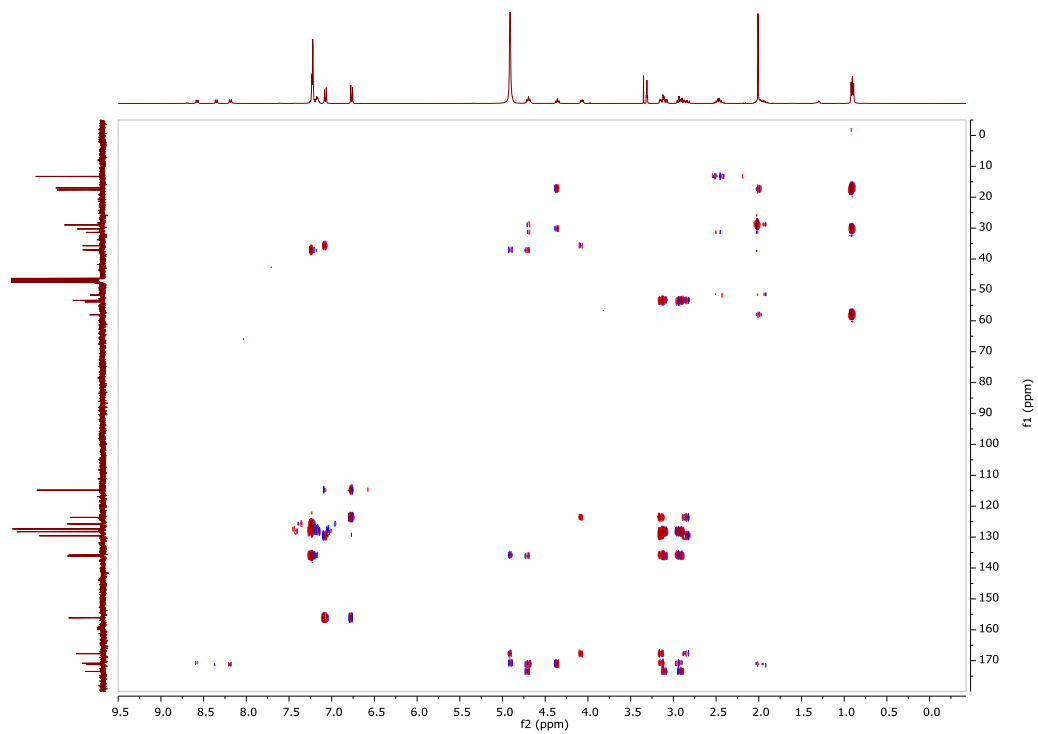
¹H-NMR spectrum of Pep₁ in methanol-*d*₄.



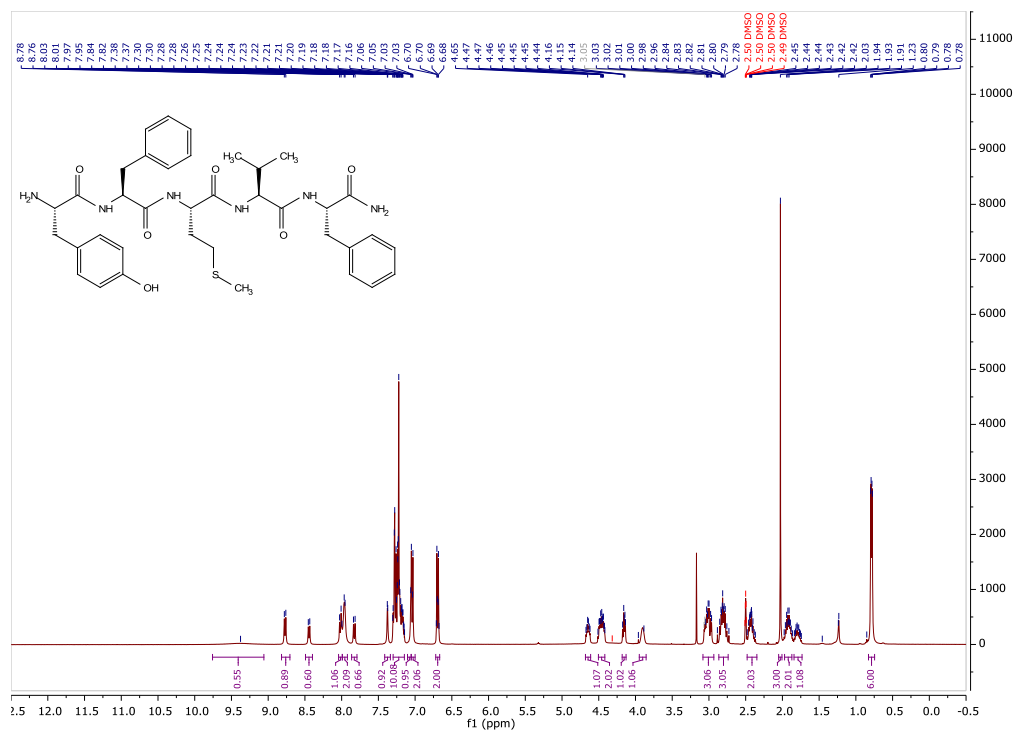
¹³C-NMR spectrum of Pep₁ in methanol-*d*₄.



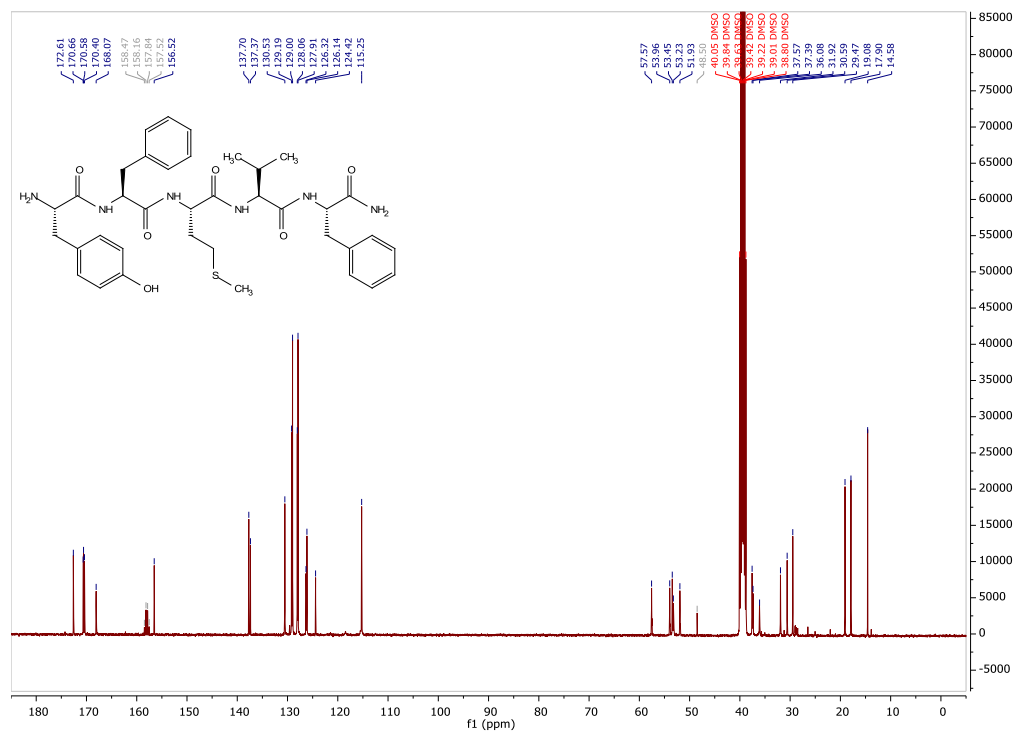
HSQC spectrum of **Pep1** in methanol- d_4 .



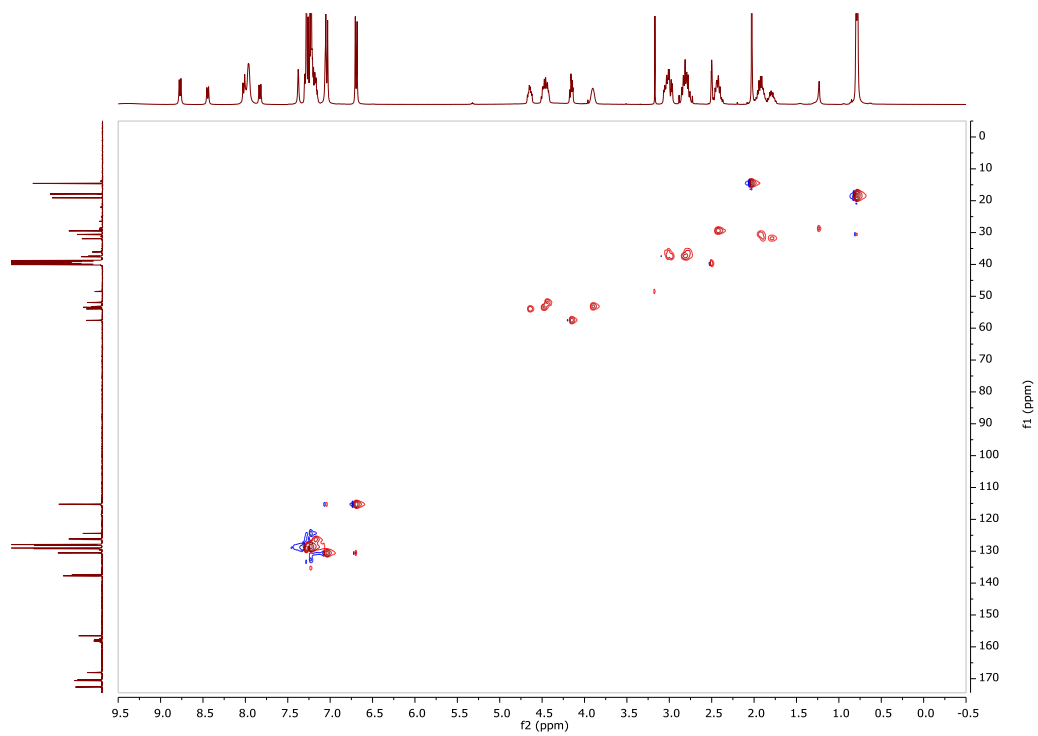
HMBC spectrum of **Pep1** in methanol- d_4 .



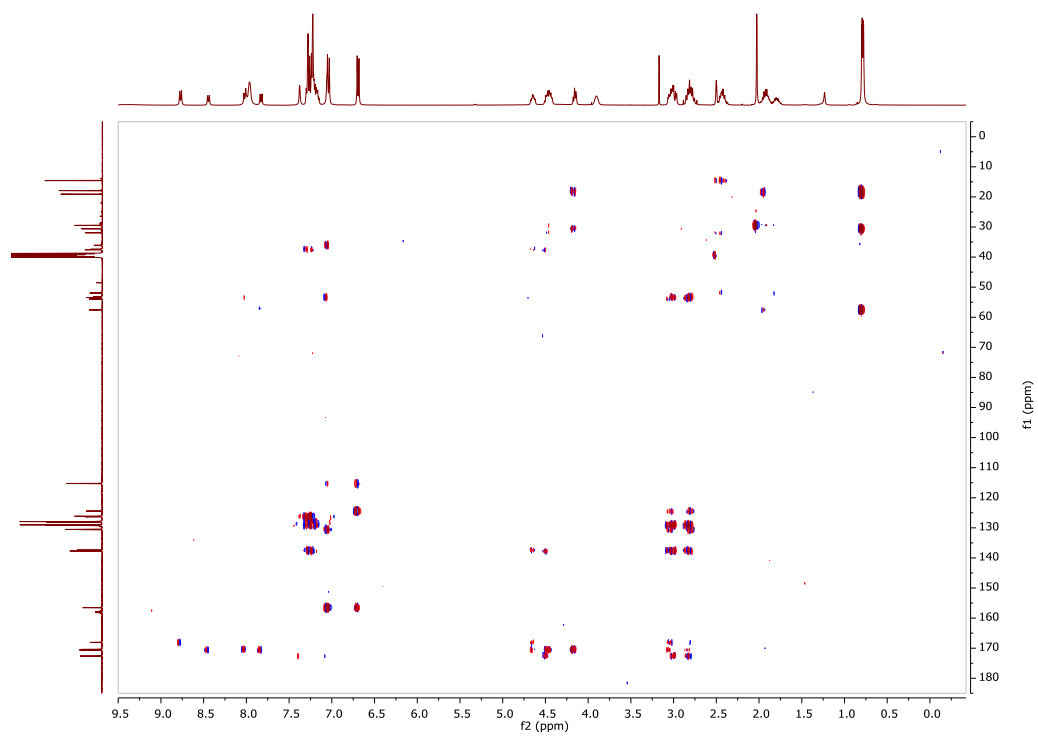
¹H-NMR spectrum of Pep1 in DMSO-*d*₆.



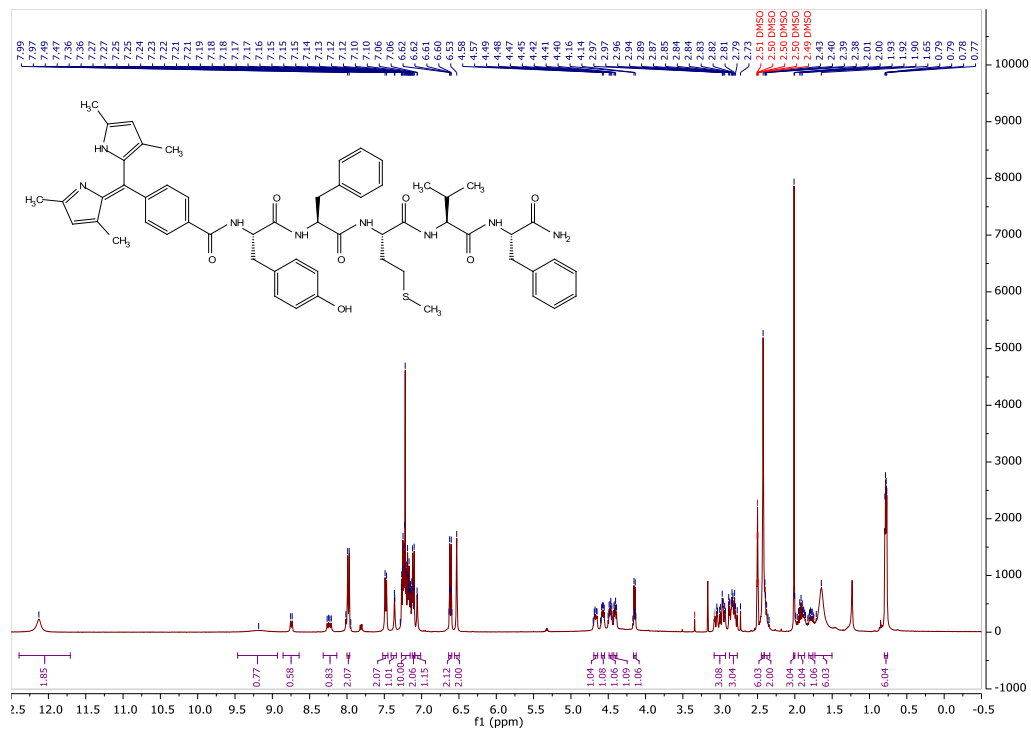
¹³C-NMR spectrum of Pep1 in DMSO-*d*₆.



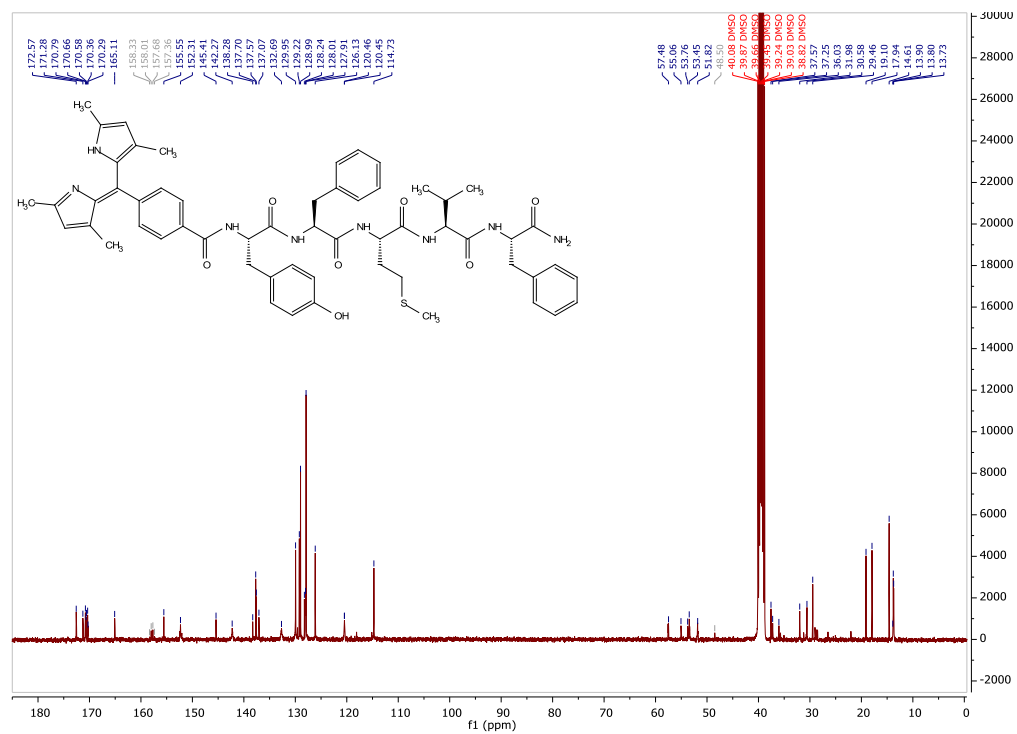
HSQC spectrum of **Pep1** in DMSO-*d*₆.



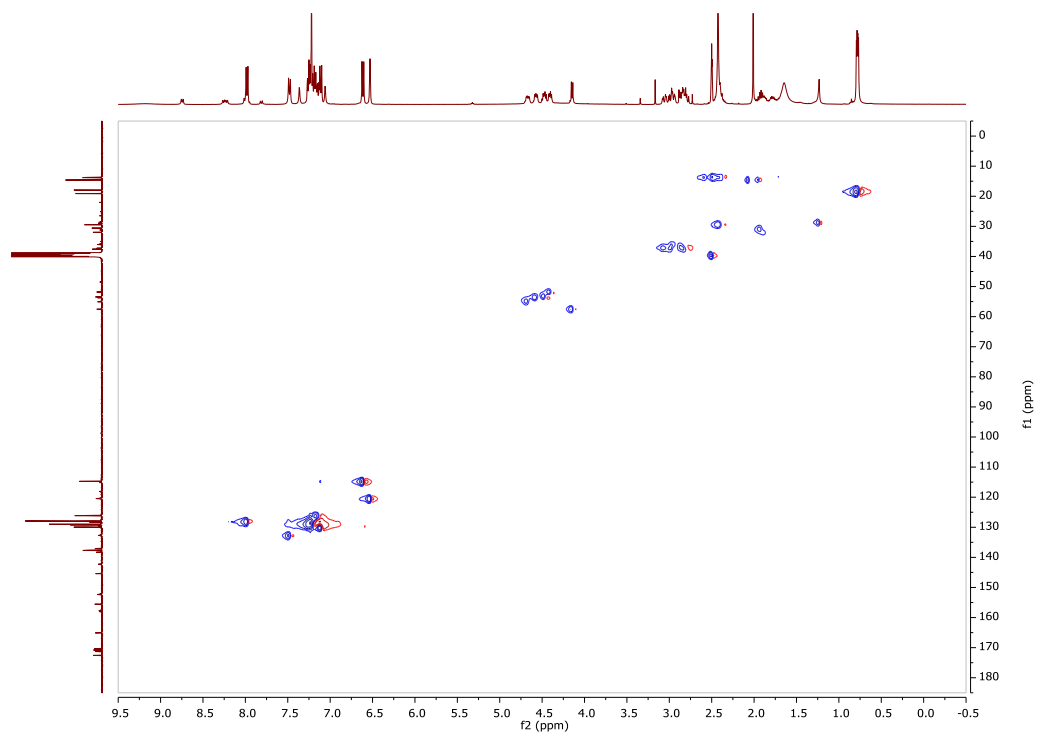
HMBC spectrum of **Pep1** in DMSO-*d*₆.



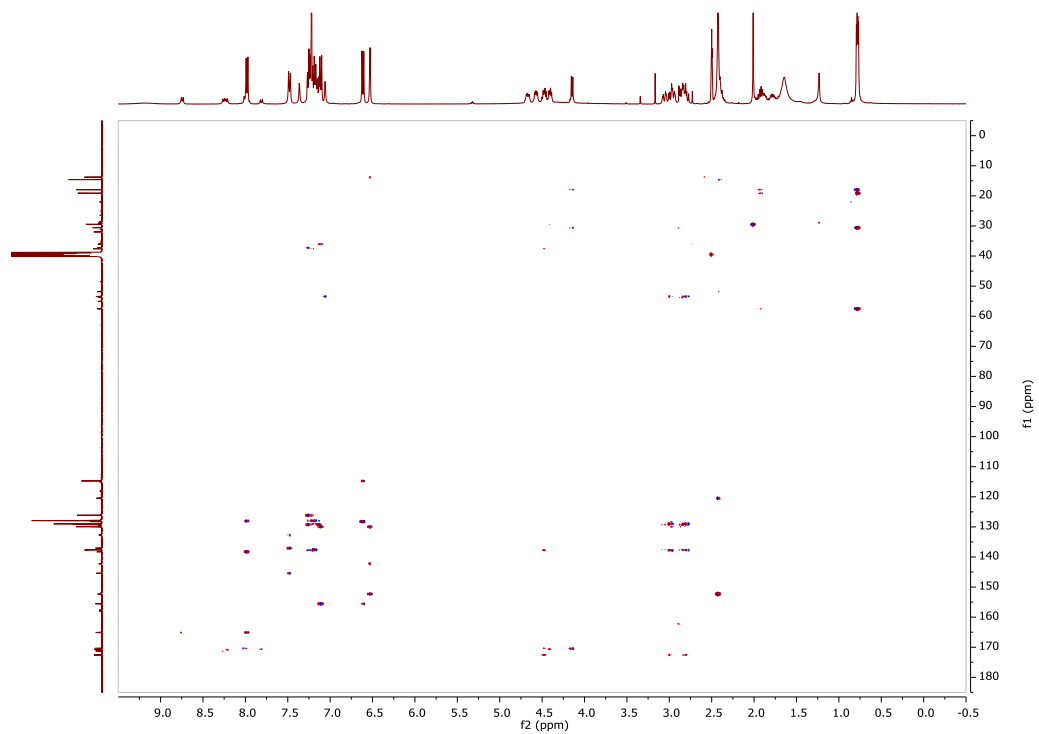
¹H-NMR spectrum of DP₁-Pep₁ in DMSO-*d*₆.



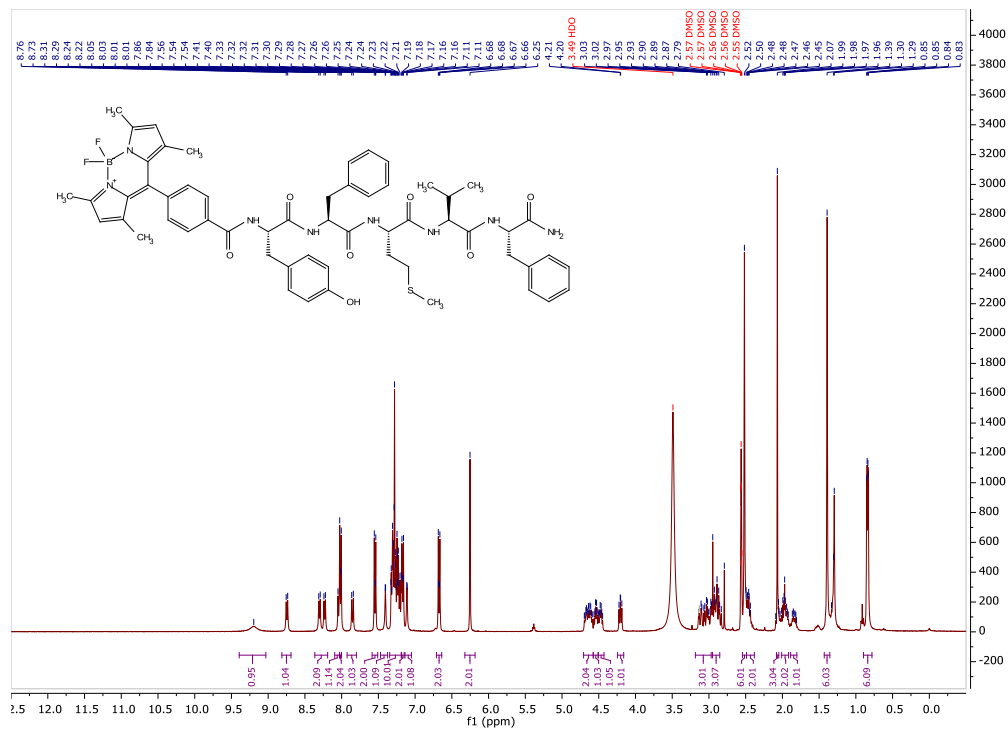
¹³C-NMR spectrum of DP₁-Pep₁ in DMSO-*d*₆.



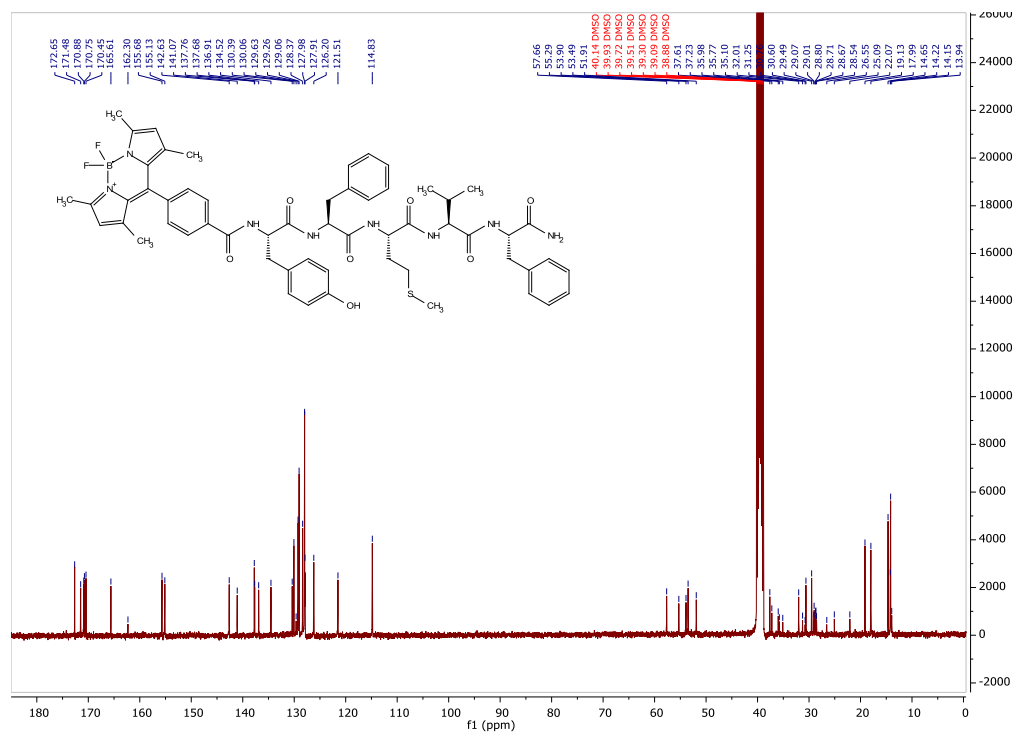
HSQC spectrum of **DP1-Pep1** in DMSO-*d*₆.



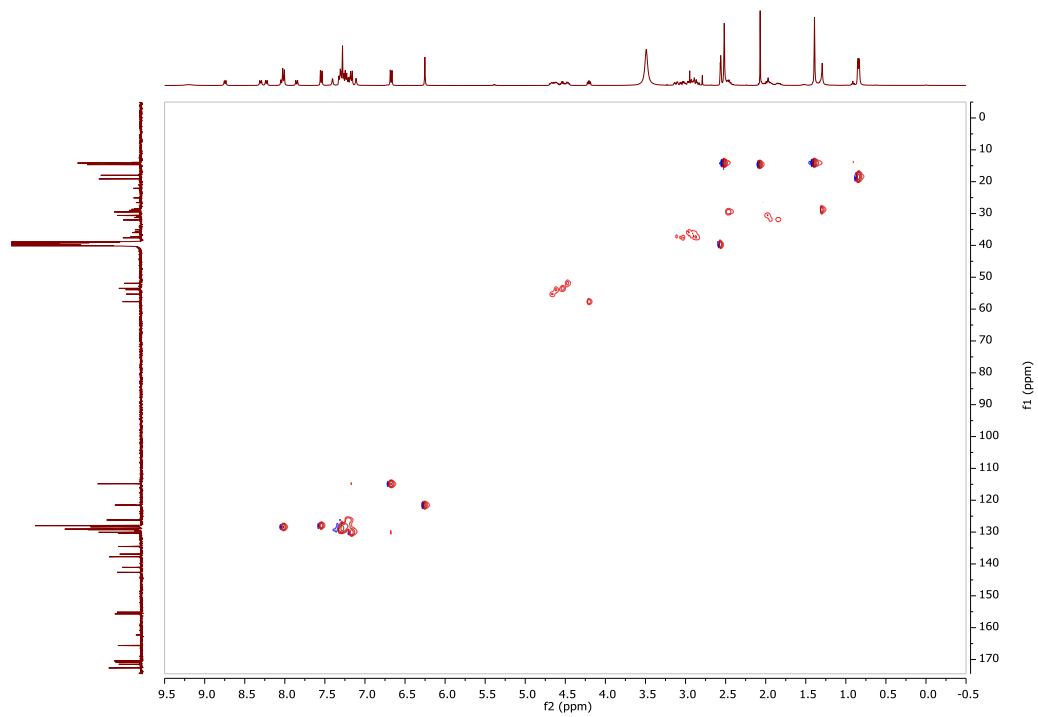
HMBC spectrum of **DP1-Pep1** in DMSO-*d*₆.



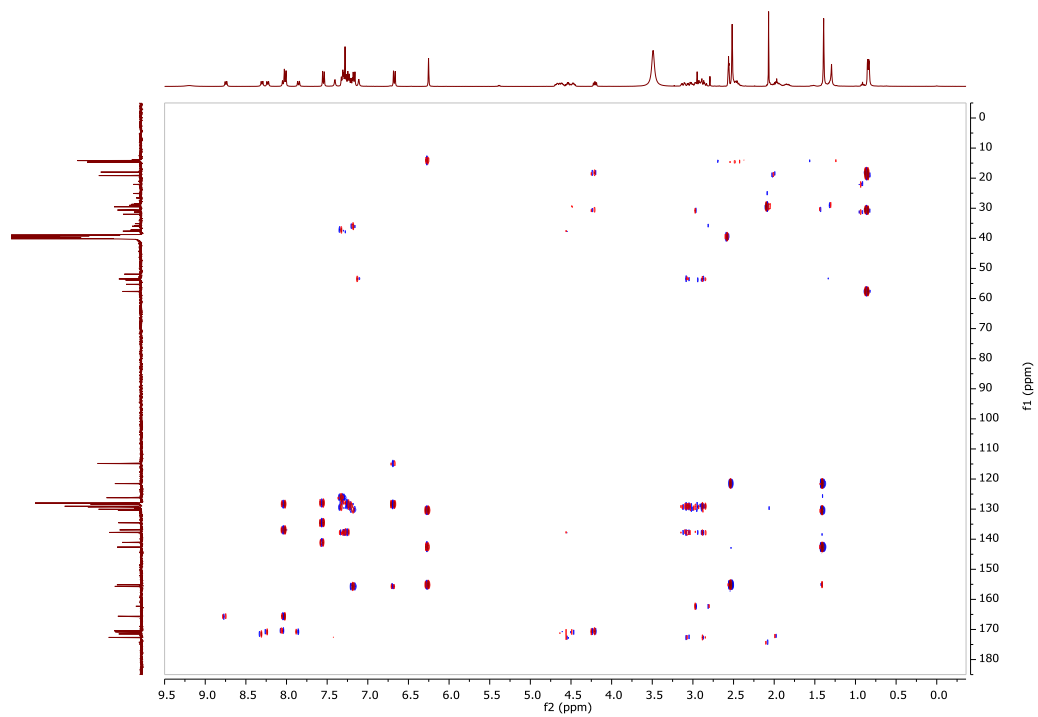
¹H-NMR spectrum of **BODIPY₁-Pep₁** in DMSO-*d*₆.



