

Supplemental Files

A spontaneous multifunctional hydrogel vaccine amplifies the innate immune response to launch a powerful antitumor adaptive immune response

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Table S1. Antibodies list used in flow cytometric analysis

| Antibody | Clone | Supplier |
|-----------------------------|-----------|-----------|
| PE anti-mouse CD80 | 16-10A1 | BioLegend |
| PE-Cy7 anti-mouse CD83 | Michel-19 | BioLegend |
| FITC anti-mouse CD86 | GL-1 | BioLegend |
| FITC anti-mouse CD80 | 16-10A1 | BioLegend |
| APC anti-mouse CD86 | GL-1 | BioLegend |
| APC anti-mouse CD40 | 3/23 | BioLegend |
| APC anti-mouse CD206 | C068C2 | BioLegend |
| FITC anti-mouse CD8a | 53-6.7 | BioLegend |
| PE anti-mouse IFN- γ | XMG1.2 | BioLegend |
| PE-Cy7 anti-mouse CD3 | 17A2 | BioLegend |
| FITC anti-mouse CD4 | GK1.5 | BioLegend |
| APC anti-mouse CD8a | 53-6.7 | BioLegend |
| PE anti-mouse CD11c | N418 | BioLegend |
| FITC anti-mouse CD45 | 30-F11 | BioLegend |
| FITC anti-mouse CD11b | M1/70 | BioLegend |
| PE anti-mouse F4/80 | BM8 | BioLegend |
| PE-Cy7 anti-mouse CD86 | GL-1 | BioLegend |
| PE anti-mouse CD4 | GK1.5 | BioLegend |
| FITC anti-mouse CD25 | PC61.5 | BioLegend |

| | | |
|-------------------------|---------|-------------|
| PE-Cy5 anti-mouse Foxp3 | FJK-16s | eBioscience |
| APC anti-mouse Gr-1 | RB6-8C5 | BioLegend |
| APC anti-mouse CD103 | 2E7 | BioLegend |
| APC anti-mouse CD8a | 53-6.7 | BioLegend |

Table S2. The grafting ratio of CpG on NOCC

| Sample ID | NOCC concentration ($\mu\text{g/mL}$) | Detecting concentration of CpG ($\mu\text{g/mL}$) | Theoretical concentration of CpG ($\mu\text{g/mL}$) |
|-----------|---|---|---|
| S1 | 6000 | 47.179 | 48 |
| S2 | 6000 | 47.952 | 48 |
| S3 | 6000 | 49.014 | 48 |

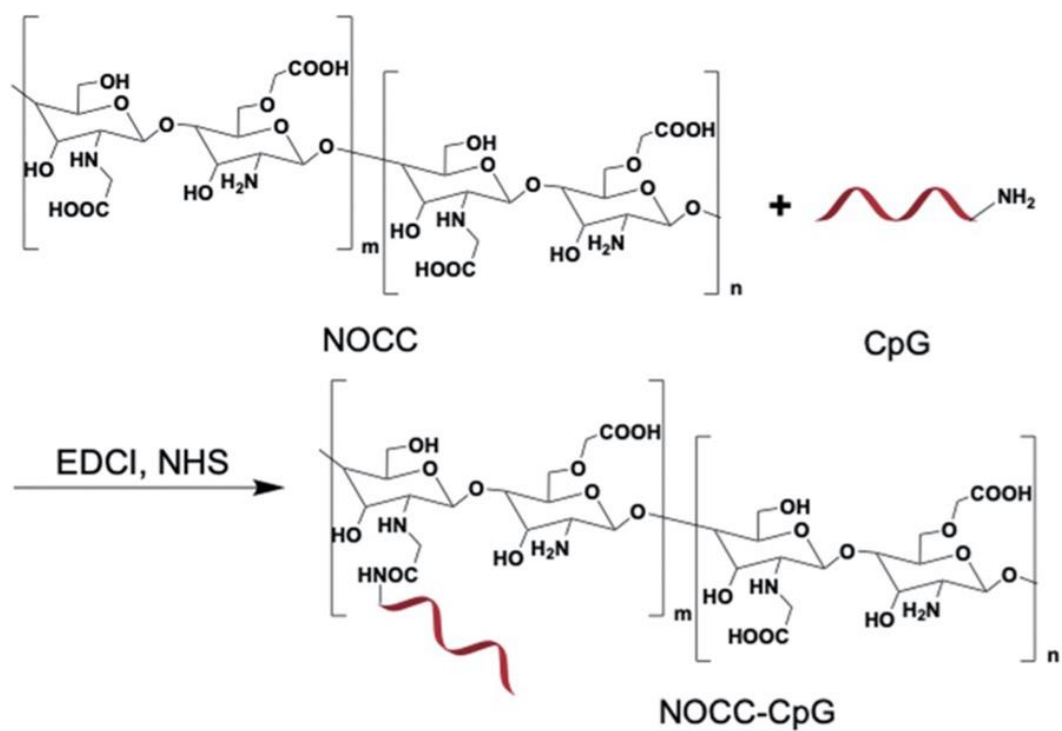


Figure S1. The scheme of synthesizing NOCC-CpG.

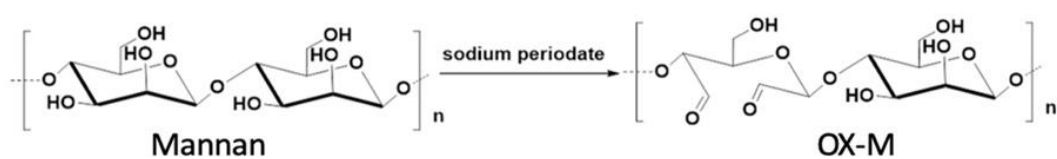


Figure S2. The scheme of synthesizing OX-M.

