

FtsZ induces membrane deformations via torsional stress upon GTP hydrolysis.

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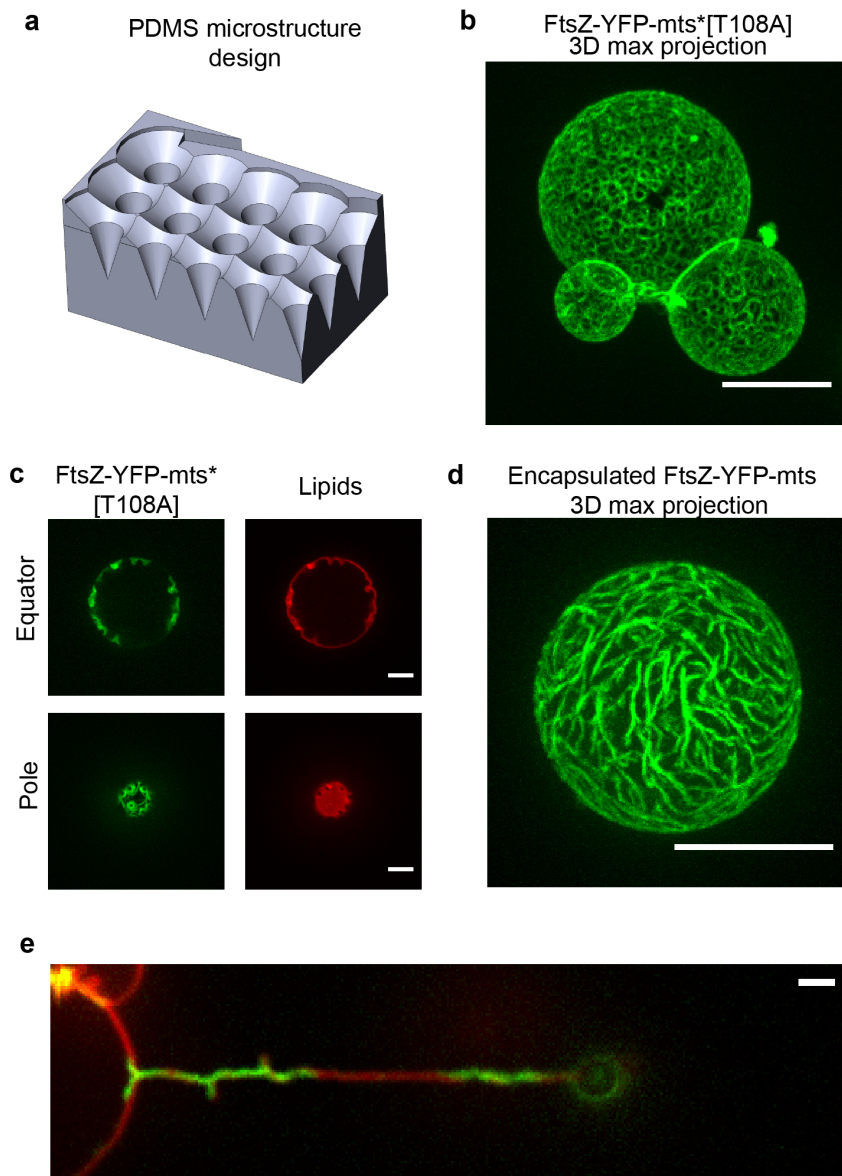
Supplementary Materials

This pdf contains:

Fig. S1 to S4

Supplementary Table 1-2 (Cloning vectors and primers)

