Intrapulmonary administration of purified NEIL2 abrogates NF-κB-mediated inflammation

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List of Supporting Material

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Exon Primers	Sequence
mCxcll-exon-For	5' CCTATCGCCAATGAGCTGC 3'
mCxcl1-exon-Rev	5' ACTTGGGGACACCTTTTAGC 3'
mCxcl2-exon-For	5' CACCAACCACCAGGCTACA 3'
mCxcl2-exon-Rev	5' GGATGATTTTCTGAACCAGGGG 3'
mCxcl10-exon-For	5' GATCCCTCTCGCAAGGACG 3'
mCxcl10-exon-Rev	5' TCAACACGTGGGCAGGATAG 3'
mIl6-exon-For	5' GTCCTTCCTACCCCAATTTCCA 3'
mIl6-exon-Rev	5' AGGTTTGCCGAGTAGATCTCAA 3'
mTnfa-exon-For	5' AAATGGTTCGTACAGCAGCG 3'
mTnfa-exon-Rev	5' GCACGGAAGTAGTCTCCACG 3'
mIL1β-exon-For	5' CATCAGCACCTCACAAGCAG 3'
mIL1β-exon-Rev	5' TAAGTGGTTGCCCATCAGAGG 3'
mGapdh-exon-For	5' GTCACCAGGGCTGCCATTTG 3'
mGapdh-exon-Rev	5' GATGTTAGTGGGGTCTCGCT 3'
Promoter Primers	Sequences
mCxcl1-Pro-For	5' ATCCTTGGGAGTGGAGCAAG 3'
mCxcl1-Pro-Rev	5' GGAGTCTGGAGTGCTGGAAC 3'
mCxcl2-Pro-For	5' AGCGCAGACATCACTTCCTT 3'
mCxcl2-Pro-Rev	5' TGAAGTGTGGCTGGAGTCTG 3'
mCxcll0-Pro-For	5' GCTCACGCTTTGGAAAGTGAA 3'
mCxcl10-Pro-Rev	5' ATGTCTCTCAGCGGTGGATG 3'
mIl6-Pro-For	5' CCCCACCCTCCAACAAAGATT 3'
mIl6-Pro-Rev	5' CAGAGAGGAACTTCATAGCGGT 3'
mTNFa-Pro-For	5' TCTCAAGCTGCTCTGCCTTC 3'
mTNFa-Pro-Rev	5' GACCATGCCTGTGTCTATTTCC 3'

 Table S1. List of primers used in qPCR assays.

Table S2. List of the mouse DNA oligonucleotides used in EMSA analysis. For clarity, the NF- κ B motif sequences are shown in red and bases marked in blue represent the alterations in sequence to generate mutation.

0	DNA Digos	Label	Duplex Sequence
mC≯ mC M	xcll-WT Cxcll- Nutant	C1-WT (Cxcl1) C1-M	NF-KB Motif 5' GAAACACCCTGTACTCCGGGAATTTCCCTGGCCCGGAGCTCTG 3' 3' ATTTGTGGGACATGAGGCCCTTAAAGGGACCGGGCCTCGAGAC 5' 5' GAAACACCCTGTACTCCGAAAAGGTCCCTGGCCCGGAGCTCTG 3' 3' CTTTGTGGGACATGAGGCTTTTCCAGGGACCGGGGCTCGAGAC 5'
mC≯ m M	xcl2-WT MCxcl2- Mutant	C2-WT (Cxcl2) C2-M	NF-kB Motif 5' GGACCCTGAGCTCAGGGAATTTCCCTGGTCCCCGGGCT 3' 3' CCTGGGACTCGAGTCCCTTAAAGGGACCAGGGGGCCCGA 5' 5' GGACCCTGAGCTCAGAAAAGGTCCCTGGTCCACGGGCT 3' 3' CCTGGGACTCGAGTCTTTTCCAGGGACCAGGTGCCCGA 5'
m] n M	Il6-WT mIl6- Mutant	IL6-WT (Il6) IL6-M	NF-kB Motif 5' TTTTTATCAAATGTGGGGATTTTCCCATGAGTCTCAAAA 3' 3' AAAAATAGTTTACACCCTAAAAGGGTACTCAGAGTTTT 5' 5' TTGCTATTAAATGTGTTCGTTTGTACAGAGTCTCAAAA 3' 3' AACGATAATTTACACAAGCAAACATGTCTCAGAGTTTT 5'
mT	'nfα-WT	Tnfα	NF-κB Motif 5' AAGAACTCAAACAG <mark>GGGGGCTTTCC</mark> CTCCTCAATATCAT 3' 3' TTCTTGAGTTTGTC <mark>CCCCGAAAGG</mark> GAGGAGTTATAGTA 5'





(**A** and **B**) Real-Time quantitative reverse transcription polymerase chain reaction analysis with total RNA isolated from brain (A) and skeletal muscles (B) of mock- (lower panel) or TNF α -treated (upper panel) (intraperitoneal) *Neil2*^{+/+} or *Neil2*^{-/-} mice relative to mock-treated *Neil2*^{+/+} groups. Results are normalized to *Gapdh*. Error bars represent ± standard deviation from the mean. n=2 independent experiments from samples prepared by pooling tissues from n=3 mice per group. *=P<0.05; **=P<0.01; ***=P<0.005 vs. *Neil2*^{+/+} groups.