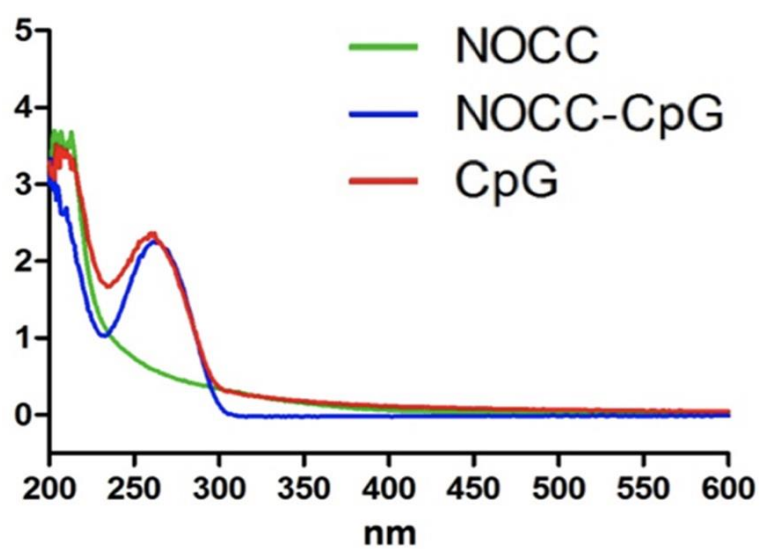


Figure S3. UV-Vis absorption spectra of CpG, NOCC, and NOCC-CpG.

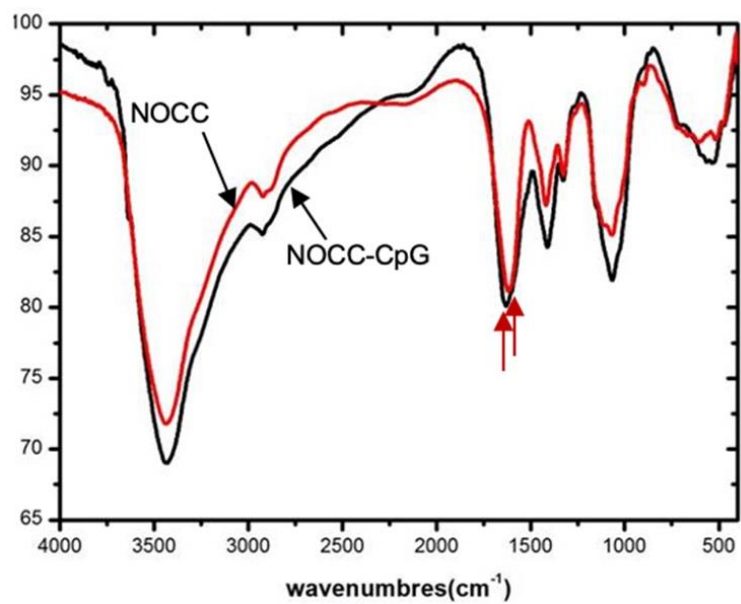


Figure S4. FT-IR spectra of the NOCC and NOCC-CpG.

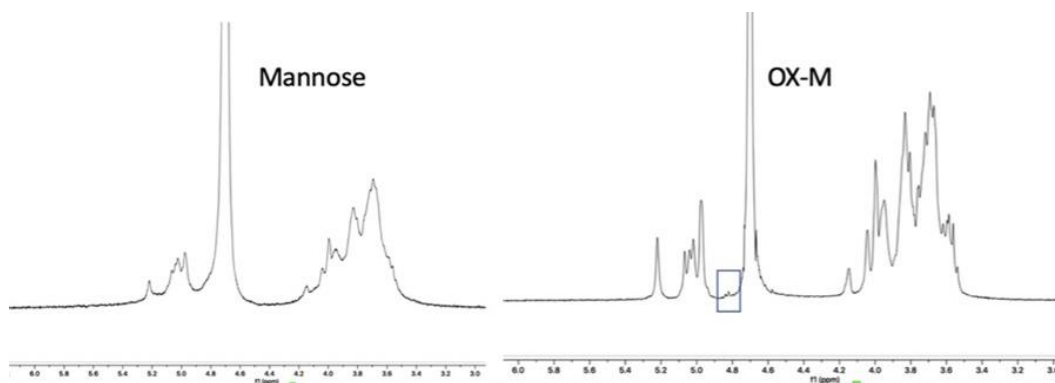


Figure S5. The ^1H -NMR spectra of the mannose and OX-M.

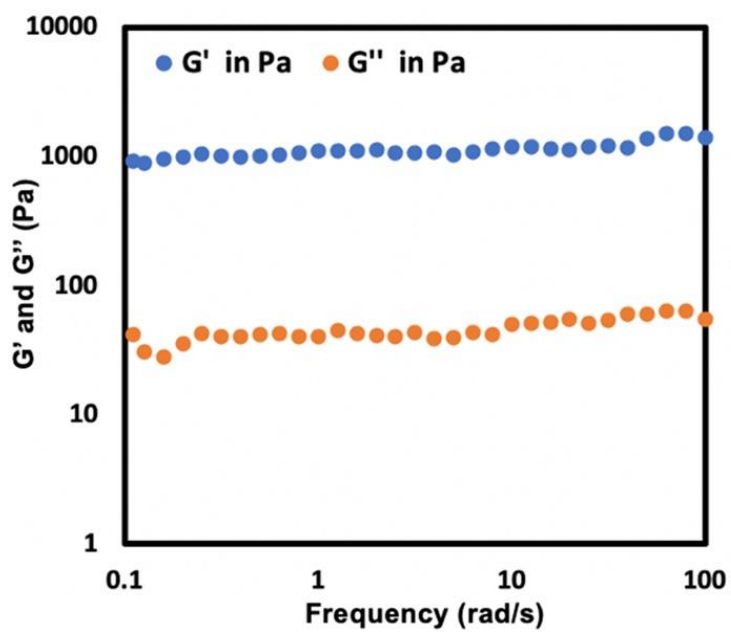


Figure S6. Dynamic frequency sweep of Ncom Gel at 37 °C.