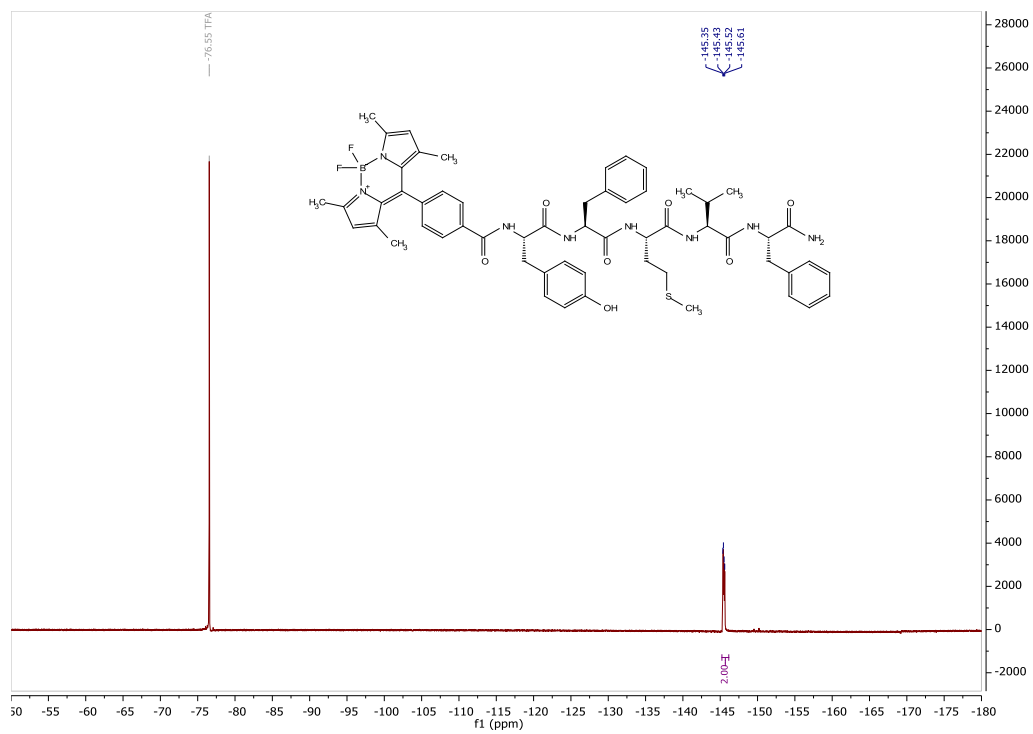
¹³C-NMR spectrum of **BODIPY₁-Pep₁** in DMSO-*d*₆.



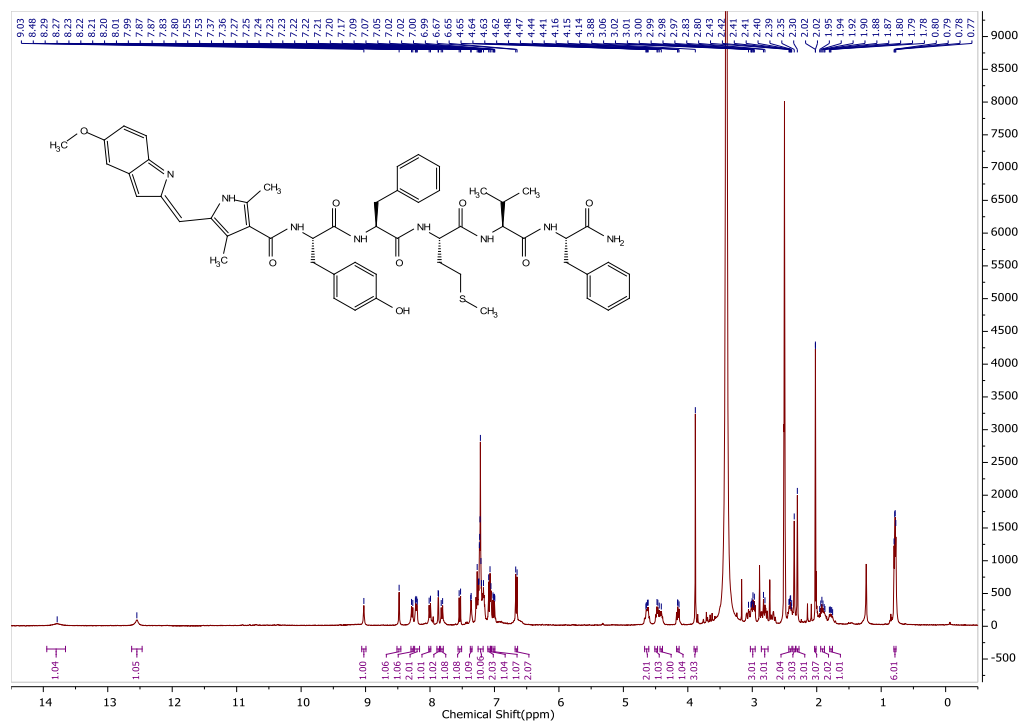
HSQC spectrum of **BODIPY₁-Pep₁** in DMSO-*d*₆.



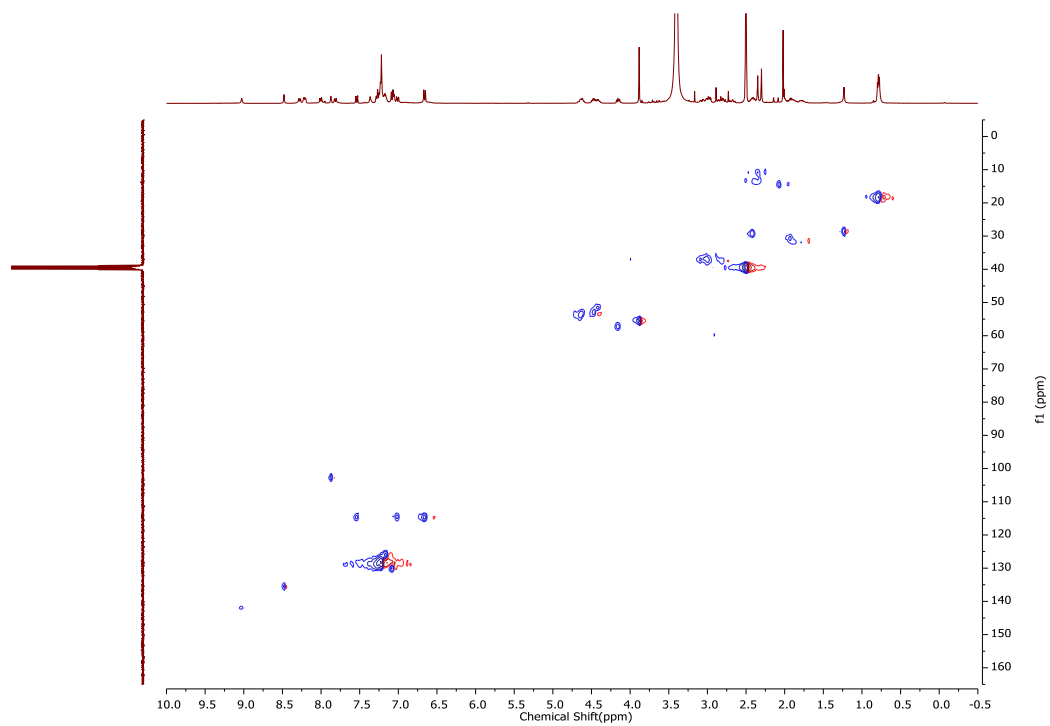
HMBC spectrum of **BODIPY₁-Pep₁** in DMSO-*d*₆.



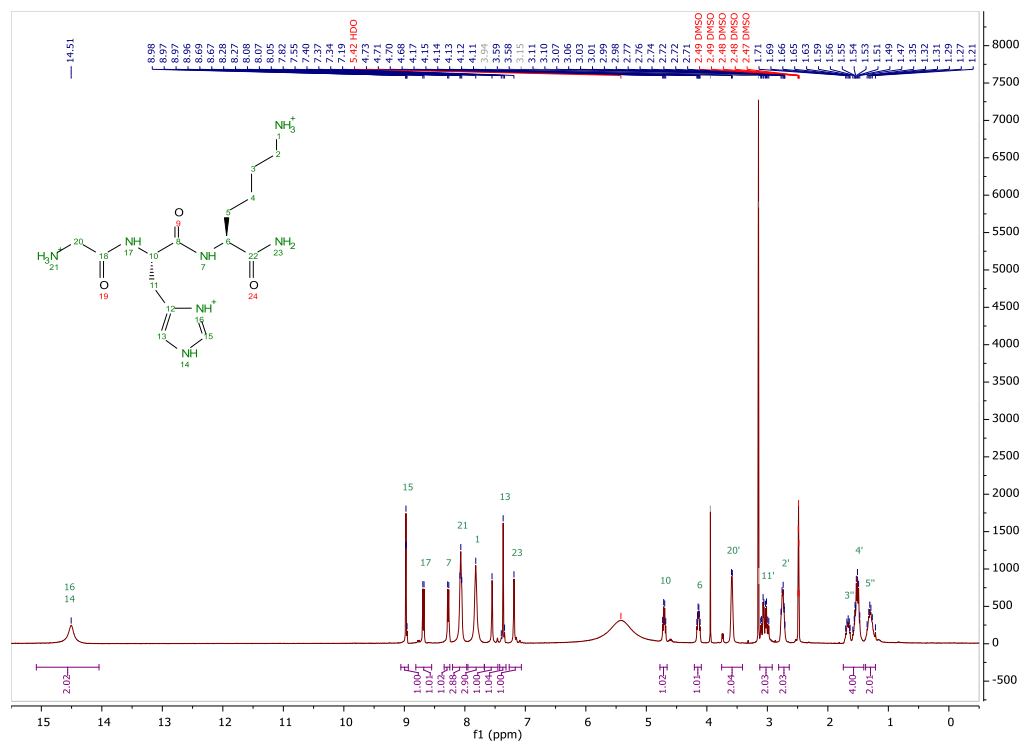
^{19}F -NMR spectrum of **BODIPY₁-Pep₁** in DMSO-*d*₆.



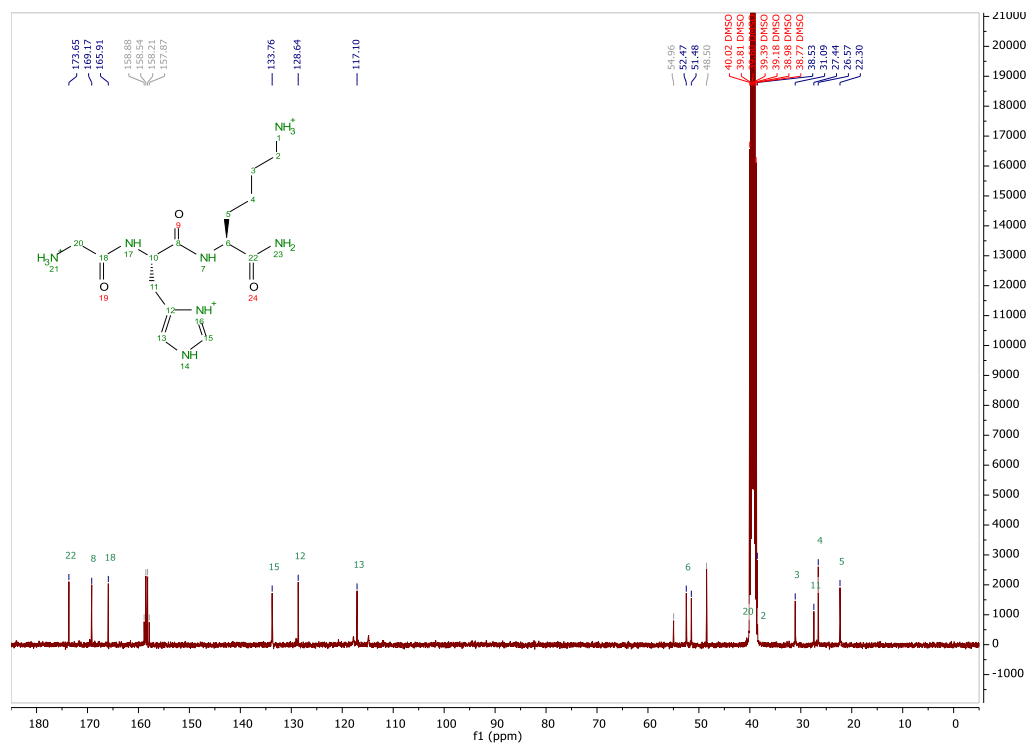
$^1\text{H-NMR}$ spectrum of DP16-Pep1 in DMSO- d_6 .



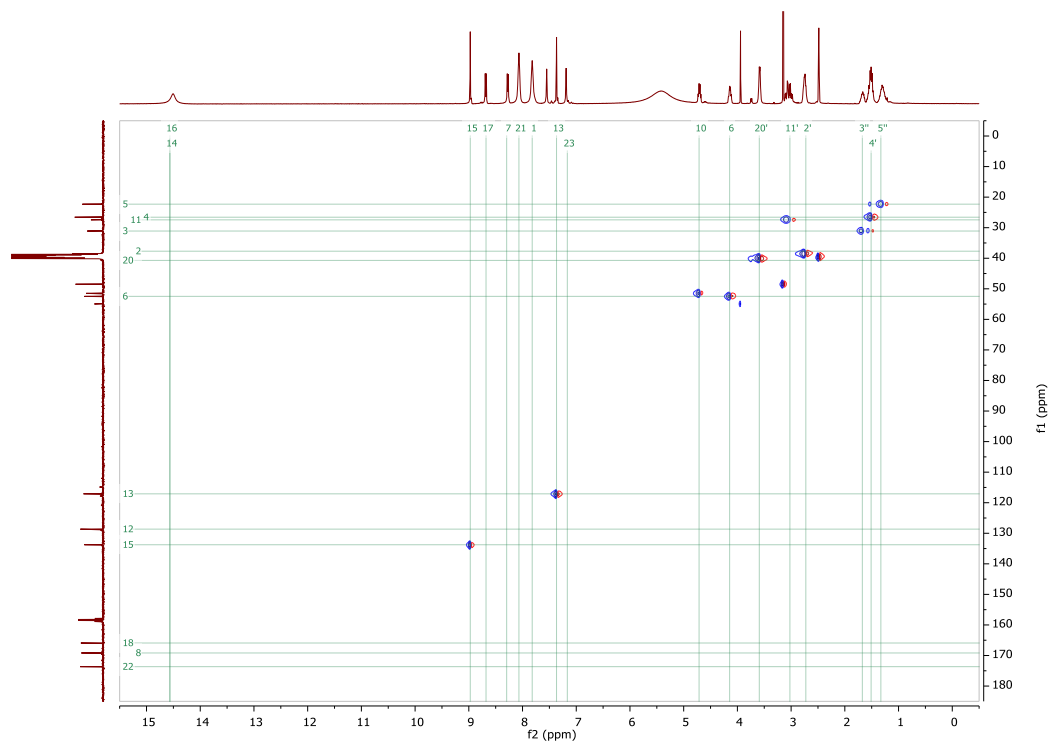
HSQC spectrum of DP16-Pep1 in DMSO- d_6 .



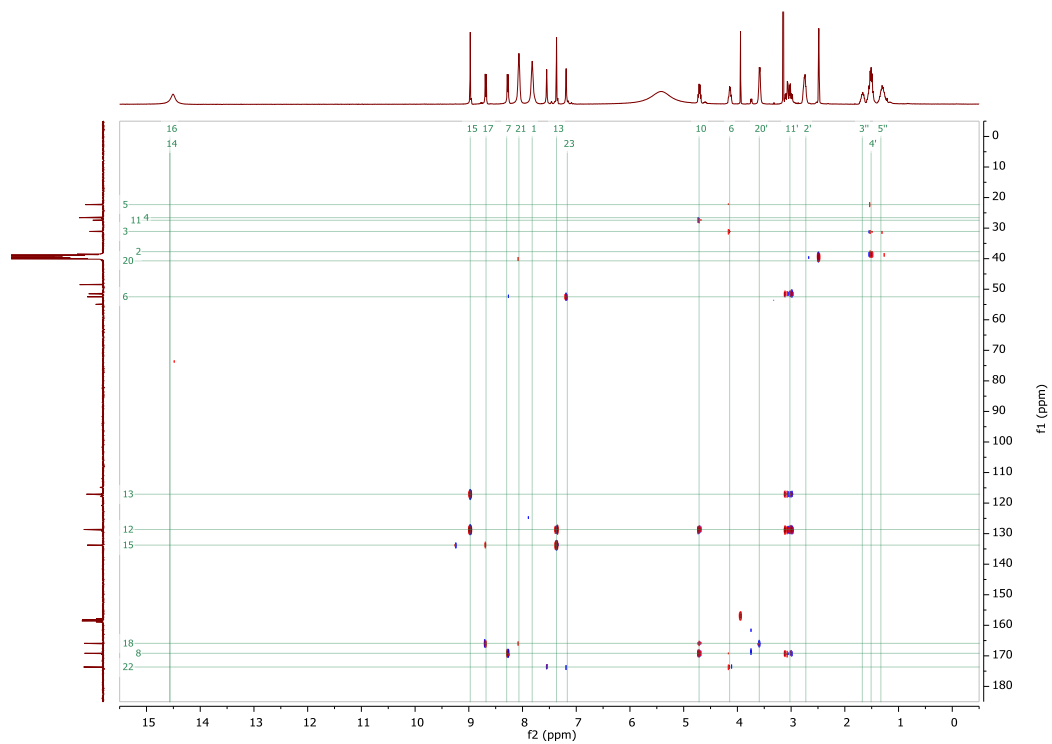
¹H-NMR spectrum of Pep₁₁ in DMSO-*d*₆.



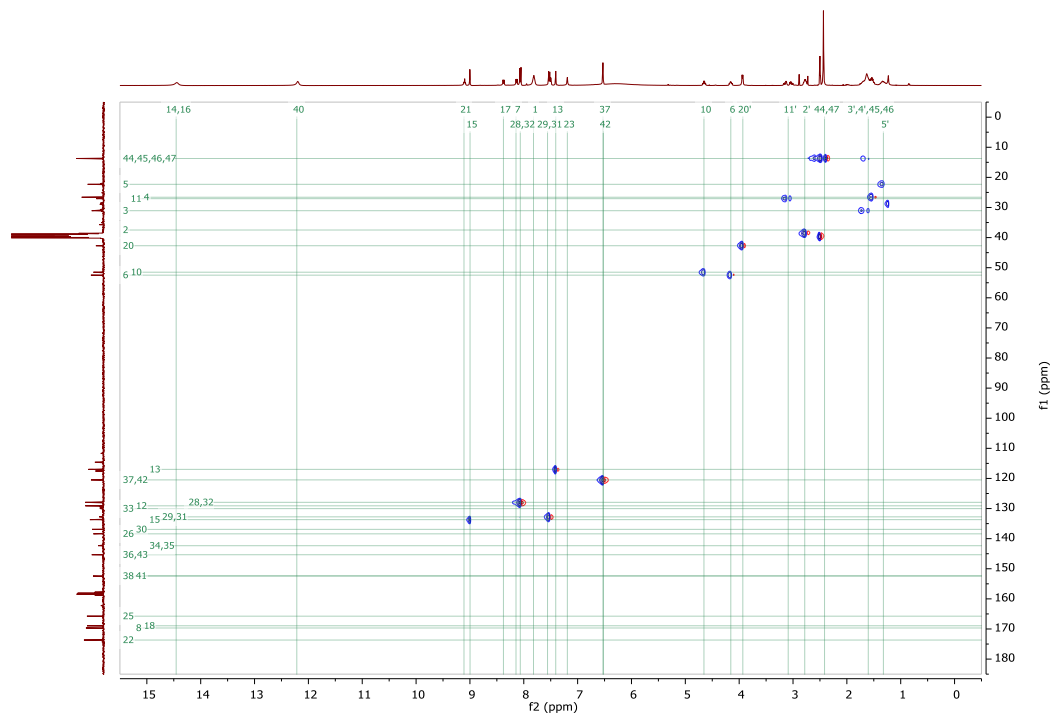
¹³C-NMR spectrum of Pep₁₁ in DMSO-*d*₆.



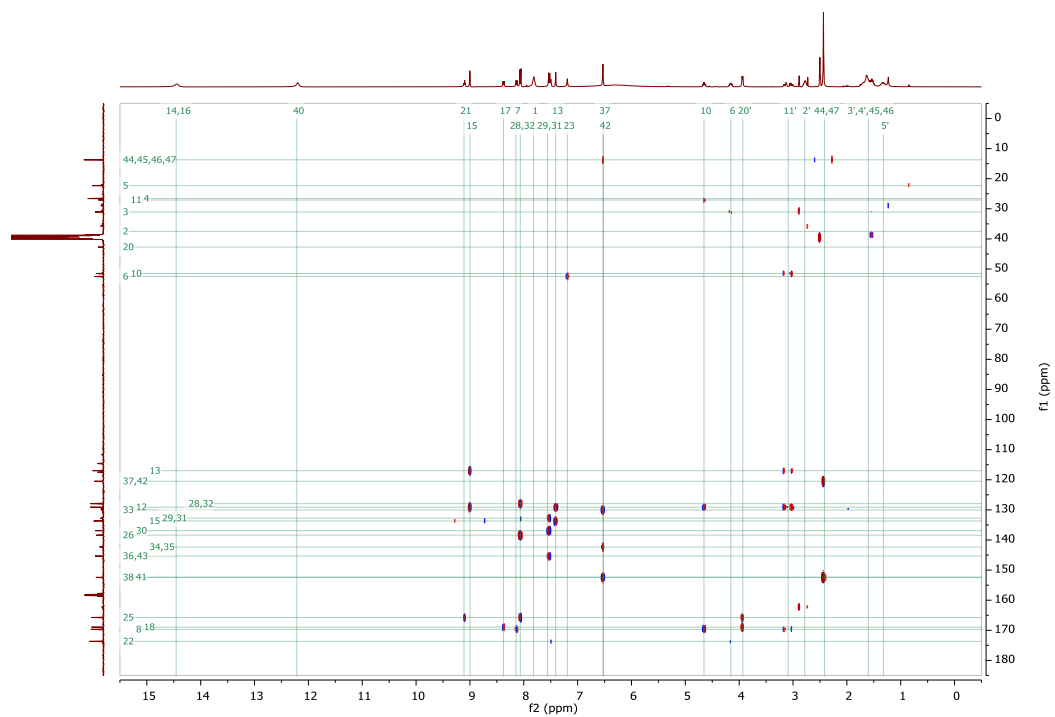
HSQC spectrum of **Pep₁₁** in DMSO-*d*₆.



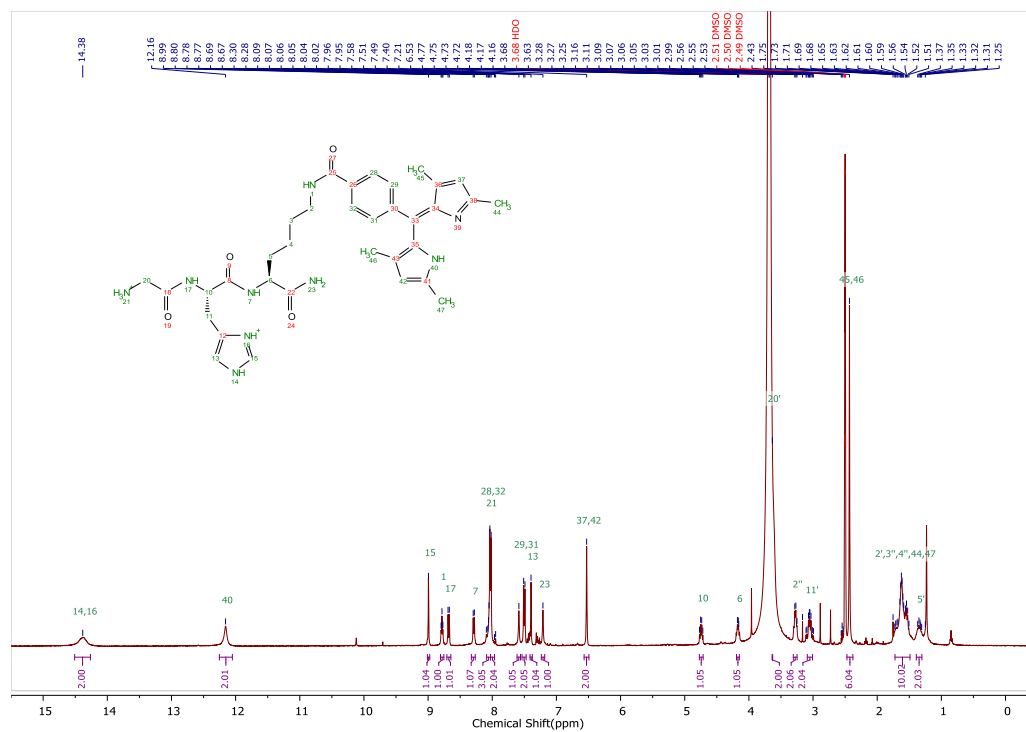
HMBC spectrum of **Pep₁₁** in DMSO-*d*₆.



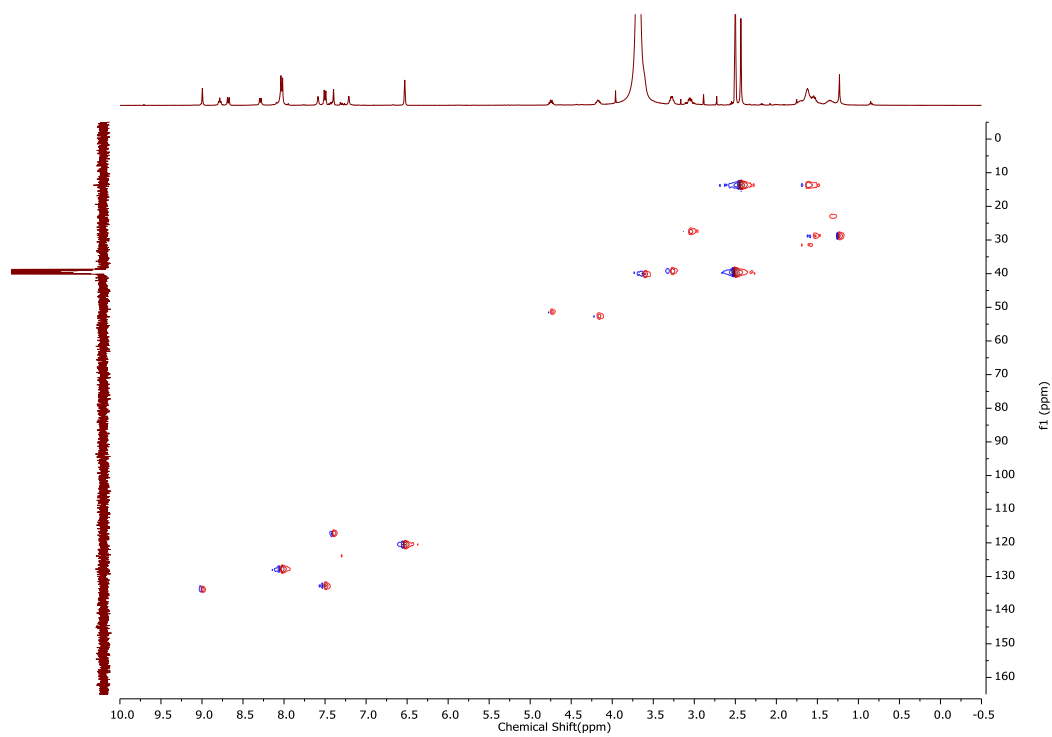
HSQC spectrum of **DP1-Pep11** in DMSO-*d*₆.



HMBC spectrum of **DP1-Pep11** in DMSO-*d*₆.



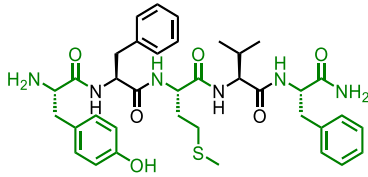
¹H-NMR spectrum of DP1-Pep11* in DMSO-*d*₆.



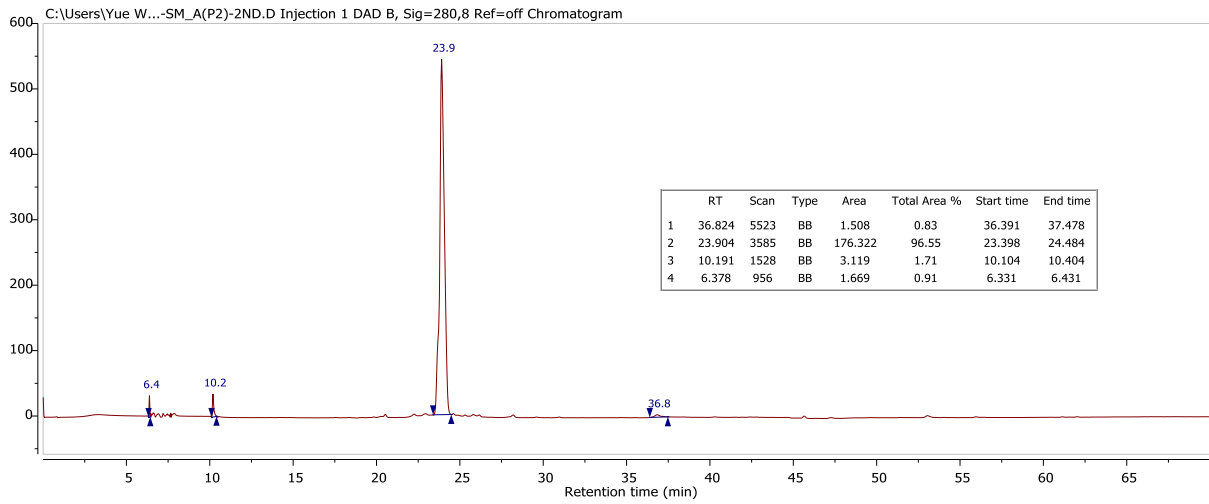
HSQC spectrum of DP1-Pep11* in DMSO-*d*₆.

