

Supplemental information

**Analysis of SARS-CoV-2 variant mutations reveals
neutralization escape mechanisms and the ability
to use ACE2 receptors from additional species**

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Supplemental Information titles and legends

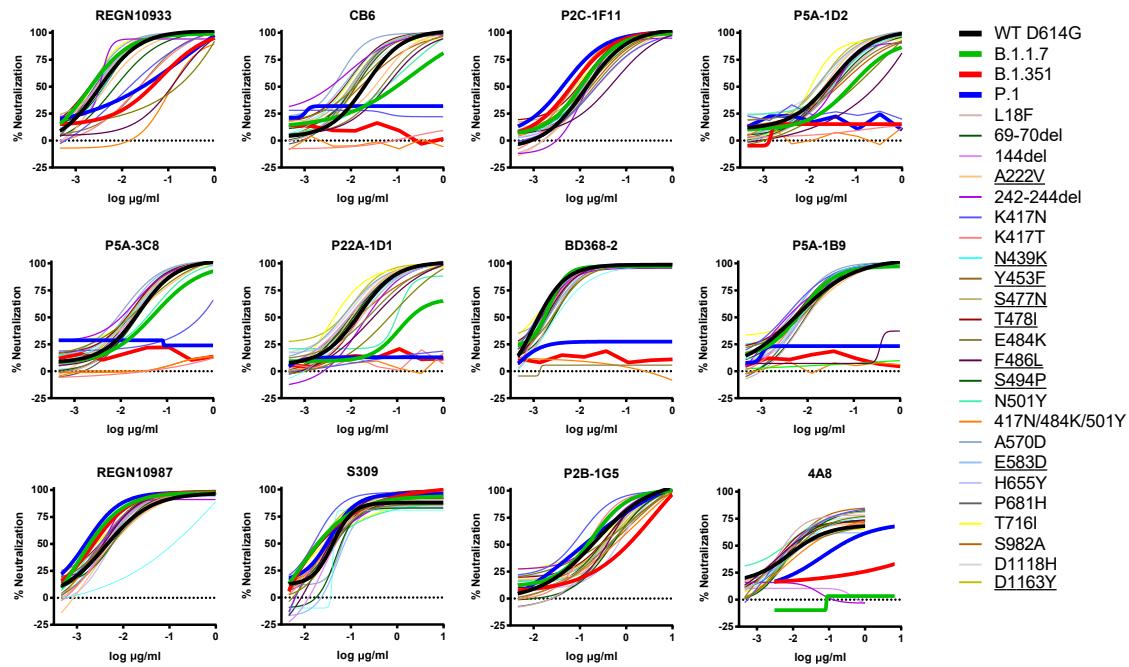


Figure S1.

Neutralization of SARS-CoV-2 variants by each antibody, related to

Figure 1. Pseudoviruses bearing the indicated mutations were tested against serial dilutions of each mAb. Neutralizing activity was defined as the percent reduction in luciferase activities compared to no antibody controls. Levels of resistance were calculated as the -fold change in IC₅₀ between each mutant and WT D614G, as presented in Figure 1C. Results were calculated from three independent experiments.