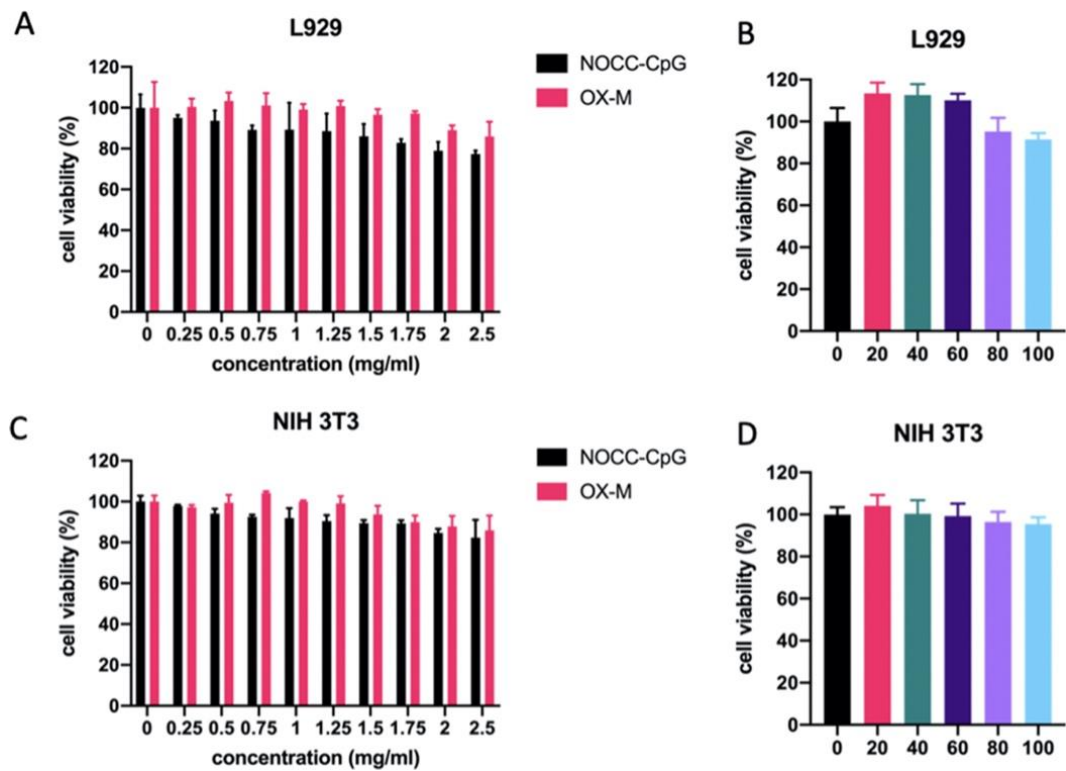


Figure S7. Cytotoxicity assessment by MTT assay for L929 and NIH3T3 cells incubated with various concentrations of NOCC-CpG and OX-M (A) and (C) and different Ncom Gel extract solutions (B) and (D) (n=3, all data are represented as means \pm s.d.).



Figure S8. Evaluation of degradation behavior of Ncom Gel in mice.

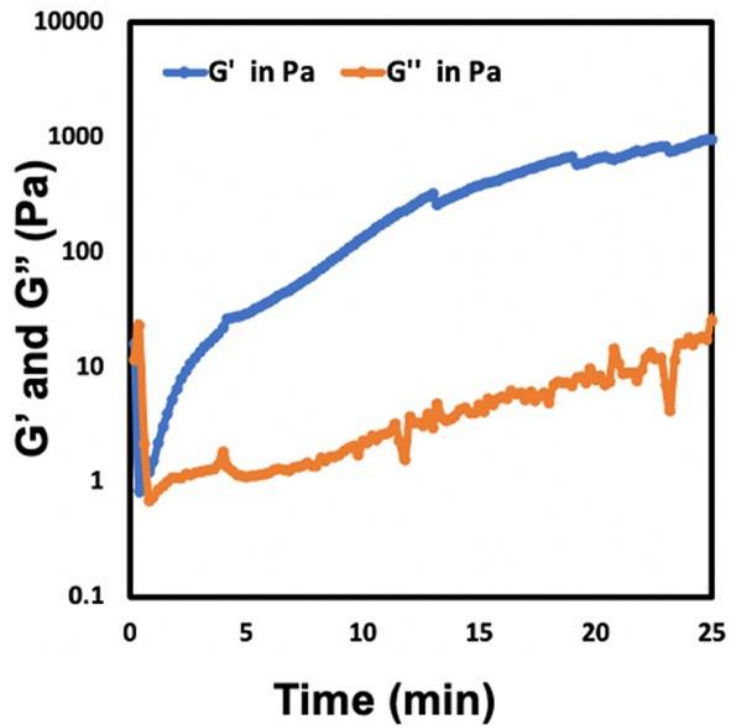


Figure S9. Rheological analysis of the NOCC-CpG/OX-M hydrogel after loading OVA (OVA/Ncom Gel).

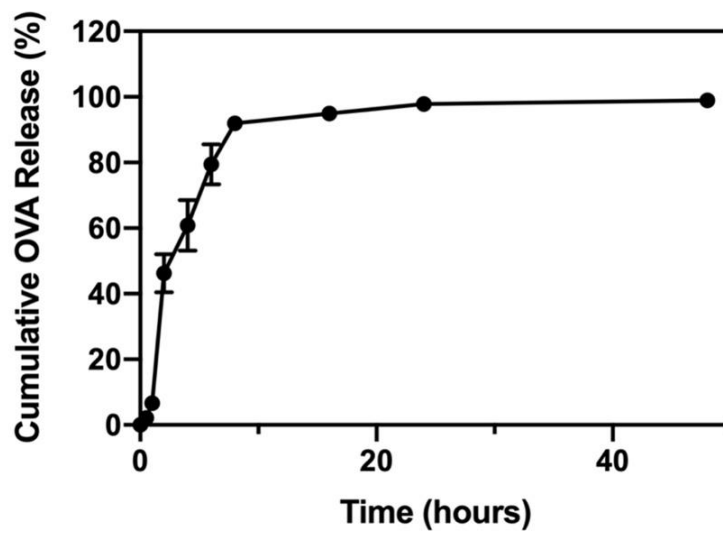


Figure S10. Cumulative release profiles of OVA-FITC from Ncom Gel in PBS.

