Supplementary information for

Structure and mechanism of the human NHE1-CHP1 complex

Yanli Dong, Yiwei Gao, Alina Ilie, DuSik Kim, Annie Boucher, Bin Li, Xuejun C. Zhang, John Orlowski, and Yan Zhao

This file contains Supplementary Figure 1–9 and Supplementary Table 1–2.



Supplementary Figure 1. Reconstitution of NHE1-CHP1 complex in nanodiscs.

a. Size-exclusion chromatogram (Superose 6 increase) of the purified sample of NHE1-CHP1 complex. Peak fractions (marked within black dashed lines) were used for nanodiscs reconstitution. **b.** Size-exclusion chromatogram of reconstituted NHE1-CHP1 complex into lipid nanodiscs. Peaks of NHE1-CHP1-MSP1D1 nanodiscs, empty nanodiscs, and cleaved GFP are indicated. **c.** Coomassie-blue-stained SDS-PAGE gel of the NHE1-CHP1-MSP1D1 nanodiscs. Components of the complex are labeled. NHE1 appeared in two bands due to different extent of glycosylation. The experiments were repeated independently with more than 3 times with similar results.



Supplementary Figure 2. Cryo-EM analysis of NHE1-CHP1^{Na/7.5} complex.

a. Flow chart of cryo-EM data processing. A total of 2,462 movie stacks were collected (Bar = 40 nm). A representative motion-corrected micrograph of this dataset is shown here. Particles were picked using Gautomatch, Topaz, and Template Picker in cryoSPARC, and were then submitted to several rounds of 3D classification, Bayesian polish, and CTF and 3D-auto refinement. The resulting map was reported at 3.3 Å. The CHP1 density is weak due to conformational flexibility. The same map of the lower threshold (transparent) and the higher threshold were overlaid to show CHP1 and TM helices features, respectively. Masks used for 3D classification and refinement were also shown as transparent surfaces alongside the arrows. **b.** Sharpened map of the NHE1-CHP1^{Na/7.5} complex, colored according to local resolution. **c.** Angular distribution of the particles in the designated orientation. **d.** The half-map and model-map Fourier shell correlation (FSC) curves of NHE1-CHP1^{Na/7.5} complex. **e.** Representative EM maps for the NHE1-CHP1^{Na/7.5} complex. Solution is the complex as well as lipid molecules.



Gautomatch



Supplementary Figure 3. Cryo-EM analysis of NHE1-CHP1^{Na/6.5} complex.

a. Flow chart of cryo-EM data processing. A total of 2,205 movie stacks were collected and motioncorrected (Bar = 40 nm). A representative motion-corrected micrograph of this dataset is shown here. Particles were picked using Gautomatch and Template Picker in cryoSPARC. Junk particles were removed using several rounds of 2D and 3D classification. The NHE1-focused reconstruction yielded a map with a reported resolution of 3.4 Å. The CHP1-focused classification resulted in a 4.0-Å map, composed of the NHE1 dimer and one CHP1 molecule. The same maps of the lower threshold (transparent) and the higher threshold were overlaid to show CHP1 and TM helix features, respectively. Masks used for 3D classification and refinement were also shown as transparent surfaces alongside the arrows. **b.** Sharpened map of the NHE1-CHP1^{Na/6.5} complex, colored according to local resolution estimation. **c**. Angular distribution of particles used in the final reconstruction. The length of each spike indicates of the number of particles in the designated orientation. **d–e.** FSC curves of the NHE1 dimer and NHE1-CHP1 complex, respectively, calculated between two independently refined half-maps before (red) and after (blue) post-processing, overlaid with an FSC curve calculated between the cryo-EM density map and the structural model shown in black. **f.** Representative EM maps for the NHE1-CHP1^{Na/6.5} complex.



Supplementary Figure 4. Cryo-EM analysis of NHE1-CHP1^{K/cariporide} complex.

