

## **Supporting Information**

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# Hypermethylation of hepatic mitochondrial *ND6* provokes systemic insulin resistance

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#### Supporting Information

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Figure S1 Linear correlations between clinical parameters and *D-loop* methylation level. A-D, BMI, fasting glucose level, fasting insulin level, and HOMA-IR index in T2DM and non-T2DM subjects. E, Relative mRNA levels of 13 mtDNA-encoded OXPHOS complex subunits; F, nDNA-encoded OXPHOS complex subunits and mtDNA-related transcription factors; G, *D-loop* methylation level in human peripheral leukocytes of T2DM and non-T2DM subjects, in A-G, n=39 for each group. H-L, Liner correlations between *ND6* mRNA level, BMI, fasting glucose, fasting insulin, HOMA-IR, and ND6 methylation level in human peripheral leukocytes of non-T2DM subjects, n=39; M-Q, Liner correlations between *ND6* mRNA level, BMI, fasting glucose, fasting insulin, HOMA-IR, and ND6 methylation level in human peripheral leukocytes of T2DM subjects, n=39; R, Linear correlations between BMI and *D-loop* methylation level; S, fasting glucose and *D-loop* methylation level; T, fasting insulin and *D-loop* methylation level; U, HOMA-IR index and *D-loop* methylation level in human peripheral leukocytes, n=78. Values are mean  $\pm$  SEM. \*\*p< 0.01.



**Figure S2** *ND6* is a primary target of mtDNA methylation in the liver of db/db mice. A-C, Fasting glucose, fasting insulin, and HOMA-IR index in HFD and db/db mice. **D**, HE staining of liver tissues. **E**, Confirmation the purity of extracted mtDNA via PCR with specific mitochondrial and nuclear primers. NC1-4, negative control of *ND1*, *ND6*, *ATP8* and *Tubulin*, respectively. Predicted PCR products were *ND1*, 152 bp; *ND6*, 199 bp; *ATP8*, 113 bp; *Tubulin*, 294 bp; *Sdha*, 247 bp. **F**, Total methylated CG, CHG and CHH frequencies; **G-I**, Methylated CG, CHG, and CHH frequencies of peptides, tRNAs and rRNAs on liver mtDNA. **J-M**, Methylated CHH frequencies on H-strand, methylated CHH, CHG, and CG frequencies on Lstrand of 13 oxidative phosphorylation (OXPHOS) complex subunits in the liver of control and

db/db mice. N, Relative *ND6* mRNA level; O, Relative mtDNA copy number; P, Relative complex I activity in the liver of the control and db/db mice. Values are mean  $\pm$  SEM. *n*=7 for the control and 4 for the HFD group, \*p< 0.05, \*\*p< 0.01.



**Figure S3 Correlations between D-loop methylation and complex I activity and HOMA-IR. A-C**, Liner correlations between *D-Loop* methylated CG, CHG, CHH frequencies and complex I activity in the liver of control and db/db mice; **D-F**, Liner correlations between *D-Loop* methylated CG, CHG, CHH frequencies and HOMA-IR index in the liver of control and db/db mice. N=15.



Figure S4 ND6 knockdown promotes mitochondrial dysfunction in murine cell lines and hepatic lipid accumulation in mice. A, Relative ND6 mRNA levels; B, mitochondrial OCR; C, fluorescence of mitochondrial superoxide in 3T3-L1 and HT22 cells transfected with ND6 siRNA (n=3). D, Relative mRNA levels of 13 mtDNA-encoded OXPHOS complex subunits in the liver of ND6 siRNA treated mice (n=6). E, Relative mRNA levels of nDNA-encoded OXPHOS complex subunits in the liver of ND6 siRNA treated mice (n=6). F, Body and tissue weight; G, Relative liver TG; H, Relative liver T-CHO; I, Serum HDL-c; J, serum LDL-c of mice; K, Serum TG levels in mice treated with ND6 siRNA for one week ( $n\geq6$ ). Values are mean  $\pm$  SEM. \*p< 0.05, \*\*p< 0.01.