HPLC chromatographs and MALDI-TOF HRMS spectra of products

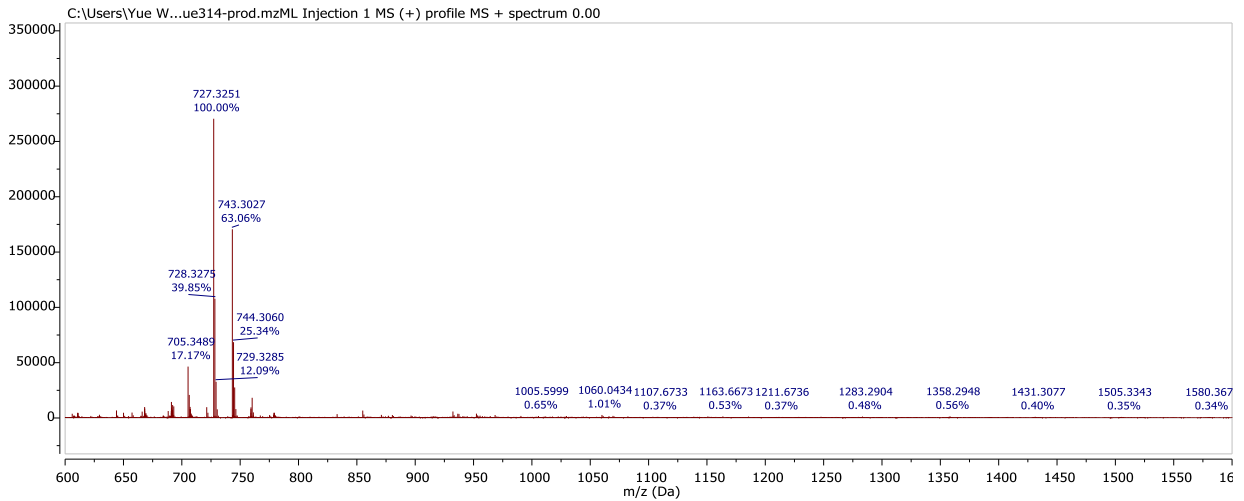
Pep₁: H₂N-YFMVF-CONH₂



Chemical Formula: C₃₇H₄₈N₆O₆S
Exact Mass: 704.3356

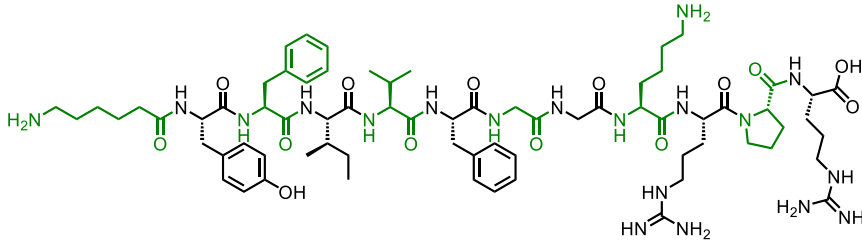


Analytic HPLC of Pep₁

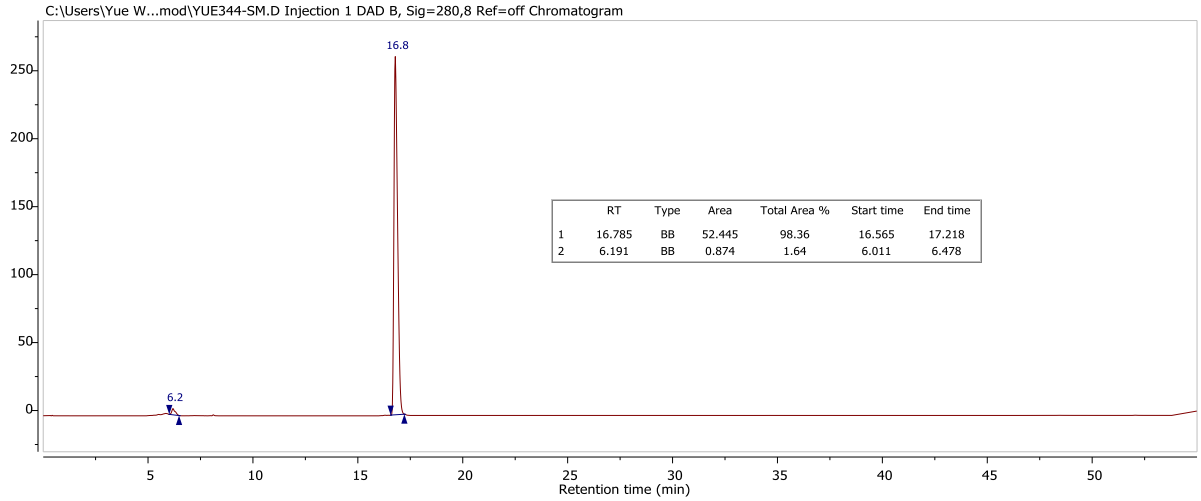


HRMS(MALDI-TOF) of Pep₁

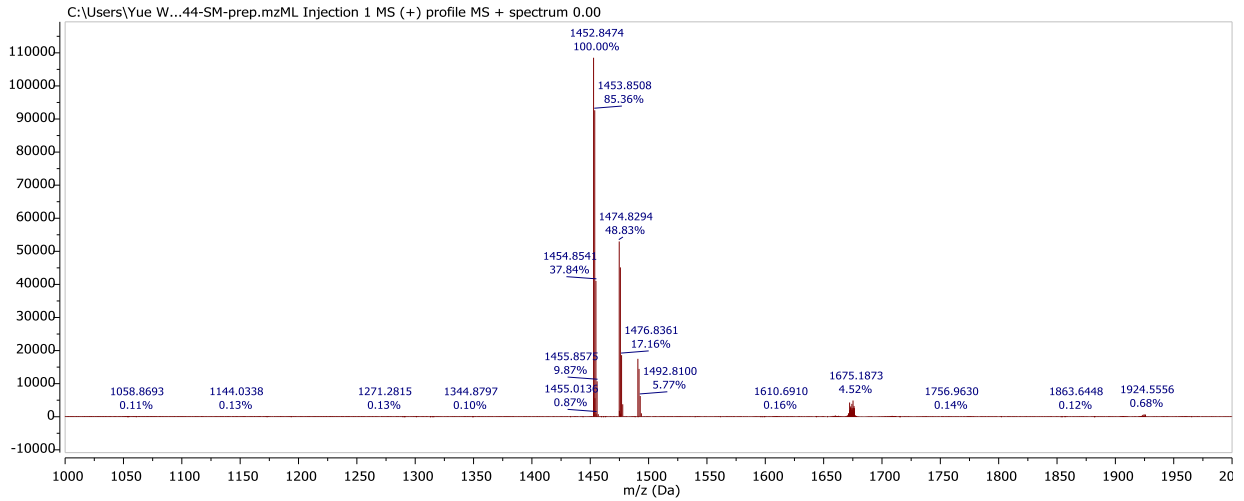
Pep₂: H₂N-Ahx-YFIVFGGKRPR-COOH



Chemical Formula: C₇₁H₁₀₉N₁₉O₁₄
Exact Mass: 1451.8401

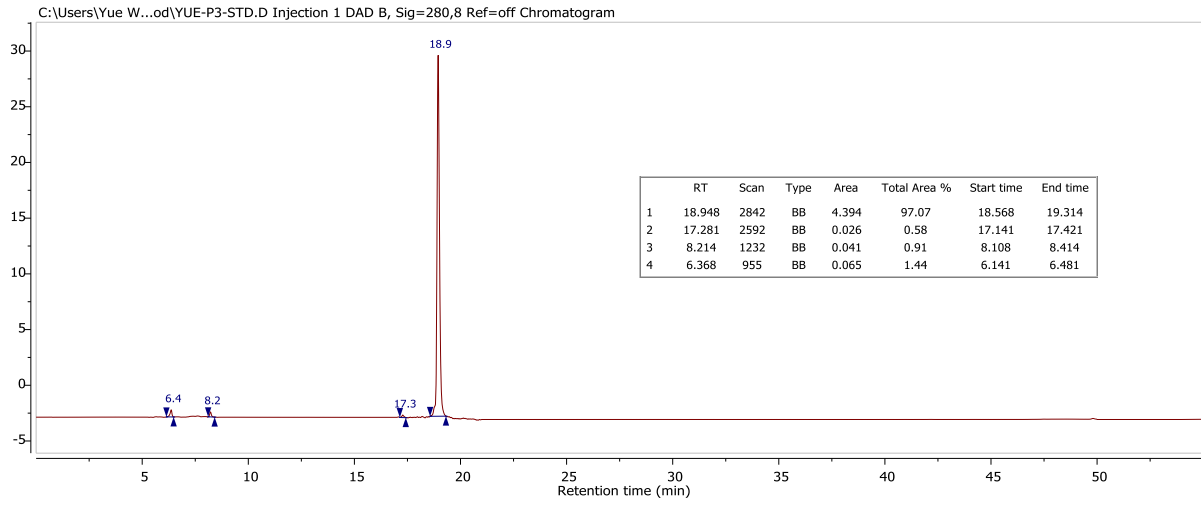
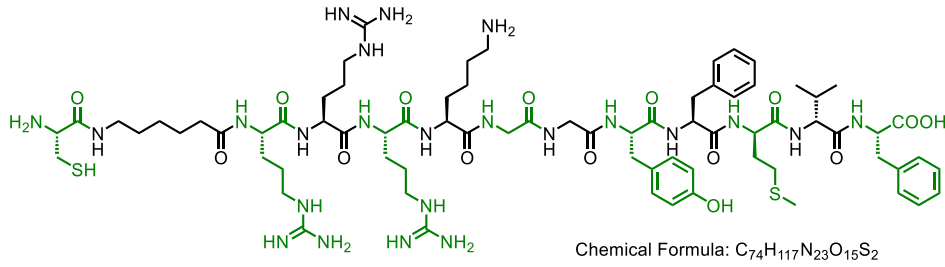


Analytic HPLC of Pep₂

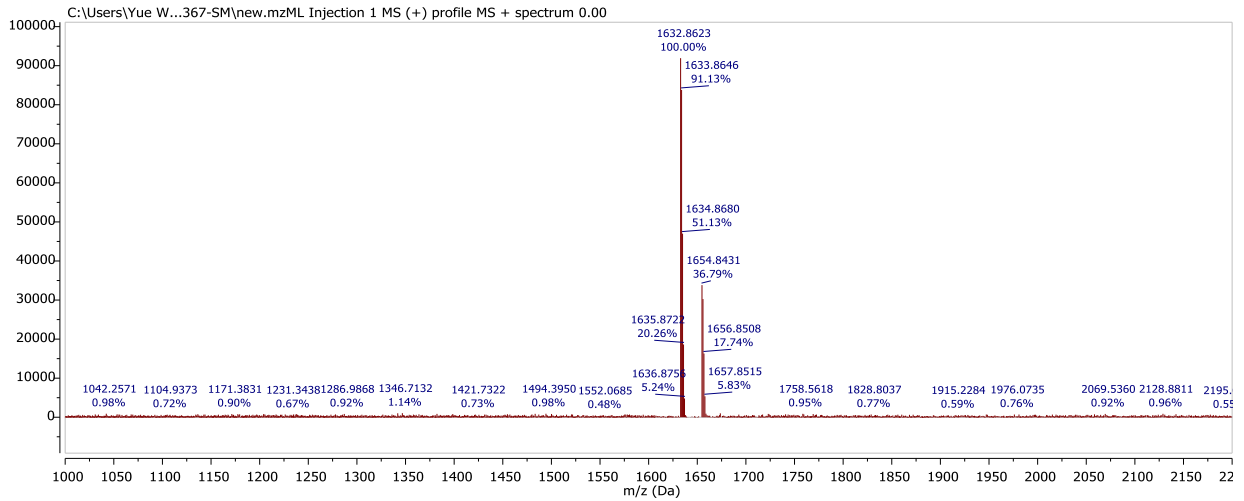


HRMS(MAIDL-TOF) of Pep₂

Pep₃: H₂N-C-Ahx- RrRK-GG-YFMVF-COOH

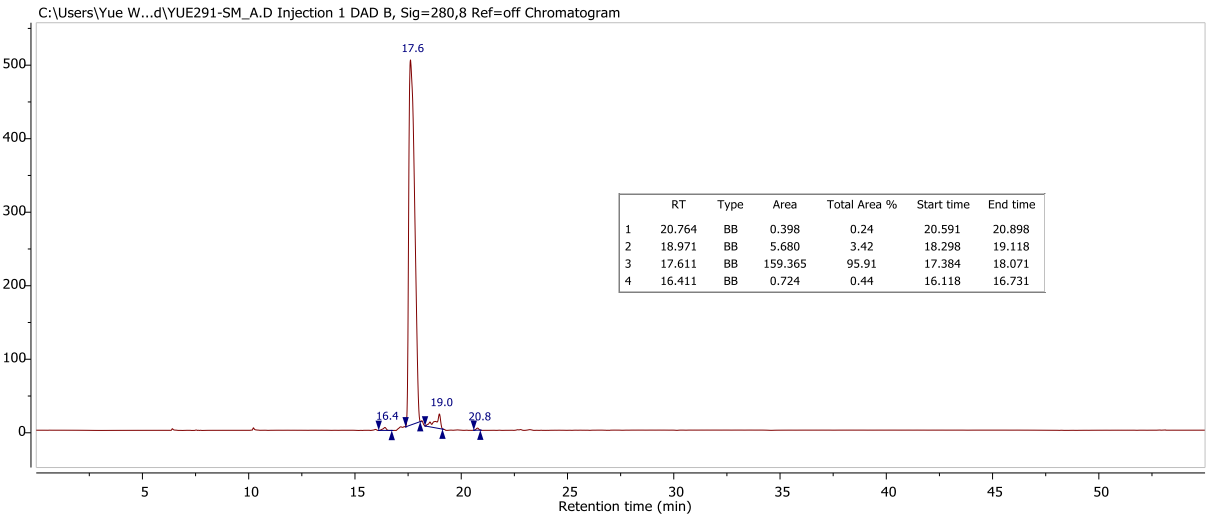
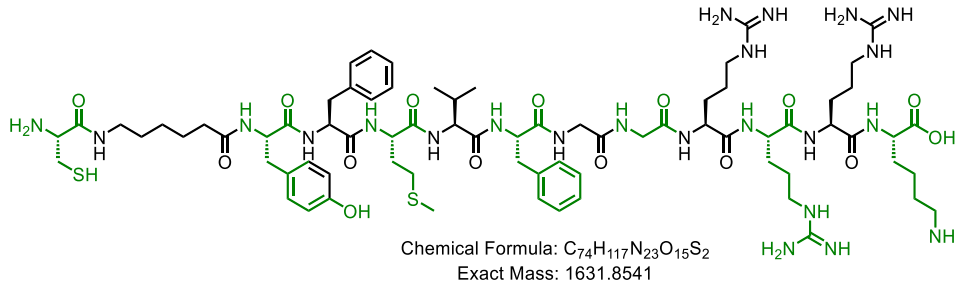


Analytic HPLC of Pep₃

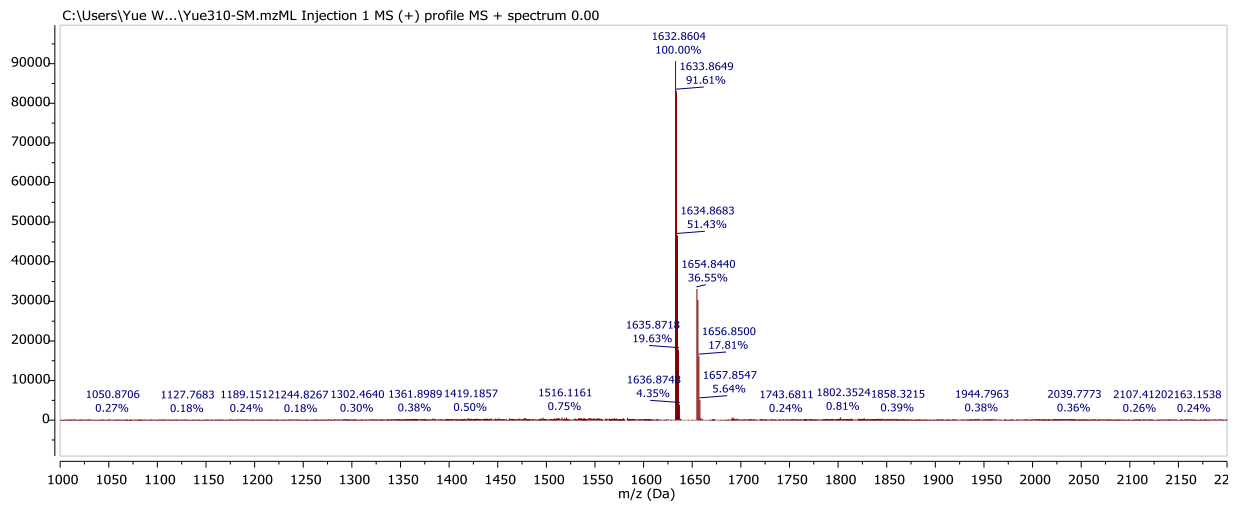


HRMS(MAIDL-TOF) of Pep₃

Pep₄: H₂N-C-Ahx-YFMVF-GG-RrRK-COOH

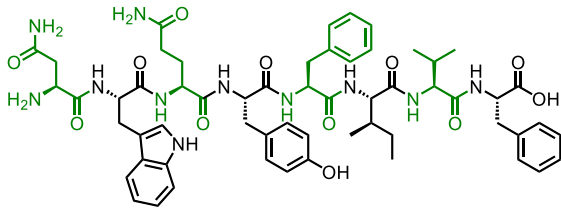


Analytic HPLC of Pep₄

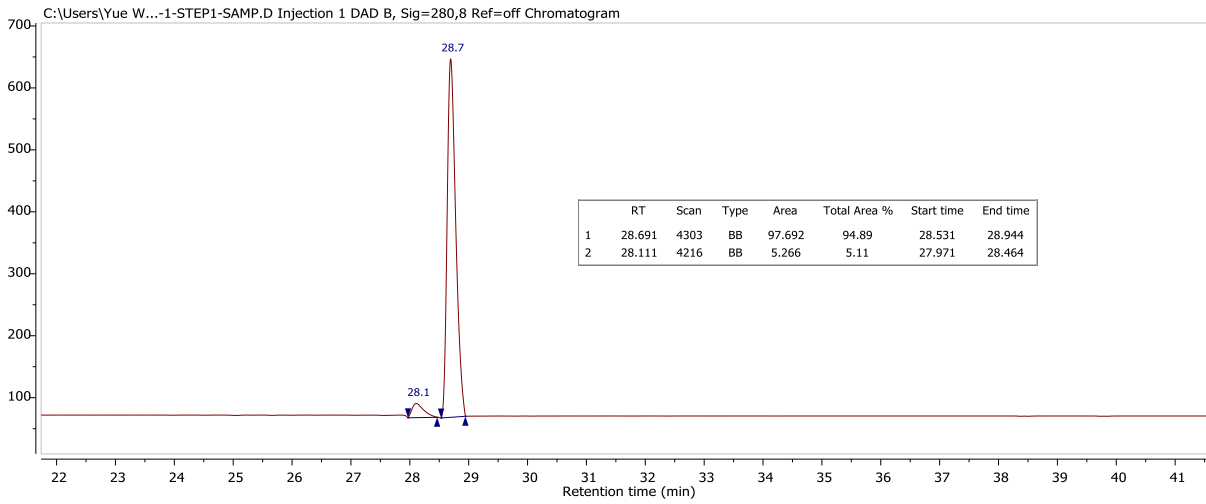


HRMS(MAIDL-TOF) of Pep₄

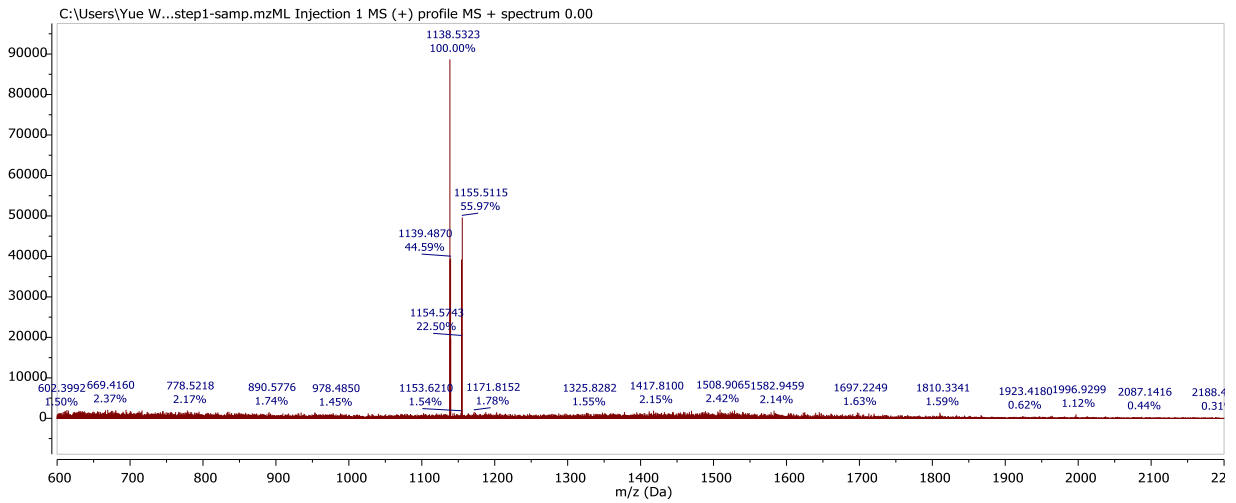
Pep₅: H₂N-NWQYFIVE-COOH



Chemical Formula: C₅₈H₇₃N₁₁O₁₂
Exact Mass: 1115.5440

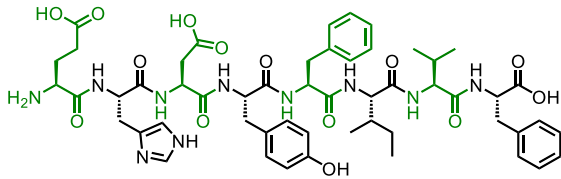


Analytic HPLC of Pep₅

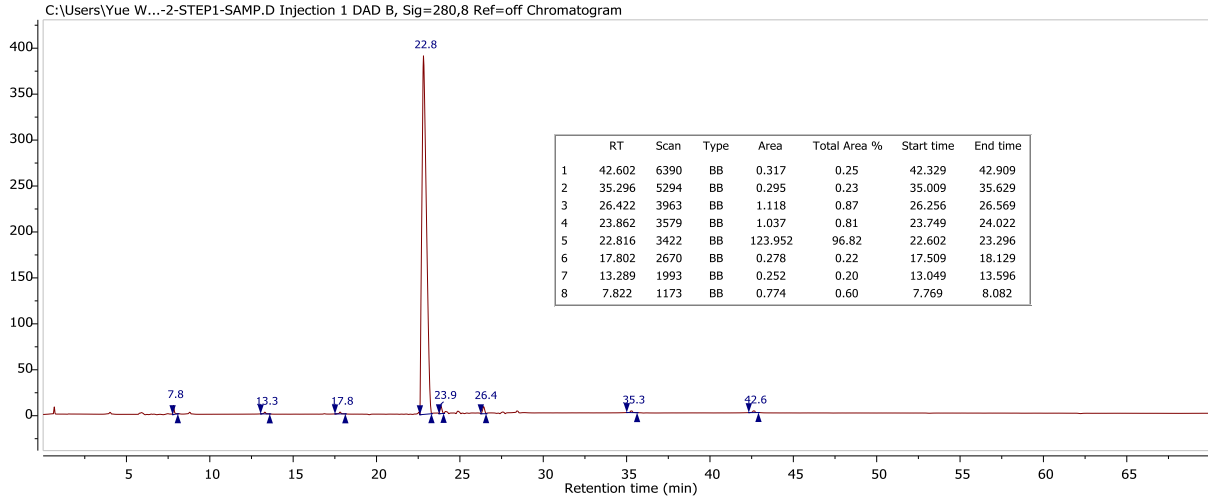


HRMS(MAIDL-TOF) of Pep₅

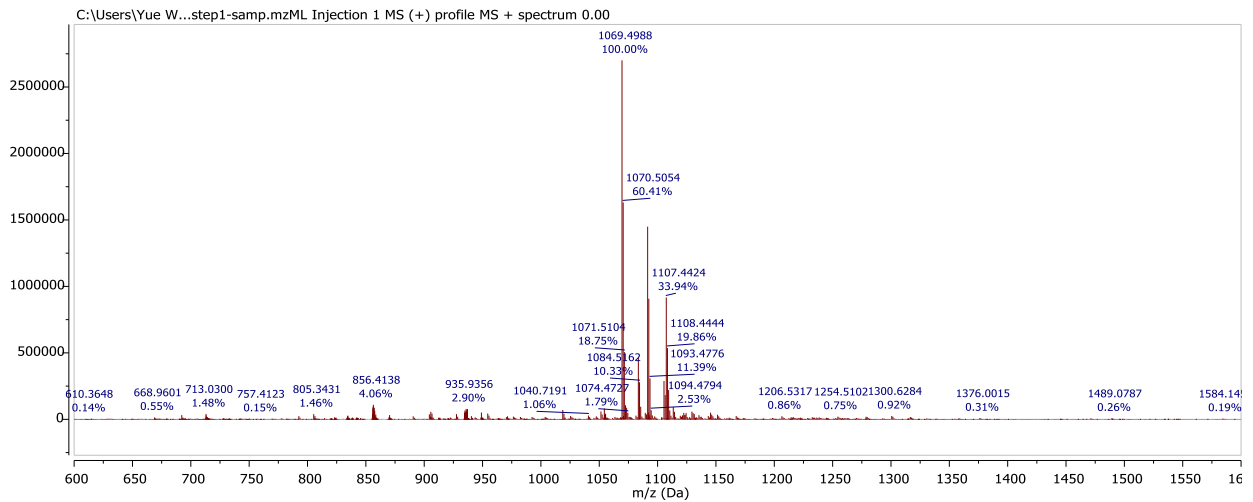
Pep₆: H₂N-EHDYFIVF-COOH



Chemical Formula: C₅₃H₆₈N₁₀O₁₄
Exact Mass: 1068.4916

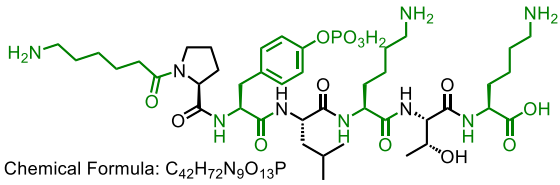


Analytic HPLC of Pep₆

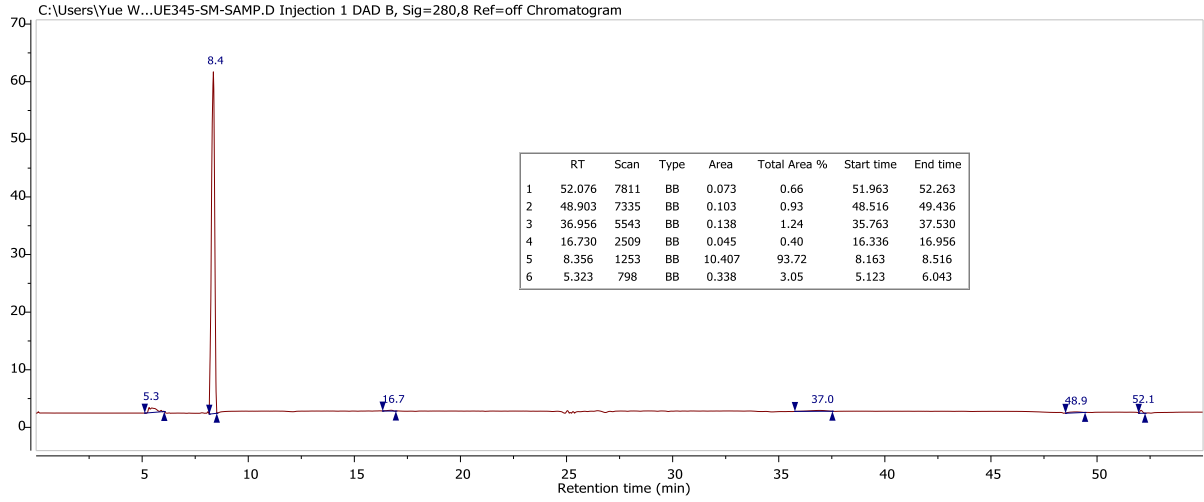


HRMS(MAIDL-TOF) of Pep₆

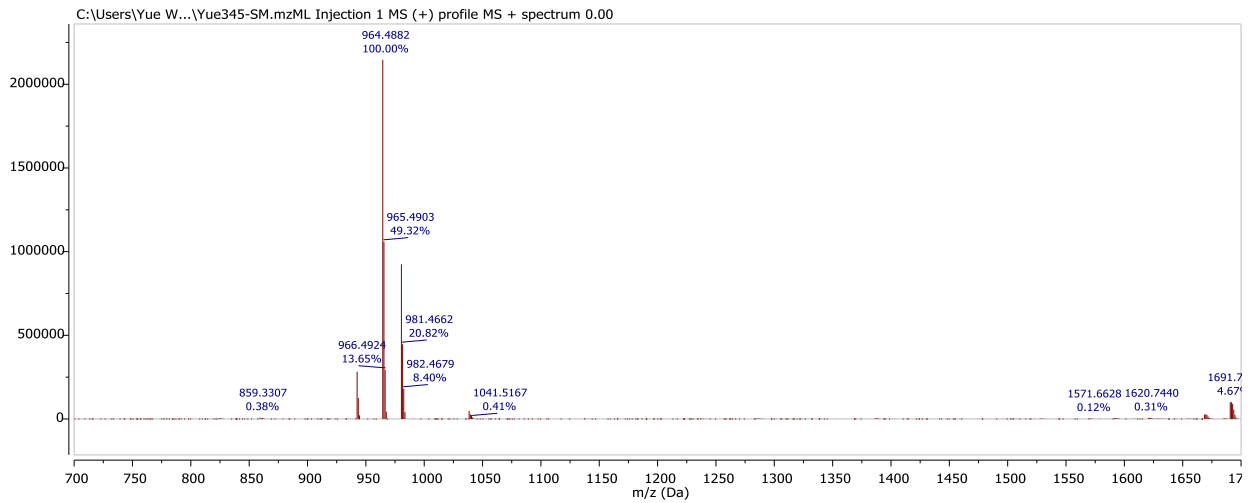
Pep7: H₂N-Ahx-P-(pTyr)-LKTK-COOH



Chemical Formula: C₄₂H₇₂N₉O₁₃P
Exact Mass: 941.4987

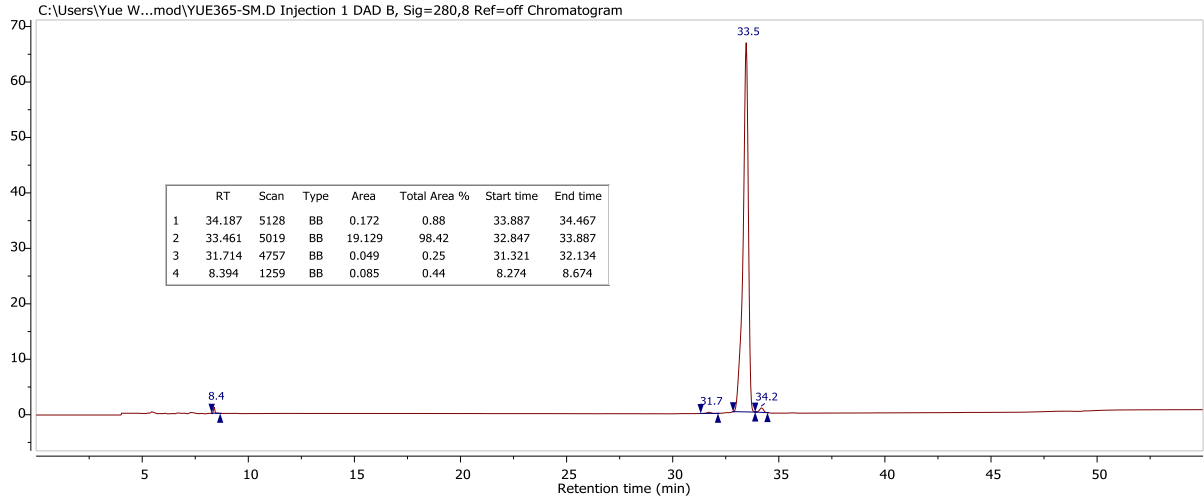
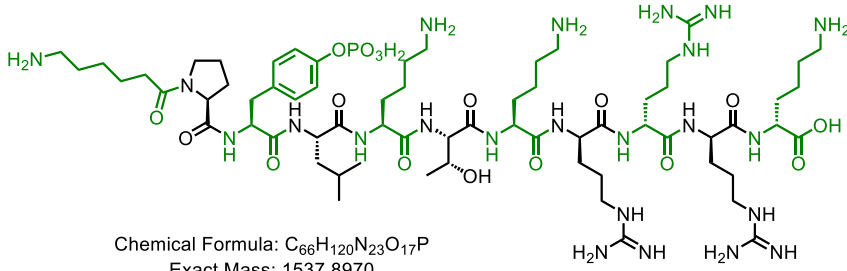