3.5 Å

a. Flow chart of cryo-EM data processing. A total of 3,855 movie stacks were collected (Bar = 40 nm). A representative motion-corrected micrograph of this dataset is shown here. Particles were picked using Gautomatch and Topaz, and was subjected to 3D classification. One of the resulting class displayed a discernable shape of CHP1 and was thus selected and submitted to further 3D classification, Bayesian Polish, CTF refinement and 3D auto refinement. The resulting map was reported at 3.6 Å. Masks used for 3D classification and refinement were shown as transparent surfaces alongside the arrows. **b.** Sharpened map of NHE1-CHP1^{Na/7.5} complex, colored according to local resolutions. **c.** Angular distribution of the particles contributing to the final reconstitution. The length of each spike indicates of the number of particles in the designated orientation. **d.** The half-map and model-map FSC curves of the NHE1-CHP1^{K/cariporide} complex. **e.** Representative EM maps for the NHE1-CHP1^{Na/7.5} complex and lipid molecules.



Supplementary Figure 5. Architecture of the NHE1-CHP1 complex.

a. A topology model of secondary structures of the NHE1-CHP1 complex. **b.** Superposition of the complex structures of NHE1-CHP1^{Na/6.5} (grey) and NHE1-CHP1^{Na/7.5} (red). **c.** Structural comparison of the NHE1-CHP1^{K/cariporide} (blue) and NHE1-CHP1^{Na/6.5} (grey) complexes. **d.** Structural comparison of the NHE1-CHP1^{Na/6.5} and horse NHE9 (PDB ID: 6Z3Z), which colored in grey and magenta, respectively. **e.** Structural

comparison of the NHE1-CHP1^{Na/6.5} complex with PaNhaP (PDB ID: 4CZA). The NHE1-CHP1^{Na/6.5} complex and PaNhaP are colored in grey and green, respectively. **f.** TM helix bundle composing of TMs 7–9. G305 is shown as a sphere.