Figure S5 Increased mitochondrial DNMT1 localization in the liver of HFD mice and db/db mice. A-B, Western blot analysis of liver DNMT1, DNMT3a and DNMT3b protein levels in total, mitochondrial and cytosolic fractions of HFD mice and (b) db/db mice. Values are mean  $\pm$  SEM. *n*=6, \*p< 0.05, \*\*p< 0.01.



Figure S6 Characterization of mouse adenovirus infection through tail vein injection. A, *GFP* mRNA expression in the tissues of mice infected with pAd/CMV/V5-*GFP* adenovirus via the tail vein injection (n=3). B, Relative *Dnmt1* mRNA level in the liver of mice infected with pAd/CMV/V5-*Dnmt1*(Ad-*Dnmt1*), pAd/CMV/V5-mt*Dnmt1*(Ad-mt*Dnmt1*) or pAd/CMV/V5-NC (Ad-NC) adenovirus (n=6). C-D, Protein levels of DNMT1 and ND6 in the liver of mice infected with adenovirus (n=6). E, Body and tissue weight of mice after *Dnmt1* overexpression (n=8). F, Relative mRNA level of nDNA-encoded OXPHOS complex subunits (n=6). G, Serum TG; H, Serum T-CHO; I, Serum HDL-c; J, Serum LDL-c of mice infected with pAd/CMV/V5-*GFP* adenovirus via the tail vein injection (n=6). Values are mean ± SEM. \*p< 0.05, \*\*p< 0.01.



Figure S7 AMPK activation is associated with PA induced *ND6* hypermethylation by DNMT1. A, Relative *ND6* mRNA level in HepG2 cells under dose dependent PA treatment for 24 h (n=5). B, Relative *ND6* mRNA level in HepG2 cells under time dependent PA treatment at 300 µM (n=5). C, Western blot analysis of DNMT1 protein subcellular localization in mouse primary hepatocytes under 300 µM PA treatment. D, Methylation levels of *D-loop* and *ND6* in PA treated cells with or without 2 µM 5-Aza for 24 h (n=5). E, Western blot of p-AMPK, AMPK, p-Akt, Akt, p-ERK1/2, ERK1/2, p-JNK, JNK, p-p38 and p38 in HepG2 cells under time dependent PA treated cells (n=4). G, Relative *ND6* mRNA level in cells after prolonged AICAR treatment (n=4). H, Relative *ND6* mRNA level in HepG2 cells under time dependent 2 µM SC79 treatment (n=3). J, *ND6* methylation and mRNA levels in PA treated cells with or without 10 µM LY294002 for 24 h (n=5). Values are mean ± SEM. \*p< 0.05, \*\*p< 0.01.



Figure S8 Hepatic knockdown of *Dnmt1* ameliorates metabolic abnormality induced by HFD. A-B, Relative mRNA levels of mtDNA-related transcription factors and nDNA-encoded OXPHOS complex subunits in the liver of mice under diets and AAV intervention (n=6). C, Relative complex II-V activities in liver (n=8). D-E, Relative TG and T-CHO level in the liver (n=8). F-K, Serum ALT activity, AST activity, TG level, T-CHO level, LDL-c level, and HDL-c level in mice (n=6). L-M, Fasting glucose and fasting insulin level in mice after diets and AAV intervention (n=6). Values are mean ± SEM. \*p< 0.05, \*\*p< 0.01.