Analytic HPLC of **Pep7**

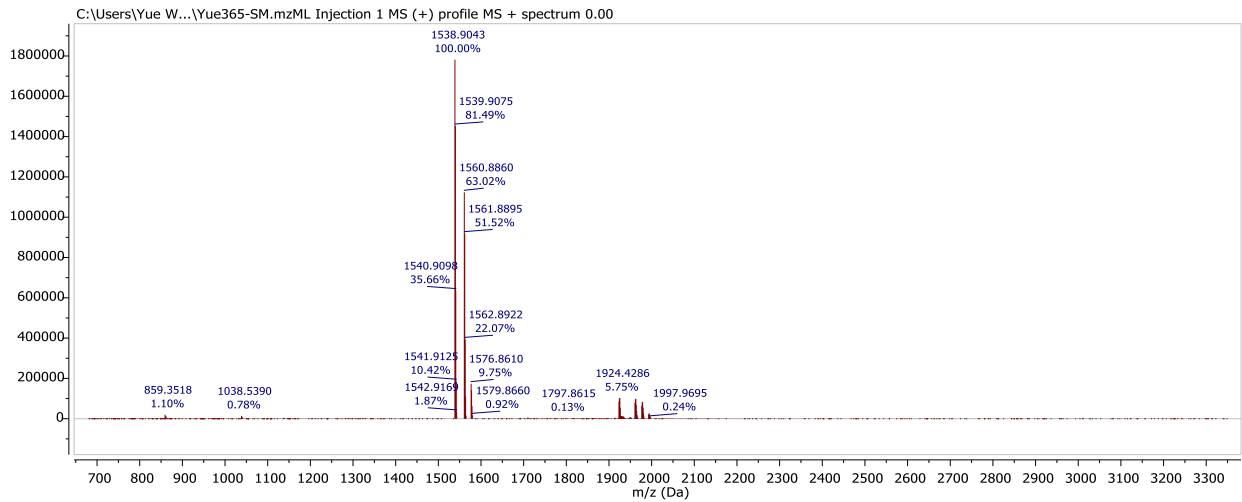


HRMS(MAIDL-TOF) of **Pep7**

Pep₈: H₂N-Ahx-P-(pTyr)-LKTKRrRK-COOH

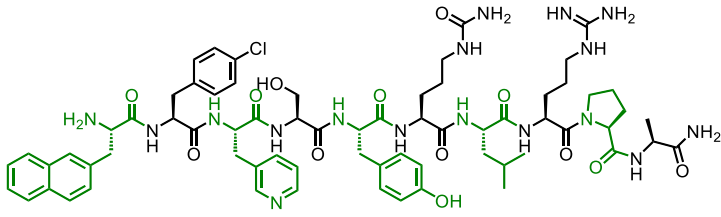


Analytic HPLC of Pep₈

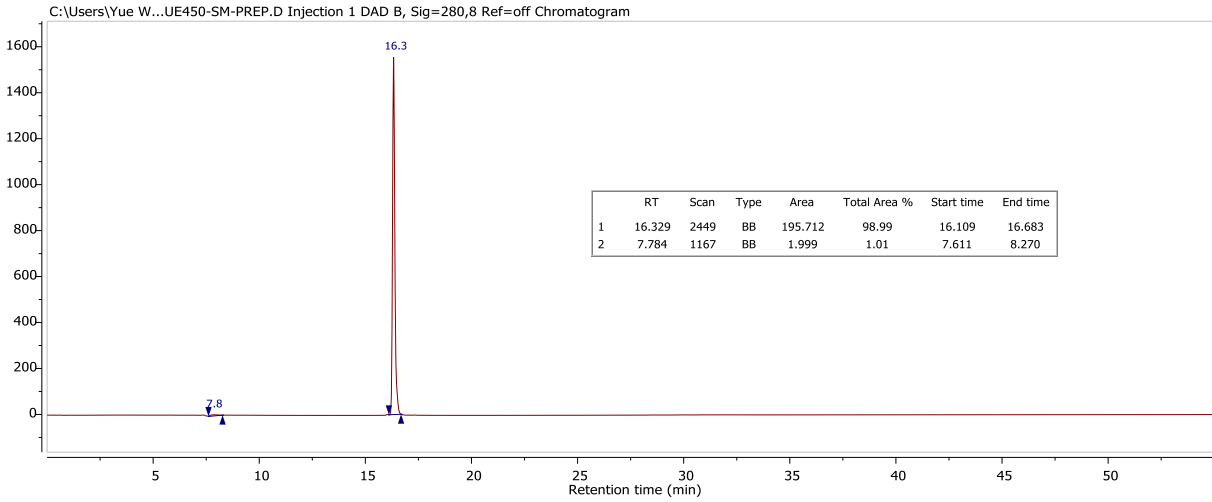


HRMS(MAIDL-TOF) of Pep₈

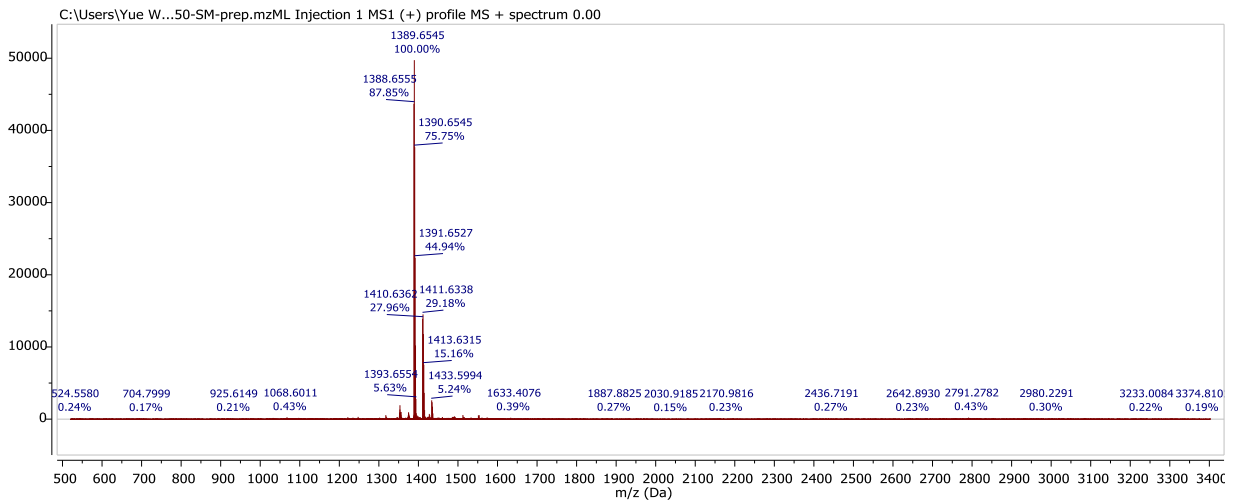
Pep₉: H₂N-Nal-Cpa-Pal-SY-Cit-LRPG-CONH₂



Chemical Formula: C₆₆H₉₀ClN₁₇O₁₃
Exact Mass: 1387.6593

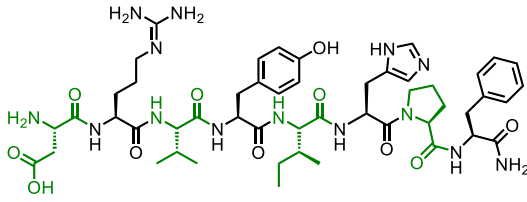


Analytic HPLC of **Pep₉**

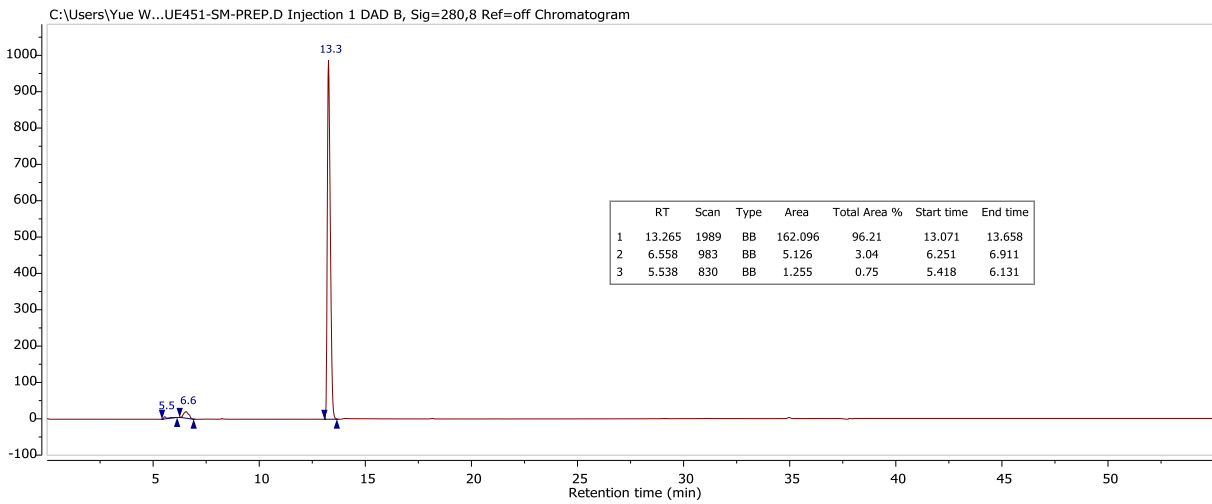


HRMS(MAIDL-TOF) of **Pep₉**

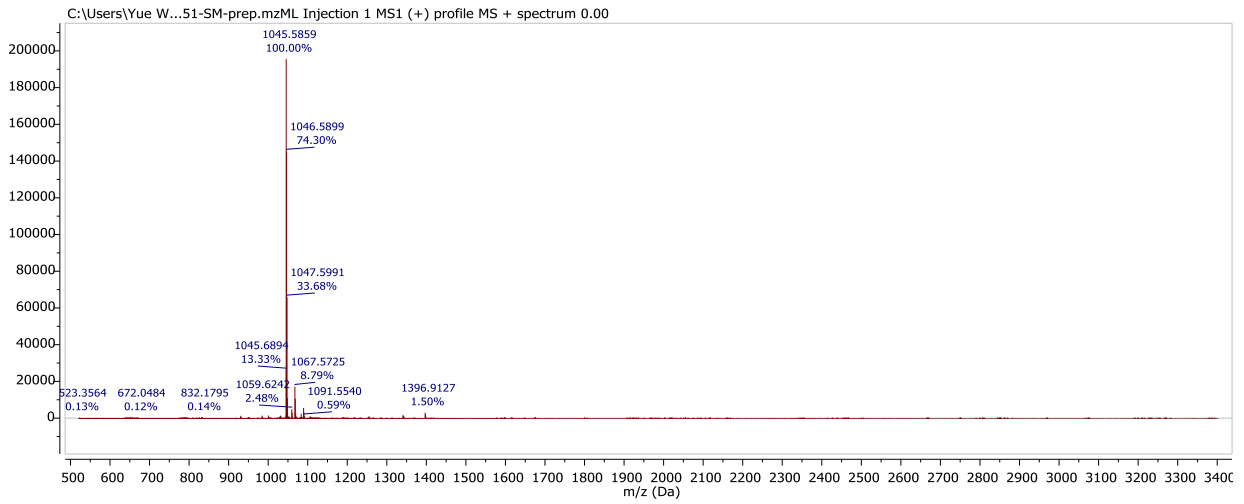
Pep₁₀: H₂N-DRVYIHPF-CONH₂



Chemical Formula: C₅₀H₇₂N₁₄O₁₁
 Exact Mass: 1044.5505

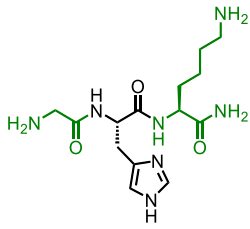


Analytic HPLC of Pep₁₀

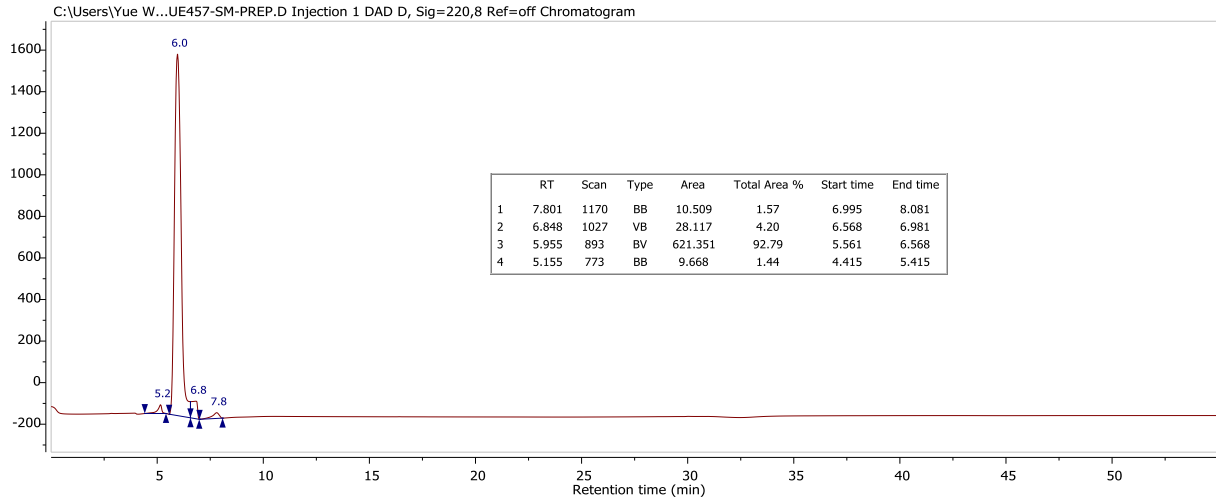


HRMS(MAIDL-TOF) of Pep₁₀

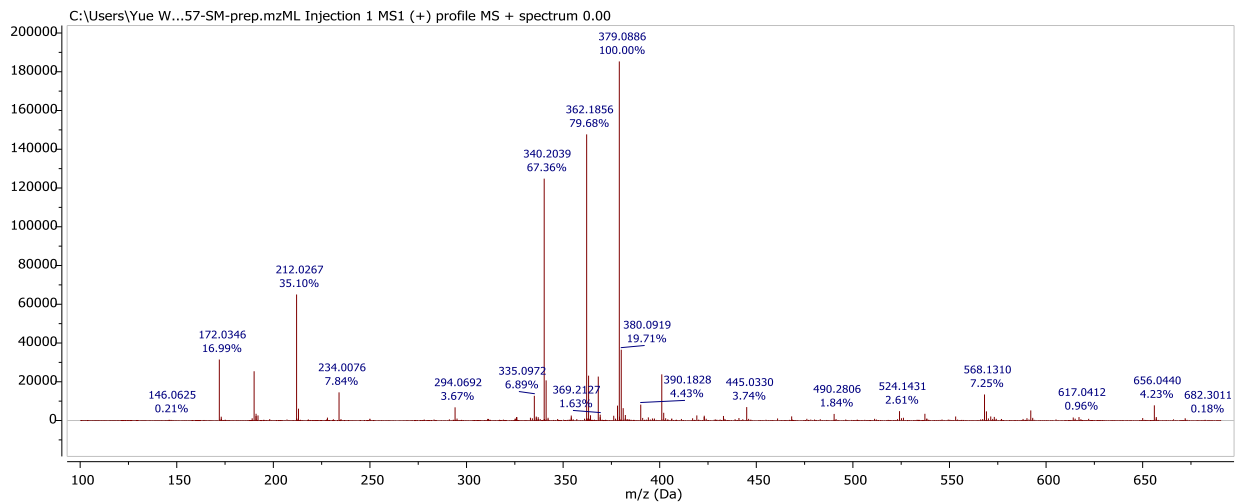
Pep₁₁: H₂N-GHL-CONH₂



Chemical Formula: C₁₄H₂₅N₇O₃
Exact Mass: 339.2019

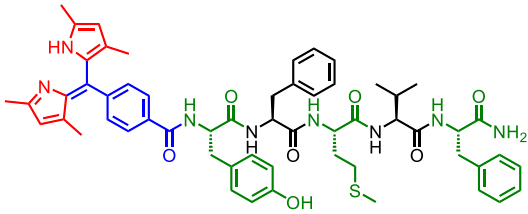


Analytic HPLC of Pep₁₁

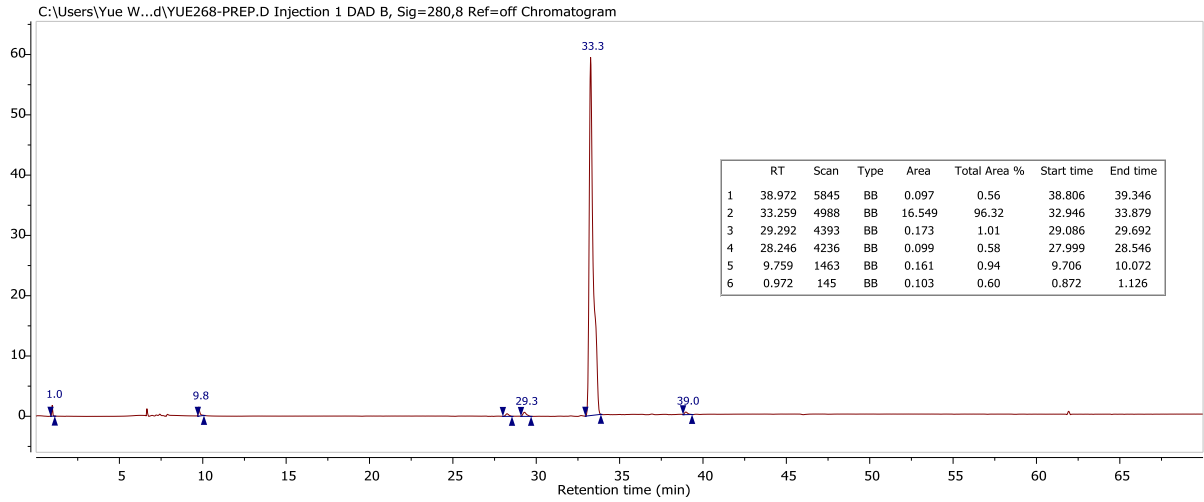


HRMS(MAIDL-TOF) of Pep₁₁

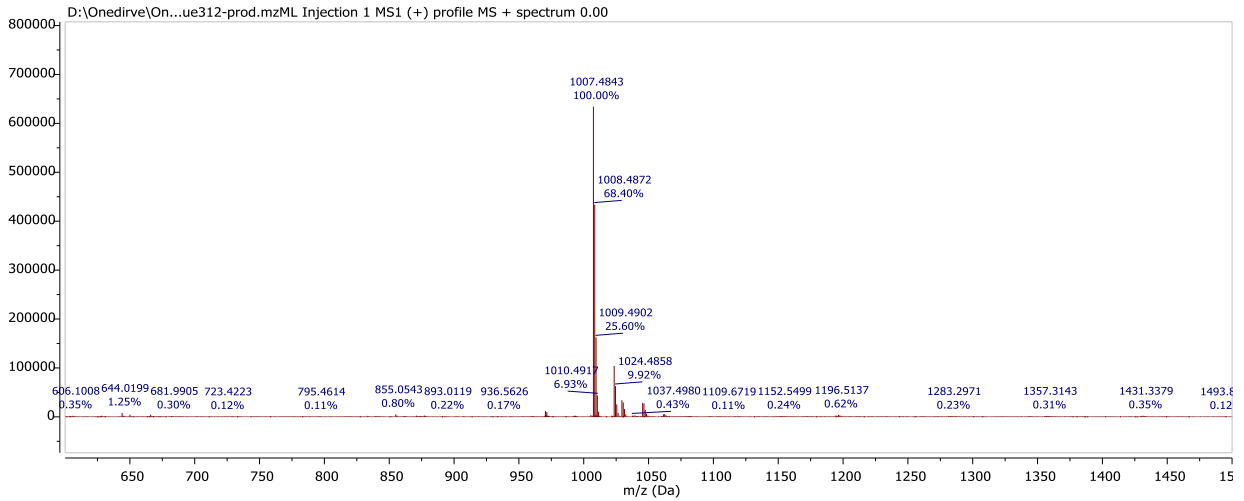
DP₁-Pep₁: DP₁-NH-YFMVF-CONH₂



Chemical Formula: C₅₇H₆₆N₈O₇S
Exact Mass: 1006.4775

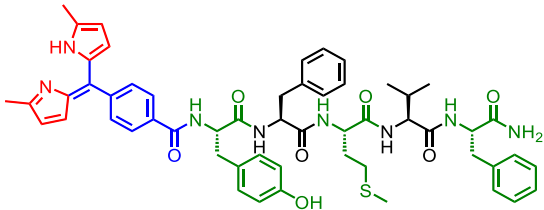


Analytic HPLC of DP₁-Pep₁

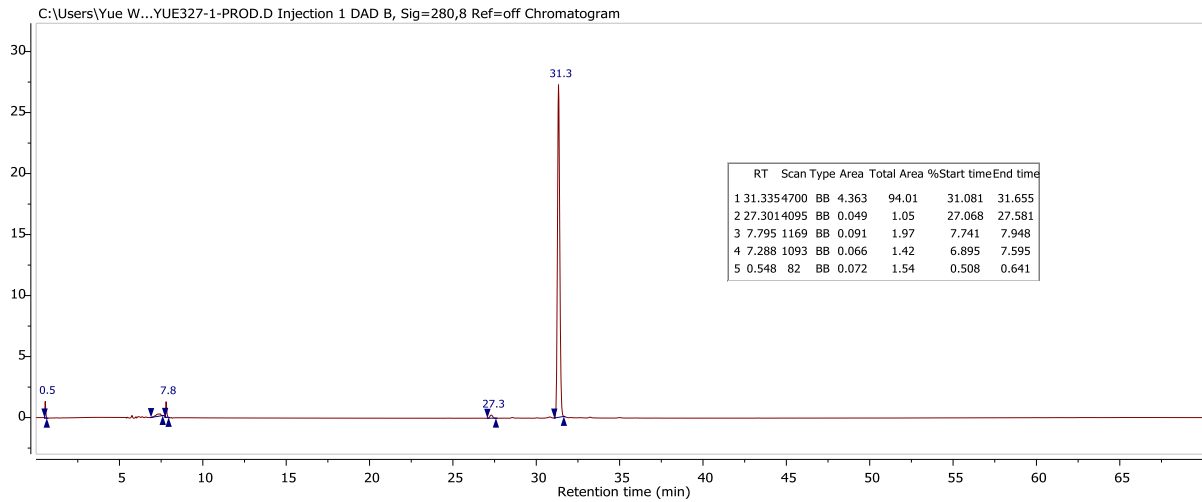


HRMS(MAIDL-TOF) of DP₁-Pep₁

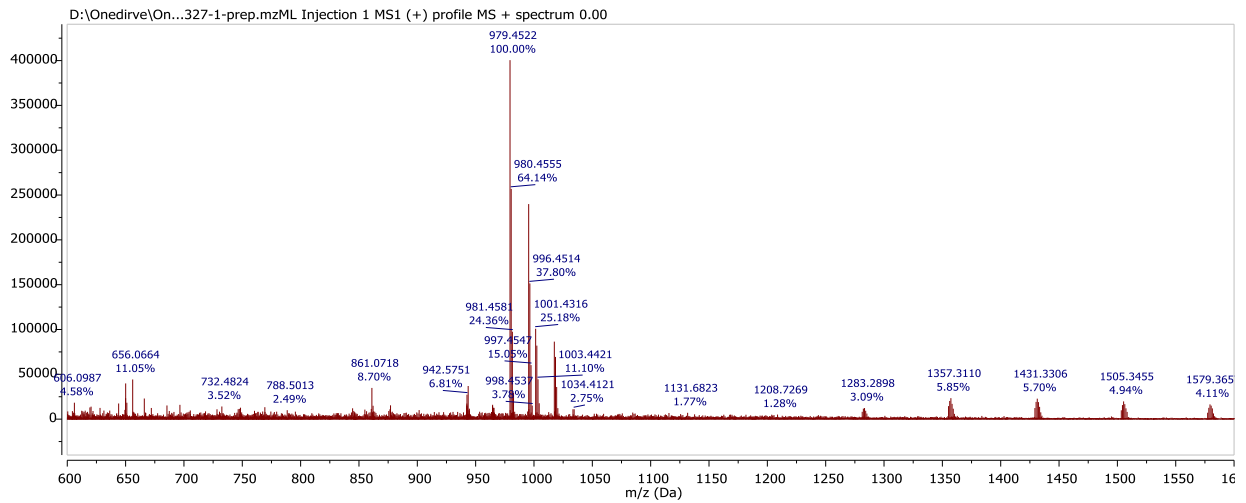
DP₂-Pep₁: DP₂-NH-YFMVF-CONH₂



Chemical Formula: C₅₅H₆₂N₈O₇S
Exact Mass: 978.4462

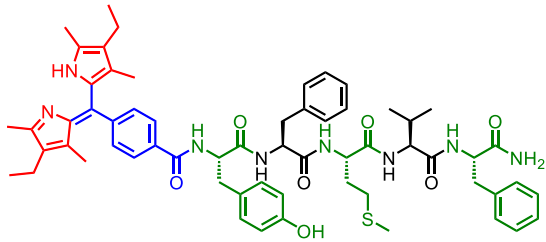


Analytic HPLC of DP₂-Pep₁



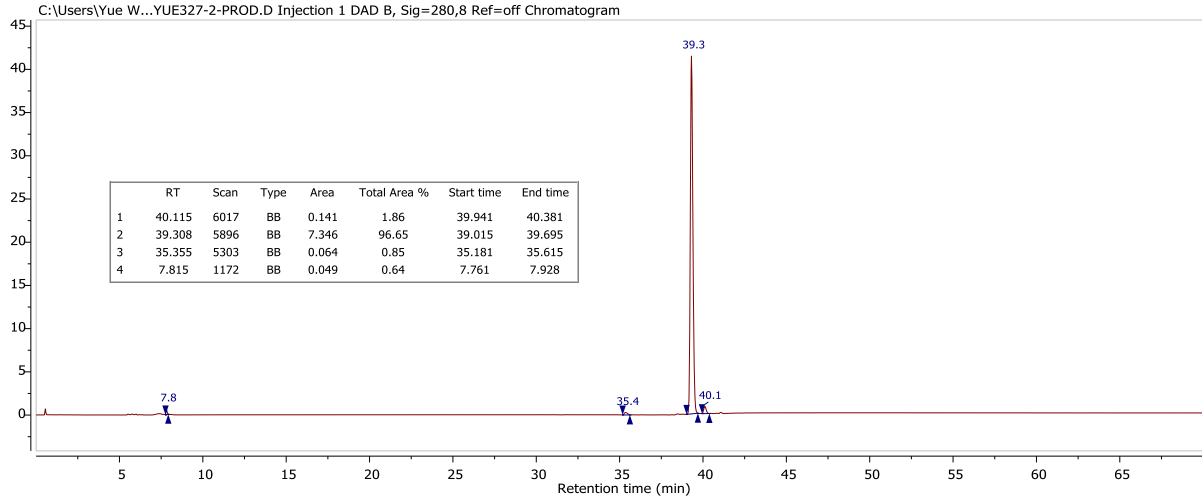
HRMS(MAIDL-TOF) of DP₂-Pep₁

DP₃-Pep₁: DP₃-NH-YFMVF-CONH₂

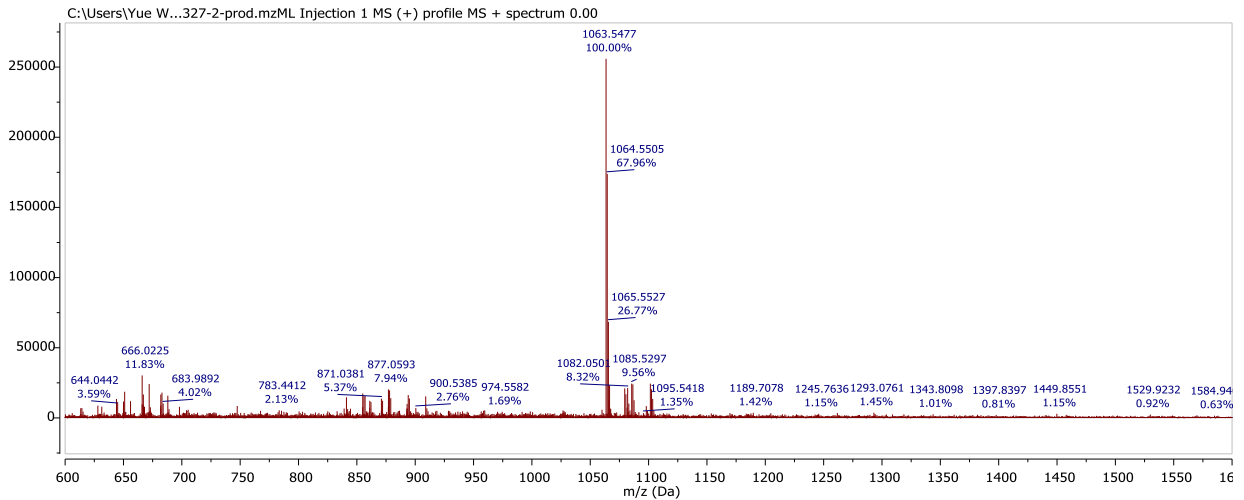


Chemical Formula: C₆₁H₇₄N₈O₇S

Exact Mass: 1062.5401

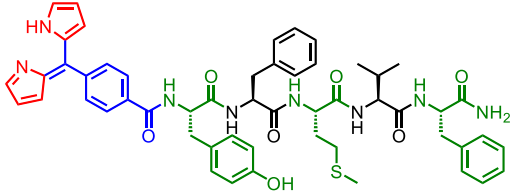


Analytic HPLC of DP₃-Pep₁

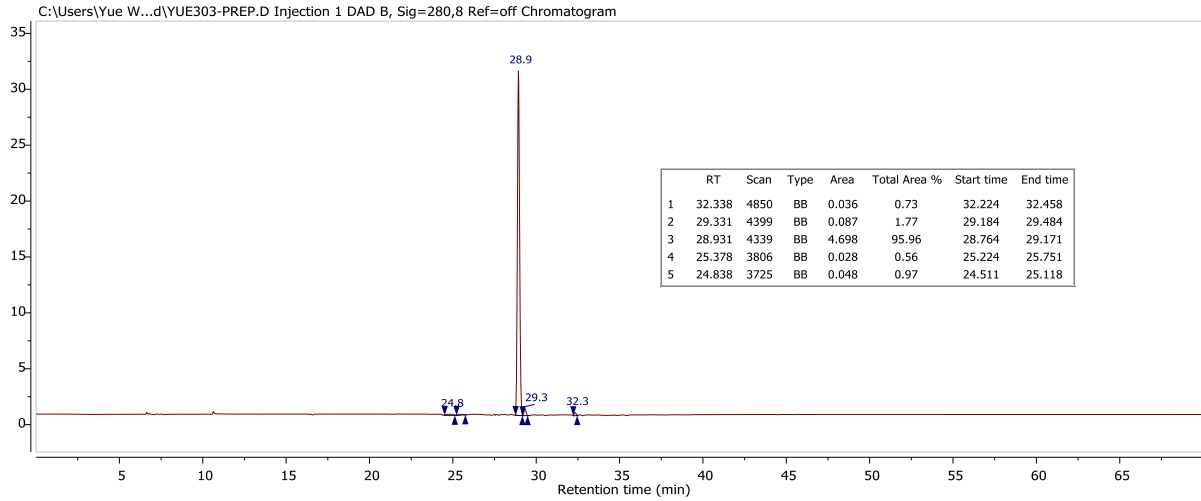


HRMS(MAIDL-TOF) of DP₃-Pep₁

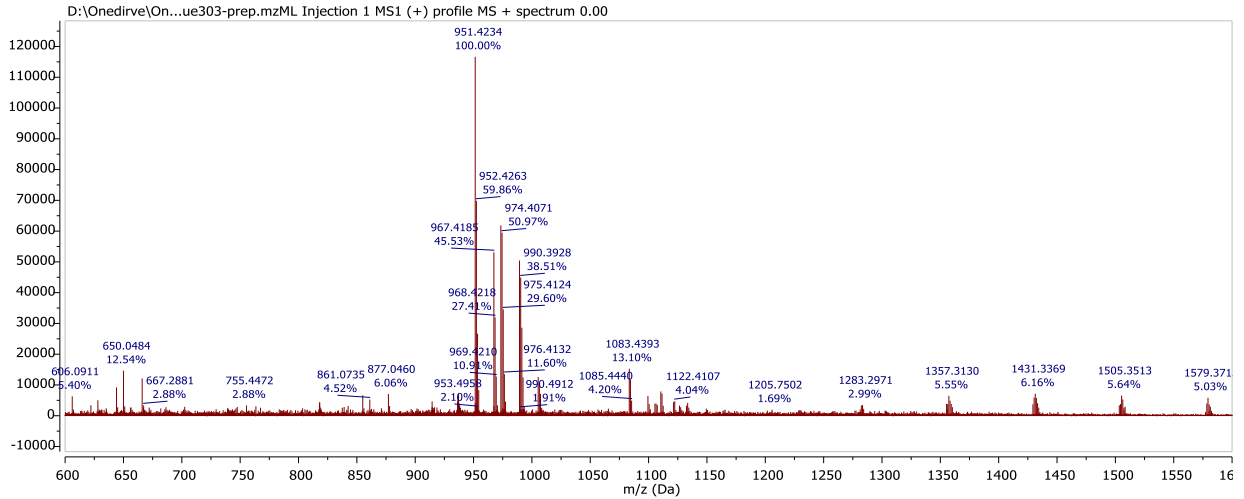
DP₄-Pep₁: DP₄-NH-YFMVF-CONH₂



Chemical Formula: C₅₃H₅₈N₈O₇S
 Exact Mass: 950.4149

