

## Electronic Supplementary Information

# Unlocking the PIP-box: A peptide library reveals interactions that drive high affinity binding to human PCNA

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### ABBREVIATIONS

**ACN**, acetonitrile; **Ala (A)**, alanine; **Arg (R)**, arginine; **Asn (N)**, asparagine; **Asp (D)**, aspartic acid; **Boc**, tert-butoxycarbonyl; **Cys (C)**, cysteine; **DCM**, dichloromethane; **DEAE**, diethylethanolamine; **DIC**, *N,N'*-diisopropylcarbodiimide; **DIPEA**, *N,N'*-diisopropylethylamine; **DMF**, *N,N'*-dimethylformamide; **DDOT**: 2,2'-(ethylenedioxy)diethanethiol; **DTT**, dithiothreitol; **E. Coli**, Escherichia Coli; **EDC**, 1-ethyl-3-(3-diethylaminopropyl)carbodiimide; **EDTA**, ethylenediaminetetraacetic acid; **ESI-TOF**, Electrospray Ionisation Time of Flight; **Fmoc**, 9-fluorenylmethoxycarbonyl; **Gln (Q)**, glutamine; **Glu (E)**, glutamic acid; **Gly (G)**, glycine; **HATU**, (1-[bis(dimethylamino)methylene]-1H-1,2,3-triazolo[4,5-b]pyridinium 3-oxide hexafluorophosphate; **HEPES**, 4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid; **His (H)**, histidine; **HOBt**, 1-hydroxybenzotriazole; **HPLC**: High Performance Liquid Chromatography; **HRMS**: High Resolution Mass Spectrometry; **Ile (I)**, isoleucine; **IPTG**, isopropyl  $\beta$ -D-1-thiogalactopyranoside; **LB**: Lennox broth; **LCMS**: Liquid Chromatography Mass Spectrometry; **Leu (L)**, leucine; **Lys (K)**, lysine; **Met (M)**, methionine; **NHS**, N-hydroxysuccinimide; **Pbf**: 2,2,4,6,7-pentamethylidihydrobenzofuran-5-sulfonyl; **PCNA**, Proliferating Cell Nuclear Antigen; **Phe (F)**, phenylalanine; **Pro (P)**, proline; **RP-HPLC**: Reverse-Phase High Performance Liquid Chromatography; **SDS**, Sodium Dodecyl Sulfate; **Ser (S)**, serine; **SPPS**: Solid-Phase Peptide Synthesis; **tBu**: tert-butyl; **TFA**: trifluoroacetic acid; **Thr (T)**, threonine; **TIPS**: triisopropylsilane; **TNBS**, 2,4,6-trinitrobenzenesulfonic acid (picrylsulfonic acid); **Tris**, trisaminomethane; **Trp (W)**, tryptophan; **Trt**, trityl; **Tyr (Y)**, tyrosine; **Val (V)**, valine;

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## SYNTHESIS & CHARACTERISATION OF PEPTIDES

Unless otherwise indicated, all starting materials were purchased from commercial sources and used without further purification. All peptides were synthesised by the Fmoc/tBu solid-phase peptide synthesis using one of three protocols detailed below, with all L-amino acids (unless otherwise specified) purchased from Chem-Impex Int'l.: Fmoc-Ala-OH, Fmoc-Arg(Pbf)-OH, Fmoc-Asp(tBu)-OH, Fmoc-Asn(Trt)-OH, Fmoc-Cys(Trt)-OH, Fmoc-Gln(Trt)-OH, Fmoc-Glu(tBu)-OH, Fmoc-Gly-OH, Fmoc-His(Trt)-OH, Fmoc-Ile-OH, Fmoc-Leu-OH, Fmoc-Lys(Boc)-OH, Fmoc-Met-OH, Fmoc-Thr(tBu)-OH, Fmoc-Tyr(tBu)-OH, Fmoc-Trp(Boc)-OH, Fmoc-Ser(tBu)-OH, Fmoc-Phe-OH, Fmoc-Pro-OH, Fmoc-Val-OH. Peptides were subsequently cleaved from the resin (and simultaneously globally deprotected) and purified per '*Cleavage from solid-support, isolation & purification*'. Purity of all compounds was confirmed by analytical RP-HPLC on an Agilent 1260 HPLC equipped with a Phenomenex Luna C18(2) column (250 x 4.6 mm) over a gradient of 5-50% B (15 min) and visualised at 220 nm. High-resolution mass spectra were collected using an Agilent 6230 ESI-TOF via direct injection in ACN with 0.1% formic acid as the running buffer. Characterisation data for all peptides is recorded in Table S1.