Table S1 Probabilities of DNMT1 isoforms to mitochondria	predicted by	y MitoProt	Π.
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Species	Isoform	Length (amino acid)	Probability to mitochondria	Mitochondrial leader peptide
Human	NM_001130823.3 NP_001124295.1 DNMT1 isoform a	1632	47.94%	MPARTAPARV
Human	NM_001318730.2 NP_001305659.1 DNMT1 isoform c	1619	48.33%	MPARTAPARV
Human	NM_001318731.2 NP_001305660.1 DNMT1 isoform d	1511	38.96%	MADANSPPKPLSKPRTPRRS
Human	NM_001379.4 NP_001370.1 DNMT1 isoform b	1616	47.33%	MPARTAPARV
Human	DNMT1 isoform b supplemented with an additional MTS reported by Shock et al. (mtDNMT1)	1717	85.54%	MNECLGHRTHLPANRGAWTS PLLRVGGVCARLAHACSLGM AGSVPSFCTGYRLSPFGTSPPP PRPDWGGRRRLRSSPLPIGFRA KSRG
Human	NM_003201.3 NP_003192.1 TFAM isoform 1 precursor	246	92.26%	MAFLRSMWGVLSALGRSGAE LCTGCGSRLRSPFSFVYLPRWF SSVLASC
Mouse	NM_001199431.1 NP_001186360.2 Dnmt1 isoform 1	1620	36.13%	MPARTAPARV
Mouse	NM_001199432.1 NP_001186361.1 Dnmt1 isoform 3	1501	72.45%	MADSNRSPRSRPKPRGPRRS
Mouse	NM_001199433.1 NP_001186362.1 Dnmt1 isoform 4	1502	72.45%	MADSNRSPRSRPKPRGPRRS
Mouse	NM_001314011.1 NP_001300940.1 Dnmt1 isoform 5	1627	36.42%	MPARTAPARV
Mouse	NM_010066.4 NP_034196.5 Dnmt1 isoform 2	1619	36.13%	MPARTAPARV
Mouse	Dnmt1 isoform 1 supplemented with an additional MTS reported by Shock et al. (mtDnmt1)	1673	91.70%	MRTPFGHSMVFPHSLALCGT CCFRLRRPLPIGFRAREKAGV SFRAVLSSATCKMPARTAPARV

Table S2 Primers used in methylation-specific PCR.

Species	Region	Primer (5'-3')
Human	D-loop-M	Forward: CGTTTTTTTTAAATAAGATATTACGA
		Reverse: AAAAATCAAAAACAAATACTACGAC
Human	D-loop-U	Forward: TGTTTTTTTTAAATAAGATATTATGA
		Reverse: AAAAATCAAAAACAAATACTACAAC

Human ND6-M	Forward: TTTCGTATTAATAGGATTTTTTCGA
	Reverse: AATTATCTTTAAATATACTACAACGAT
Human ND6-U	Forward: TTTTGTATTAATAGGATTTTTTTGA
	Reverse: ATAATTATCTTTAAATATACTACAACAAT
Mouse <i>D-loop-M</i>	Forward: GTTTTTTTTAAATAAGATATTTCGA
	Reverse: TACTATCCTTTCATACCTTAACGAC
Mouse <i>D-loop-U</i>	Forward: GTTTTTTTTAAATAAGATATTTTGA
	Reverse: TACTATCCTTTCATACCTTAACAAC
	Forward: GATATTTTTTAGTAGTTATAGTAGTCGT
MD0-111	Reverse: TAATTTTAAAAATTTAATAAATCGTT
NDC U	Forward: TGGATATTTTTTAGTAGTTATAGTAGTTGT
MD0-0	Reverse: TAATAATTTTAAAAATTTAATAAATCATT
	ND6-M ND6-U D-loop-M D-loop-U ND6-M ND6-U

#### Table S3 Primers used in mtDIP.

Species	Region	Primer (5'-3')
Human	Dloop	Forward: TGCTTGTAAGCATGGGGAGG
	D-100p	Reverse: ACATTACTGCCAGCCACCAT
Human	ND6	Forward: GGGGTTTGTGGGGGTTTTCTTC
		Reverse: ACCCATATAACCTCCCCAA