bNHE1	1	MUL BSGTCGL					BBUNUSUTDU
hNHE2	1	MEPLGNWR	SLRAPLPPMLLLLLL	VGLLF VLKSHGLQLS VAGPVGALAETLLN	APRAMGTSSSPPSPASVV	APGTTLFEESR	
hNHE3	1	MWGL0	GARGPDRGLLLALAL	GLARAG	GVEVEPGGAHGE	ESGG	
hNHE4	1	FV	TY-SPWNCLLLLVAL	CSEASSDLNESAN	STAQYASNAWF	AAASSEPEEG	
hNHE5	1	MLRA	ALSLLALPL	AGAAEEP	TQKPESPGEP	-PPG	
hNHE6	1	MARRGWRRAPLRRGVGSSP	RARRLMRPLWLLLAV	SVFDWAGASD-GGGGB	EARAMDEEIVSEKQAEESH	HRQDSANLLIFII	LLTLT
hNHE7	1	MEPGDAARPGSGRATGAPP	-PRLLLLPLLLGWGL	RVAAAASASSSGAAAH	EDSSAMEELATEKEAEESH	RQDSVSLLTFII	LLTLT
hNHE8	1		MGE	KMAEEERFPNTTHEGH	FNVTLHTTLVVTTKLVLP	FPGKPILPVQTG-	
hNHE9	1	MER			QSRVMSEKDEYQFÇ	QHQGAVELLVFN	LLILT
MJNnaP1 DoNihoD	1						
Familar	1						
		EL1	TM 1	IL1	TTM 2	EL:	2
			TWT		172		
LATTE 1	07						
NHE1	82 67		IPFEISLWILLACLM IPFEITUWILLASIAI	CIGFHVIPTISSIVPI	SCLLIVVGLLVGGLIKG	G-ETPP	
hNHE3	41	FOVVTFEWAHVO	DPYVTALWILLASLA	CIGFHLSHKVTSVVPI	SSCHEIMVGHEHGGIIVGV	ADHIASF	
hNHE4	56	ISVFELDYDYVO	IPYEVTLWILLASLA	KIGFHLYHRLPGLMPH	ESCLLILVGALVGGIIFG	DHKSPP	
hNHE5	33	LELFRWOWHEVE	APYLVALWILVASLA	(IVFHLSRKVTSLVPI	ESCLLILLGLVLGGIVLA	/AKKAEY	
hNHE6	84	ILTIWLFKHRRAR	FLHETGLAMIYGLLV	JLVLRYGIHVPSDVN	NVTLSCEVQS-SPTTLLV-		
hNHE7	84	ILTIWLFKHRRVR	FLHETGLAMIYGLIV	WILRYGTPATSGRD	-KSLSCTQEDRAFSTLLVN	NVSGKFFEYTLKO	EISPGKINSV
hNHE8	48	EQAQQEEQSSG	MTIFFSLLVLAICII	LVHLLIRYRLH-FLPH	ESVAVVSLGILMGAVIKI	EFKKLAN	
hNHE9	34	ILTIWLFKNHRFR	FLHETGGAMVYGLIM	LILRYATAPTDIES	GTVYDCVKLTFSPSTLLVN	NITDQVYEYKYKI	REISQHNINPH
MjNhaP1	1	ME	LMMAIGYLGLALVLG	SLVAKIAEKLKIPI	DIPLLLLLGLIIGPFLQII	IPSDSAM	
PaNhaP	1	MI	ELSLAEALFLILFTG	/ISMLISRRTGIS	VVPIFILTGLVIGPLLKL	IPRDLAH	
						EL3	
			тмз —	[L2a —	TM4		- TM5a
							-
hNHE1	154	FLQSDVFFLF	LLPPIILDAGYFLPL	ROFTENLGTILIFAV	VGTLWNAFFLGGLMYAVCI	LVGGEQ-IN	IGLLDNLLFG
hNHE2	134	AMKTDVFFLY	LLPPIVLDAGYFMPT	RPFFENIGTIFWYAVV	VGTLWNSIGIGVSLFGICÇ	QIEAFG-LSI	DITLLQNLLFG
hNHE3	108	TLTPTVFFFYLLPPIVLDAGYFMPNRLFFGNLGTILLYAVVGTVWNAATTGLSLYGVFLSGLMG-DLQIGLLDFLLFG					
INHE4	123			RPFFENIGSILWWAVI	LGALINALGIGLSLYLICÇ	2VKAFG-LGI	DVNLLQNLLFG
hNHE6	144						
hNHE7	168	EONDMLRKVTFDPEVFFNILLPPITFHAGYSLKKRHFFRNIGSTLAIAFLGTAISUFVIGSIMIGUVILMKVIGULA-GDFIFTDULLFG					
hNHE8	114	WKEEEMFRPNMFFLLLLPPIIFESGYSLHKGNFFQNIGSITLFAVFGTAISAFVVGGGIYFLGOADVISKLNMTDSFAFG					
hNHE9	119	QGNAILEKMTFDPEIFFNVLLPPIIFHAGYSLKKRHFFQNLGSILTYAFLGTAISCIVIGLIMYGFVKAMIHAGQLKNGDFHFTDCLFFG					