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Figure. S2. TNF α -induced inflammatory gene expression in the lungs of *Neil2^{-/-}* vs. *Neil2^{+/+}* mice. Area plot displays mRNA expression of *Cxcl1*, *Cxcl2*, *Cxcl10*, *Il6*, *Tnf* α and *Il1* β in *Neil2^{+/+}* or *Neil2^{-/-}* mice lung at 15-, 30-, 60- and 120-min post TNF α intranasal treatment as analyzed by Real-Time quantitative reverse transcription polymerase chain reaction analysis. Results are normalized to *Gapdh* expression and mock-treated *Neil2^{+/+}* groups; n=2 independent experiments, with data collected from n=4 mice per group; p-values for *Neil2^{-/-}* vs. *Neil2^{+/+}* groups are shown.



Figure. S3. Electrophoretic mobility-shift assays (EMSAs) of NF-κB's binding to its cognate motif derived from target gene promoters.

(**A-C**) EMSA using nuclear extracts (NEs) from mock- or TNFα-treated intranasally *Neil2*^{+/+} or *Neil2*^{-/-} male mice lung (pooled, n=3) with ³²P-labeled probe containing wild-type κ B-motif from mouse *Cxcl2*-(C2-WT, A and B) and *Il6*- promoter (IL6-WT, C); competition analysis in B, lanes 1-3, and in C, lanes 6-8 with 100-fold molar excess of wild-type or mutant competitor as indicated. (**D**) EMSAs of recombinant RelA protein complex using ³²P-labeled wild-type C1-WT, C2-WT and IL6-WT or mutant C1-M, C2-M and IL6-M NF- κ B-motifs; Lanes 1, 3, 5, 7, 9 and 11 show 'probe only' lanes. (**E**)

Immunoblot of NEIL2 in nuclear extracts (NEs) from untreated (-) or TNF α -treated (+) MLE12 cells transfected with control siRNA (si*Cont.*) or NEIL2-specific siRNA (si*Neil2*). HDAC2 was used as loading control; Quantification shows NEIL2 band intensities normalized to HDAC2 and represented relative to untreated *siCont.* group arbitrarily taken as 100. Representative data images from three independent experiments are shown. > and ^O denote RelA-DNA complexes; • denotes nonspecific complex; FP represents free probe. Error bars represent \pm standard deviation from the mean. n=3 independent experiments. ***=*P*<0.005 vs. *siCont.* groups.



Figure. S4. NEIL2 overexpression in human and mouse cell lines.

(A) Immunoblot of cell extracts from h358 cells expressing control (Empty)- or NEIL2-FLAG-vectors, using anti -FLAG or -Tubulin antibodies. (B) Immunoblot of cell extracts from untreated (-) or TNF α -treated (+) MLE12 cells transfected with control or NEIL2-FLAG vectors, using anti -FLAG or -GAPDH antibodies. Un-transfected/untreated MLE12 cell extract was loaded as negative control. Representative data images from two independent experiments are shown.



Figure. S5. Intrapulmonary rNEIL2 delivery in *Neil2^{+/+}* or *Neil2^{-/-}* male and female mice.

(A) Analysis of glycosylase activity of HI-rNEIL2 or Act-rNEIL2 with ³²P-labeled 5-hydroxyuracil DNAbubble substrate (51-mer). The substrate and the cleaved 25-mer product are indicated by arrows. (**B** and **C**) Immunoblots of whole cell extracts from -/+TNF α treated *Neil2*^{+/+} male (B) or female (C) mice lung, 72 h post-transduction with HI-rNEIL2 or Act-rNEIL2 using anti-His and GAPDH antibodies. Act-rNEIL2 protein (15 ng) was loaded as positive control. (**D**) Neutrophil counts post 16 h mock-challenge in broncho-alveolar lavage fluid of *Neil2*^{+/+}/*Neil2*^{-/-} male and female mice lung transduced with HI-rNEIL2 or Act-rNEIL2. Error bars represent ± standard deviation from the mean (n=3). ns= not significant; *=p <0.05; **=p <0.01; ***=p <0.005 vs. HI-rNEIL2 transduced group.



Figure. S6. Electrophoretic mobility-shift assays (EMSAs) with lung nuclear extract from *Neil2*^{+/+} or *Neil2*^{-/-} male mice.

Full size EMSA images for Figure 5E are displayed here. EMSAs of NF- κ B binding using nuclear extracts pooled from *Neil2*^{+/+}/*Neil2*^{-/-} male mice lung (n=3), transduced with HI-rNEIL2 or Act-rNEIL2 prior to

mock/TNF α -exposure, with radiolabeled probes derived from *Cxcl1*, *Cxcl2*, *IL6* and *Tnf\alpha* promoters as indicated on the top; Representatives of three independent gels are shown; > and ^O denote ReIA-DNA complexes; • denotes nonspecific complex; FP represents free probe. lane 5: probe only.



Figure. S7. Electrophoretic mobility-shift assays (EMSAs) with lung nuclear extract from *Neil2*^{+/+} or *Neil2*^{-/-} female mice.

EMSA of NF- κ B binding using nuclear extracts pooled from *Neil2*^{+/+}/*Neil2*^{-/-} female mice lung (n=3), transduced with HI-rNEIL2 or Act-rNEIL2 prior to mock/TNF α -exposure, with radiolabeled probes

derived from *Cxcl1*, *Cxcl2*, *IL6* and *Tnfa* promoters as indicated on the top; Representatives of three independent gels are shown; > and ^O denote RelA-DNA complexes; • denotes nonspecific complex; FP represents free probe. lane 5: probe only.