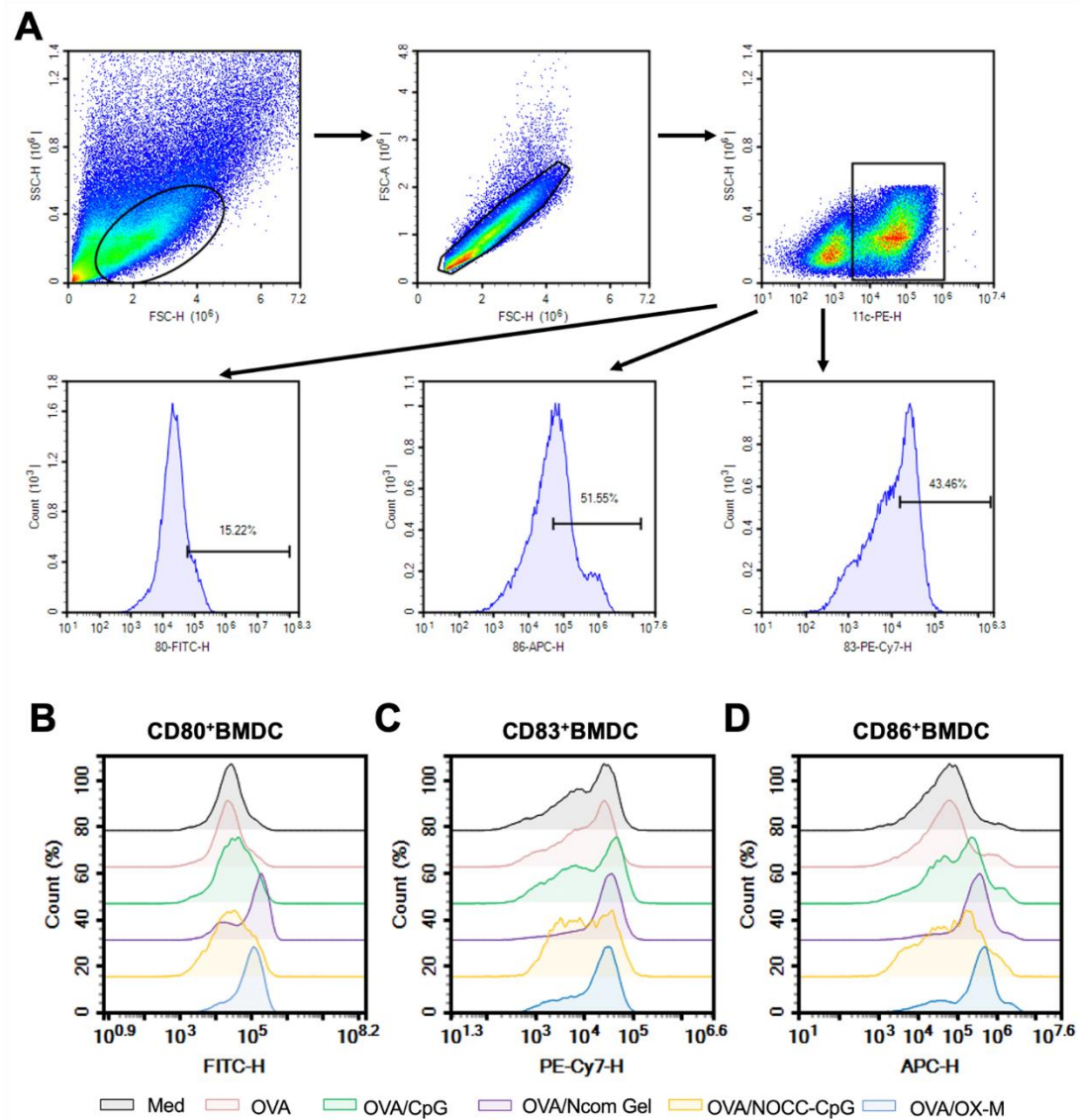


Figure S11. Evaluation of Ncom Gel vaccine on BMDC activation *in vitro*. (A) . Representative scatter plots and gating information derived from analysis of CD11c⁺ CD80⁺, CD11c⁺ CD86⁺ and CD11c⁺ CD83⁺ BMDCs. (B-D) Flow cytometry analysis for the expression of CD80, CD83 and CD86 on BMDCs after various treatments.

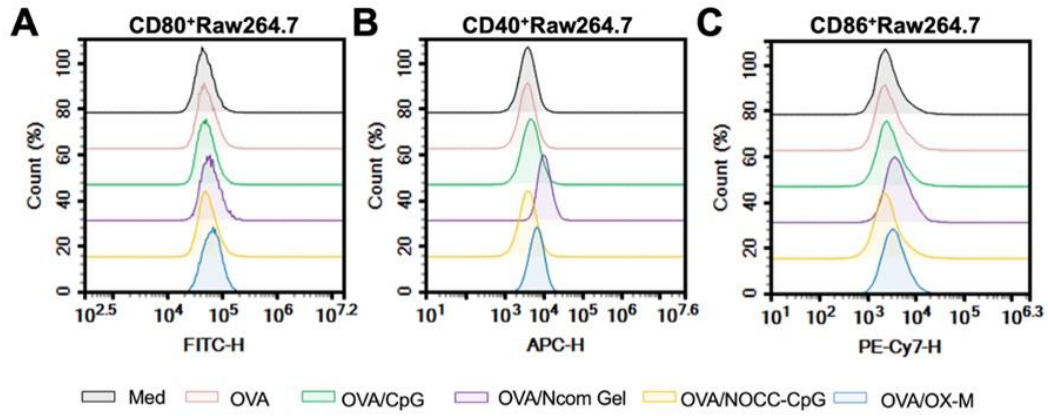


Figure S12. Evaluation of Ncom Gel vaccine on M1 polarization *in vitro*. (A-C) Flow cytometry analysis for the expression of CD80, CD40, and CD86 on Raw264.7s after different treatments.

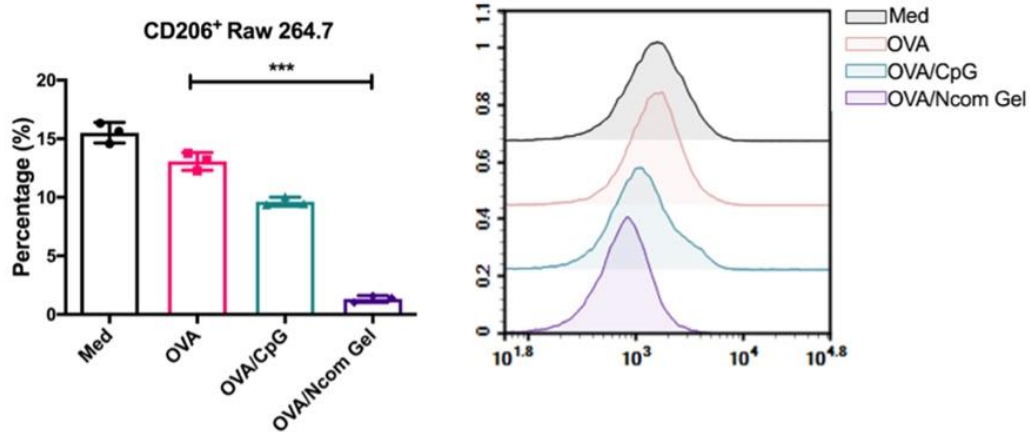


Figure S13. Evaluation of Ncom Gel vaccine on M2 polarization *in vitro*.