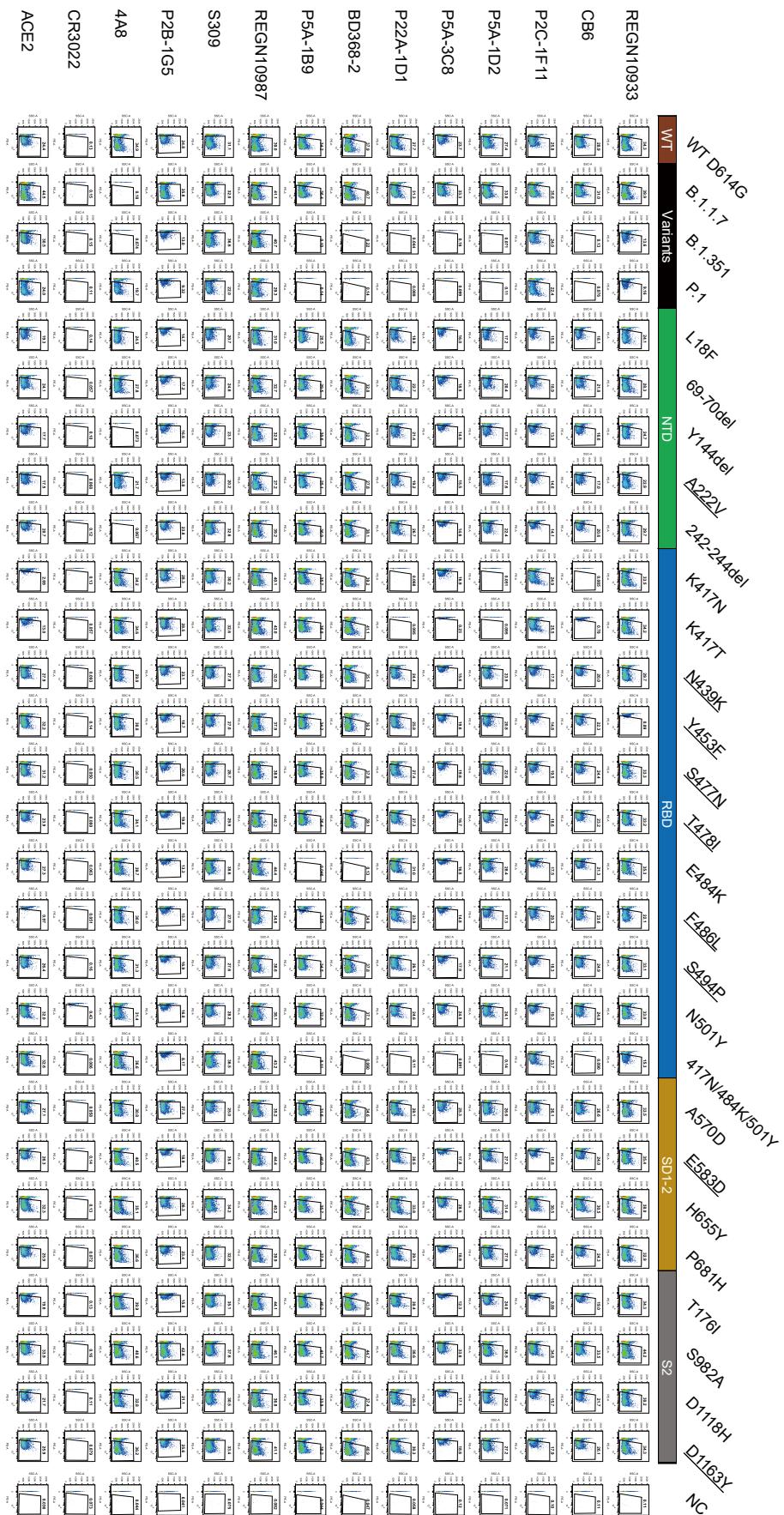


Figure S2.

Binding to cell surface expressed SARS-CoV-2 variants by each antibody, related to Figure 1. Wildtype and mutant S proteins were expressed on the surface of HEK 293T, incubated with the mAbs or human soluble ACE2 under study, followed by staining with anti-human IgG Fc PE or anti-his PE, and analyzed by FACS. The gated cell percentages are shown. The fold changes in antibody binding, as shown in Figure 1D, was determined by comparing the total MFI in the selected gate between S variants and WT D614G. Data shown were calculated from three independent experiments. CR3022 is a negative control antibody. NC is HEK 293T cells with mock transfection.

