

**Abscisic acid implicated in differential plant responses of
Phaseolus vulgaris during endophytic colonization by
Metarhizium and pathogenic colonization by *Fusarium***

Shasha Hu and Michael J. Bidochka*

Department of Biological Sciences, Brock University, St. Catharines, ON Canada L2S

3A1

*Address correspondence to

Email: mbidochka@brocku.ca

Telephone: +1 905-688-5550

Supplementary Information

Supplementary Figure S1. The concentration of important plant hormones during endophytic or pathogenic colonization.

Supplementary Figure S2. Colony morphology and conidial production of *M. robertsii* and *F. solani* under different amount of ABA.

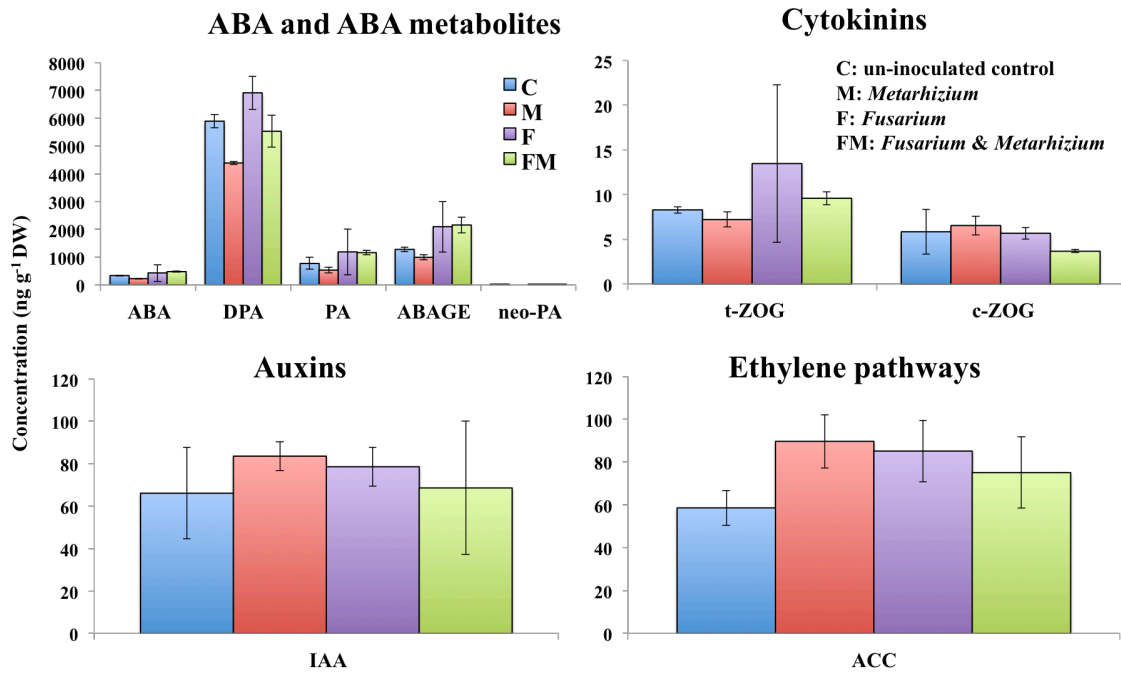
Supplementary Figure S3. Analysis of stomatal aperture during the exogenous application of ABA on bean leaves.

Supplementary Figure S4. Relative expression of genes involved in ABA biosynthetic pathway in bean leaves and roots.