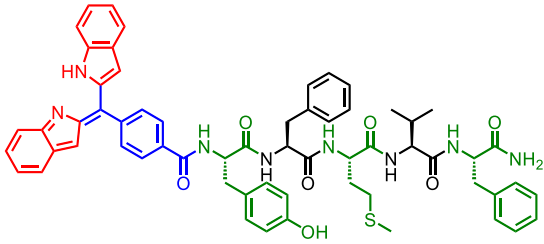


Analytic HPLC of DP₄-Pep₁

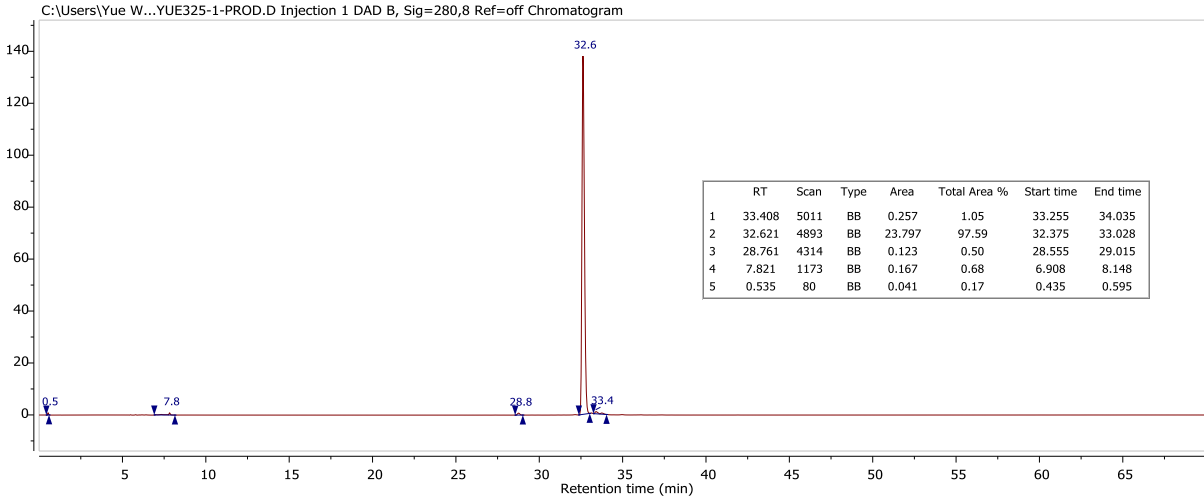


HRMS(MAIDL-TOF) of DP₄-Pep₁

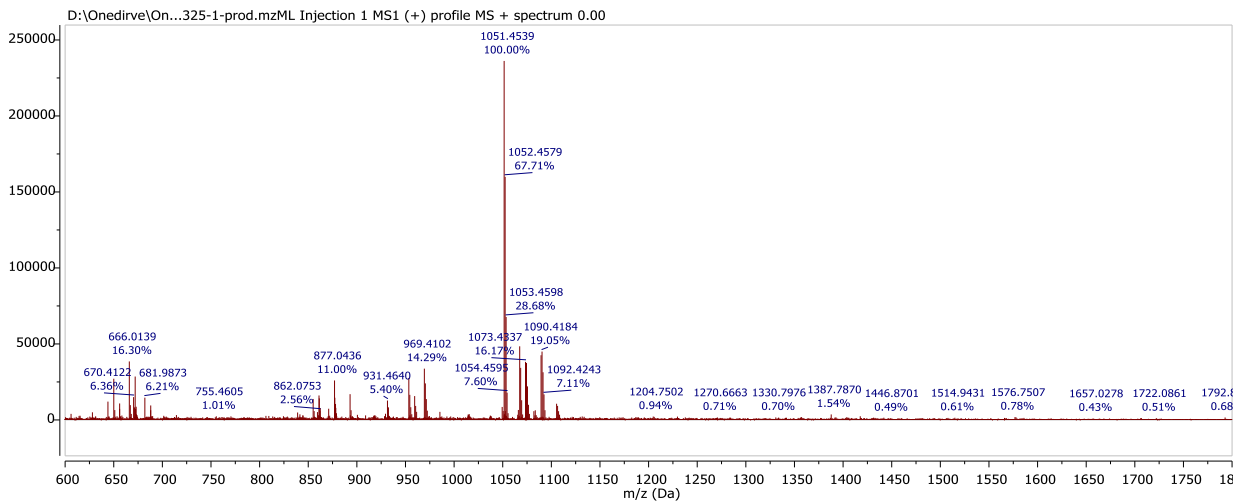
DP₅-Pep₁: DP₅-NH-YFMVF-CONH₂



Chemical Formula: C₆₁H₆₂N₈O₇S
 Exact Mass: 1050.4462

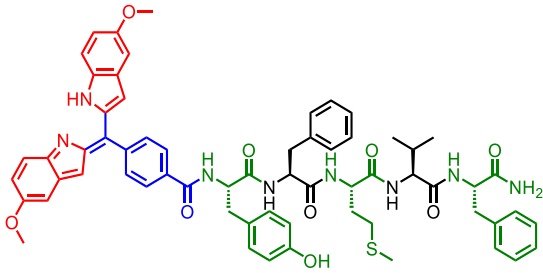


Analytic HPLC of DP₅-Pep₁

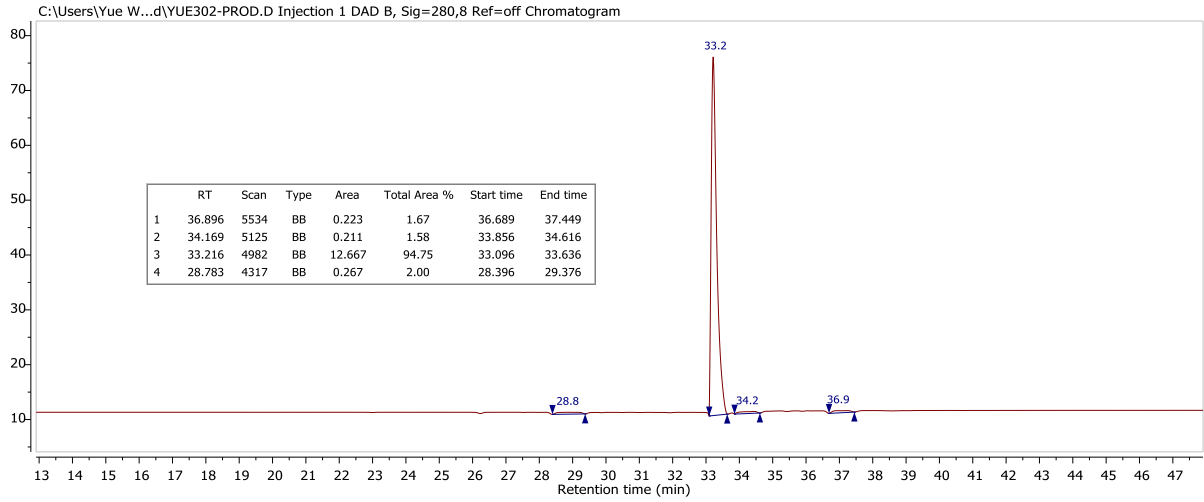


HRMS(MAIDL-TOF) of DP₅-Pep₁

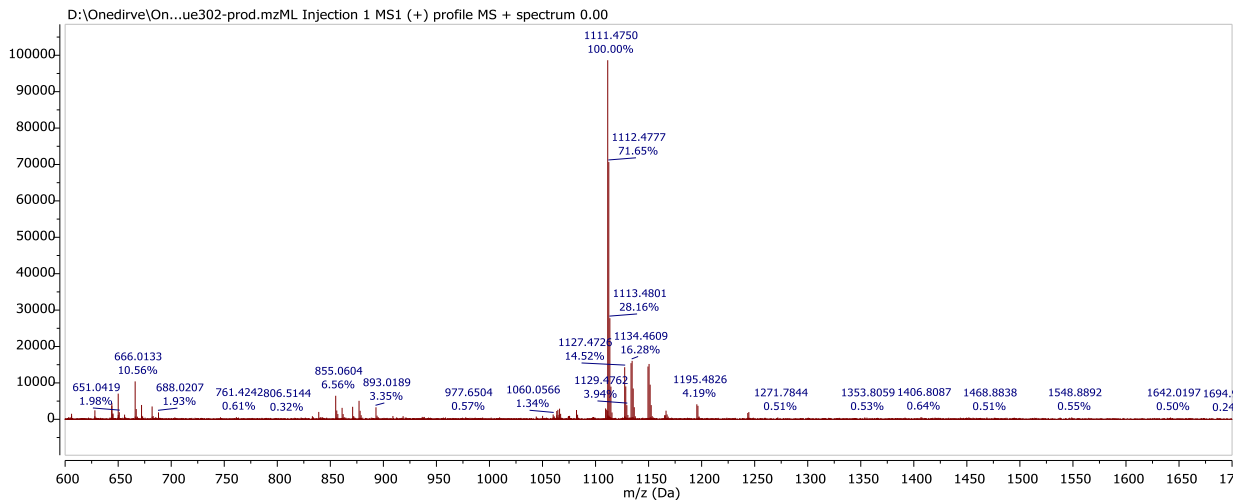
DP₆-Pep₁: DP₆-NH-YFMVF-CONH₂



Chemical Formula: C₆₃H₆₆N₉O₉S
Exact Mass: 1110.4673

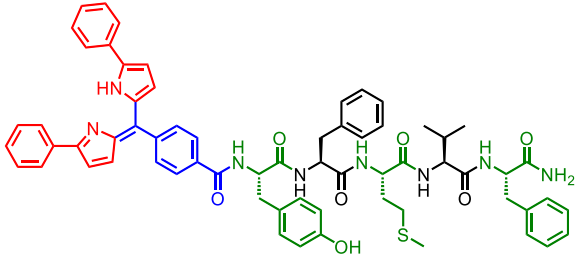


Analytic HPLC of DP₆-Pep₁

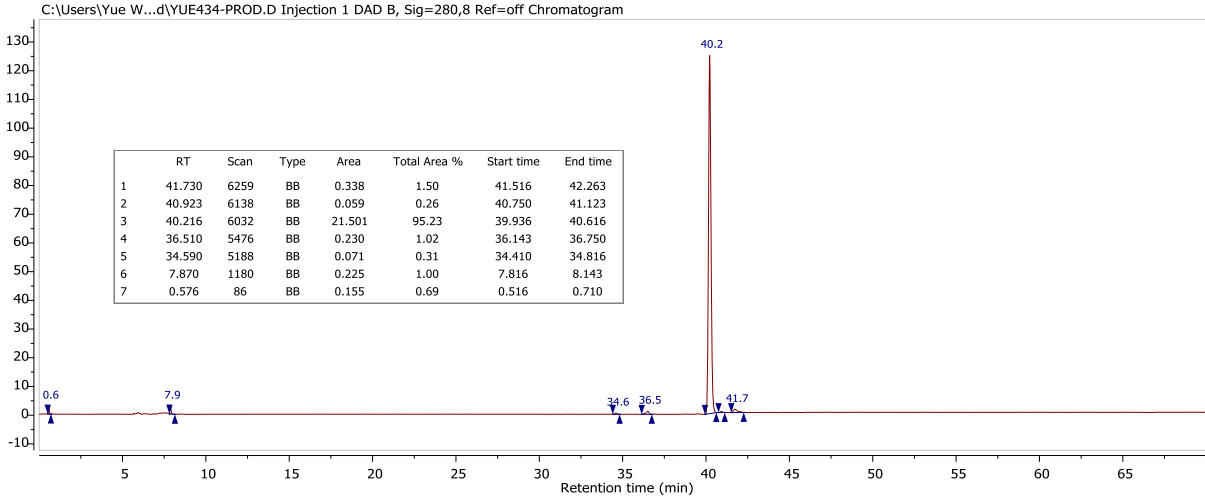


HRMS(MAIDL-TOF) of DP₆-Pep₁

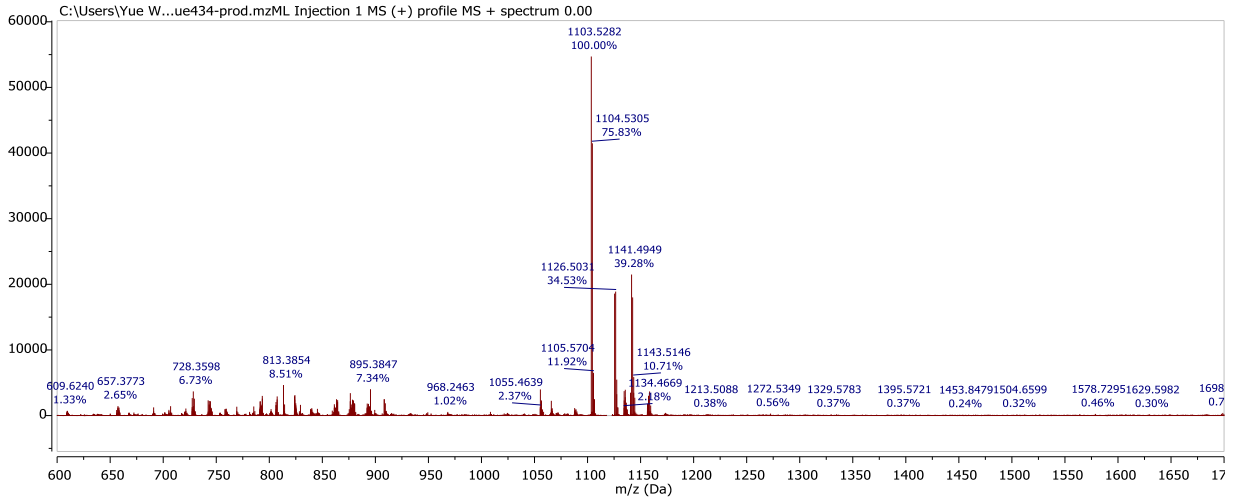
DP₉-Pep₁: DP₉-NH-YFMVF-CONH₂



Chemical Formula: C₆₅H₆₆N₈O₇S
 Exact Mass: 1102.4775

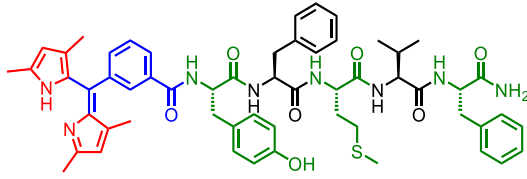


Analytic HPLC of DP₉-Pep₁

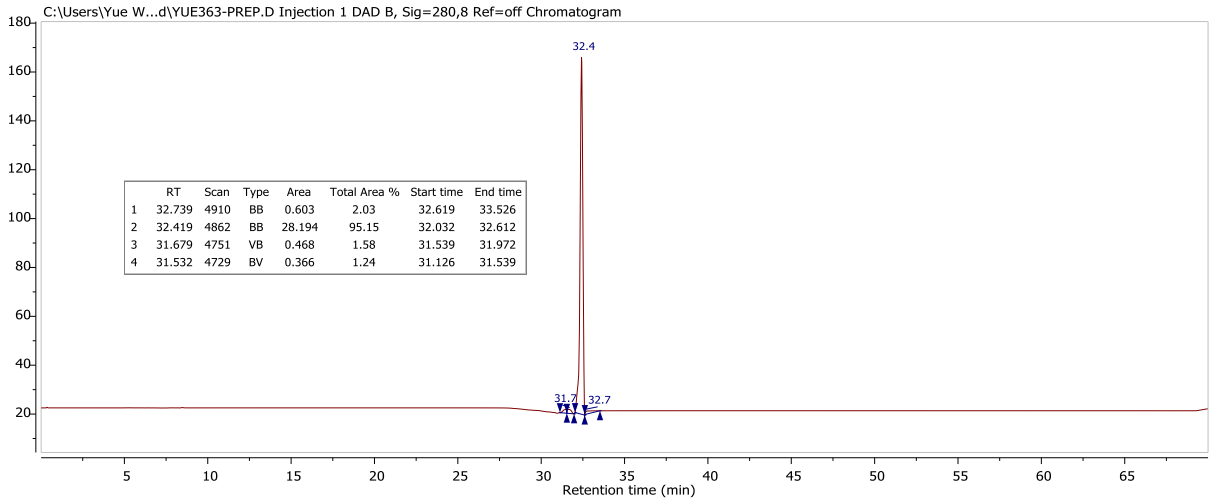


HRMS(MAIDL-TOF) of DP₉-Pep₁

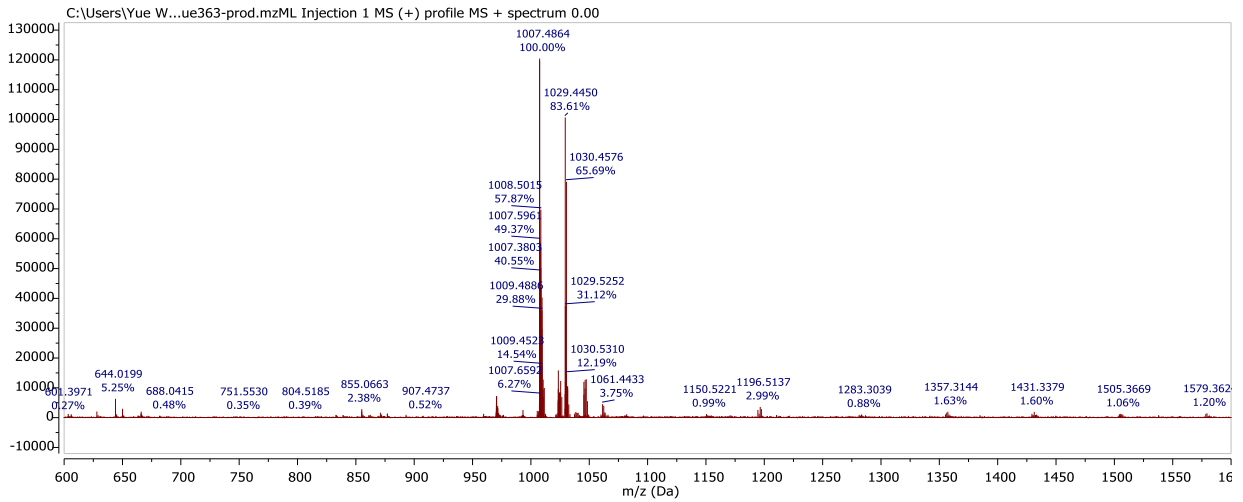
DP₁₀-Pep₁: DP₁₀-NH-YFMVF-CONH₂



Chemical Formula: C₅₇H₆₆N₈O₇S
Exact Mass: 1006.4775

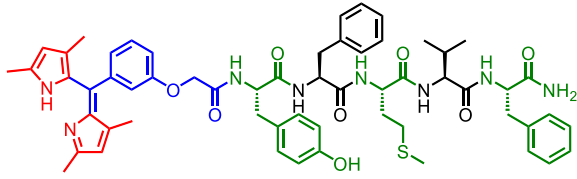


Analytic HPLC of DP₁₀-Pep₁

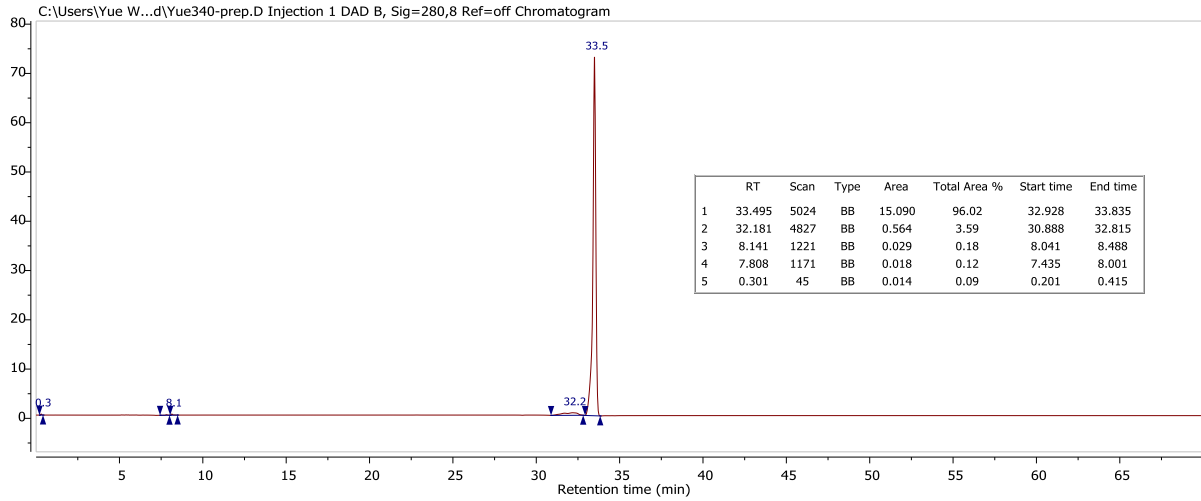


HRMS(MAIDL-TOF) of DP₁₀-Pep₁

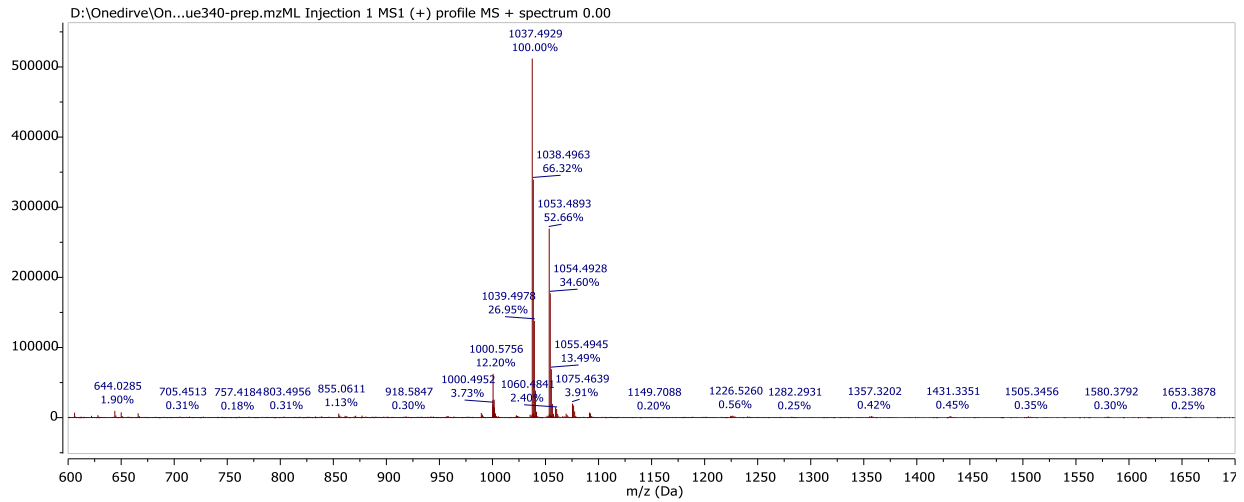
DP₁₂-Pep₁: DP₁₂-NH-YFMVF-CONH₂



Chemical Formula: C₅₈H₆₈N₈O₈S
 Exact Mass: 1036.4881

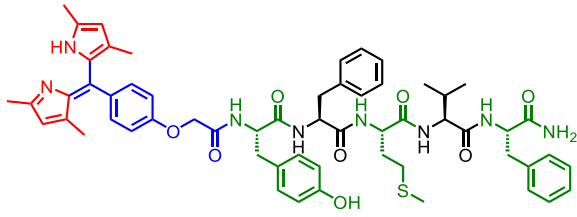


Analytic HPLC of DP₁₂-Pep₁

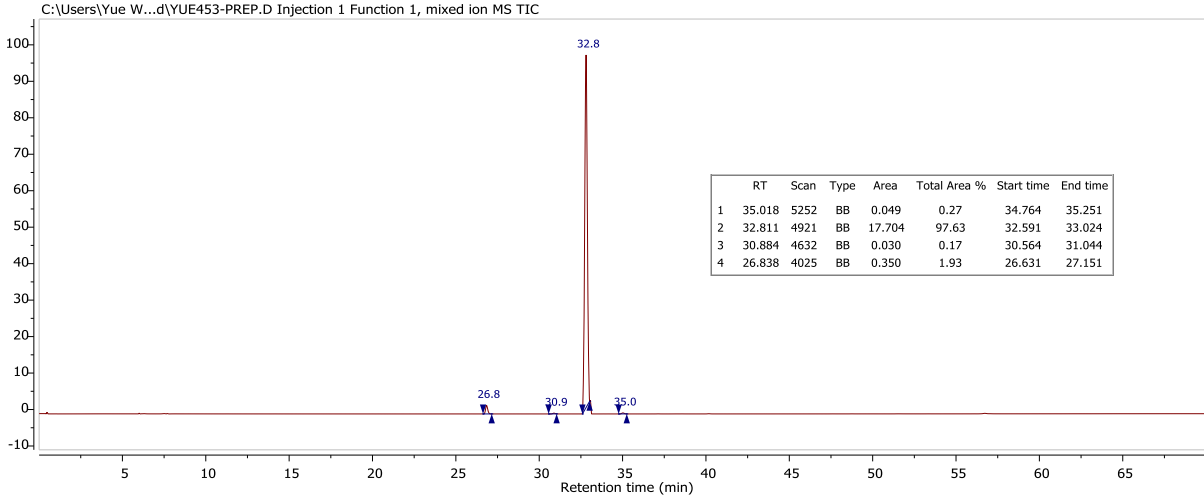


HRMS(MAIDL-TOF) of DP₁₂-Pep₁

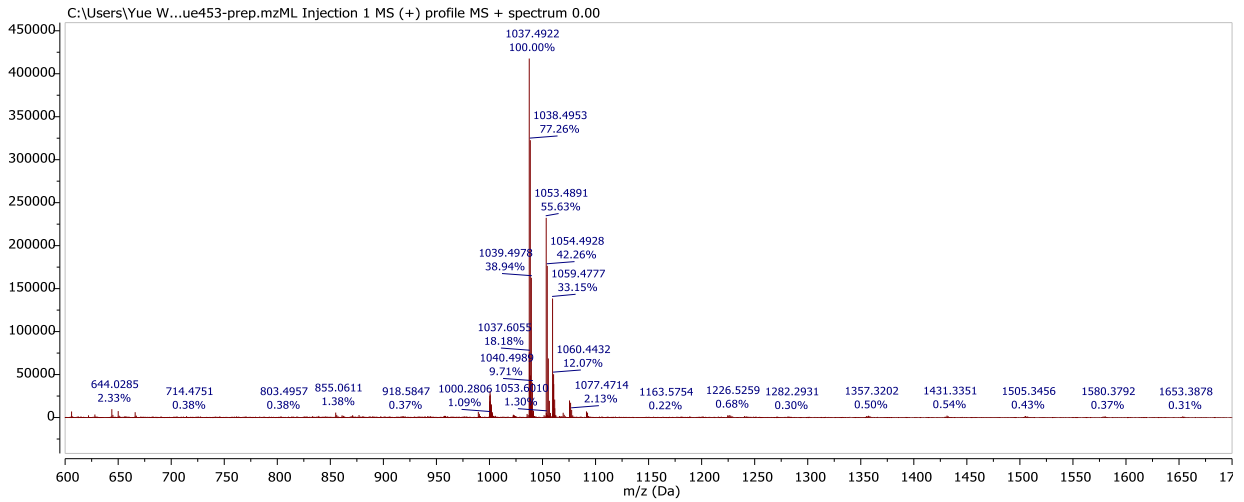
DP₁₃-Pep₁: DP₁₃-NH-YFMVF-CONH₂



Chemical Formula: C₅₈H₆₈N₈O₈S
Exact Mass: 1036.4881

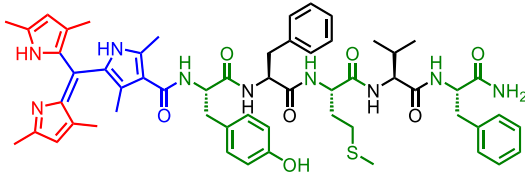


Analytic HPLC of DP₁₃-Pep₁



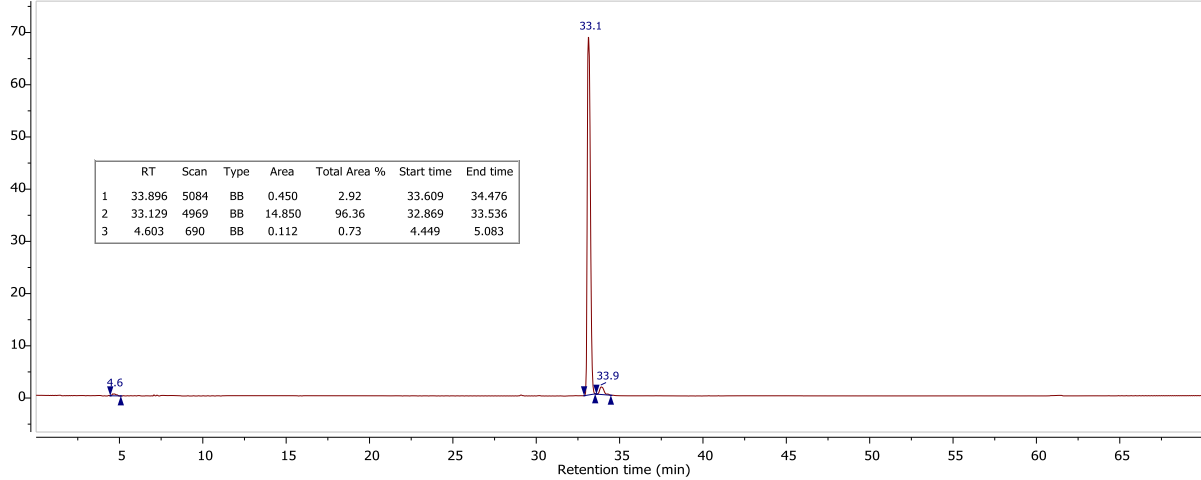
HRMS(MAIDL-TOF) of DP₁₃-Pep₁

DP₁₄-Pep₁: DP₁₄-NH-YFMVF-CONH₂



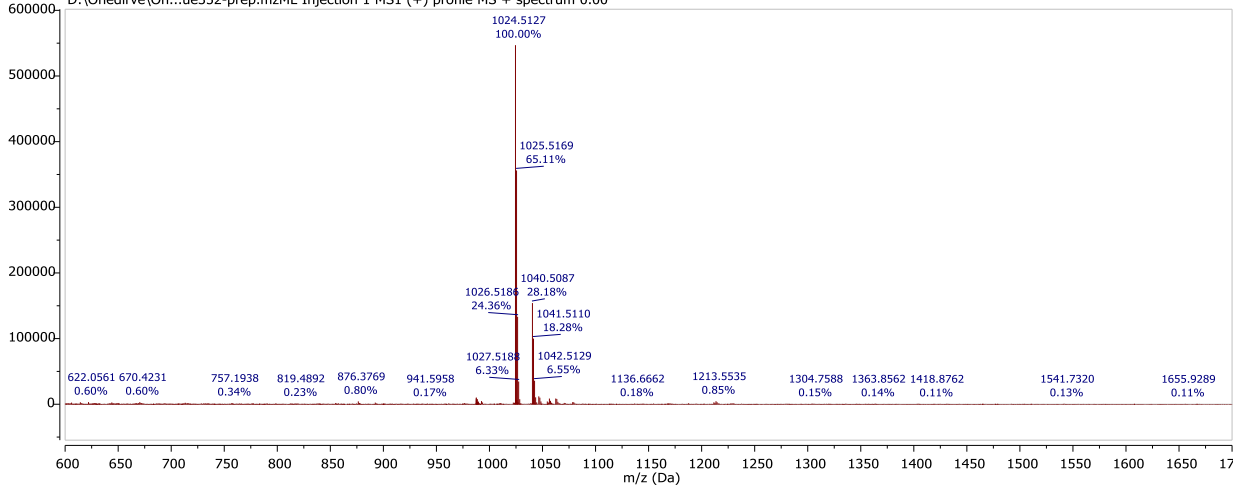
Chemical Formula: C₅₇H₆₉N₉O₇S
 Exact Mass: 1023.5041

C:\Users\Yue W...d\YUE352-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram