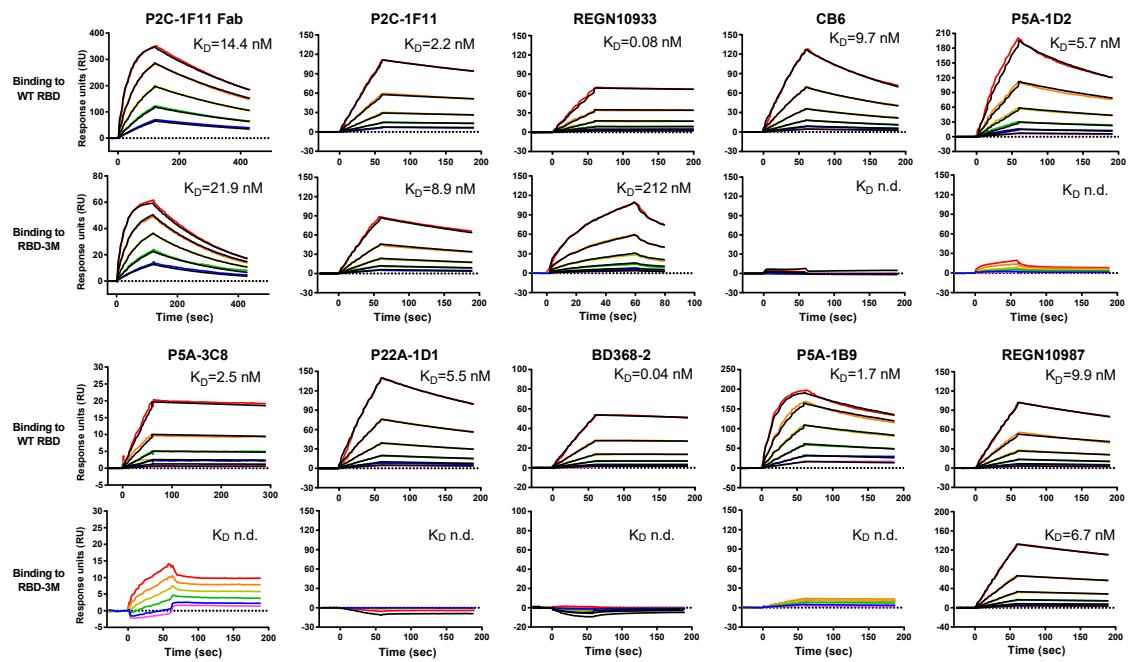


Figure S3.

Binding kinetics of mAb to WT RBD and RBD-3M measured by SPR, related to Figure 2. The Fab and IgG form of mAbs were immobilized on a CM5 sensor and serial concentrations of either wildtype RBD or mutant RBD-3M were flowed through the system. Colored lines indicate the experimentally derived curves. Black lines represent best fitted curves based on the experimental data. The calculated KDs for each mAb are shown against WT RBD and mutant RBD-3M. The names of mAb tested are indicated at the top of each graph.