MjNhaP1	55	EIFEYAGPIGLIFILLGGAFTMRISLLKRVIKTVVRLDTITFLITLLISGFIFNMVLNLPYTSPVGYLFG					
PaNhaP	55	EIFDFVRVF	GLVIILFTEGHNLSW	RLLKKNMPTIVTLDT	IGLILTALIAGFIFKVVFN	1S	-SFLLGFLFG
		T	r.3		ET.4		
		— TM5b		TM6		TM7	
				T			
hNHE1	232	SIISAVDPVAVLAVFEEIH	INELLHILVFG <mark>E</mark> SLLI	NDAVTVVLYHLFEEF#	ANYEHVGIVI	DIFLGFLSFFVV	LGGVLVGVVY
hNHE2	212	SLISAVDPVAVLAVFENIH	VNEQLYILVFGESLL	NDAVTVVLYNLFKSF(CQMKTIETII	DVFAGIANFFVV	GGVLIGIFL
hNHE3	186	SLMAAVDPVAVLAVFEEVHVNEVLFIIVFGSSLLNDAVTVVLYNVFESFVALGGDNVTGVDCVKGIVSFFVVSLGGTLVGVVF					
INHE4	201	SLISAVUFVAVLAVFEEARVNEQLYMMIFGBALLNDGITVVLYNMLIAFTKMHKFEDIETVDILAGCARFIVVGLGGVLFGIVF					
hNHE6	225	AIVSATOPVTVLAIFHELO	VDVELVALLEGESVL	DAVIVULIRVENSF	DPAGDNSHTFDVT	AMEKSIGIFI.GI	SGSFAMGAAT
hNHE7	257	AIISAT	ADVDLYALLFGSVL	DAVAIVLSSSIVAY	OPAGLNTHAFDAA	AFFKSVGIFLGI	SGSFTMGAVT
hNHE8	195	SLISAVDPVATIAIFNALH	VDPVLNMLVFG <mark>E</mark> SIL	DAVSIVLTNTAEGL	rknmsdvsGwq1	FLQALDYFLKM	FGSAALGTLT
hNHE9	209	SLMSAT <mark>D</mark> PVTVLAIFHELH	VDPDLYTLLFG <mark>E</mark> SVL	NDAVAIVLTYSISIYS	SPK-ENPNAFDAA	AFFQSVGNFLGI	AGSFAMGSAY
MjNhaP1	126	AITAAT <mark>D</mark> PATLIPVFSRVR	INPEVAITLEA <mark>E</mark> SIF	1 <mark>D</mark> PLGIVSTSVILGLI	FGLFS	SSSNPLIDLITL	GGAIVVGLLL
PaNhaP	124	AIIGAT <mark>D</mark> PATLIPLFRQYR	VKQDIETVIVT <mark>E</mark> SIF	N <mark>D</mark> PLGIVLTLIAISMI	LVPGYGGGIFSTLSEKLGI	IYAGGVIYFLYNV	/SVSISL <mark>G</mark> IFL
		TT 4		PT F		TTE	
		TM7 114	TM8	<u>- 6113</u>	тм9	<u>611</u>	TM10
hNHE1	313	GVIAAFTSRFTSHIRVI	EPLFVFLYSYMAYLS	AELFHİSGIN	MALIASĠVVMRPYVEANI-	SHKS	SHTTIŘYFLKM
hNHE2	293	GFIAAFTTRFTHNIRVI	EPLFVFLYSYLSYIT	AEMFHLSGIN	AITACAMTMNKYVEENV-	SQK8	SYTTIKYFMKM
hNHE3	269	AFLLSLVTRFTKHVRII	EPGFVFIISYLSYLT	SEMLSLSAII	LAITFCGICCQKYVKANI-	SEQS	SATTVRYTMKM
hNHE4	285	GFISAFITRFTQNISAI	EPLIVFMFSYLSYLA	AETLYLSGI1	LAITACAVTMKKYVEENV-	SQTS	SYTTIKYFMKM
hNHE5	262	AFLLALTTRFTKRVRIIEPLLVFLLAYAAYLTAEMASLSAILAVTMCGLGCKKYVEANISHKSRTTVKYTMKT			GRTTVKYTMKT		
hNHE6	310	GVVTALVTKFTKLREFQLL	STGLFFLMSWSTFLL	LEAWGFTGV	VAVLFCGITQAHYTYNNL-	STES	SQHRTKQLFEL
NNHE7	342	G-VNANVTKFTKLHCFPLL	ETALFFLMSWSTFLL	ALACGFTGW	VAVLFCGITQAHYTYNNL-	SVES	OT NOOT PT
hNHEQ	219	ATTTALL TKETKI CEEDMI	ETGI.FFI.I.SWEAFI C	XEAAGITCI	VAVLECGUTOAHVTVNNT -		KIRTKOLEFE
MiNhaP1	202	AKIYEKIIIHCDFHEYV	APLVLGGAMI.I.I.VVC	DLLPSICGYGFSCV	AVAIMGLYLGDALFRADI	 TT	YKYIVSFCDD
		CTI CYKETKETCIEDEET	FARST START CEFTCI	PIDACCVI	WATUTCI VI CNYKI I KR		TEVENUENDE

