

# The SARS-unique domain (SUD) of SARS-CoV and SARS-CoV-2 interacts with human Paip1 to enhance viral RNA translation

Jian Lei, Yue Ma-Lauer, Yinze Han, Matthias Thoms, Robert Buschauer, Joerg Jores, Volker Thiel, Roland Beckmann, Wen Deng, Heinrich Leonhardt, Rolf Hilgenfeld, and Albrecht von Brunn **DOI: 10.15252/embj.2019102277** 

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Corresponding Author Name: Albrecht von Brunn, Rolf Hilgenfeld

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#### porting Checklist For Life Sciences Articles (Rev. June 2017)

This checklist is used to ensure good reporting standards and to improve the reproducibility of published results. These guidelines are consistent with the Principles and Guidelines for Reporting Preclinical Research issued by the NIH in 2014. Please follow the journal's authorship guidelines in preparing your manuscript.

#### A- Figures

#### 1. Data

#### The data shown in figures should satisfy the following conditions:

- the data were obtained and processed according to the field's best practice and are presented to reflect the results of the experiments in an accurate and unbiased manner.
   figure panels include only data points, measurements or observations that can be compared to each other in a scientifically
- meaningful way.

  graphs include clearly labeled error bars for independent experiments and sample sizes. Unless justified, error bars should not be shown for technical replicates.
- → if n< 5, the individual data points from each experiment should be plotted and any statistical test employed should be
- justified Source Data should be included to report the data underlying graphs. Please follow the guidelines set out in the author ship guidelines on Data Presentation.

#### 2. Captions

#### Each figure caption should contain the following information, for each panel where they are relevant:

- a specification of the experimental system investigated (eg cell line, species name).
   the assay(s) and method(s) used to carry out the reported observations and measurements
   an explicit mention of the biological and chemical entity(ies) that are being measured.
- an explicit mention of the biological and chemical entity(ies) that are altered/varied/perturbed in a controlled manner.
- → the exact sample size (n) for each experimental group/condition, given as a number, not a range;
   → a description of the sample collection allowing the reader to understand whether the samples represent technical or biological replicates (including how many animals, litters, cultures, etc.).
   → a statement of how many times the experiment shown was independently replicated in the laboratory.
   → definitions of statistical methods and measures:
   common tests, such as t-test (please specify whether paired vs. unpaired), simple x2 tests, Wilcoxon and Mann-Whitney

- - tests, can be unambiguously identified by name only, but more complex techniques should be described in the methods
  - · are tests one-sided or two-sided?

  - are there adjustments for multiple comparisons?
    exact statistical test results, e.g., P values = x but not P values < x;

    deficition of (authors plant a profile to the pr
  - definition of 'center values' as median or average;
  - · definition of error bars as s.d. or s.e.m.

Any descriptions too long for the figure legend should be included in the methods section and/or with the source data.

n the pink boxes below, please ensure that the answers to the following questions are reported in the manuscript itsel ncourage you to include a specific subsection in the methods section for statistics, reagents, animal models and hi

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# **B- Statistics and general methods**

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1.a. How was the sample size chosen to ensure adequate power to detect a pre-specified effect size?	NA .
1.b. For animal studies, include a statement about sample size estimate even if no statistical methods were used.	NA .
2. Describe inclusion/exclusion criteria if samples or animals were excluded from the analysis. Were the criteria pre- established?	NA .
3. Were any steps taken to minimize the effects of subjective bias when allocating animals/samples to treatment (e.g. randomization procedure)? If yes, please describe.	NA .
For animal studies, include a statement about randomization even if no randomization was used.	NA .
4.a. Were any steps taken to minimize the effects of subjective bias during group allocation or/and when assessing results (e.g. blinding of the investigator)? If yes please describe.	NA .
4.b. For animal studies, include a statement about blinding even if no blinding was done	NA .
5. For every figure, are statistical tests justified as appropriate?	Yes
Do the data meet the assumptions of the tests (e.g., normal distribution)? Describe any methods used to assess it.	Student's T-Test, Excel
Is there an estimate of variation within each group of data?	Standard deviation

Is the variance similar between the groups that are being statistically compared?  6. To show that artibodies were profiled for use in the system under study (assay and species), provide a citation, catalog rambur analyse cione number, supplementary information or reference to an antibody validation profile e.g.,  7. Infortify the source of cell lines and export if they were exceedly authericated (e.g., by STR profiling) and tested for mycoplasma contamination.  * for all hyperinks, please see the table at the top right of the document.  **Imal Models  5. Report species, strain, gender, age of animals and genetic modification status where applicable. Please detail housing and husbandry conditions and the source of animals.  9. For experiments involving live vertebrates, include a statement of compliance with ethical regulations and identify the committee(s) approving the experiments.  10. We recommend consulting the ARRVIT guidelines [see link list at top right] Place Biols. 8(d), e1000412, 2010) to ensure that other relevant aspects of animal studies are adequately reported. See author guidelines, under Reporting Guidelines. See blow little link list at top right] and MMC (bee link list at top right) recommendations. Please confirm (compliance.  11. Moretify the committee(s) approving the study protocol.  12. Include a statement confirming that informed consent was obtained from all subjects and that the experiments confirmed to the principles set on in the WMA Decisionation of Helinia and the Department of Intellin and Human dates or samples.  13. Report the clinical first registration on number (of CinicalTrials gover experiments), where applicable.  14. Report any restrictions on the availability (antifor on the use of human data or samples.  15. Report the clinical first registration number (of CinicalTrials gover experiment), where applicable.		
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# F- Data Accessibility

18: Provide a "Data Availability" section at the end of the Materials & Methods, listing the accession codes for data	Data availability:
generated in this study and deposited in a public database (e.g. RNA-Seq data: Gene Expression Omnibus GSE39462,	b. Atomic coordinates and structure factors for the Mac2:Paip1M complex have been deposited in
Proteomics data: PRIDE PXD000208 etc.) Please refer to our author guidelines for 'Data Deposition'.	the Protein Data Bank with accession code 6YXJ.
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c. Crystallographic data for small molecules	
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ethical obligations to the patients and relevant medical and legal issues. If practically possible and compatible with the	
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controlled repositories such as dbGAP (see link list at top right) or EGA (see link list at top right).	
21. Computational models that are central and integral to a study should be shared without restrictions and provided in a	NA
machine-readable form. The relevant accession numbers or links should be provided. When possible, standardized forma	
(SBML, CellML) should be used instead of scripts (e.g. MATLAB). Authors are strongly encouraged to follow the MIRIAM	
guidelines (see link list at top right) and deposit their model in a public database such as Biomodels (see link list at top	
right) or JWS Online (see link list at top right). If computer source code is provided with the paper, it should be deposited	
in a public repository or included in supplementary information.	

16. For phase II and III randomized controlled trials, please refer to the CONSORT flow diagram (see link list at top right) and submit the CONSORT checklist (see link list at top right) with your submission. See author guidelines, under 'Reporting Guidelines'. Please confirm you have submitted this list.

17. For tumor marker prognostic studies, we recommend that you follow the REMARK reporting guidelines (see link list at top right). See author guidelines, under 'Reporting Guidelines'. Please confirm you have followed these guidelines.

# G- Dual use research of concern

22. Could your study fall under dual use research restrictions? Please check biosecurity documents (see link list at top	NA
right) and list of select agents and toxins (APHIS/CDC) (see link list at top right). According to our biosecurity guidelines,	
provide a statement only if it could.	