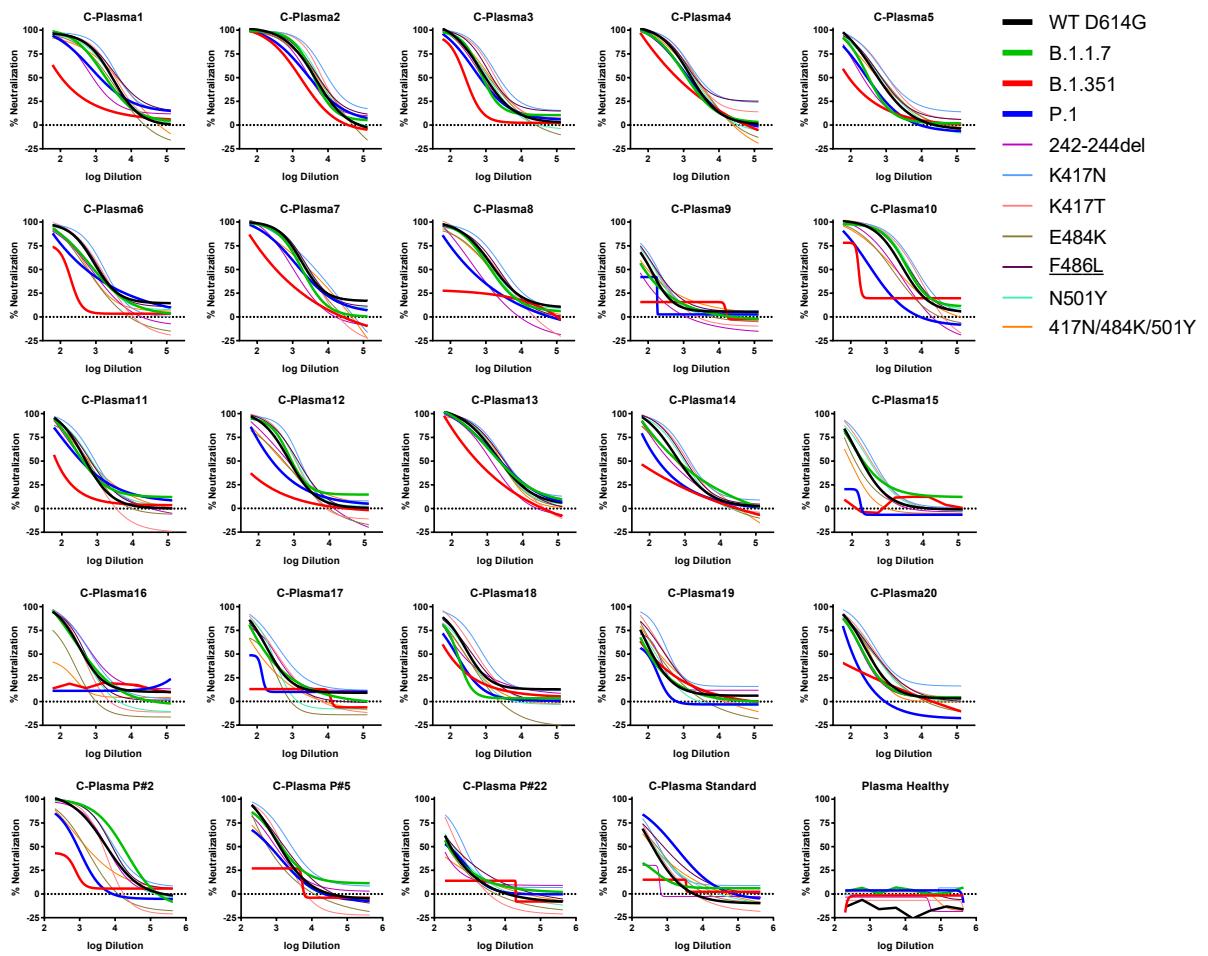


Figure S4.

Neutralization of SARS-CoV-2 variants by each convalescent plasma, related to Figure 3. Pseudoviruses bearing the indicated mutations were tested against serial dilutions of convalescent plasma. Neutralization activity was defined as the percent reduction in luciferase activity relative to no serum control. The actual ID₅₀ and the -fold changes between each mutant and WT D614G pseudovirus were calculated to estimate the resistance levels shown in Figure 3. Results were calculated from three independent experiments.

Table S1. Data collection and refinement statistics (molecular replacement), related to Figure 2.

RBD-K417N-E484K-N501Y-P2C-1F11 complex	
Data collection	
Space group	C2
Cell dimensions	
a, b, c (Å)	195.844, 85.973, 57.872
α, β, γ (°)	90, 99.75, 90
Resolution (Å)	50.00-2.094 (2.15-2.094) ^a
Unique reflections	50710 (3033)
R_{sym} or R_{merge}	0.088(0.384)
I / sI	17.2(3.2)
Completeness (%)	90.92 (54.79)
CC _{1/2} (%)	99.7 (76.2)
Redundancy	5.3 (2.3)
Refinement	
Resolution (Å)	20.98-2.094
No. reflections	50684
$R_{\text{work}} / R_{\text{free}}$	16.9/19.7
No. atoms	
Protein	4728
Ligand/ion	14
Water	505
B-factors	
Protein	41.18
Ligand/ion	81.75
Water	46.02
R.m.s. deviations	
Bond lengths (Å)	0.007
Bond angles (°)	0.89
Ramachandran statistics (%)	
Favored	97.19
Allowed	2.64
Disallowed	0.17

^a One crystal for the data, values in parentheses are for highest-resolution shell.

Table S2. Molecular interaction between P2C-1F11 and mutant RBD-K417N-E484K-N501Y, related to Figure 2.