		TM10	EL6	TM11	IL6	- TM12a	<u>——тм12b_егл</u>
bNHE1	396	WSSVEFTT TETEL CUSTU	ACS_HHWNW			TRKDOPTTA	VCCI PCATAFSI GVI I DKKNE
hNHE2	366	I.SSVSETI.IFIFMGVSTV	CKN-HEWNW	AFVCETLAFCLMWRALC	VEGLINF INRFRITRI	UTEKDOETTA	VGGLEGATCEALVELLPAAVE
hNHE3	342	LASSAETITEMELGISAV	NPFTWTWNT	AFVILTI.VETSVVRATC	WVI.OTWI.I.NRVRMVOI	LEPTDOVVI.S	VGGLEGAVAFALVVLLDGDKV
hNHF4	358	LASSALTITA DOIDAV	GKN_HEWNW	AFTCETLAFCOTWRATS	VFALEVISNOFRTEP	ESTRDOCTTR	VSGVRGAGSESLAFLLPLSLF
hNHE5	335	LASCAETVIEMLIGISAV	DSSKWAWDS	CLVLGTLIFTLFFRALG	VVI.OTWVI.NOFRI.VPI		VGGLEGAVAFALVILLDETKV
hNHE6	385	LNFLAENFIFSYMGLTLF	TFONHVENP	TEVVGAEVATELGRAAN	TYPLSLLLNLGRRSK	GSNFOHMMM	FAGLEGAMAFALATEDTAT
hNHE7	416	LHFLAENFIFSYMGLALF	TFOKHVFSP	IFIIGAFVAIFLG <mark>R</mark> AAH	IYPLSFFLNLGRRHK	LGMNFOHMMM	FSGLRGAMAFALAIRDTAS
hNHE8	354	VAFLCETCVFAFLGLSIF	SFP-HKFEI	SFVIWCIVLVLFG <mark>R</mark> AVN	IFPLSYLLNFFRDHK	TPKMMFIMW	FSGLRGAIPYALSLHLDLEPM
hNHE9	368	MNFLAENVIFCYMGLALF	TFONHIFNA	LFILGAFLAIFVARACN	IYPLSFLLNLGRKOK	IPWNFOHMMM	FSGLRGAIAFALAIRNTES
MjNhaP1	280	LSLLARVFIFVFLGACIK	LSMLENYFI	PGLLVALGSIFLAR PLG	VFLGLIGSKHS	FKEKLYFA	LEGP <mark>R</mark> GVVPAALAVTVGIEIL
PaNhaP	297	LAALATIFIFVLLGAEMN	LEVIWSNLG	KGLLVALGVMILA <mark>R</mark> PLA	TLPLLKWWN	FREYLFIA	LEGP <mark>R</mark> GVVPSALASLP
		_ тм1	3	IL7	HC1	IL8	нс? —
			3				
		·	:	• • • • • • • • • • • • • • • • • • • •	······································		·
hNHE1	475	PMCDLFLTAIITVIFFTV	FVQGMTIRP	LVDLLAVKKKQETKRSI	NEEIHTQFLDHLLTG	LEDICGHYGH	HHWKDKLNRFNKKYVKKCLIA
hNHE2	455	PRKKLFITAAIVVIFFTV	FILGITIRP	LVEFLDVKRSNKKQQAV	SEEIYCRLFDHVKTG	LEDVCGHWGH	NFWRDKFKKFDDKYLRKLLIR
INHE3	432	KERNLFVSTT11VVFFTV	TFQGLTIKP	LVQWLKVKRSEHREPRI	NEKLHGRAFDHILSA.	LEDISGQIGH	NYLRDKWSHFDRKFLSRVLMR
INHE4	447	PRKKMFVTATLVVIYFTV	FIQGITVGP	LVRYLDVKKTNKKE-SI	NEELHIRLMDHLKAG.	LEDVCGHWSH	YQVRDKFKKFDHRYLRKILIR
INHES	425	PARDYFVATTIVVVFFTV	IVQGLTIKP	LVKWLKVKRSEHHKPTL	NQELHEHTFDHILAA	VEDVVGHHGY	HIWRDRWEOFDRRILSOLLMR
IINTIE0	4/3	YAROMMF STILLIVFFTV	WVF GGGTTAI	MLSCLHIKVGVDSDQE-	HLGV		SEOVI OCDORDS A DONDWOR
IIINFIE / hNILIES	504 112	IARQMMFTTTLLIVFTVMIIGGGTTPMLSWLNIRVGVEEPSEEDQNEHHWQYFRVGVDPDQDPPPNNDSFQVLQGDGPDSARGNRTKQE					
hNHE0	445	ERRULIGITTI VIVLTILLIGGSTMPLI KLMU JEDAKAHKKN					
MiNhaP1	364	QPRQMMFTTTLLLVFFTVNVFGGGTPMLTWLQIRVGVDLDENLKEDPSSQHQEANNLDKNMTKAE					
PaNhaP	373	NNAUKIFASIINIIFIDIANTIIIGIFMIILS-					
i ur thur	575			1111110			1112
		IL9 UC	•				
		HC.	•				
hNHE1	565	GERSK-ÉPQLIAFYHKME	MKQAIELVĖ	815			
hNHE2	545	ENQPKSSIVSLYKKLEIKHAIEMAE812					
hNHE3	522	RSAQKSRDRILNVFHELNLKDAISYVA834					
hNHE4	536	KNLPKSSIVSLYKKLEMKQAIEMVE798					
hNHE5	515	RSAYRIRDQIWDVYYRLNIRDAISFVD896					
hNHE6	531	SAWLFRMWYNFDHNYLKPLLT669					
hNHE7	594	SAWIFRLWYSFDHNYLKPILT725					
hNHE8	493	KTEKMGNTVESEHLSELTE581					
hNHE9	522	SARLFRMWYSFDHKYLKPILT645					
MjNhaP1	402	ASWAGMLALKLL	GEYKPKYKE	443			
PaNhaP	409	TLWIPILKDKLD		426			