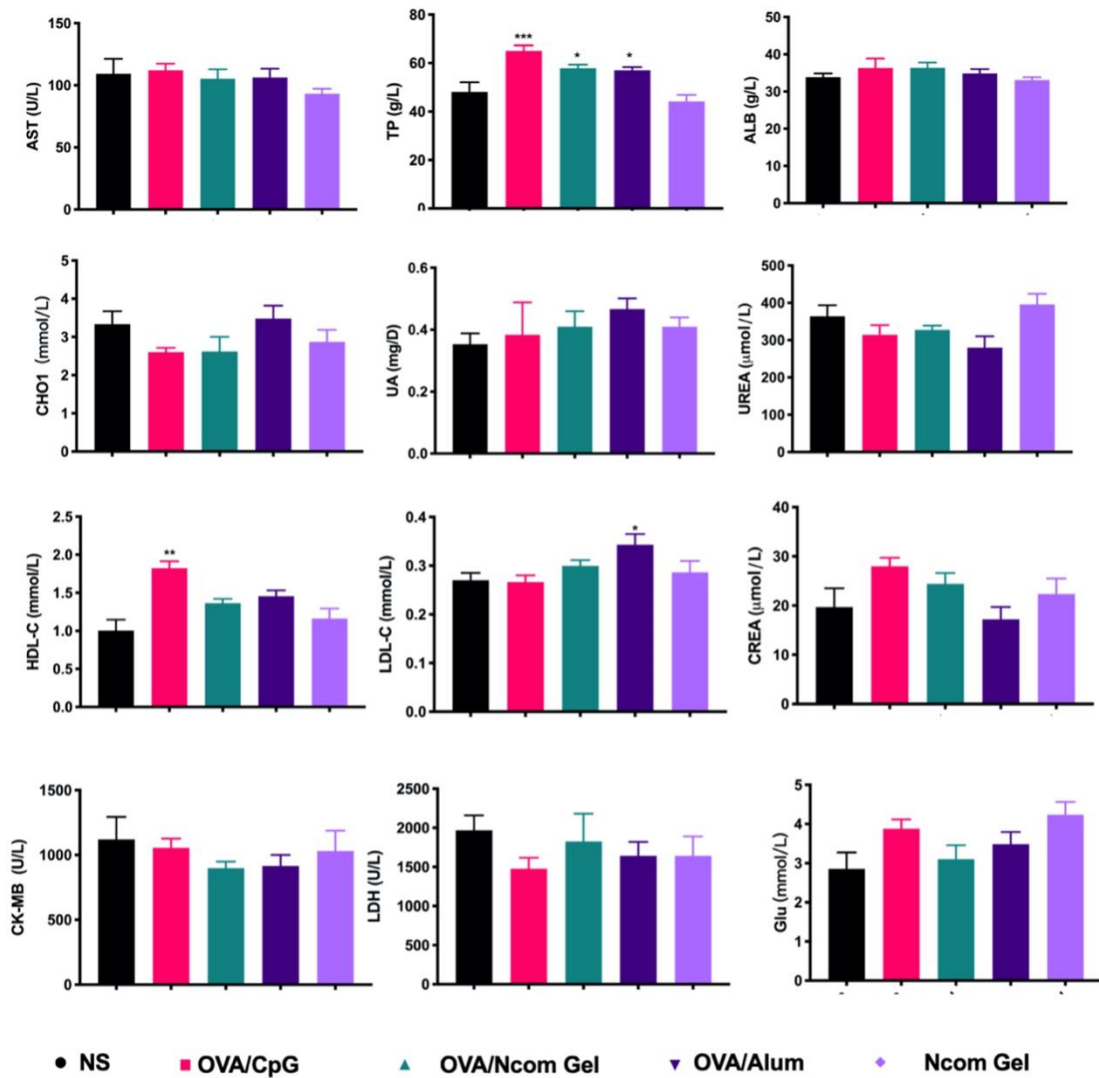


Figure S14. Blood chemistry profile analysis after treatment with various formulations

(n=6 biologically independent samples, all data are represented as means \pm s.d.)