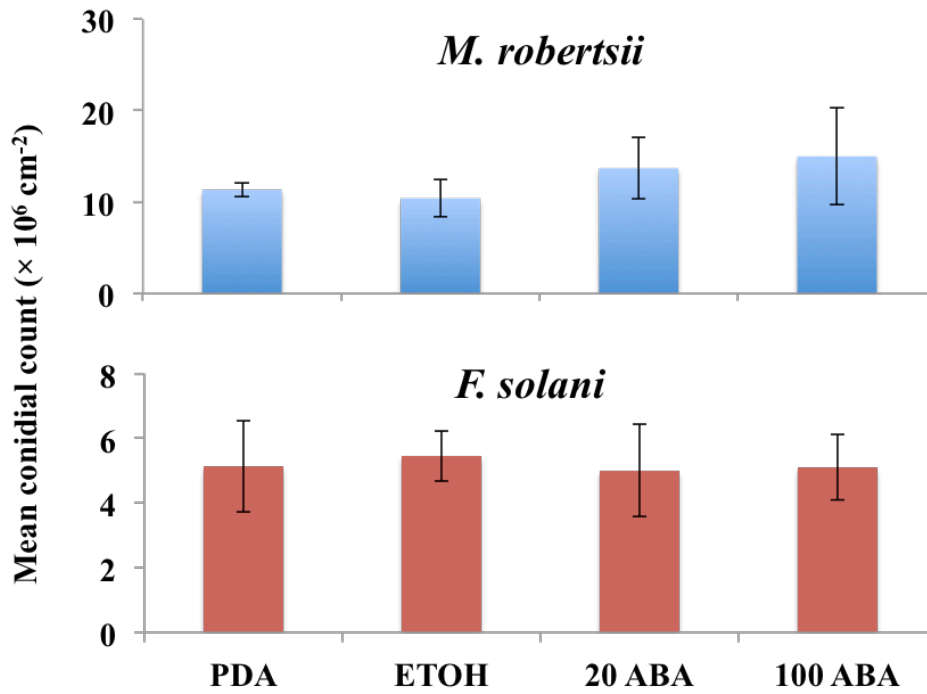
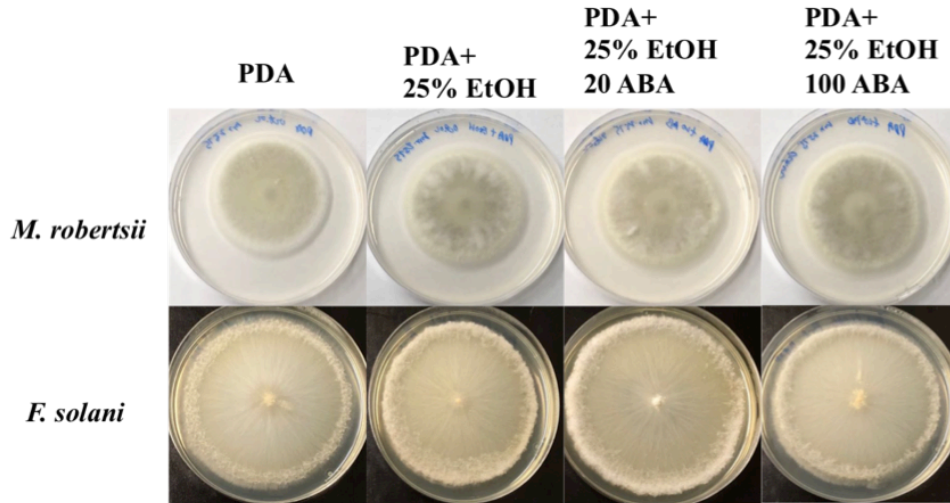
Supplementary Figure S5. CFU of *M. robertsii* or *F. solani* in bean root homogenate after exogenous application of 25% ethanol.

Supplementary Table S1. Plant hormones tested in this study

Supplementary Table S2. Primer sequences of *Phaseolus vulgaris* used for RT-PCR analysis.

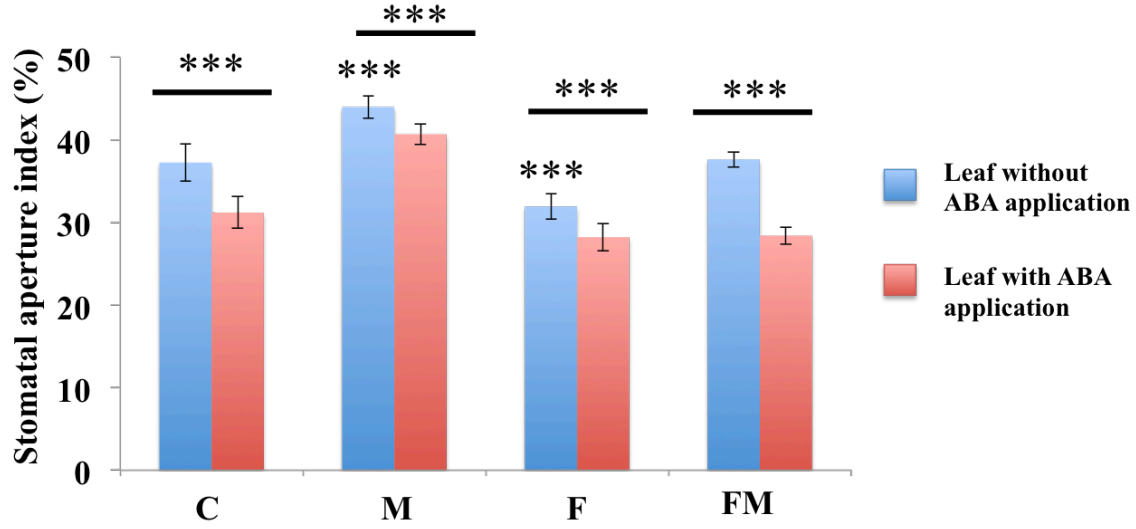


Supplementary Figure S1. The concentration of important plant hormones during endophytic or pathogenic colonization. *cis*-Abscisic acid (ABA), Dihydrophaseic acid (DPA), Phaseic acid (PA), Abscisic acid glucose ester (ABAGE), neo-PA (neo-Phaseic acid), (*trans*) zeatin-O-glucoside (*t*-ZOG), (*cis*) zeatin-O-glucoside (*c*-ZOG), indole-3-acetic acid (IAA), 1-aminocyclopropane-1-carboxylic acid (ACC).