	P2C-1F11	Length (Å)	Interactions
P2C-1F11/RBD	E/K417/N[N]	H/Y52/CE2[C] 3.84	
		H/Y52/OH[O] 3.87	Hydrogen bond
	E/K417/CG[C]	H/Y52/CE2[C] 3.85	
		H/Y52/CZ[C] 3.97	
		H/Y52/OH[O] 3.32	
	E/K417/CE[C]	H/Y52/OH[O] 3.69	
	E/K417/NZ[N]	H/Y52/OH[O] 3.19	Hydrogen bond
	E/K417/CD[C]	H/Y33/OH[O] 4.02	
	E/K417/CE/[C]	H/Y33/OH[O] 4.03	
	P2C-1F11/RBD-3M	E/N417/N[N]	H/Y52/CE2[C] 3.82
		H/Y52/OH[O] 3.53	Hydrogen bond
	E/Y453/CE1[C]	L/Y33/OH[O] 3.50	
	E/Y453/CZ[C]	L/Y33/OH[O] 3.43	
	E/Y453/OH[O]	L/Y33/CE1[C] 3..58	
		L/Y33/CZ[C] 3..47	
		L/Y33/OH[O] 2.52	
	E/Q493/CG[C]	L/Y33/OH[O] 3.62	
	E/Q493/NE2[N]	L/S32/CB[C] 3.96	
		L/S32/OG/[O] 3.77	
	E/T500/C[C]	L/S28/OG[O] 3.90	
	E/T500/O[O]	L/S28/CB[C] 3.30	
		L/S28/OG[O] 3.01	
	E/Y501/CA[C]	L/S28/OG[O] 3.87	
	E/Y501/CE1[C]	L/V29/O[O] 3.79	
	E/Y501/CZ[C]	L/S30/CA[C] 3.94	
	E/Y501/OH[O]	L/S30/CA[C] 3.82	
		L/S30/CB[C] 3.85	
		L/S30/OG[O] 3.85	

Table S3. Information of the study subjects, related to Figure 3.

Patients	Gender	Age	Severity
C-Plasma1	male	60	severe
C-Plasma2	male	57	severe
C-Plasma3	male	57	severe
C-Plasma4	female	81	severe
C-Plasma5	female	62	mild
C-Plasma6	male	35	mild
C-Plasma7	male	55	mild
C-Plasma8	female	52	mild
C-Plasma9	female	36	mild
C-Plasma10	male	63	mild
C-Plasma11	female	63	severe
C-Plasma12	male	75	mild
C-Plasma13	male	78	severe
C-Plasma14	male	62	mild
C-Plasma15	male	30	mild
C-Plasma16	male	30	mild
C-Plasma17	female	29	mild
C-Plasma18	female	43	mild
C-Plasma19	male	58	mild
C-Plasma20	female	67	severe
C-Plasma P#2	female	65	severe
C-Plasma P#5	female	63	severe
C-Plasma P#22	male	62	mild

Table S4. Primers used in the construction of mutated clones of SARS-CoV-2 S, related to Figure 1.

No.	Primer Name	sequence
1	Mut-1stMS-F	TTGGTACCGAGCTCGGATCCATGTTCTGCTGACCACCA AGAGAACCATGTTCGTGTTCTGGTG
2	Mut-S-R	CCACTGTGCTGGATATCTGCAGAATTCTCAGGTGTAGTG CAGCTTCAC
3	nCov-SL18F-F	GTGAGCAGCCAGTGCCTGAATTCCACCACAGAAC
4	nCov-SL18F-R	GTTCTGGTGGTCAAATTACCGCACTGGCTGCTCAC
5	nCov-SHV69-70del-F	CGTGACCTGGTCCACGCCATCAGCGGCACCAATGGCAC CAAGAG
6	nCov-SHV69-70del-R	CTCTGGTGCCATTGGTGCCGCTGATGGCGTGGAAACAG GTCACG
7	nCov-SY144del antisense	CTCTTGTGTTCTTATGATAAACACCCAGGAAAGGGTCAT
8	nCov-SY144del	ATGACCCCTTCCTGGGTGTTATCATAAGAACACAAGAG
9	nCov-SA222V-F	CAGGGCTTCAGCGTCTGGAGCCTCTG
10	nCov-SA222V-R	CAGAGGCTCCAGGACGCTGAAGCCCTG
11	nCov-S242-244del-F	ACCAGATTCCAGACCCTGCACAGATCATATCTTACAC
12	nCov-S242-244del-R	GTGTAAGATATGATCTGTGCAGGGTCTGGAATCTGGT
13	nCov-SK417N-Fg1251t	CAGGGCAGACCGGCAATATGCCGAC
14	nCov-SK417N-R	GTCGGCGATATTGCCGGTCTGCCCTG
15	nCov-SK417T-Fa1250c	GGGCAGACCGGCACGATGCCGACTAC
16	nCov-SK417T-R	GTAGTCGGCGATCGTGCCGGTCTGCC
17	nCov-SN439K-F	CGTGATCGCGTGGAACTCTAAGAATCTAGATTGAAA
18	nCov-SN439K-R	TTTCGAATCTAGATTCTAGAGTCCACGCGATCAG
19	nCov-SY453F-Fa1358t	GGCAATTACAATTACCTGTTCAGACTGTTCAGAAAGAGC
20	nCov-SY453F-R	GCTCTTCTGAACAGTCTGAACAGGTAATTGTAATTGCC
21	nCov-SS477N-FG1430A	CTACCAGGCCGGCAACACACCCGTGTAATG
22	nCov-SS477N-R	CATTACACGGTGTGTTGCCGGCTGGTAG
23	nCov-ST478I-F	TACCAAGGCCGGCAGCATACCGTGTAAATGG
24	nCov-ST478I-R	CCATTACACGGTATGCTGCCGGCTGGTA
25	nCov-SE484K-Fg1450a	CACCGTGTAAATGGCGTGAAGGGCTCAATTGCTAC
26	nCov-SE484K-R	GTAGCAATTGAAGCCCTCACGCCATTACACGGTG