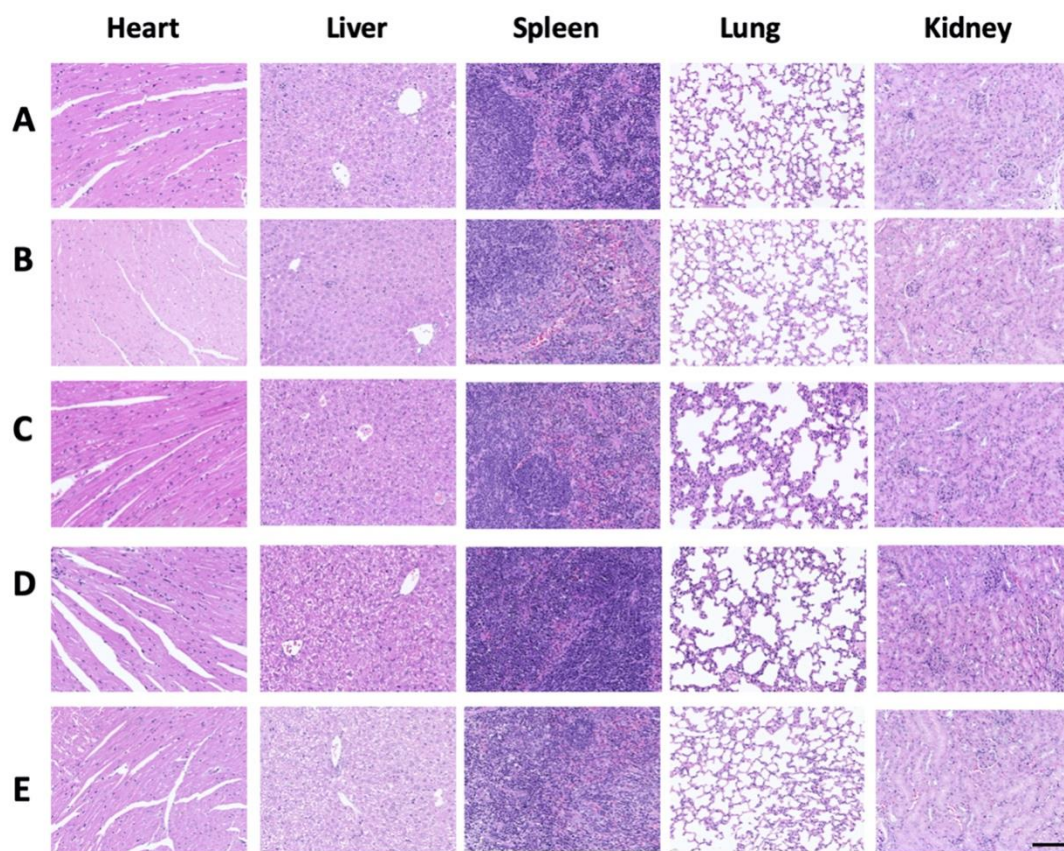


Figure S15. H&E staining of vital organ sections. PBS (A), OVA/CpG (B), OVA/Ncom Gel (C), OVA/Alum (D), and Ncom Gel (E) (Scale bar = 100 μ m).

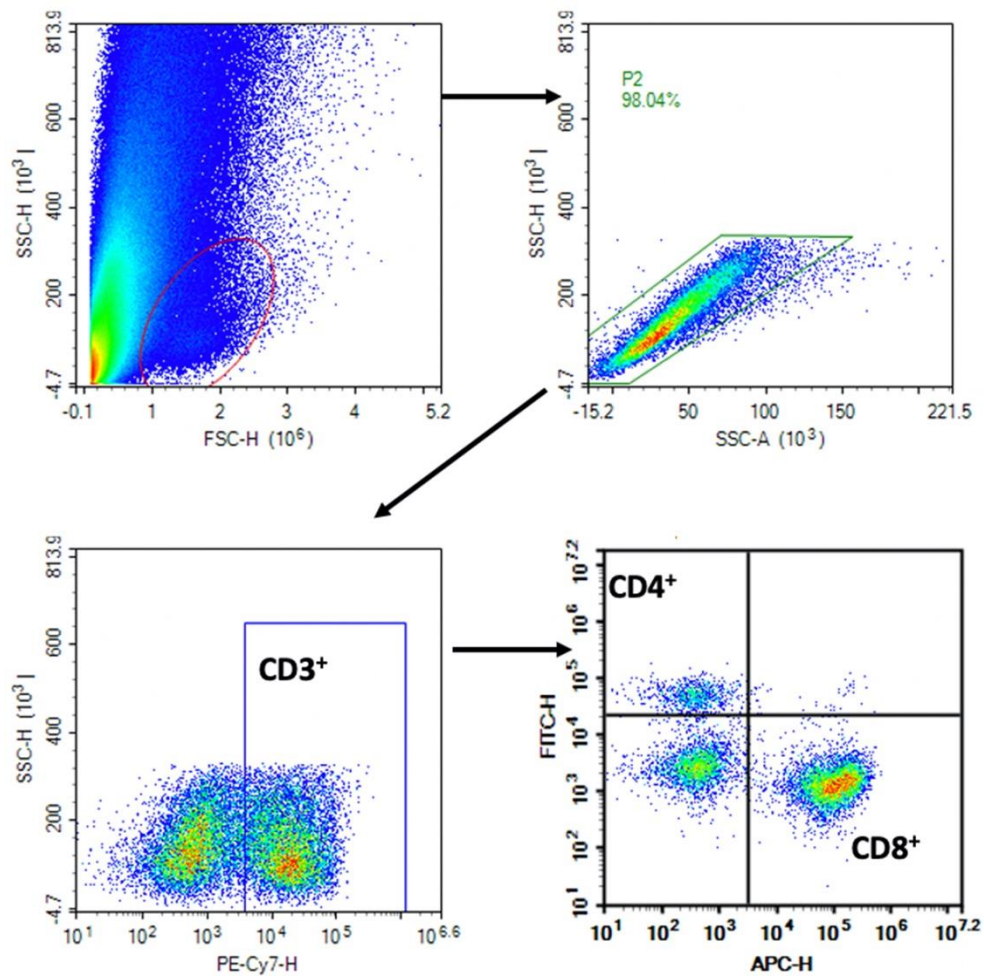


Figure S16. Representative scatter plots and gating information derived from analysis of CD3⁺ CD4⁺ and CD3⁺ CD8⁺ T cells in the tumor.