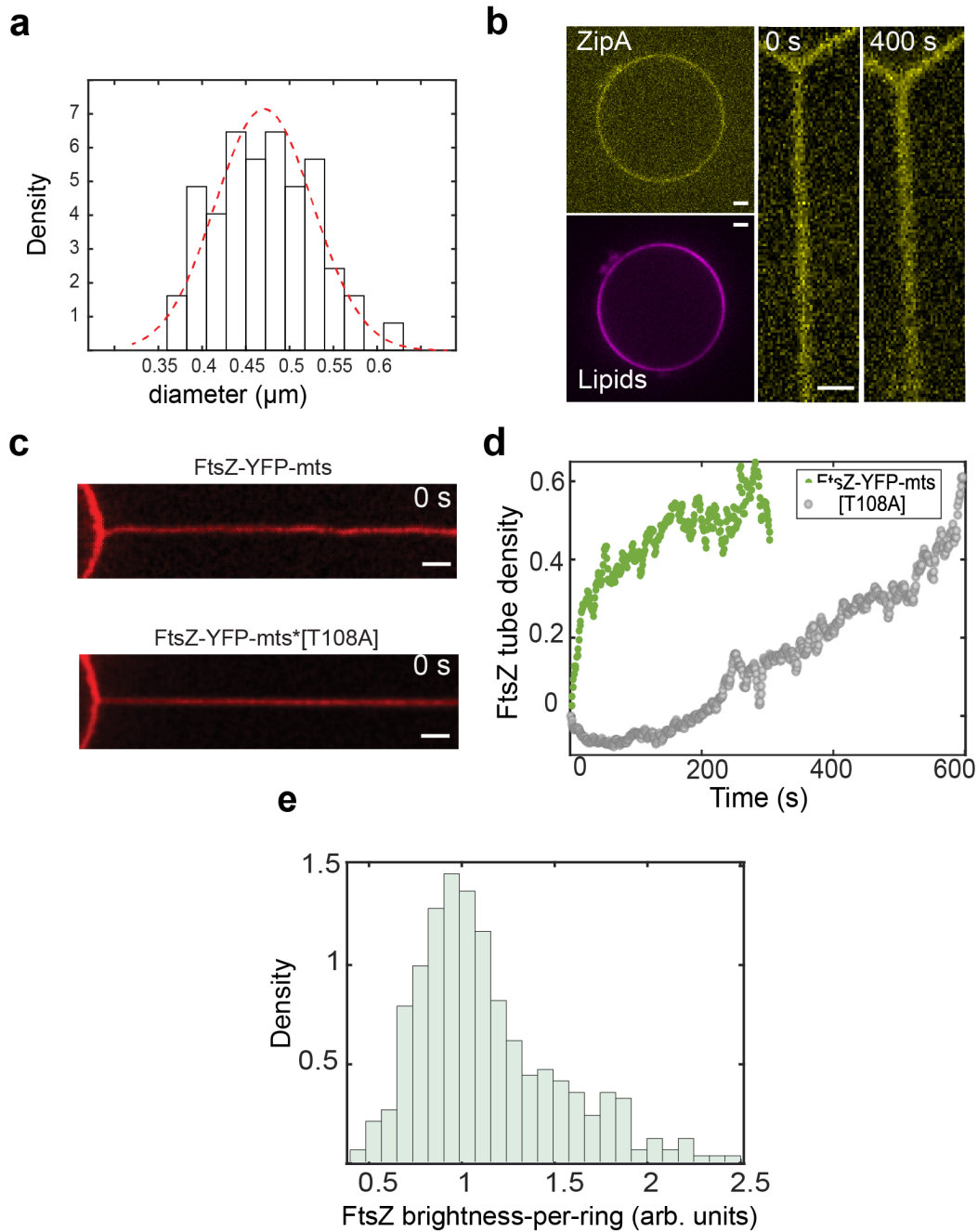
Fig. S1.



Design of microstructure, FtsZ-YFP-mts*[T108A] deformations, high density encapsulated FtsZ-YFP-mts and FtsZ-YFP-mts supercoiling. a) 3D sketch of the PMDS microstructure with inwards cone-like shapes. b) FtsZ-YFP-mts*[T108A] also self-assembled into ring-like structures on GUVs (GUVs:N>20) (Scale bar = 10 μ m). c) After deflation, FtsZ-YFP-mts*[T108A] induced inwards conical deformations emerging from before mentioned rings (GUVs:N>20) (Scale bar = 2 μ m). d) By encapsulating FtsZ-YFP-mts in GUVs, Mg⁺² and GTP content-conditions were fine-tuned to obtain either ring structures (Fig. 5) or nematic phases at a higher membrane protein density (GUVs:N>10) (Scale bar = 10 μ m). e) FtsZ-YFP-

mts torsion over lipid tubes can be found in (less frequent) experiments displaying plectonic/supercoiled regions (N=2). (Scale bar = 2 μm).