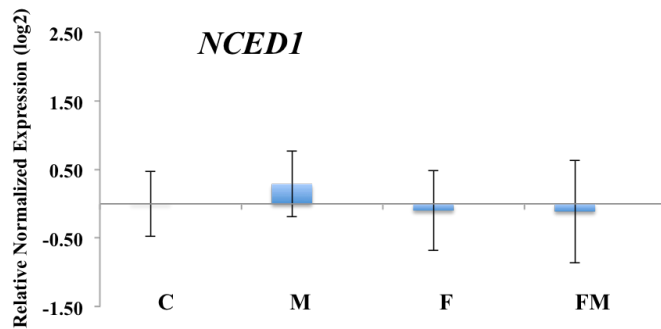
Supplementary Figure S2. Colony morphology and conidial production of *M. robertsii* and *F. solani* under different amount of ABA.

ABA, abscisic acid. X-axis represents values of ABA μg per PDA plate.

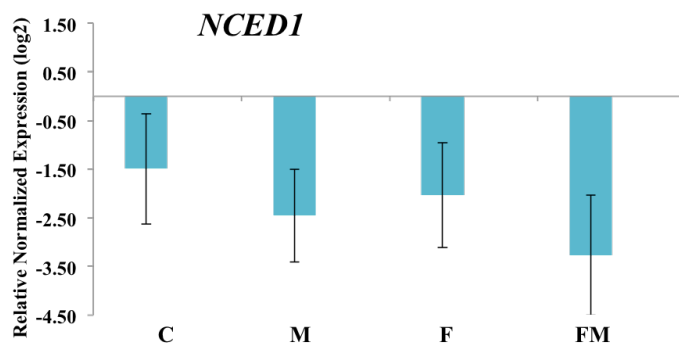


Supplementary Figure S3. Analysis of stomatal aperture during the exogenous application of ABA on bean leaves. The stomatal aperture index in the bean leaves after 14 days of un-inoculated control (C) and bean colonized by *M. robertsii* (M), *F. solani* (F), and *M. robertsii* together with *F. solani* (FM). Standard deviations are shown. Data were analyzed by one-way ANOVA with Fisher's least-significant difference (LSD) test. Statistical differences shown; * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$. Asterisks alone indicate statistically significant differences relative to un-inoculated control. Asterisk above black line indicate the significant differences of stomatal aperture index with exogenous ABA application compared to that in corresponding plant without ABA application.

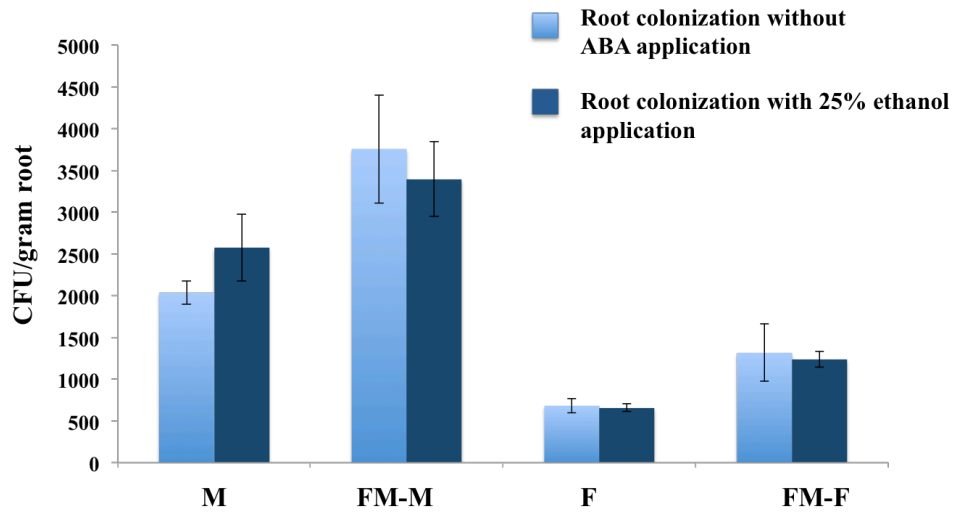
Gene Expression in Bean Leaves



Gene Expression in Bean Roots



Supplementary Figure S4. Relative expression of genes involved in ABA biosynthetic pathway in bean leaves and roots. RNA was extracted at 14 days bean leaf and root in un-inoculated control (C), colonized by *M. robertsii* (M), *F. solani* (F), and *M. robertsii* together with *F. solani* (FM). For each gene, the expression level in un-inoculated control bean leaf was set to 1. Standard errors are shown. Figure Data were analyzed with standard t-test in Bio-Rad CFX Manager software. No statistical differences are shown nine-*cis* epoxy-carotenoid dioxygenases 1(NCED1).