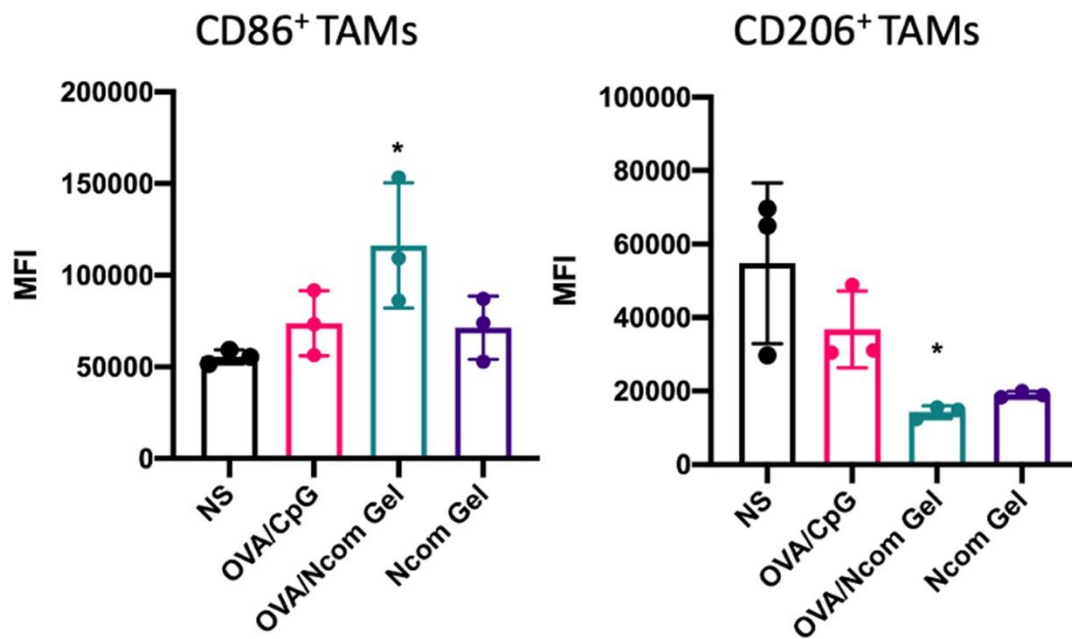


Figure S17. Fluorescence intensity of M1 TAMs (left) and M2 TAMs (right) were analyzed after various treatments (n=3 biologically independent samples, all data are represented as means \pm s.d. and analyzed with one-way ANOVA with Tukey test. * P < 0.05, **P < 0.01, *** P < 0.001 and **** P < 0.0001).

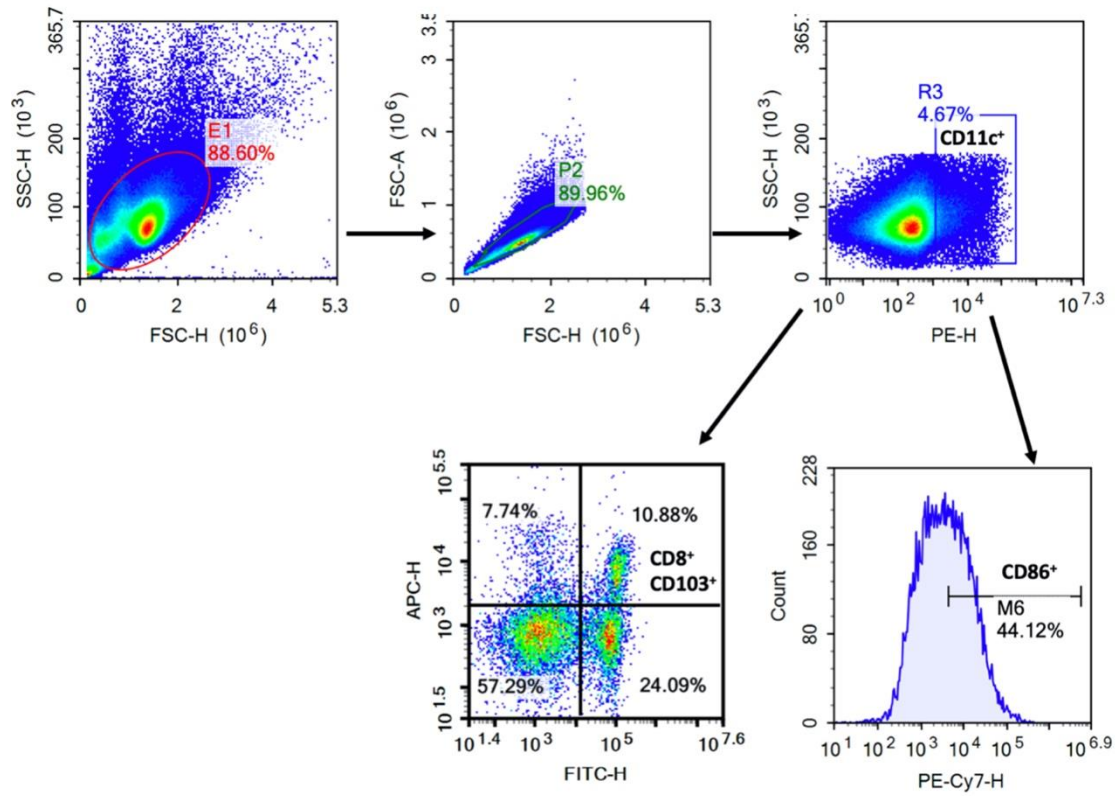


Figure S18. Representative scatter plots and gating information derived from analysis of CD11c⁺ CD86⁺ and CD11c⁺ CD8⁺ CD103⁺ DCs in TDLN.

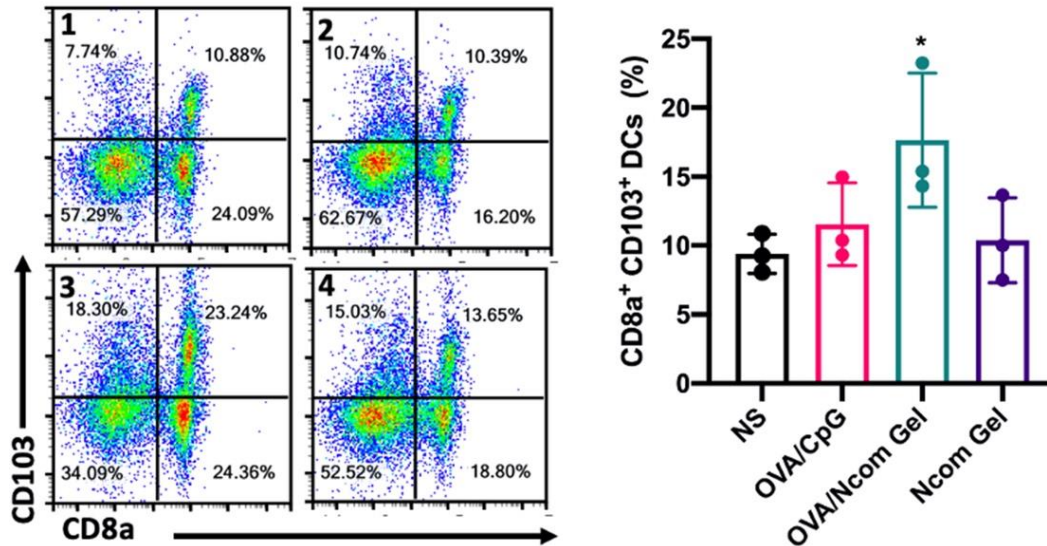


Figure S19. The representative flow cytometry dot plots of CD8⁺CD103⁺DCs in tumor-draining lymph node (TDLN) and quantitative data were shown (n=3 biologically independent samples, all data are represented as means ± s.d. and analyzed with one-way ANOVA with Tukey test. * P < 0.05, **P < 0.01, *** P < 0.001 and **** P < 0.0001).