#### Table S4 Primers used in real-time PCR.

Species	Gene	Primer (5'-3')
Mouse ND6		Forward: CTACCCCAATCCCTCCTT
	ND6	Reverse: GGTTTGGTGGATCGTTTT
N		Forward: GAGGCTGATGATGATGTGTC
Mouse	Mouse Dnmt1	Reverse: GGAATCATCTGGAATGACCG
		Forward: ATGTGGTTCGAGATTCTCCC
Mouse	Mouse Ndufa1	Reverse: GCAACTCGTTTTTCCTTGC
М		Forward: ATCCCATAGGTCAGACAAGC
Mouse	Mouse Ndufs1	Reverse: CAGTTGTGCGAACATATCCT
Mouse Sdha	C 11	Forward: AAGATTACAAAGTGCGGGTC
	Sana	Reverse: TTTCATCAGTAGGAGCGGAT
Mouse	Sdhb	Forward: GTTCTCGGCAGAGTCGG

		Reverse: TCCGCACTTATTCAGATCCA
Mouse	I I a sub	Forward: CACTCTCAGGTCAAAATGGC
	Uqcrb	Reverse: ATCTCGCATTAACCCCAGT
Mouse	I I a conce	Forward: GCGTCTATCTTCTGTCCCA
	Uqcrq	Reverse: ACCACTACAAACGGCGG
Mouse	Confe	Forward: CATTGGCTACCATGAGTTCC
	Coxoc	Reverse: CAGCCACGCCAAACTTATAG
Mouse	Cou7o	Forward: GAGCAAGGTCGTGTAGAAAG
	Cox/c	Reverse: GAAAATGGCAAATTCTTCCCC
Mauga	1 to 5 a 1	Forward: CATTTTGTGCCAGTCGTCC
Mouse	Агрэат	Reverse: ATTTTTGGAGACCAGTCCCG
Mouse	1 to 5 h 1	Forward: AATTCCTGTTGGTCCTGAGA
	Alpsol	Reverse: CCAAAGAGTCCGATTTTCCC
Mouso	Tfam	Forward: AGACTACACTGGGAAACCAC
wiouse	1 jum	Reverse: GTTAGGCTTATAGGGACCCA
Mouso	Teb Im	Forward: CCCAAACCAAAAGTTGATGT
wouse	IJDIM	Reverse: TTTCCTTCGAAACTGAAACG
Mouso	TA 2m	Forward: GGCATTACTTAAAGCTGGTG
wiouse	1jD2m	Reverse: TTTATAGGAACACCTGCTGAC
Mouso	Dolumet	Forward: TTCCCTCAGGAGTTTGTCTG
wiouse	FOIrmi	Reverse: TGATAGGGCTGTATGATGGG
Mouso	188 "DNA	Forward: GTAACCCGTTGAACCCCATT
wiouse	105 / 104	Reverse: CCATCCAATCGGTAGTAGCG
Mouso	D-loop	Forward: AGGCATGAAAGGACAGCA
wiouse	(for DNA)	Reverse: TTGGCATTAAGAGGAGGG
Mouso	18S rRNA	Forward: GAGAAACGGCTACCACATCC
wiouse	(for DNA)	Reverse: CACCAGACTTGCCCTCCA
Human	ND I	Forward: TTCGCCCTATTCTTCATAGC
Tuman		Reverse: GGAGGTTAGAAGTAGGGTCT
Human	ND2	Forward: ACACCCTTAATTCCATCCAC
		Reverse: GAGATAGGTAGGAGTAGCGT
Human	ND 3	Forward: AGAAAAATCCACCCCTTACG
riulliall	IND J	Reverse: CATGGTAGGGGTAAAAGGAG
Human	ND4	Forward: CTCTCTGTGCTAGTAACCAC