Supplementary Figure S5. CFU of *M. robertsii* or *F. solani* in bean root homogenate after exogenous application of 25% ethanol. M: *M. robertsii*, F: *F. solani*, FM: *F. solani* and *M. robertsii* treatment group. Standard error bars are shown.

Supplementary Table S1 Plant hormones tested in this study

Group	Abbreviation	Full Name
ABA and metabolites	ABA	<i>cis</i> -Abscisic acid
	ABAGE	Abscisic acid glucose ester
	DPA	Dihydrophaseic acid
	PA	Phaseic acid
	7'OH-ABA	7'-Hydroxy-abscisic acid
	neo-PA	<i>neo</i> -Phaseic acid
	<i>t</i> -ABA	<i>trans</i> -Abscisic acid
Auxins	IAA	Indole-3-acetic acid
	IAA-Asp	N-(Indole-3-yl-acetyl)-aspartic acid
	IAA-Glu	N-(Indole-3-yl-acetyl)-glutamic acid
	IAA-Ala	N-(Indole-3-yl-acetyl)-alanine
	IAA-Leu	N-(Indole-3-yl-acetyl)-leucine
	IBA	Indole-3-butyric acid
Cytokinins	<i>t</i> -ZOG	(<i>trans</i>) Zeatin-O-glucoside
	<i>c</i> -ZOG	(<i>cis</i>) Zeatin-O-glucoside
	<i>t</i> -Z	(<i>trans</i>) Zeatin
	<i>c</i> -Z	(<i>cis</i>) Zeatin
	dhZ	Dihydrozeatin
	<i>t</i> -ZR	(<i>trans</i>) Zeatin riboside
	<i>c</i> -ZR	(<i>cis</i>) Zeatin riboside
	dhZR	Dihydrozeatin riboside
	iP	Isopentenyladenine
	iPR	Isopentenyladenosine
	KIN	Kinetin
	Ethylene Gibberellins	ACC
GA1		Gibberellin 1
GA3		Gibberellin 3
GA4		Gibberellin 4
GA7		Gibberellin 7
GA8		Gibberellin 8
GA9		Gibberellin 9
GA19		Gibberellin 19
GA20		Gibberellin 20
GA24		Gibberellin 24
GA29		Gibberellin 29
GA34		Gibberellin 34
GA44		Gibberellin 44
GA51		Gibberellin 51
GA53		Gibberellin 53

Supplementary Table S2 Primers sequences of *Phaseolus vulgaris* used for RT-PCR

analysis

Gene	NCBI ID	Sequence	Size (bp)	Forward Sequence	Reverse Sequence
<i>Act</i>	XM_007162263		126	CACCGAGGCACCGC TTAATC	CGGCCACTAGCGTA AAGGGAA
<i>ERF1</i>	XM_007144028		161	CGCTCTCAAGAGGA AACACTCC	TGAATCAGAAGGAG GAGGGAAAT
<i>ERF5</i>	XM_007157197		151	GGCTCCAAGTGGAT TGAGAAC	TCAGAATCAGATAA CTACAAAGCACAA
<i>HPL</i>	XM_007149930		153	TCAAGGCTACATTT GTATTTCCA	TGGTGCACATTTCTT AGTAGCAA
<i>PR1</i>	XM_007154263		98	TGGTCCTAACGGAG GATCAC	TGGCTTTTCCAGCT TTGAGT
<i>PR2</i>	XM_007154264		246	GTGAAGGACGCCGA TAACAT	ACTGAGTTTGGGGT CGATTG
<i>PR3</i>	XM_007137247		111	TGGAGTTGGTTATG GCAACAA	ATTCTGATGGGATG GCAGTGT
<i>PR4</i>	XM_007147114		140	CGCAGTGAGTG CAT ATTGCT	TGGTGCACATTTCTT AGTAGCAA
<i>NCED1</i>	AF190462		162	CTCCTTTTCTACGCT CGCAG	GGTGGTTAAGTCGC CGTTAG
<i>PvCYP707A1</i>	DQ352541		111	GCCTTTTGGCAGTG GGATTCAT	GCACCCACAACAG ACCACCT
<i>PvCYP707A3</i>	DQ352543		121	AGACTACCACACTC GCCTCA	GCCATCACCCCAAG AGTTCA