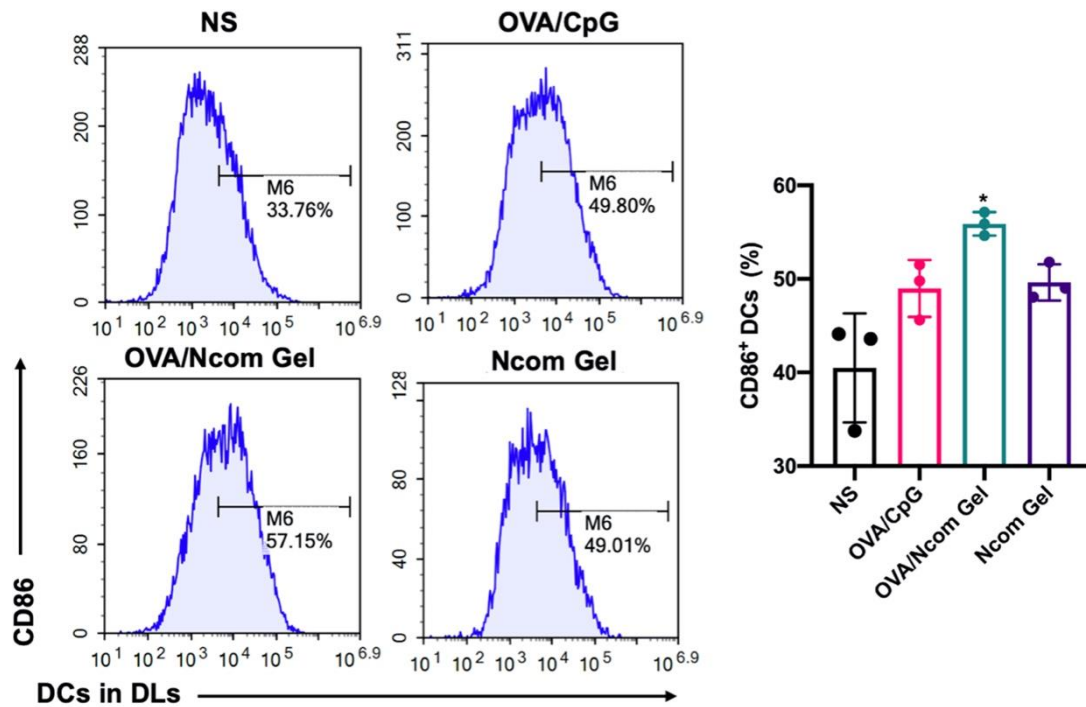


Figure S20. The representative flow cytometry histogram (left) and quantitative data (right) of CD86⁺DCs in TDLN were analyzed after different treatments (n=3 biologically independent samples, all data are represented as means ± s.d. and analyzed with one-way ANOVA with Tukey test. * P < 0.05, **P < 0.01, *** P < 0.001 and **** P < 0.0001).

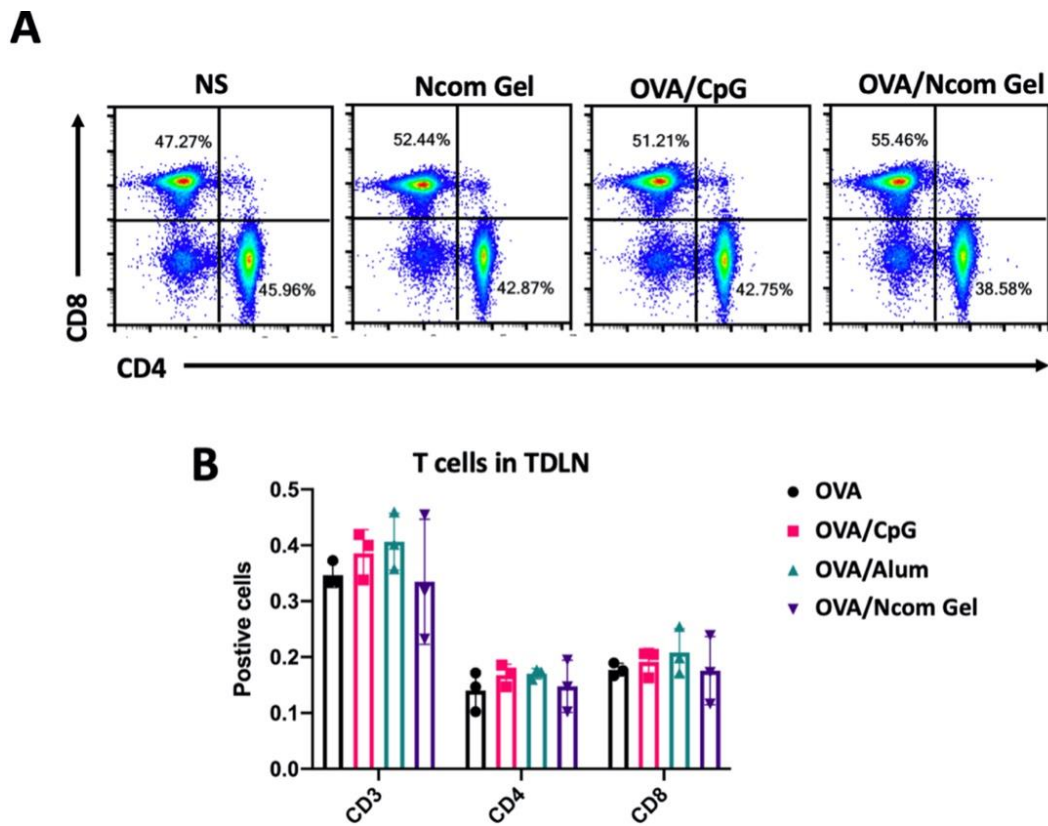


Figure S21. The representative flow cytometry dot plot of CD8⁺ T cells and CD4⁺ T cells in TDLN (top) and quantitative data (bottom) were examined after 2 days of treatment (n=3 biologically independent samples, all data are represented as means ± s.d.).