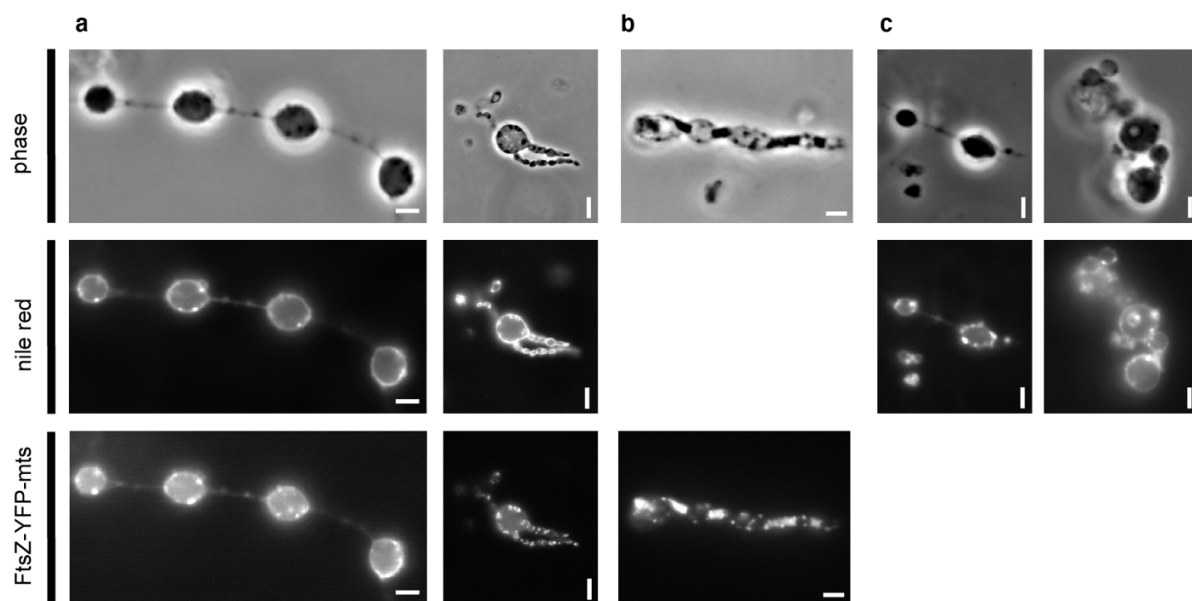
Fig. S2.



Tube diameter distribution, ZipA control experiments and ring-unit-brightness distribution. a) The diameter distribution (N=55) showed a Gaussian distribution with a mean of 0.47 μm . This implied that membrane tensions equivalent to this mean (\pm std) were highly

frequent despite of precise control on the vesicle membrane tension. b) Vesicles decorated with ZipA and imaged under deflation conditions exhibited no deformations (GUVs:N=14). In addition, we examined lipid tubes only coated with siZipA-Alexa 488. No deformations (N=10) were observed in the range of 400-600 seconds. c) Similar initial lipid tube diameter (0.44 μm) for experiments shown in Figure 3 a-b d) For experiments shown in Fig. 3 a-b, FtsZ-YFP-*mts* entered rapidly to the lipid tube while the mutant without GTPase activity (slower). e). Distribution of FtsZ brightness-per-ring (N=412 analyzed rings). The distribution's mode value was chosen as the value for FtsZ brightness-per-ring. (Scale bar = 2 μm).

Fig. S3.