### Peptide Synthesis:

All peptides were synthesised using one of the following methods, excepting ten peptides that were purchased from Shanghai Royobiotec at >95% purity, with purity & identity confirmed on receipt by HPLC and HRMS. *Purchased peptides*: p21<sub>μ</sub>-M147L, p21<sub>μ</sub>-M147A, p21<sub>μ</sub>-M147W, p21<sub>μ</sub>-M147V, p21<sub>μ</sub>-Q144D, p21<sub>μ</sub>-Q144M, p21<sub>μ</sub>-Q144S, p21<sub>μ</sub>-FY150/151YF, p21<sub>μ</sub>-p15, and p21<sub>μ</sub>-Cdt1.

### Liberty Blue Synthesis:

All peptides were synthesised by the Fmoc/tBu solid-phase peptide synthesis protocol on a CEM Liberty Blue Automated Microwave Peptide Synthesiser (CEM Corp., Matthews, NC, USA) using the standard manufacturer's conditions. The peptides were assembled on 0.1 mmol scale on Chem Impex Rink Amide AM resin (0.47 mmol/g) or Mimotopes Rink Amide resin (0.456 mmol/g). The resin was initially swollen in DCM (10 mL, 15 min) and then the resin washed with DMF (2 × 5 mL) and transferred to the microwave reaction vessel. The resin-bound Fmoc-groups were deprotected with a mixture of 20% piperidine and 0.1 M OxymaPure in DMF using the standard microwave deprotection method with a maximum temperature of 90°C. Couplings were performed with Fmoc-protected amino-acids (0.2 M in DMF, 5 equiv), OxymaPure (1 M in DMF, 5 equiv) and DIC (0.5 M in DMF, 5 equiv) under the 'Standard Coupling' microwave method with a maximum temperature of 90°C, except for coupling of Fmoc-His(Trt)-OH which was coupled using a maximum 50°C 10 min coupling procedure; and Fmoc-Arg(Pbf)-OH which used the default 'Arginine Double Coupling' microwave method which included two couplings steps – the first at room temperature and the second at a maximum of 75°C. Following assembly of the desired sequence the N-terminal protecting group was removed. The resin was then removed from the synthesiser, washed with DCM (3 × 5 mL) and then diethyl ether (3 × 5 mL) and air dried with suction. The peptides were then cleaved from the resin according to the protocol '*Cleavage from solid-support, isolation & purification*' detailed below.

*Peptides synthesised by this method*: p21<sub>139-160</sub><sup>‡</sup>, p21<sub>μ</sub>, p21<sub>μ</sub>-F150Y, p21<sub>μ</sub>-Y151F, p21<sub>μ</sub>-F150H, p21<sub>μ</sub>-M147I

<sup>‡</sup> The N-terminal glycine was subjected to a standard double coupling cycle as initial syntheses indicated incomplete incorporation at this position. Please note, this sequence is particularly prone to aspartimide formation.

### Prelude Synthesis:

Peptides were assembled using a Protein Technologies Prelude® Peptide synthesiser. Chem Impex Rink Amide resin (0.1 mmol, 0.47 mmol/g) was preswollen in DCM for 15 min, and washed with DMF (3 × 10 mL, 1.5 min). The Fmoc-protecting group was removed by treatment with 20% piperidine (3 × 8 mL, 7 min) and the resin washed with DMF (5 × 10 mL, 1.5 min). An amino-acid was then coupled by addition of the required Fmoc-amino-acid (7.5 equiv, 150 mM in DMF), HCTU (5 equiv, 0.5 M in DMF) and DIPEA (10 equiv, 1 M in DMF) and bubbled with nitrogen for 15 min before the solution was drained, and a fresh coupling solution added to the resin and bubbled with nitrogen for 10 min. The solution was drained and the resin washed with DMF (4 × 8 mL, 1.5 min). Fmoc deprotection and coupling steps were repeated until the desired sequence was achieved, and the final N-terminal Fmoc group deprotected. The resin was then removed from the synthesiser and washed