Supplementary Figure 6. Sequence alignment of sodium/proton exchangers.

Secondary structural elements of hNHE1 are marked above the sequence alignment, and unmodeled loops are represented as dashed lines. Helices of the dimerization domain, core domain, and from cytoplasmic region are colored in grey, purple, and pink. Mutation sites are marked as red triangles. Conservative residues among these proteins are highlighted in red, blue and grey for acidic, basic, and others, respectively.



Supplementary Figure 7. Structural analysis of core domains.

a–b. Structural comparison of the core domains viewed parallel to the membrane and from the intracellular side of the membrane, respectively. The core domain of NHE1-CHP1^{Na/7.5} complex and NHE1-CHP1^{K/cariporide} complex are colored in red and blue, respectively. Sidechains of the E262 and R425 are shown in sticks. **c–d.** Closeup view of the model around TM5b and IL3 in the inward- and outward-facing conformations, respectively. Selected residues D238, D247, D248, and R500 are shown as sticks. TM12 is colored in a cyan-to-green spectrum. **e–f.** Local structures of TM5b, TM6, TM13, IL3, and IL7 in NHE-CHP1^{Na/7.5} complex (**e**) and NHE1-CHP1^{K/cariporide} (**f**). The corresponding densities of TM5b, IL3, and IL7 are shown in transparent grey surface.



Supplementary Figure 8. Biochemical and functional analysis of NHE1-CHP1 complex.

a. Cellular distribution of NHE1 in AP-1 cells. AP-1 cells were transiently transfected (48 h) with wild-type NHE1 tagged at its C-terminus with mCherry fluorescent protein alone or wild type or mutant NHE1 co-expressed with either wild-type CHP1 or mutant CHP1 (R192A^{CHP1}), as indicated. Scale bars represent 10 μ M. Images are representative of at least 6 different cells from two independent transfections. **b–c.** Assessment of protein expression of NHE1. Blots were stripped and reprobed with a mouse monoclonal anti- β -tubulin antibody to control for protein loading. Blots are representative images from five separate experiments. **d.** Quantitation of protein expression was determined by densitometry of X-ray films exposed within the linear range of the films. To control for subtle loading variations, NHE1 expression determined relative to endogenous β -tubulin (ratio of NHE1/ β -tubulin) and then normalized to 100%. Values represent the mean \pm S.D. (n = 5 independent blots). **e.** Measurement of cytoplasmic pH following an NH₄Cl-imposed acid-load in AP-1 cells transfected with NHE1 and CHP1. NHE1 activity was defined as the initial linear rate of pH recovery as a function of time upon reintroduction of Na⁺-rich solution. Tracings are a representative experiment (mean \pm S.E.M.; NT, n = 8 cells; NHE1, n = 3 cells, NHE1-CHP1, n = 4 cells; D238A^{NHE1}-CHP1, n = 4 cells; NHE1-R192A^{CHP1}, n = 4; Y577A/H578A^{NHE1}-CHP1, n = 5 cells) of 2 or 3 independent experiments. Source data are provided as a Source Data file.