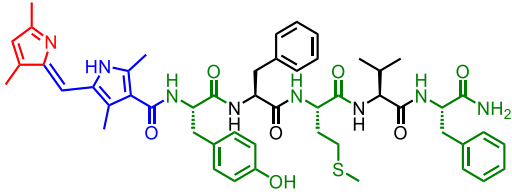
Analytic HPLC of DP₁₄-Pep₁

D:\Onedrive\On...ue352-prep.mzML Injection 1 MS1 (+) profile MS + spectrum 0.00

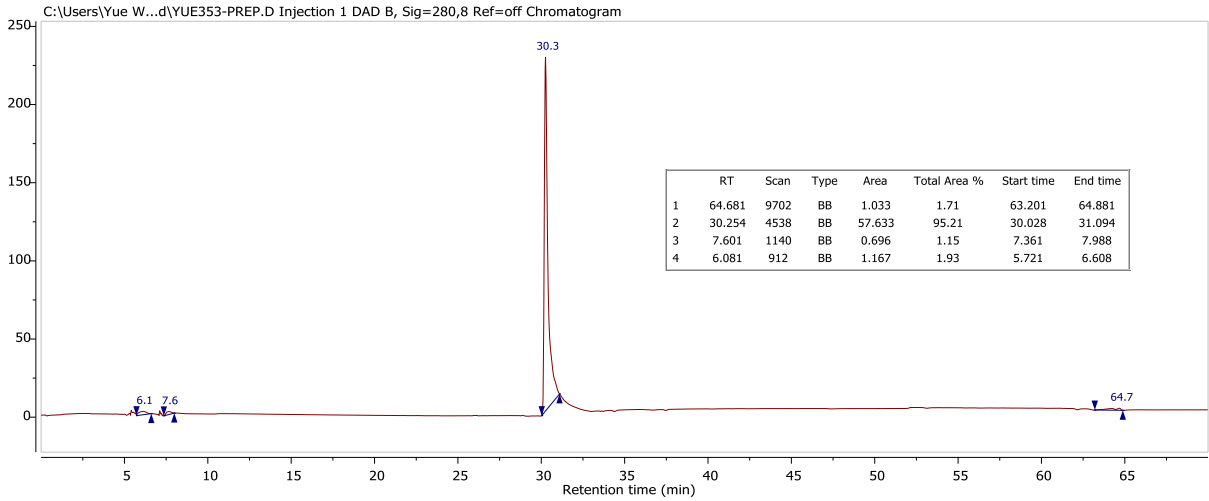


HRMS(MAIDL-TOF) of DP₁₄-Pep₁

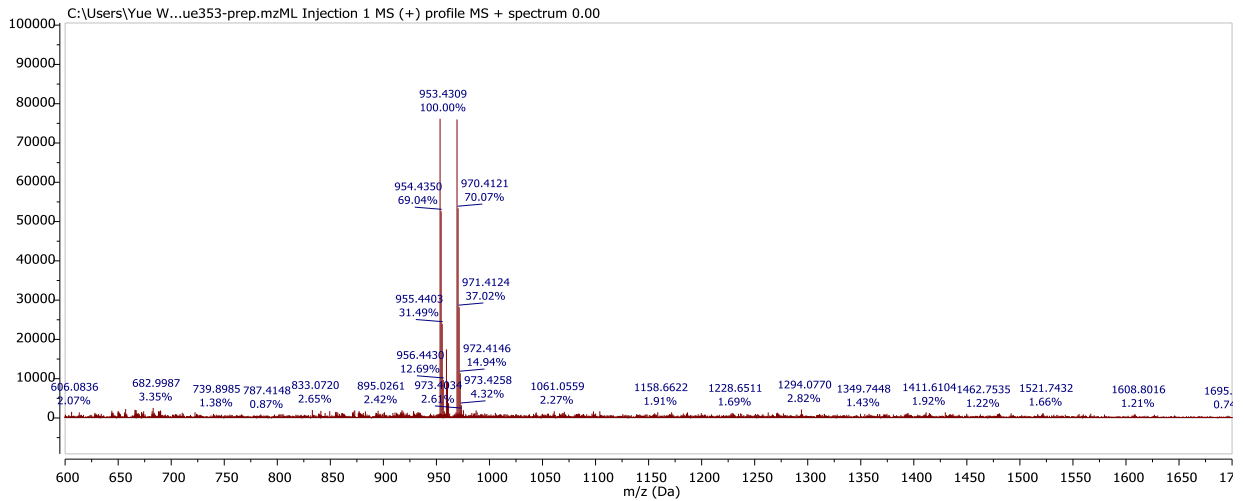
DP₁₅-Pep₁: DP₁₅-NH-YFMVF-CONH₂



Chemical Formula: C₅₁H₆₂N₈O₇S
Exact Mass: 930.4462

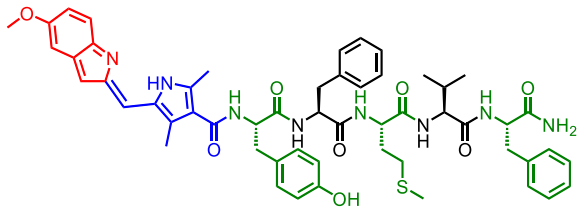


Analytic HPLC of DP₁₅-Pep₁

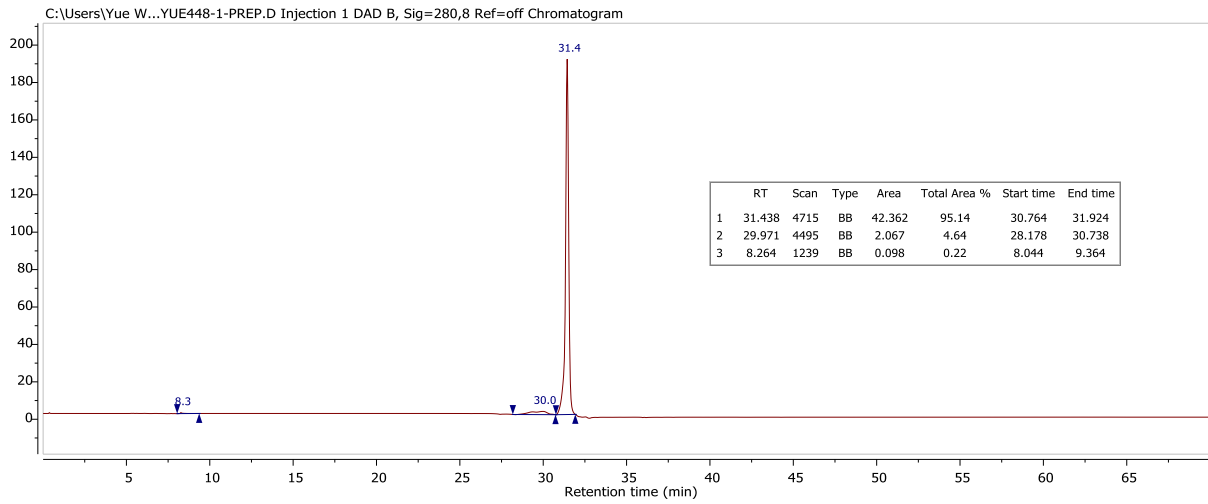


HRMS(MAIDL-TOF) of DP₁₅-Pep₁

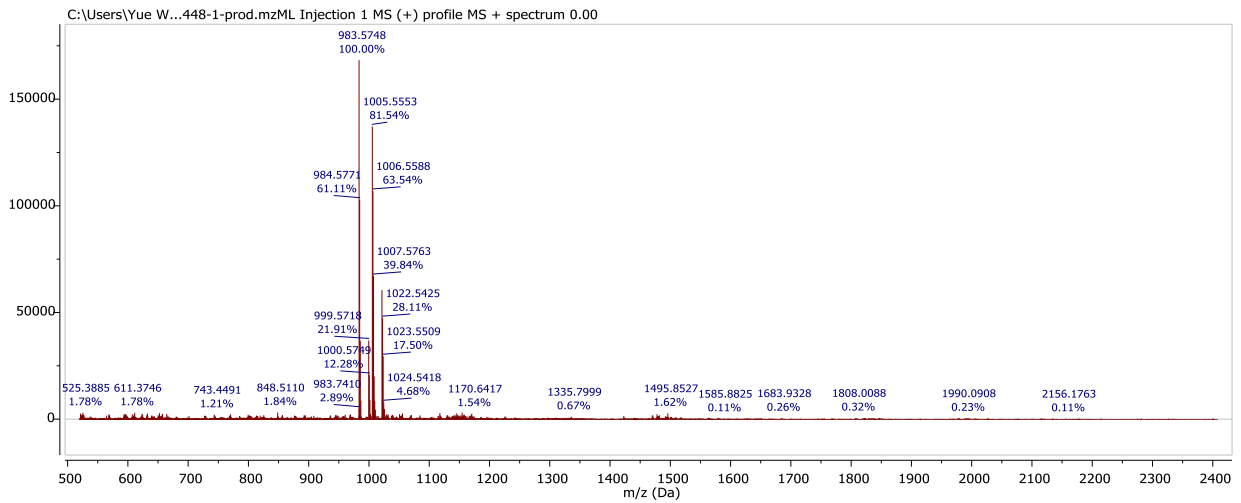
DP₁₆-Pep₁: DP₁₆-NH-YFMVF-CONH₂



Chemical Formula: C₅₄H₆₂N₈O₈S
Exact Mass: 982.4411

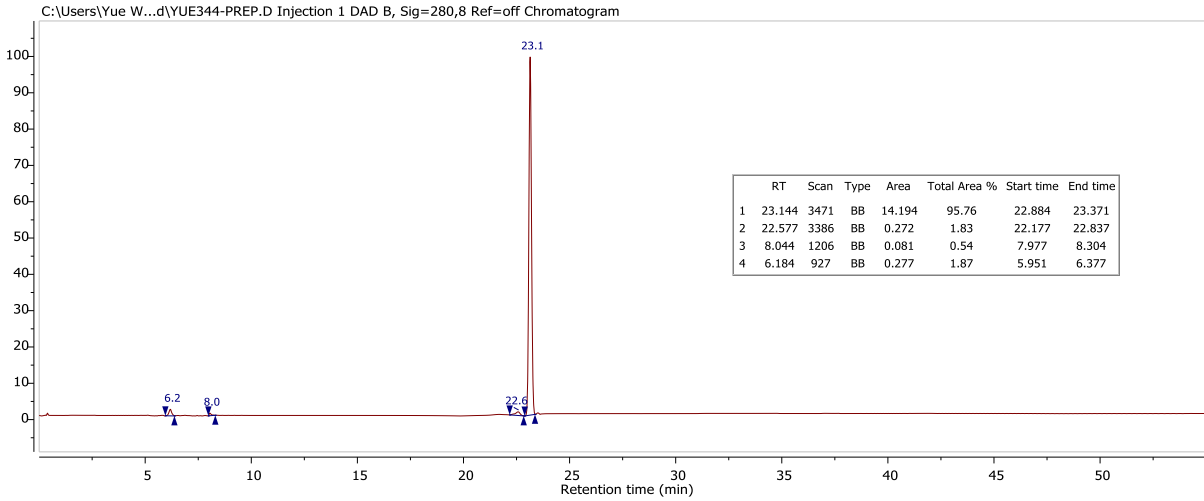
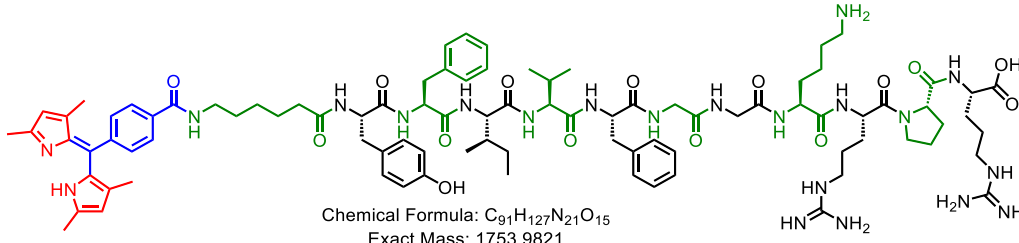


Analytic HPLC of DP₁₆-Pep₁

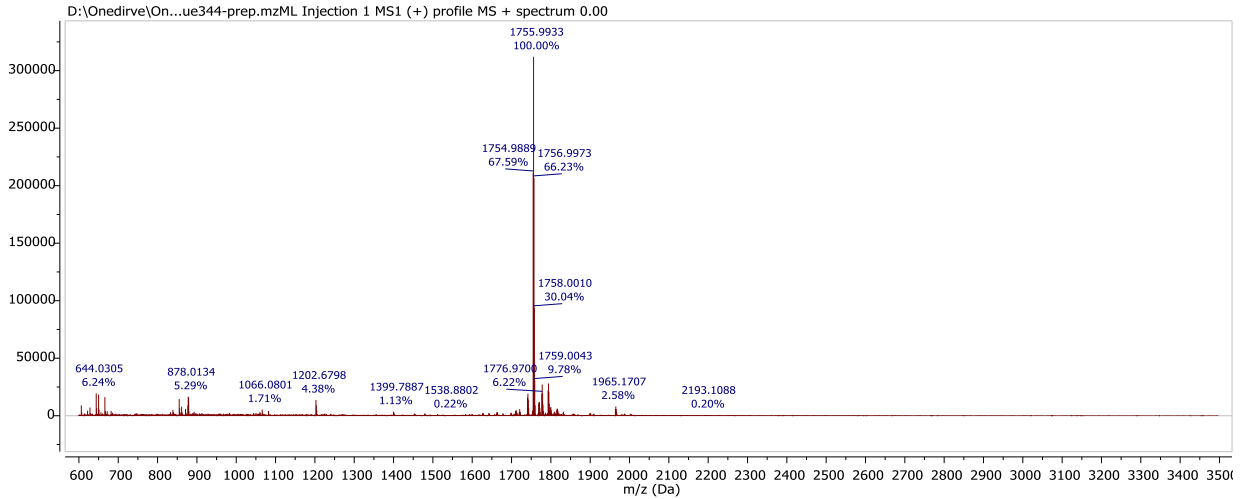


HRMS(MAIDL-TOF) of DP₁₆-Pep₁

DP₁-Pep₂: DP₁-HN-Ahx-YFIVFGGKRPR-COOH

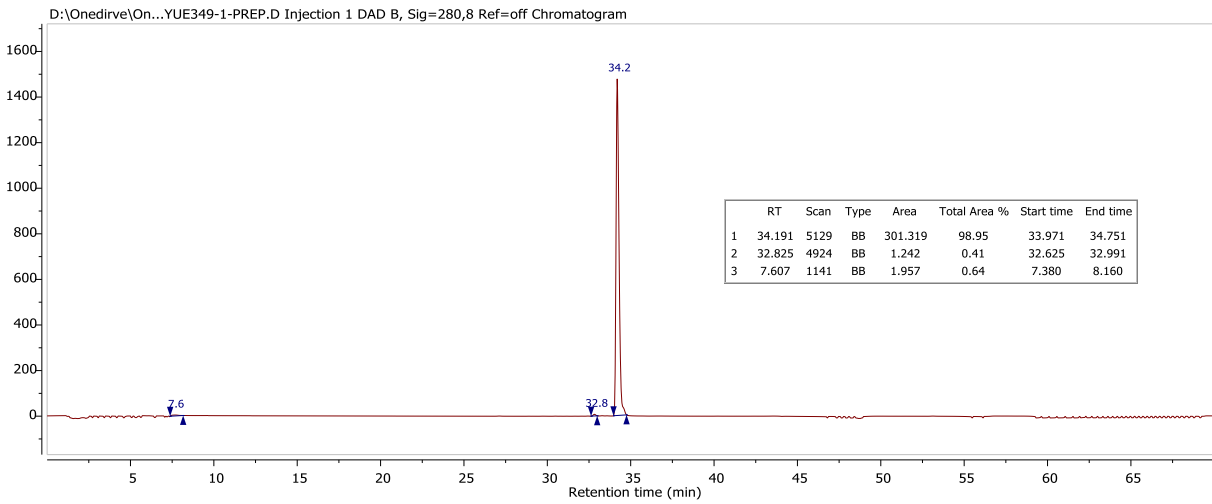
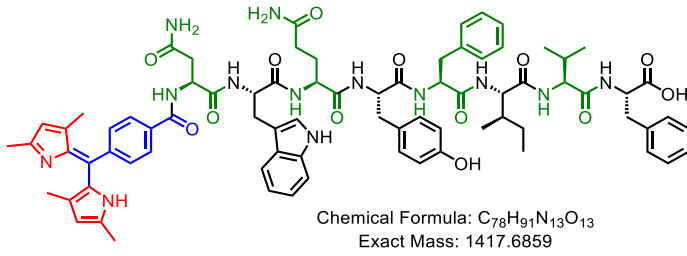


Analytic HPLC of DP₁-Pep₂

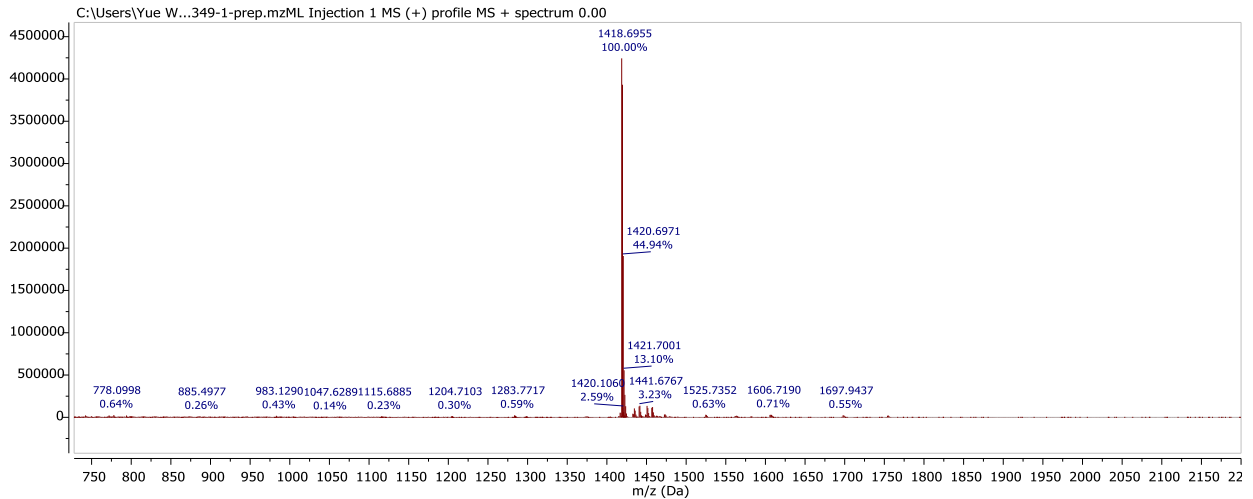


HRMS(MAIDL-TOF) of DP₁-Pep₂

DP₁-Pep₅: DP₁-HN-NWQYFIVE-COOH

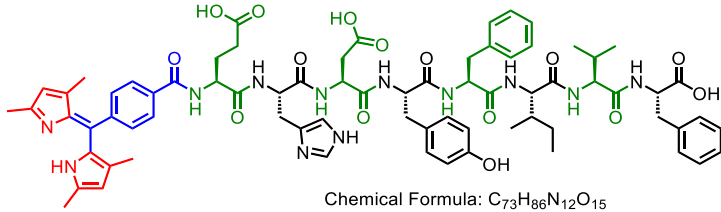


Analytic HPLC of DP₁-Pep₅

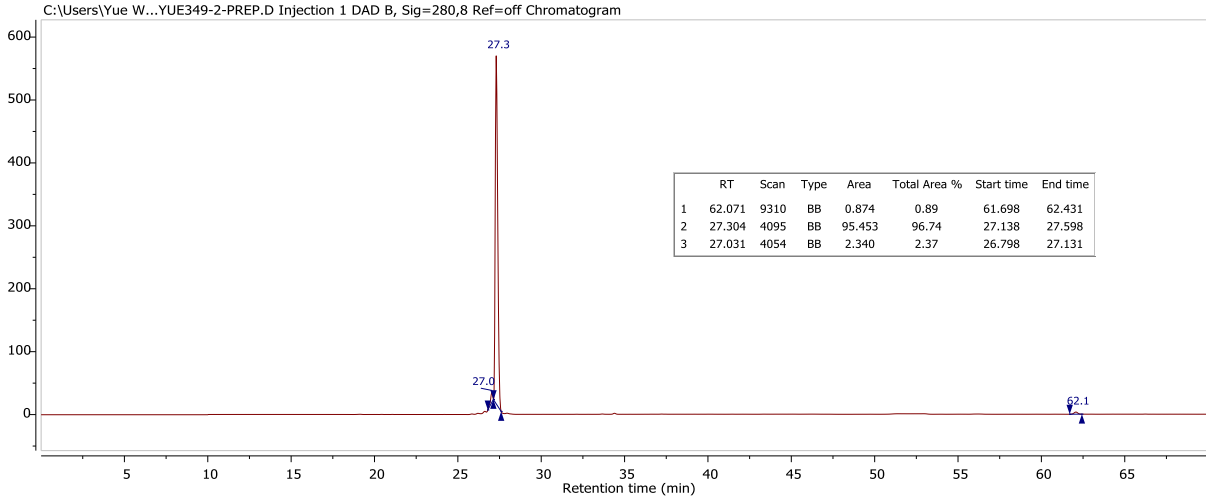


HRMS(MAIDL-TOF) of DP₁-Pep₅

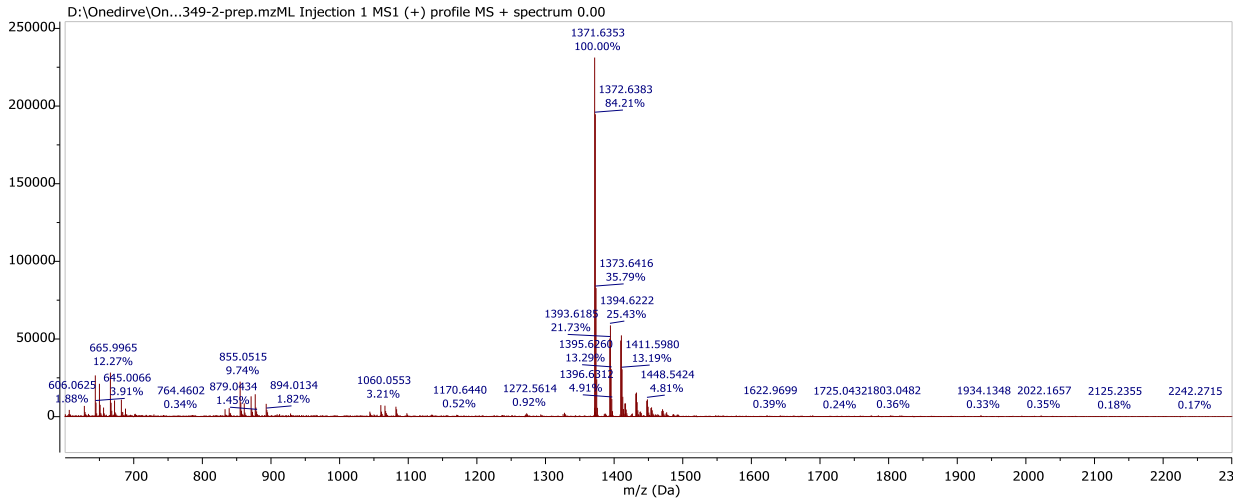
DP₁-Pep₆: DP₁-HN-EHDYFIVE-COOH



Chemical Formula: C₇₃H₈₆N₁₂O₁₅
Exact Mass: 1370.6336

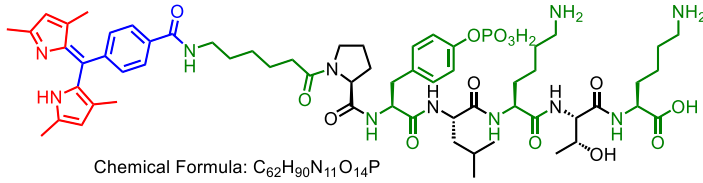


Analytic HPLC of DP₁-Pep₆

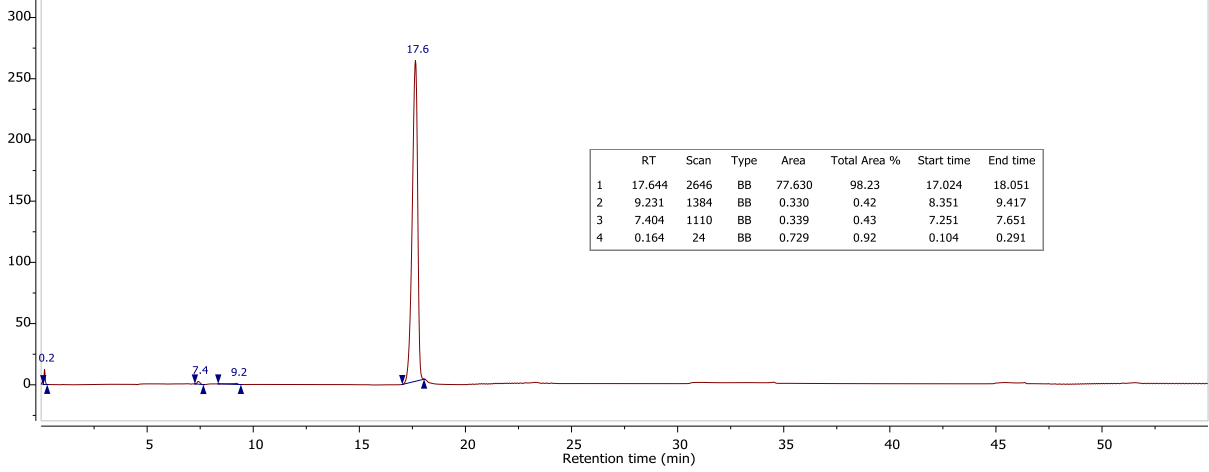


HRMS(MAIDL-TOF) of DP₁-Pep₆

DP₁-Pep₇: DP₁-HN-Ahx-P-(pTyr)-LKTK-COOH

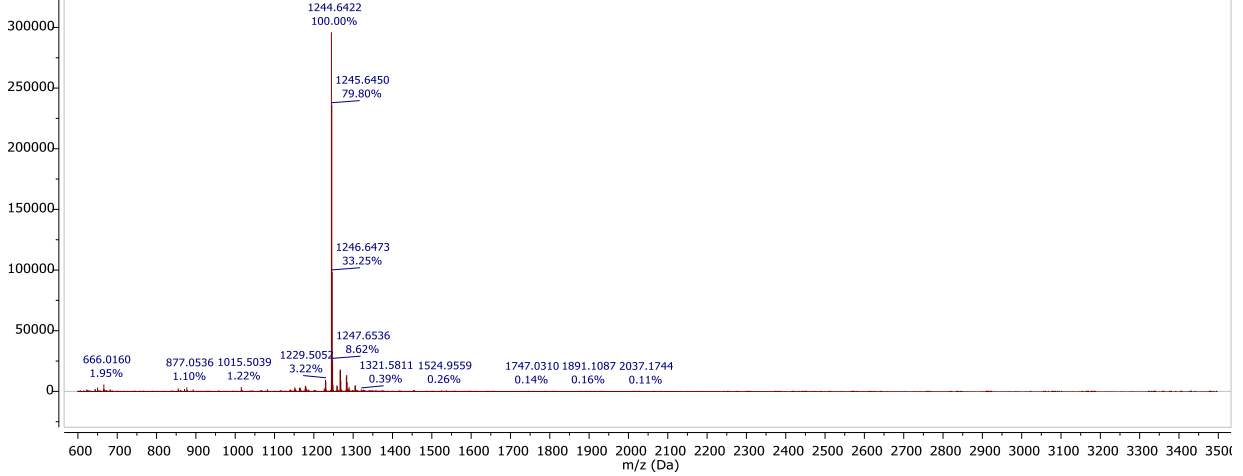


C:\Users\Yue W...d\YUE345-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram



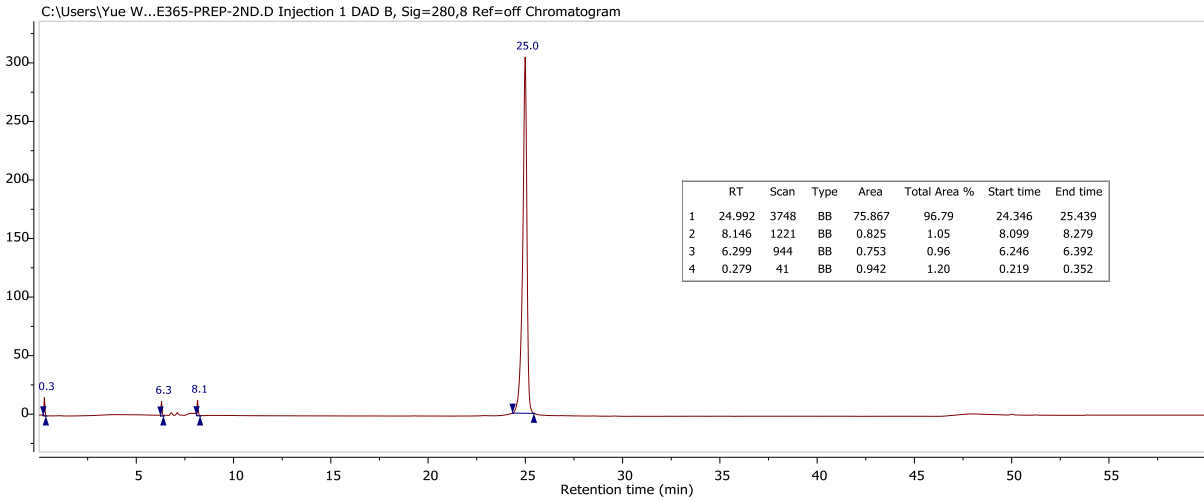
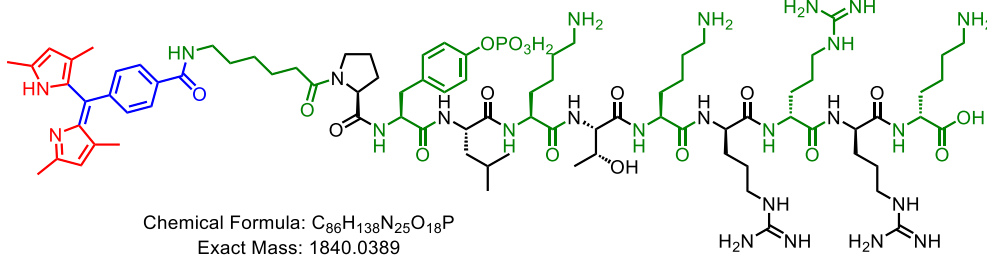
Analytic HPLC of DP₁-Pep₇