		Reverse: ATGGGGGGATAGGTGTATGAA
TT		Forward: CACACCTCATATCCTCCCTA
Human ND	ND4L	Reverse: GCATTGGAGTAGGTTTAGGT
	ND 5	Forward: CCTATTCGCAGGATTTCTCA
Human	ND5	Reverse: AATGTGCATAGTGGGGATTT
		Forward: CAACCATCATTCCCCCTAAA
Human	NDO	Reverse: GTGGGTTTAGTAATGGGGTT
	CVTD	Forward: AAACTTACTATCCGCCATCC
Human	CIIB	Reverse: GTAAGGGTGGAAGGTGATTT
11	COVI	Forward: CCTAATCACAGCAGTCCTAC
Human	COXI	Reverse: ATGGGAGATTATTCCGAAGC
Human	COV2	Forward: AAACCGTCTGAACTATCCTG
Human	COA2	Reverse: AATGGGGGAAGTATGTAGGA
Human	COY2	Forward: CGCTAAATCCCCTAGAAGTC
Human	COAS	Reverse: AGCCAATAATGACGTGAAGT
Human		Forward: GATCCCCACCTCCAAATATC
numan	AIFO	Reverse: TAAGAGATCAGGTTCGTCCT
Uumon		Forward: AACACAAACTACCACCTACC
numan	AIFO	Reverse: CAATGAATGAAGCGAACAGA
Human		Forward: ACCCAGAAGTAGGGTTTTGG
numan	NDUFAI	Reverse: TTCTCCAAACCCTTTGACAC
Uuman	NDUESI	Forward: CAAGGATTCTAGTCCCTCCG
TTuman	ND01'SI	Reverse: CTTAACATATTGCTTCTCCCCG
Uumon	SDHA	Forward: AACATGGAGGAGGACAACTG
TTuman	SDIIA	Reverse: CATATCGCAGAGACCTTCCA
Human	SDHR	Forward: TTGTTCCCGATTTGAGCAAC
Tuman	SDIID	Reverse: TCACGCTCTTCTATGGACTG
Human	UOCPR	Forward: TGGTCAAAATGGCTGGTAAG
Tuman	UQUND	Reverse: CTCGCATTAACCCCAGTTTA
Human	UOCPO	Forward: TCAGCTACAGCTTGTCACC
пuman	υψεκφ	Reverse: AAACACTACAAACTGCGGC
Human	COY6C	Forward: GCTTTGTATAAGTTTCGTGTGG
riuiiiall	CUAUC	Reverse: AACTGAGAAAGACTTACTGTCC
Human	COX7C	Forward: GATCTGCATTTGCTACACCC

	Reverse: GGCTGCACCTCTTAAAATGC
ATP5B1	Forward: CCTGTTGGTCCTGAGACTTT
	Reverse: CCAAAAAGCCCAATTTTGCC
TFAM	Forward: CCAAGAAGCTAAGGGTGATT
	Reverse: TGTTTCTTTATTGTGCGACG
TFB1M	Forward: AAGTTGTTAAGACTGCAAGC
	Reverse: TTTCAACCACCAGAAGTTCA
TFB2M	Forward: TGGATGGAAAACTACGAGTG
	Reverse: GGAACTGCTTCTATTCCCAA
POLRMT	Forward: AAGGTCAAGCAAATAGGAGG
	Reverse: GTCAGGCCCTTCCTGTA
18S rRNA	Forward: GTAACCCGTTGAACCCCATT
	Reverse: CCATCCAATCGGTAGTAGCG
D-loop	Forward: CAGTACCTAACAAACCCACA
(for DNA)	Reverse: GAGGTCGTAAACCCTATTGT
18S rRNA	Forward: TGGTGAGCTGCGAGAATAG
(for DNA)	Reverse: TTTTATGGTAATAACGCGGC
	ATP5B1 TFAM TFB1M TFB2M POLRMT 18S rRNA D-loop (for DNA) 18S rRNA (for DNA)