Supplementary Figure 9. Conformational heterogeneity and electrostatic potentials of the NHE1-CHP1 complex.

a–d. Representative conformations of CHP1 (pink) in the NHE1-CHP1^{Na/6.5} complex (**a–b**) and NHE1-CHP1^{Na/7.5} complex (**c**-**d**). **e**-**g**. The electrostatic potentials of NHE1 in NHE1-CHP1^{Na/6.5} complex (**e**) and NHE1-CHP1^{K/cariporide} complex (f). g. The electrostatic potential of CHP1. The NHE1 and CHP1 interaction areas are marked.

	NHE1-CHP1 ^{Na/7.5} complex	NHE1-CHP1 ^{Na/6.5} complex	NHE1-CHP1 ^{K/cariporide} complex		
Accession					
PDB ID	7DSW	7DSV	7DSX		
EMDB Accession	EMD-30848	EMD-30847	EMD-30849		
Data Collection					
Microscope	Titan Krios	Talos Arctica	Titan Krios		
Camera	K2 Summit	K2 Summit	K2 Summit		
Voltage (kV)	300	200	300		
Magnification	130,000 ×	130,000 ×	130,000 ×		
Pixel Size (Å)	1.04	1	1.04		
Defocus Range (µm)	-1.2 ~ -2.2	-1.2 ~ -2.2	-1.2 ~ -2.2		
Energy Filter Slit Width (eV)	20	20	20		
Total Dose (e/Å ²)	60	50	60		
Micrographs (No.)	2,462	2,205	3,855		
Final Particles (No.)	108,712	100,503	61,460		
Map Resolution (Å)	3.3	3.4	3.5		
	Model	Validation			
Number of Atoms	6,736	6,768	10,982		
Residues	840	840	1,370		
Ligands	N/A	N/A	Cariporide		
Sharpening B-factor (Å ²)	131	175	84		
R. M. S. Deviations					
Bond Lengths (Å)	0.006	0.006	0.009		
Bond Angles (°)	0.779	0.77	1.046		
Ramachandran					
Favored (%)	91.87	92.34	91.16		
Allowed (%)	8.13	7.66	8.84		
Outlier (%)	0.00	0.00	0.00		
MolProbity	2.27	2.11	2.31		
EMRinger	1.35	1.54	1.96		

Supplementary Table 2. Primers used in this study.

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Name	Sequence
NHE1-H-SalI-F	ATACGCGTCGACATGGTTCTGCGGTCTGGCAT
NHE1-H-NotI-R	AAATATGCGGCCGCCTGCCCCTTGGGGAAGGACG
CHP1-H-pET21a-F	CTGGTGGACAGCAAATGGGTCGCGGATCCGAATTCATGGGTTCTCGGGCC
	TCCA
CHP1-H-pET21a-R	TCTCAGTGGTGGTGGTGGTGGTGATGATGCTCGAGGTGAAGAAATCGGAT
	GCTCA
CHP1-H-SalI-F	ATACGCGTCGACATGGGTTCTCGGGGCCTCCA
CHP1-H-NotI-R	AAATATGCGGCCGCCTGCCCCTTGGGGAAGGACG
NHE1-CHP1-biGBac1a-D238A ^{NHE1} -F	AGCATCATCTCGGCCGTGGCACCCGTGGCGGTTCTGGCTGTCTTT
NHE1-CHP1-biGBac1a-D238A ^{NHE1} -R	AAAGACAGCCAGAACCGCCACGGGTGCCACGGCCGAGATGATGCT
NHE1-CHP1-biGBac1a-Y577A/H578A ^{NHE1} -F	GAGCCCCAGCTCATTGCCTTCGCCGCCAAGATGGAGATGAAGCAGGCCAT
	CG
NHE1-CHP1-biGBac1a-Y577A/H578A ^{NHE1} -R	CGATGGCCTGCTTCATCTCCATCTTGGCGGCGAAGGCAATGAGCTGGGGC
	TC
NHE1-CHP1-biGBac1a-R192A ^{CHP1} -F	GTAGAACAGAAAATGAGCATCGCATTTCTTCACTGATCTAG
NHE1-CHP1-biGBac1a-R192A ^{CHP1} -R	CTAGATCAGTGAAGAAATGCGATGCTCATTTTCTGTTCTAC