Table S4 continued. Primers used in the construction of mutated clones of SARS-CoV-2 S, related to Figure 1.

No.	Primer Name	sequence
27	nCov-SF486L-F	ATGGCGTGGAGGGCTAAATTGCTACTTCCCTC
28	nCov-SF486L-R	GAGGGAAAGTAGCAATTAAAGCCCTCCACGCCAT
29	nCov-SS494P-F	TACTCCCTCTGCAGCCCTACGGCTTCCAGCC
30	nCov-SS494P-R	GGCTGGAAGCCGTAGGGCTGCAGAGGGAAAGTA
31	nCov-SN501Y-Fa1501t	GCTTCAGCCTACCTATGGCGTG GGCTAC
32	nCov-SN501Y-R	GTAGCCCACGCCATAGGTAGGCTGGAAGC
33	nCov-SA570D-F	CATCTGTGGTGTGTCGTGATGTCTCTGCCG
34	nCov-SA570D-R	CGGCAGAGACATCGACGACACCACAGATG
35	nCov-SE583D-F	CCCTCAGACCCTGGATATCCTGGACATCACT
36	nCov-SE583D-R	AGTGATGTCCAGGATATCCAGGGTCTGAGGG GCCGGCTGCCTGATCGCGCCGAGTACGTGAATAATAGC
37	H655Y-F	TACGAGTGCAGACATCCCTA
38	H655Y-R	TAGGGATGTCGCACTCGTAGCTATTATTACGTACTCGGC GCCGATCAGGCAGCCGGC
39	nCov-SP681H-F	ACCCAGACCAATGCCATAGAACAGAGCCAGAACG
40	nCov-SP681H-R	GCTTCTGGCTCTTCTATGGCTATTGGTCTGGGT
41	nCov-ST716I-F	TAGCATGCCATCCCTATCAATTTCACCATCAGCG
42	nCov-ST716I-R	CGCTGATGGTGAAATTGATAGGGATGGCGATGCTA
43	nCov-SS982A-F	GTACTCAACGATATCCTGGCCAGACTGGACAAGGTGGA
44	nCov-SS982A-R	TCCACCTTGTCCAGTCTGGCCAGGATATCGTTGAGTAC
45	nCov-ST1027I-Fc3080t	CCAATCTGGCCGCCATCAAGATGAGCGAGTG
46	nCov-ST1027I-R	CACTCGCTCATTTGATGGCGGCCAGATTGG
47	nCov-SD1118H-F	GCTCACGAAGGTATTGGGGTGGTGTGATCTGAG
48	nCov-SD1118H-R	CTCAGATCATACCACCCACAATACCTCGTGAGC
49	nCov-SD1163Y-F	CAAGAACACACCAGCCCTTATGTGGACCTCGGTGATATT
50	nCov-SD1163Y-R	AAATATCACCGAGGTCCACATAAGGGCTGGTGTGATTCTG
51	nCov-SV1176F-F	TTCGGGAATCAATGCCAGCTTCGTGAATATCCAGAAGGAAA
52	nCov-SV1176F-R	TTTCCTCTGGATATTACGAAGCTGGCATTGATTCCCGAA