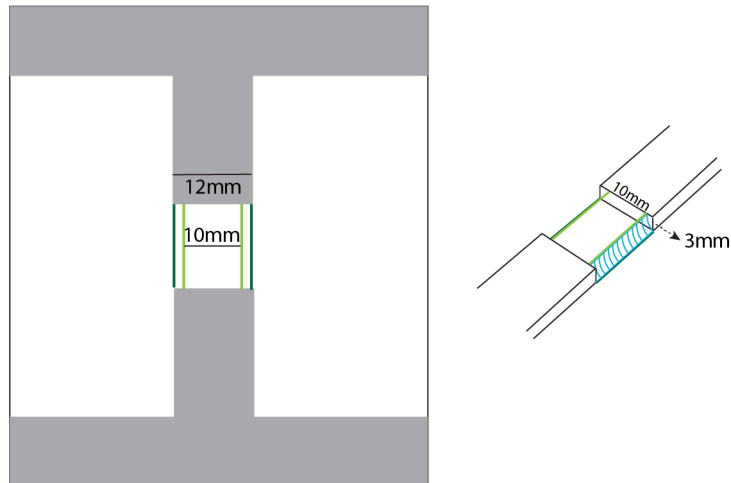
Observed phenotypes of *E. coli ftsZ-YFP-mts* after lysozyme treatment in sucrose-buffer.

The first columns in a, b & b show a pearl necklace like appearance of the cells, while the second column in b might show an earlier stage of this chaining type of vesiculation. Nile red was used to prove that the observed vesicles and tubular connections are actually phospholipid-membranes. As a control unstained cells b) and stained cells c) were also imaged separately in

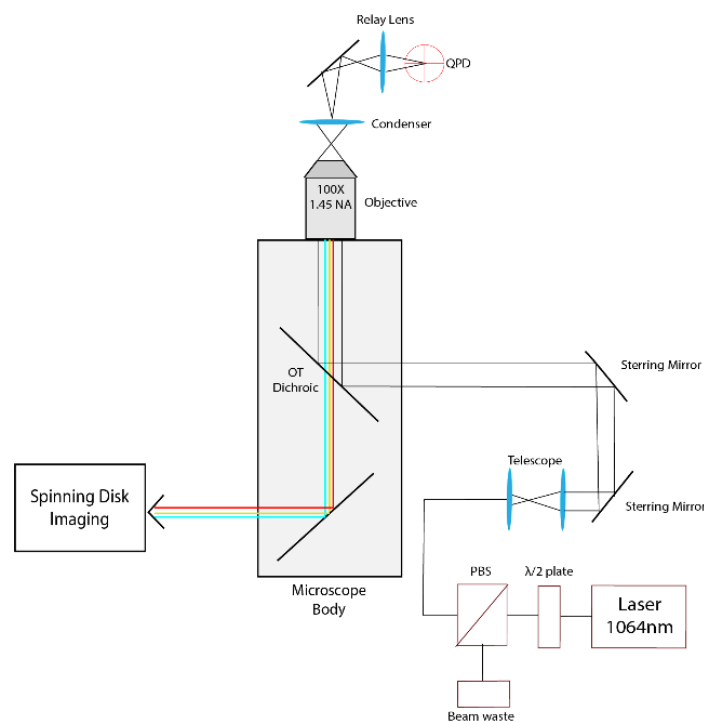
the respective channel. Example micrographs from (N=3) biological replicates. (scale bar = 2 μ m).

Fig. S4

a



b



Drawing for the experiment chamber and optical trapping device a) Sketch of imaging chamber for deflated vesicle preparation. b) Optical tweezers setup layout.

Supplementary Table 1: Cloning Vectors

Vector	Protein	Source/reference
pET-11b	FtsZ-YFP-mts	gifted by Harold Erickson, Ref. 1
pET-11b	FtsZ-YFP-mts*[T108A]	Ref. 3
pET-28a	FtsZ-WT	Ref. 31
pET-15ZIP	sZipA	Ref. 32
pEKEx2	FtsZ-YFP-mts	This publication

Supplementary Table 2: Primers

Name	Sequence 5' – 3'
T108A_RV	GGTGGTGGTGCCGGTACAGGT
T108A_FW	ACCTGTACCGGCACCACCACC
sZipAI	CATATGGCTGCCGCGCG
sZipAII	ACCAGCCGTAAAGAACG
Sall-ftsZ	CATGTCGACATGTTTGAACCAATGGA ACTTACC
SacI-mts	CATGAGCTCTTATCCTCCGAACAAGCG