D:\OneDrive\On...ue345-prep.mzML Injection 1 MS1 (+) profile MS + spectrum 0.00

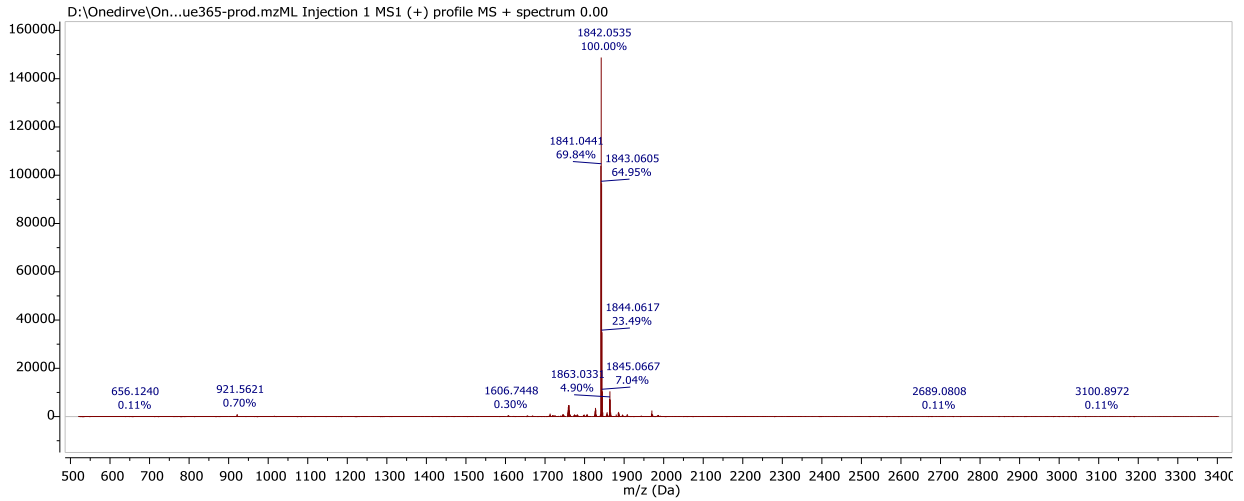


HRMS(MAIDL-TOF) of DP₁-Pep₇

DP₁-Pep₈: DP₁-HN-Ahx-P-(pTyr)-LKTkRrRK-COOH

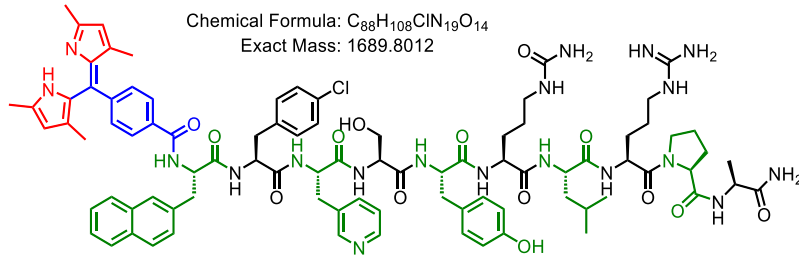


Analytic HPLC of DP₁-Pep₈

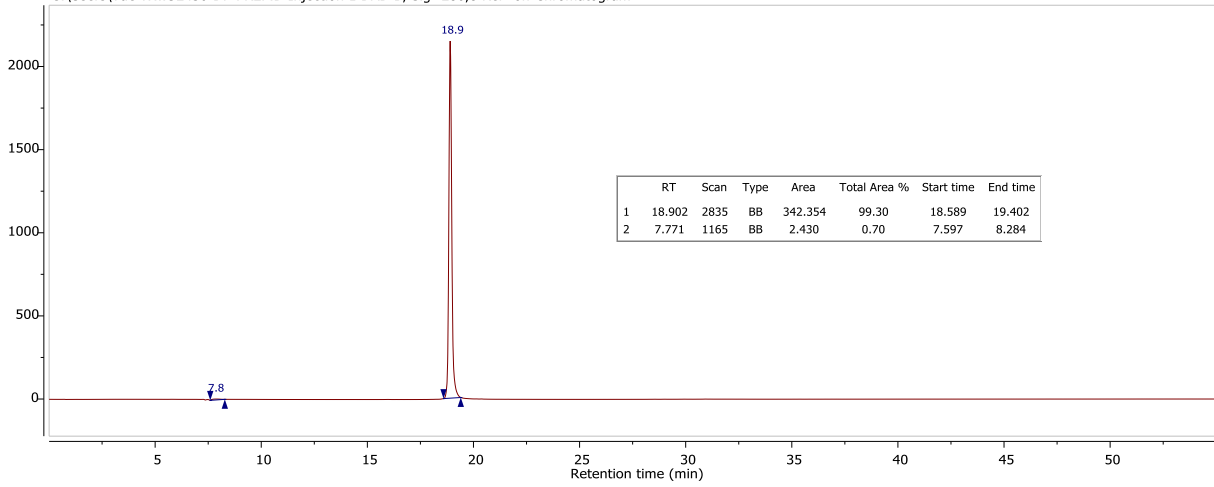


HRMS(MAIDL-TOF) of DP₁-Pep₈

DP₁-Pep₉: DP₁-HN-Nal-Cpa-Pal-SY-Cit-LRPG-CONH₂

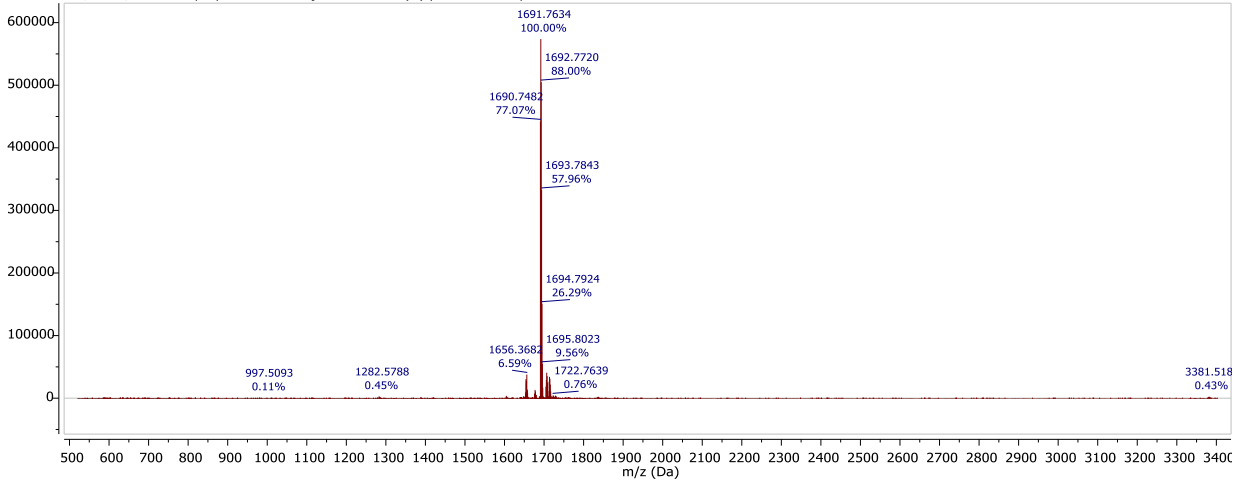


C:\Users\Yue W...UE450-DP-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram



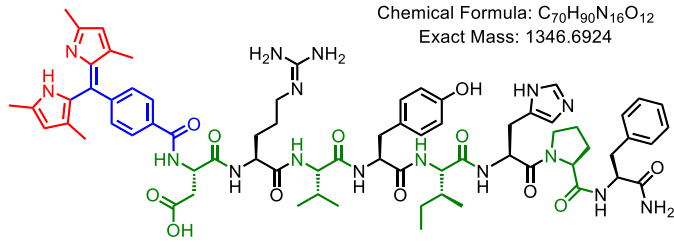
Analytic HPLC of DP₁-Pep₉

C:\Users\Yue W...P-prep-2nd.mzML Injection 1 MS1 (+) profile MS + spectrum 0.00

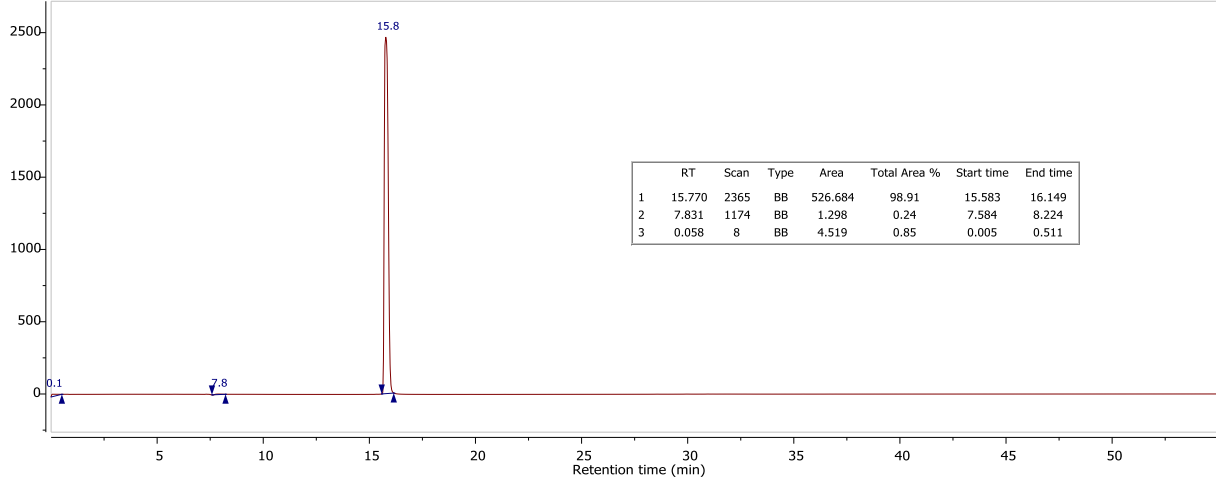


HRMS(MAIDL-TOF) of DP₁-Pep₉

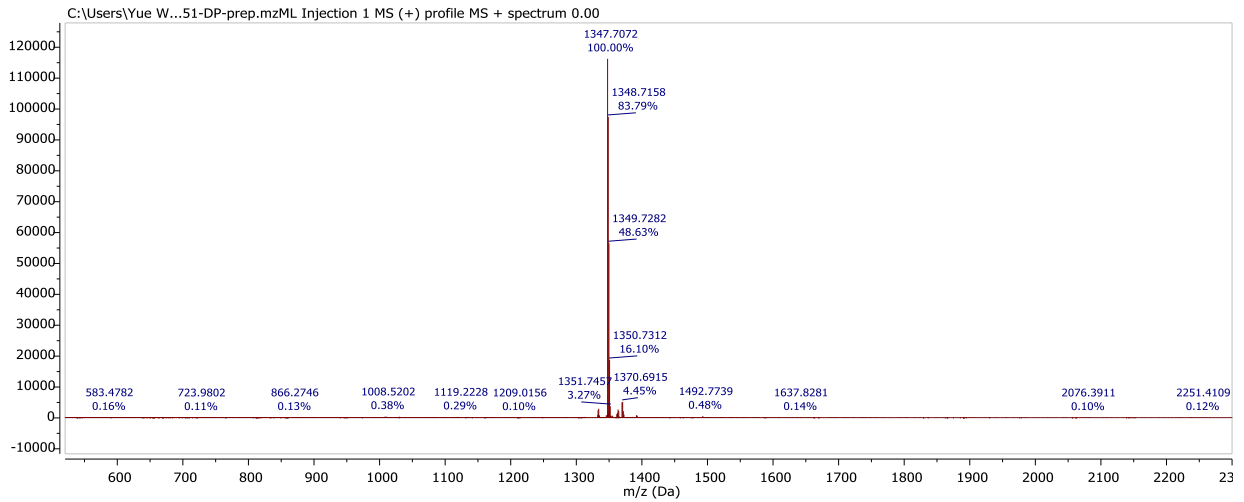
DP₁-Pep₁₀: DP₁-HN-DRVYIHPF-CONH₂



D:\Ondirve\On...UE451-DP-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram

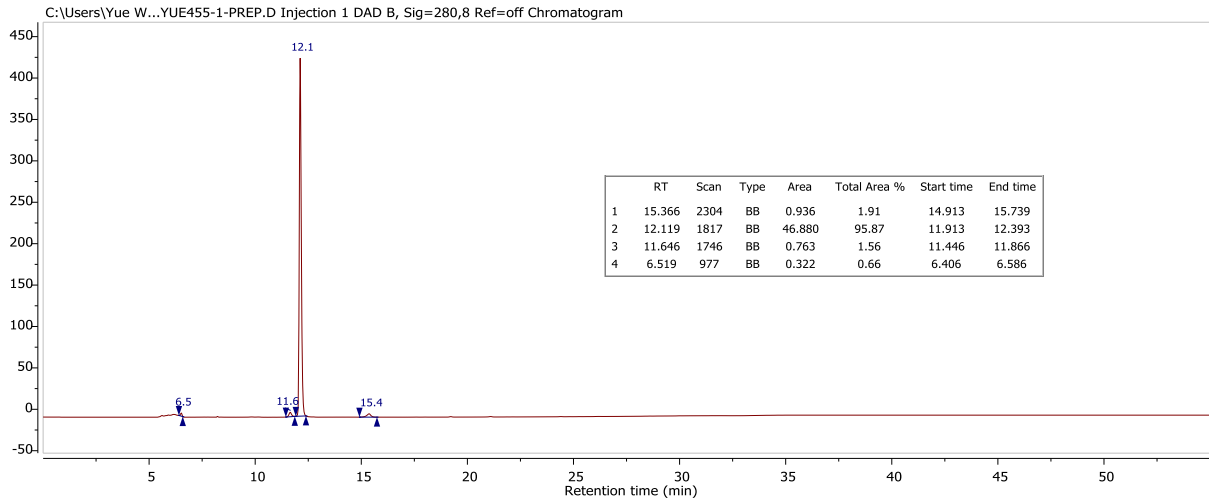
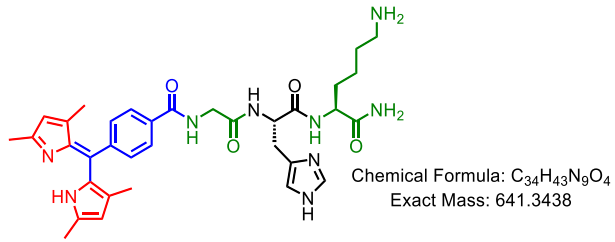


Analytic HPLC of DP₁-Pep₁₀

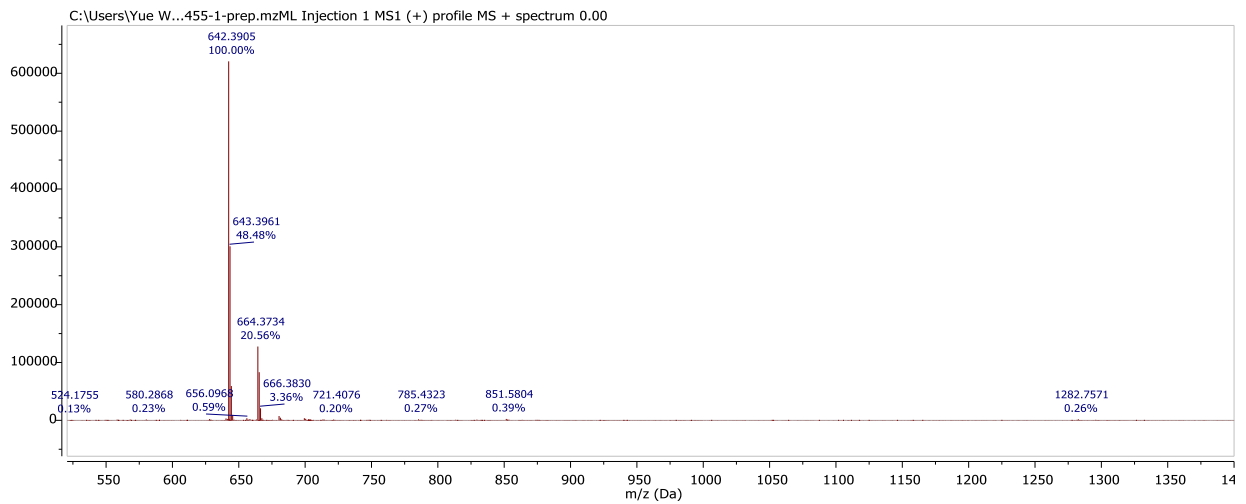


HRMS(MAIDL-TOF) of DP₁-Pep₁₀

DP₁-Pep₁₁: DP₁-HN-GHL-CONH₂

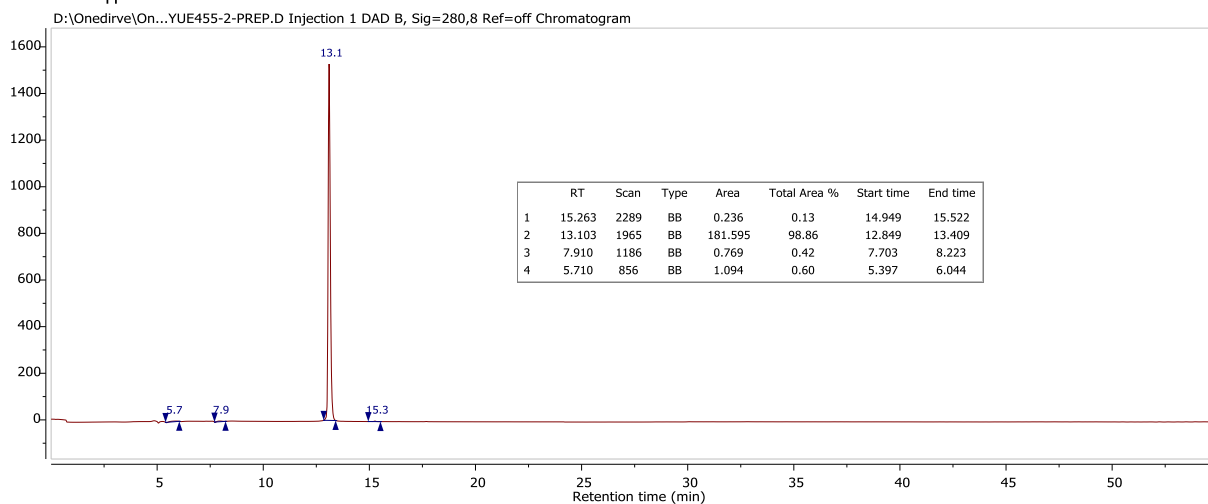
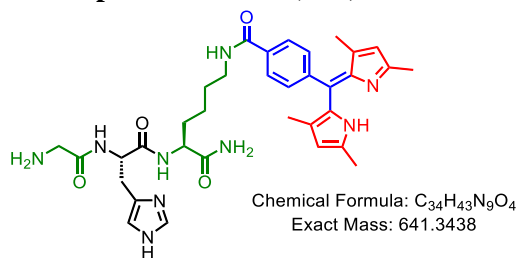


Analytic HPLC of DP₁-Pep₁₁

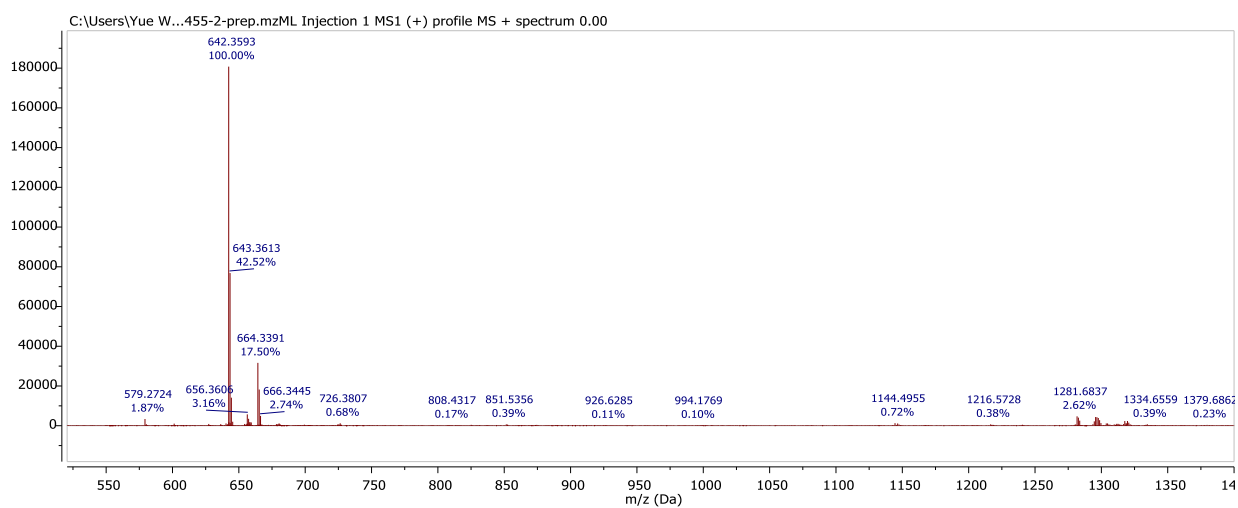


HRMS(MAIDL-TOF) of DP₁-Pep₁₁

DP₁-Pep₁₁*: H₂N-GHL(DP₁)-CONH₂

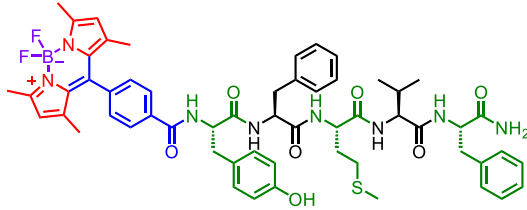


Analytic HPLC of DP₁-Pep₁₁*



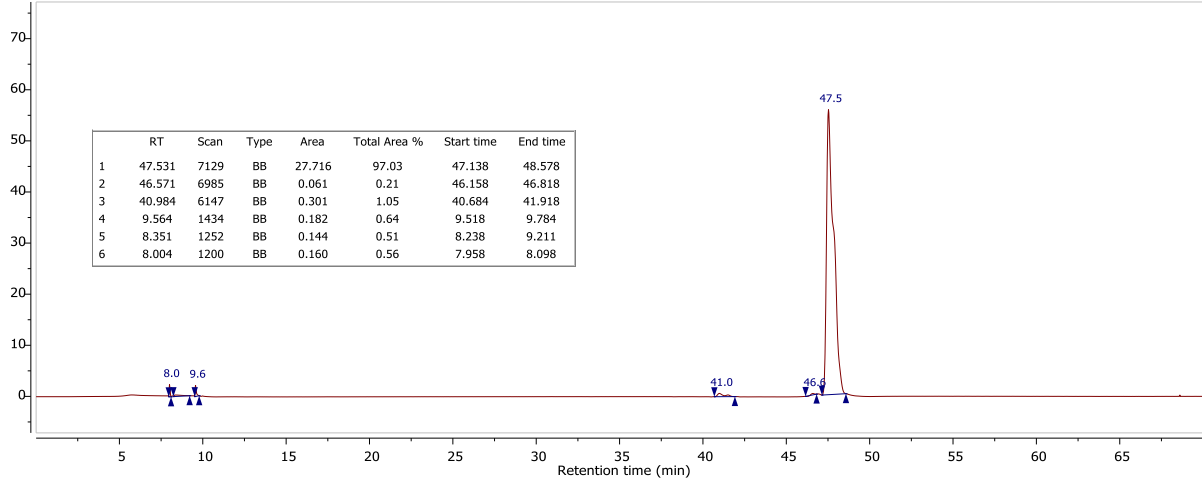
HRMS(MAIDL-TOF) of DP₁-Pep₁₁*

BODIPY₁-Pep₁: BODIPY₁-NH-YFMVF-CONH₂

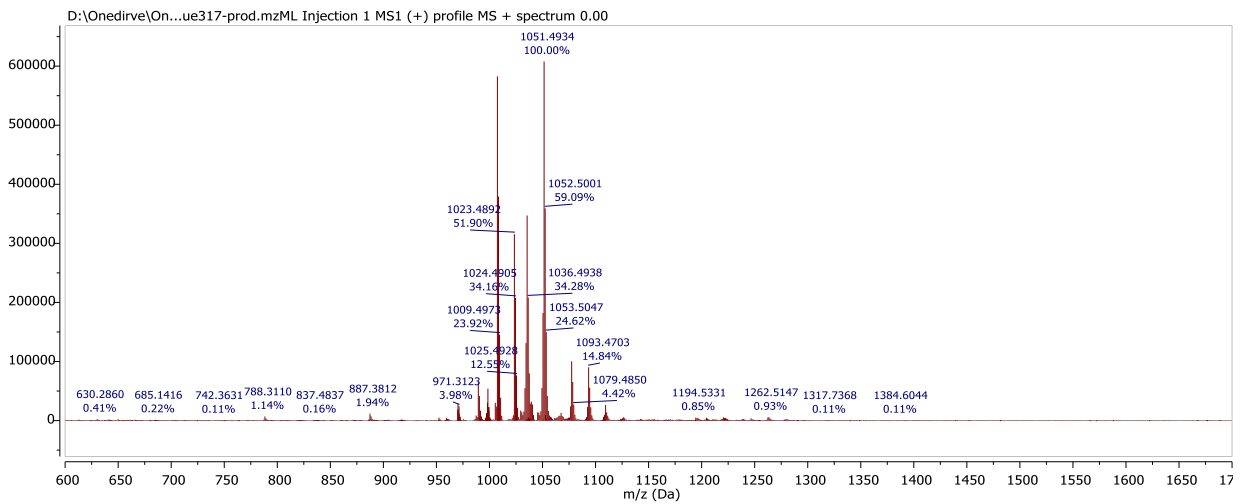


Chemical Formula: C₆₇H₆₅BF₂N₈O₇S
Exact Mass: 1054.4758

C:\Users\Yue W...d\Yue317-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram

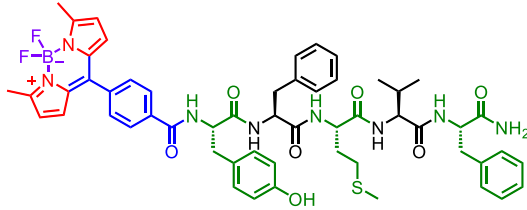


Analytic HPLC of BODIPY₁-Pep₁



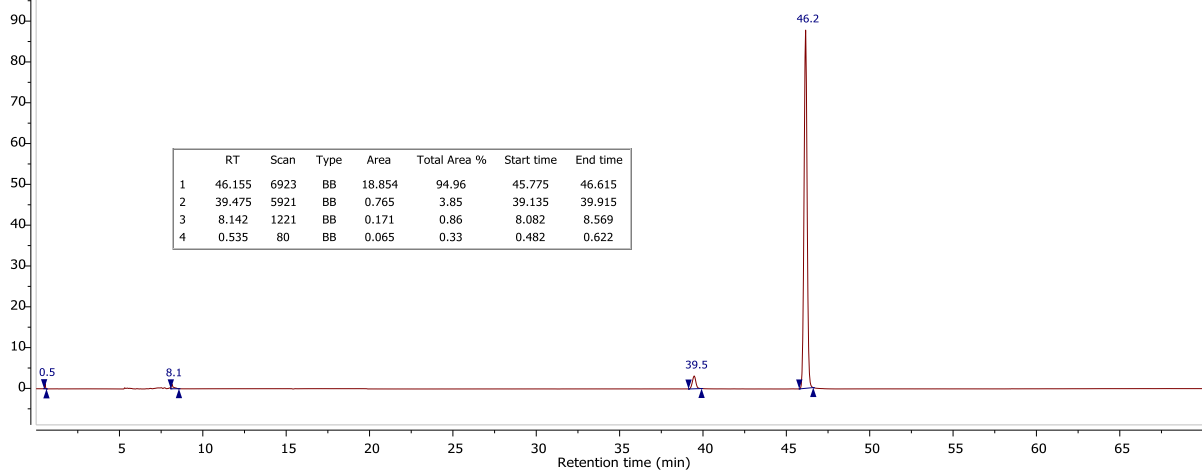
HRMS(MAIDL-TOF) of BODIPY₁-Pep₁

BODIPY₂-Pep₁: BODIPY₂-NH-YFMVF-CONH₂



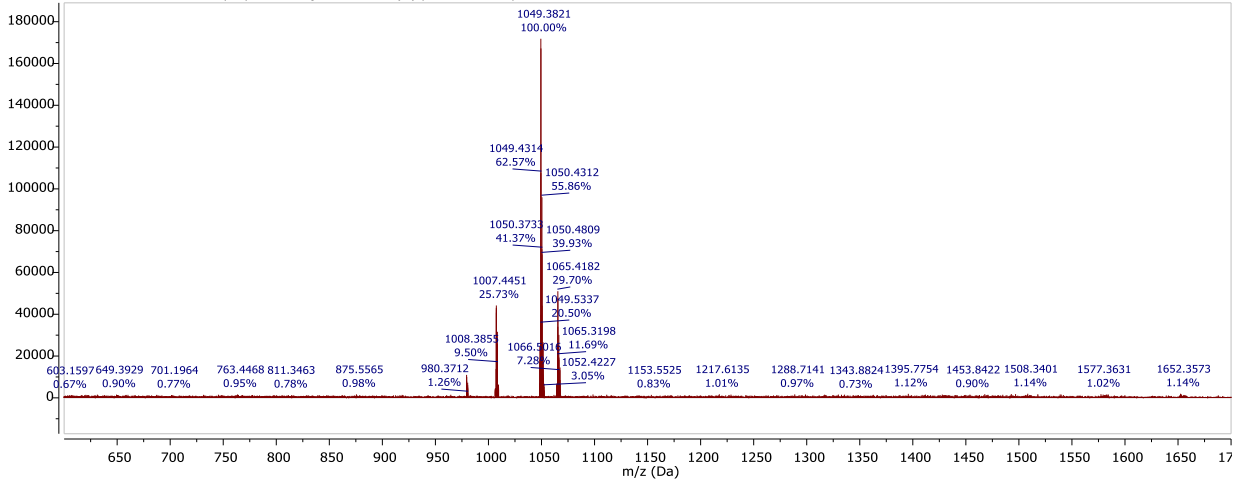
Chemical Formula: C₆₅H₆₁BF₂N₈O₇S
Exact Mass: 1026.4445

C:\Users\Yue W...YUE446-5-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram



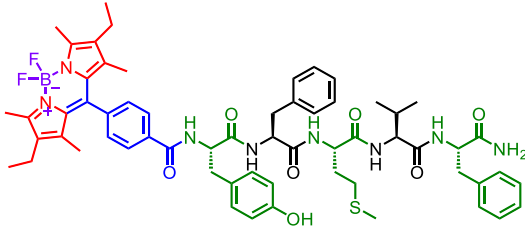
Analytic HPLC of BODIPY₂-Pep₁

C:\Users\Yue W...327-1-prep.mzML Injection 1 MS (+) profile MS + spectrum 0.00

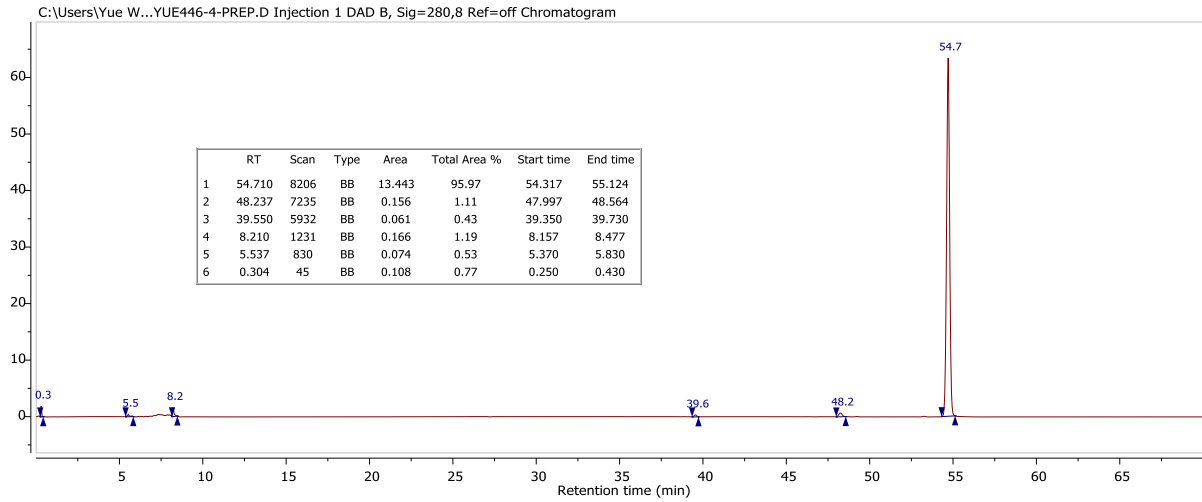


HRMS(MAIDL-TOF) of BODIPY₂-Pep₁

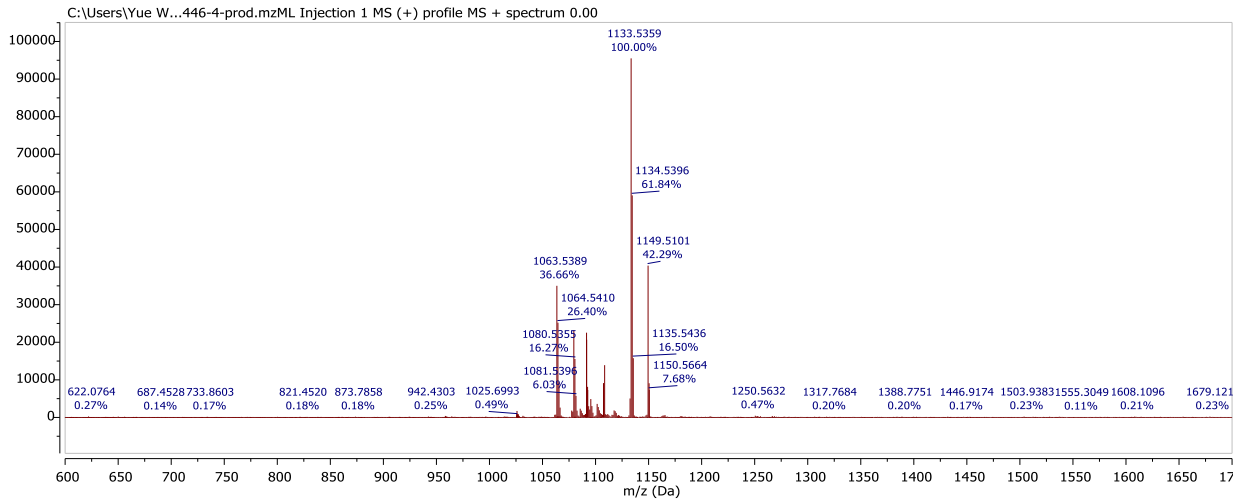
BODIPY₃-Pep₁: BODIPY₃-NH-YFMVF-CONH₂



Chemical Formula: C₆₁H₇₃BF₂N₈O₇S
Exact Mass: 1110.5384

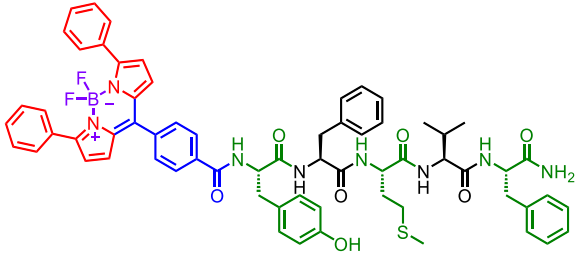


Analytic HPLC of BODIPY₃-Pep₁

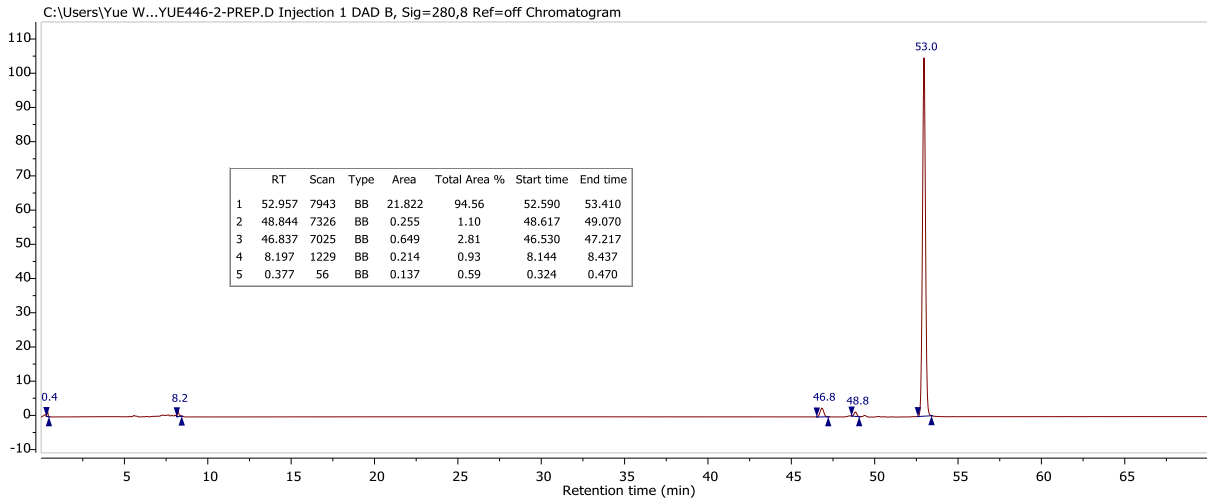


HRMS(MAIDL-TOF) of BODIPY₃-Pep₁

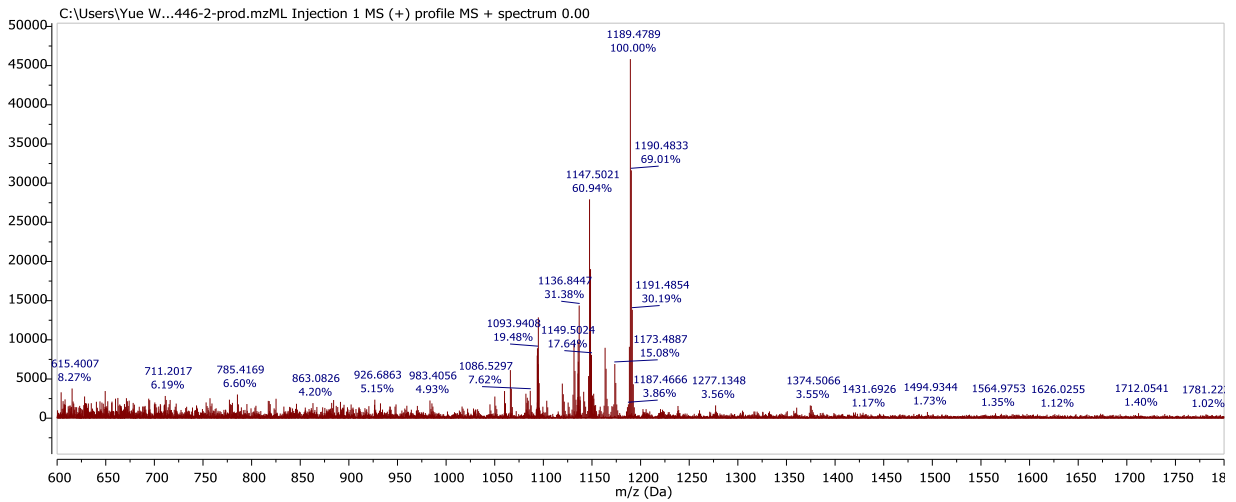
BODIPY₉-Pep₁: BODIPY₉-NH-YFMVF-CONH₂



Chemical Formula: C₆₅H₆₅BF₂N₃O₇S
Exact Mass: 1150.4758

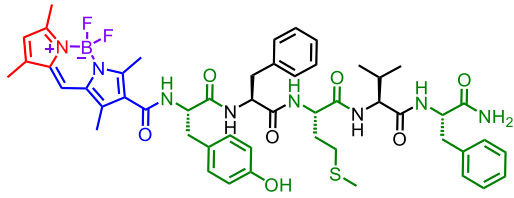


Analytic HPLC of BODIPY₉-Pep₁

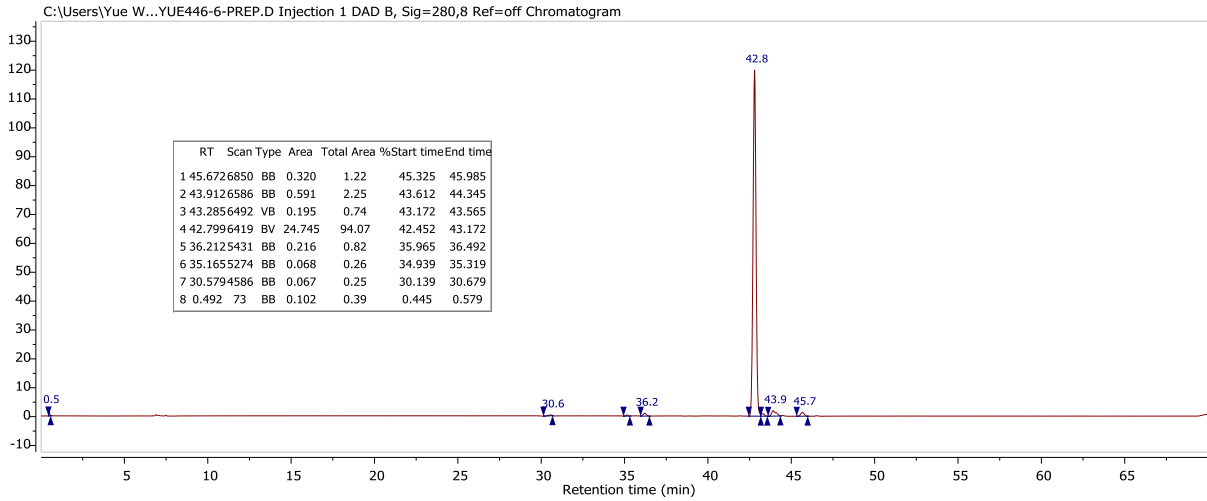


HRMS(MAIDL-TOF) of BODIPY₉-Pep₁

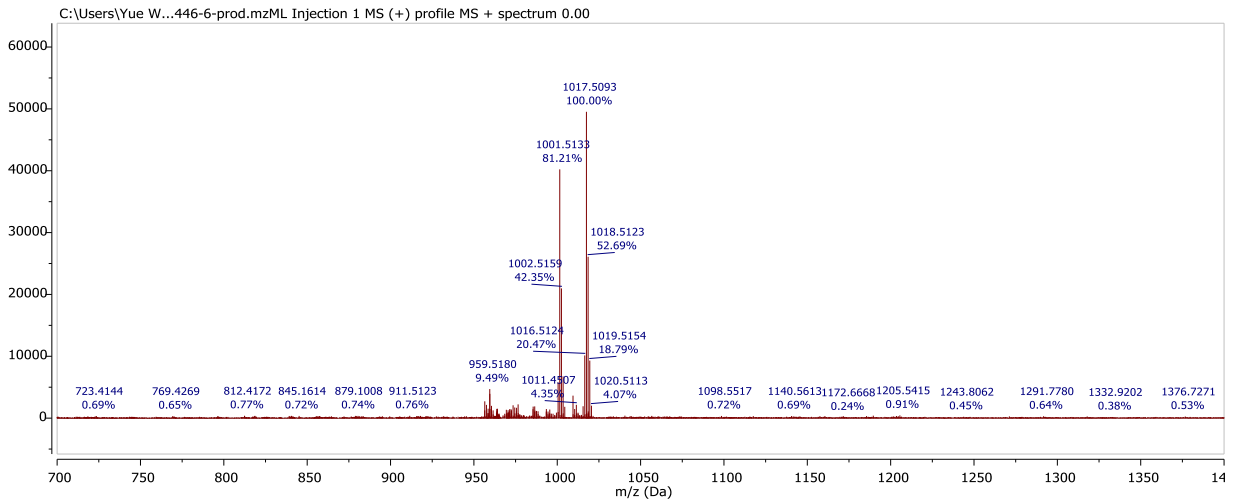
BODIPY₁₅-Pep₁: BODIPY₁₅-NH-YFMVF-CONH₂



Chemical Formula: C₅₁H₆₁BF₂N₈O₇S
Exact Mass: 978.4445

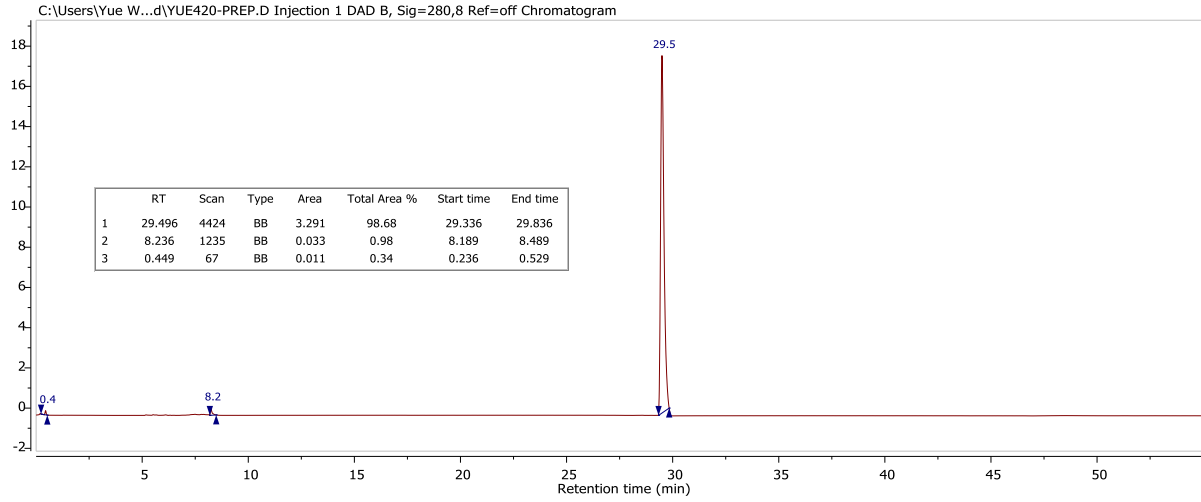
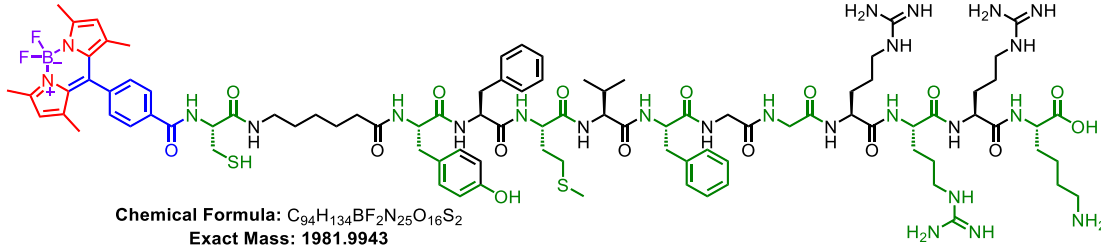


Analytic HPLC of BODIPY₁₅-Pep₁

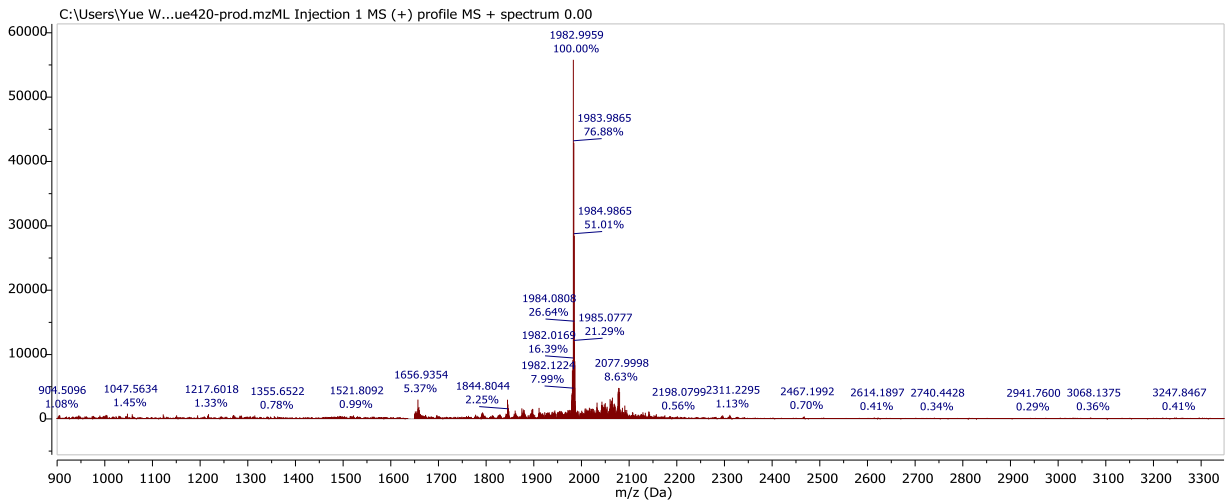


HRMS(MAIDL-TOF) of BODIPY₁₅-Pep₁

BODIPY₁-Pep₄: BODIPY₁-HN-C-Ahx-YFMVF-GG-RrRK-COOH

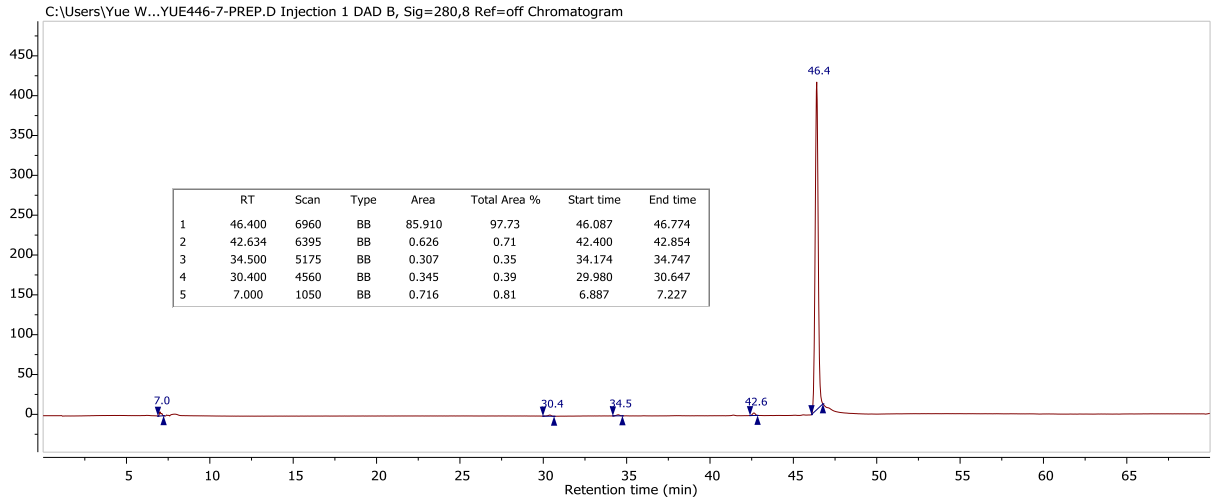
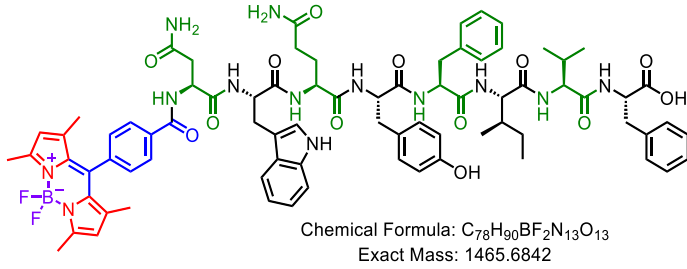


Analytic HPLC of **BODIPY₁-Pep₄**

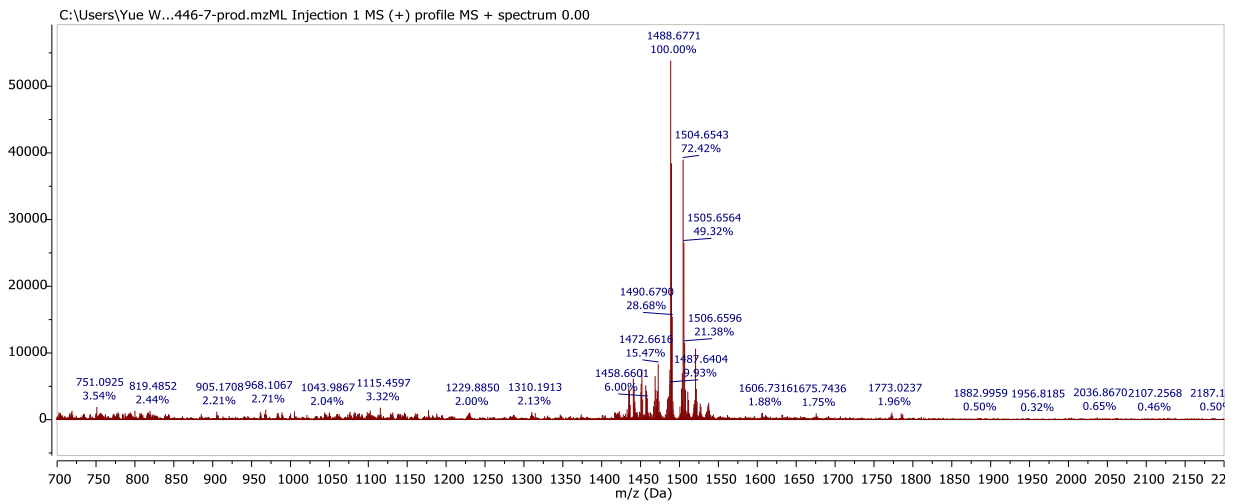


HRMS(MAIDL-TOF) of **BODIPY₁-Pep₄**

BODIPY₁-Pep₅: BODIPY₁-HN-NWQYFIVE-COOH

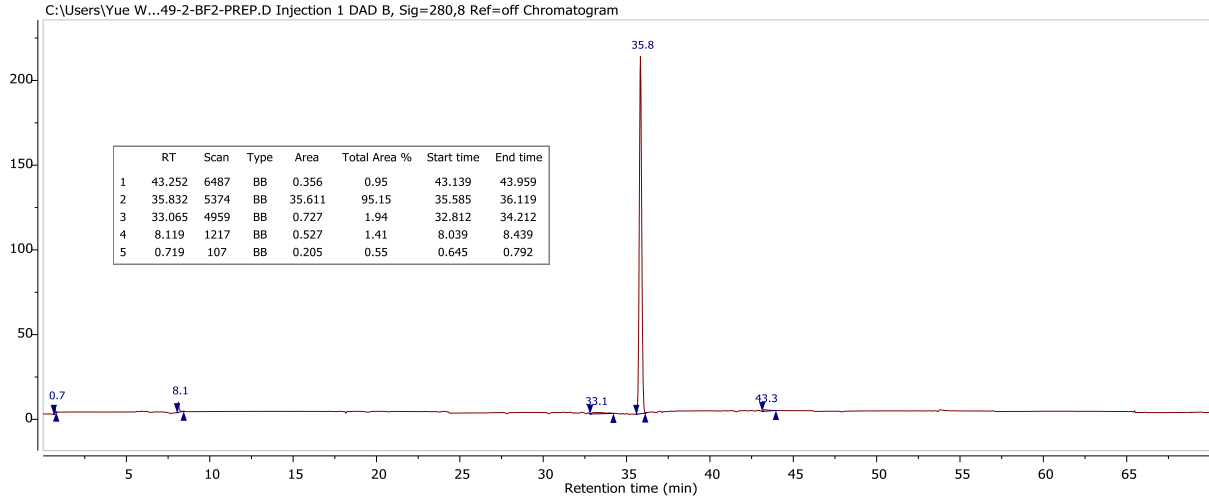
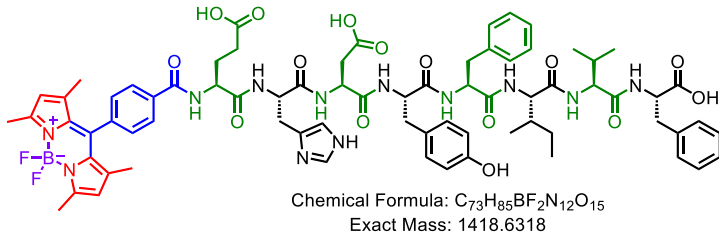


Analytic HPLC of BODIPY₁-Pep₅

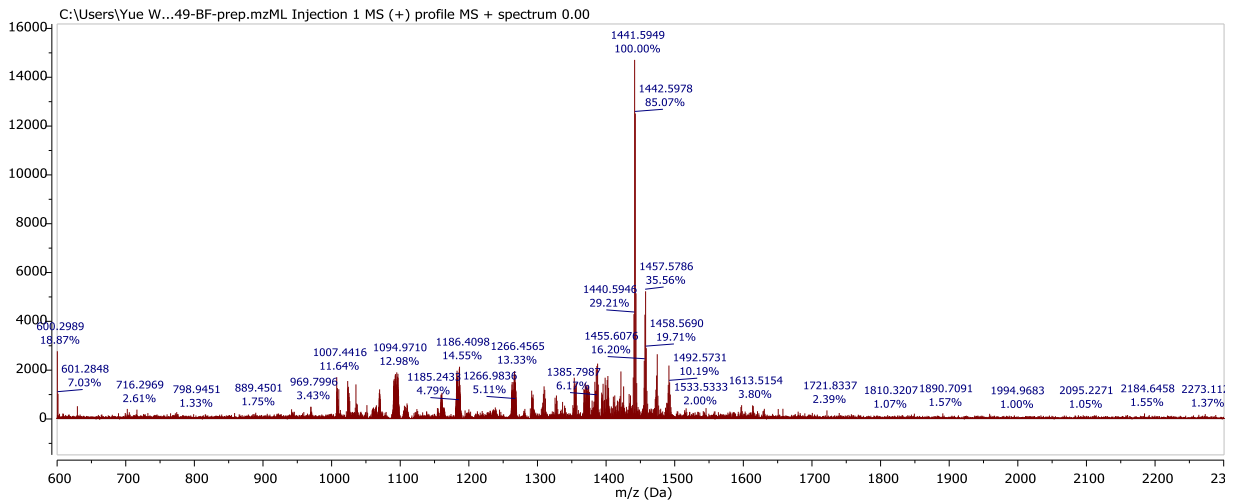


HRMS(MAIDL-TOF) of BODIPY₁-Pep₅

BODIPY₁-Pep₆: BODIPY₁-HN-EHDYFIVF-COOH

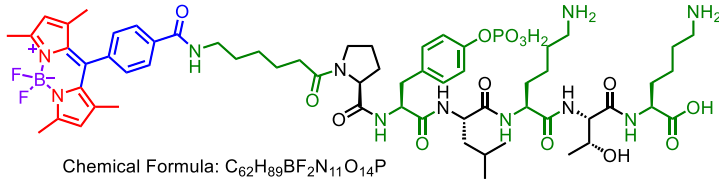


Analytic HPLC of BODIPY₁-Pep₆

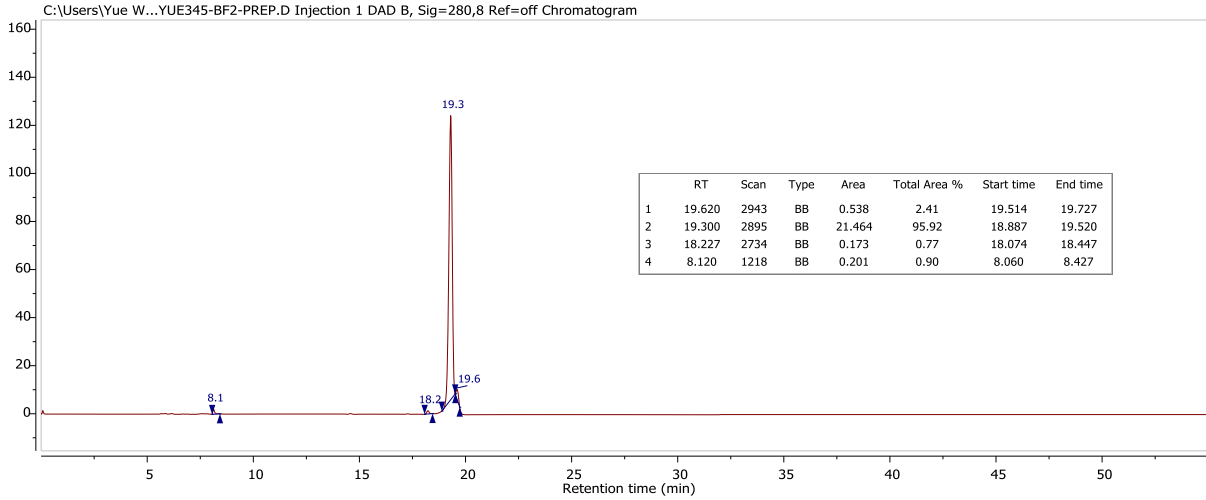


HRMS(MAIDL-TOF) of BODIPY₁-Pep₆

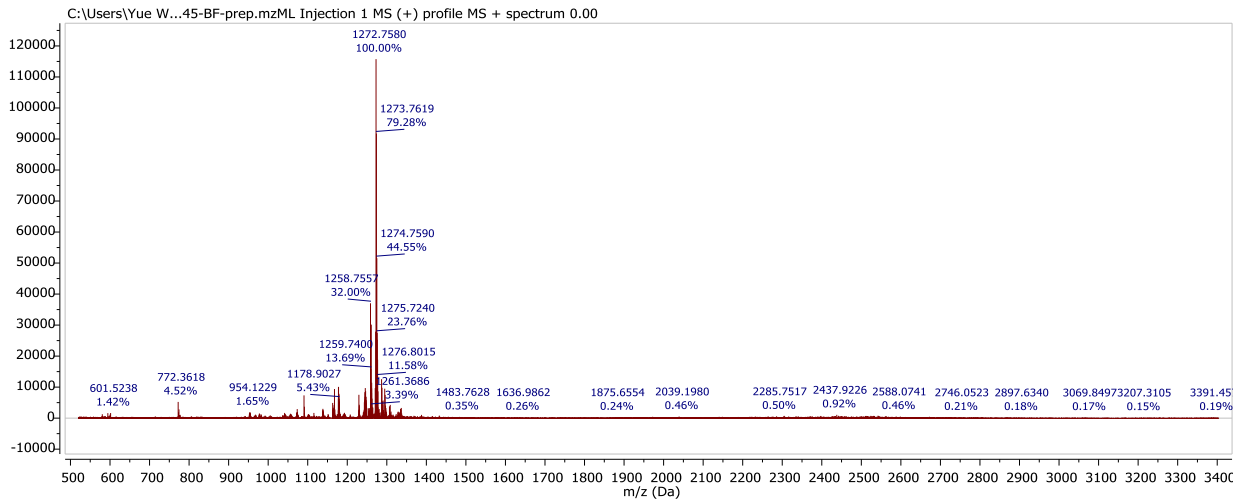
BODIPY₁-Pep₇: DP₁-HN-Ahx-P-(pTyr)-LKTK-COOH



Chemical Formula: C₆₂H₈₉BF₂N₁₁O₁₄P
 Exact Mass: 1291.6389

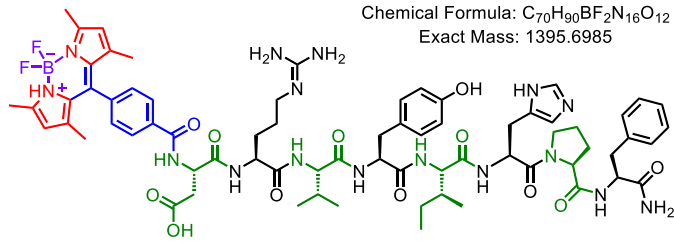


Analytic HPLC of BODIPY₁-Pep₇

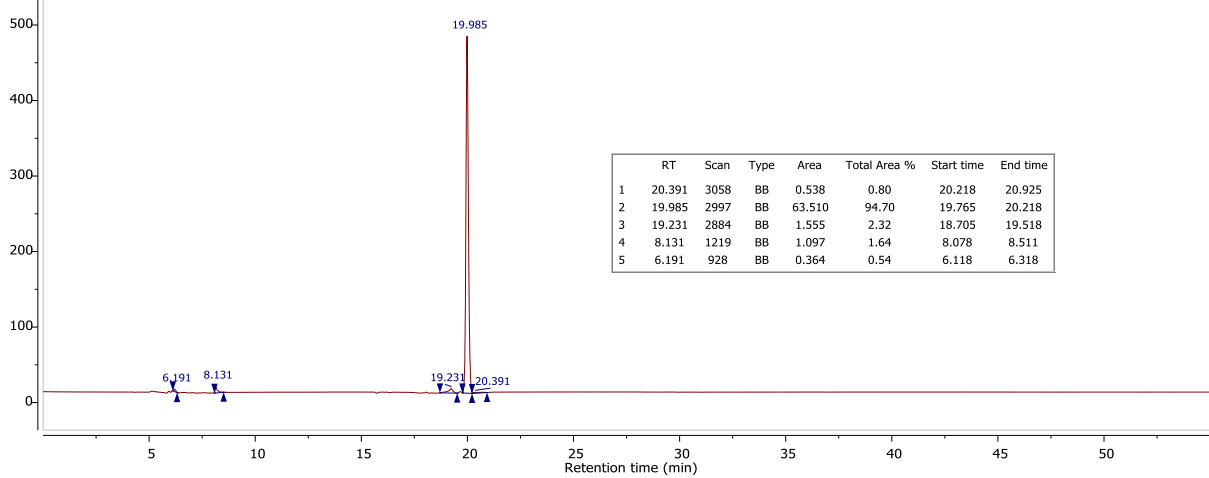


HRMS(MAIDL-TOF) of BODIPY₁-Pep₇

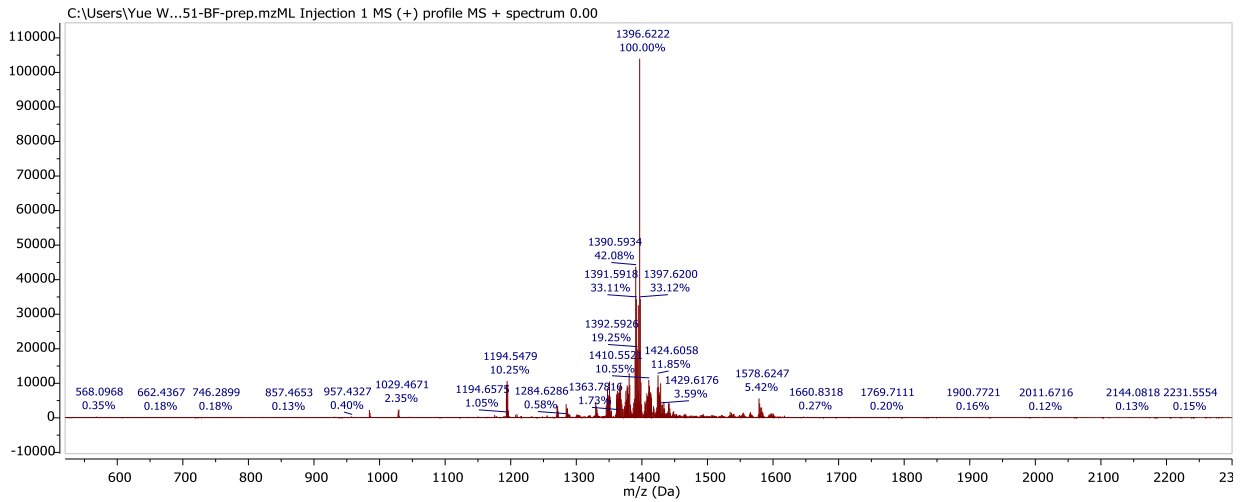
BODIPY₁-Pep₁₀: BODIPY₁-HN-DRVYIHPF-CONH₂



C:\Users\Yue W...UE451-BF-PREP.D Injection 1 DAD B, Sig=280,8 Ref=off Chromatogram



Analytic HPLC of BODIPY₁-Pep₁₀



HRMS(MAIDL-TOF) of BODIPY₁-Pep₁₀