with DCM (3 × 5 mL) and then diethyl ether (3 × 5 mL) and air dried with suction. The peptides were then cleaved from the resin according to the protocol '*Cleavage from solid-support, isolation & purification*' detailed below.

*Peptides synthesised by this method:* p21<sub>μ</sub>-Q144K, p21<sub>μ</sub>-Q144N, p21<sub>μ</sub>-T145K, p21<sub>μ</sub>-T145R, p21<sub>μ</sub>-S146K, p21<sub>μ</sub>-S146R, p21<sub>μ</sub>-T148D, p21<sub>μ</sub>-T148E, p21<sub>μ</sub>-D149E, p21<sub>μ</sub>-TS145/146KK, p21<sub>μ</sub>-TS145/146RK, p21<sub>μ</sub>-TS145/145KR, p21<sub>μ</sub>-TS145/146RR, p21<sub>μ</sub>-TD148/149EE, p21<sub>μ</sub>-TS148/149DE, p21<sub>μ</sub>-RNaseH2B, p21<sub>μ</sub>-MCMT, p21<sub>μ</sub>-pol λ, p21<sub>μ</sub>-pol β, p21<sub>μ</sub>-pol δ<sub>p66</sub>, p21<sub>μ</sub>-FEN1, p21<sub>μ</sub>-RFC, p21<sub>μ</sub>-RecQ5, p21<sub>μ</sub>-DNALig1, p21<sub>μ</sub>-WRN, p21<sub>μ</sub>-XPG, p21<sub>μ</sub>-Cdt2 p21<sub>μ</sub>-Pogo, p21<sub>μ</sub>-pol η, p21<sub>μ</sub>-pol κ.

#### *Manual Synthesis*

Rink Amide PL resin (0.1 mmol, 322 mg, 0.31 mmol/g, Agilent) was swollen in 1:1 DMF/DCM (15 mL) for 15 min. The Fmoc-protecting group was removed by treatment of the resin with a solution of 20% piperidine and 0.1 M HOBt in DMF (5 mL) for 15 min. The solution was drained and the resin washed with DMF (3 × 5 mL). Amino-acid couplings were achieved by addition of a solution of Fmoc-protected amino-acid (5 equiv), HATU (5 equiv) and DIPEA (10 equiv) in DMF (5 mL), to the resin and stirred intermittently for 1 h. The solution was drained and the resin washed with DMF (5 × 5 mL). The *N*-terminal Fmoc-protecting group was removed by treatment of the resin with a solution of 20% piperidine and 0.1 M HOBt in DMF (5 mL) for 10 min, the solution was drained and the resin washed with DMF (5 × 5 mL). A TNBS test\* was used to verify each coupling (negative/colourless) and deprotection (positive/red) step, with steps repeated as necessary. Successive couplings and Fmoc-deprotections were repeated to achieve the desired sequence. The resin was washed with DCM (3 × 5 mL) and then diethyl ether (3 × 5 mL) and air dried with suction. The peptides were then cleaved from the resin according to the protocol '*Cleavage from solid-support, isolation & purification*' detailed below.

\* *TNBS Test:* A small spatula of swollen resin taken out and 1 drop each of TNBS (100 μL 5% w/v in H<sub>2</sub>O added to 900 μL of DMF) and DIPEA solutions (100 μL in 900 μL of DMF) added and allowed to develop for 1 min. Clear/yellow beads indicated no free amine (negative), while red/orange beads showed free amine was present (positive).

*Peptides synthesised by this method:* p21<sub>μ</sub>-pol ι, p21<sub>μ</sub>-PARG, p21<sub>μ</sub>-RD1, p21<sub>μ</sub>-RD2, p21<sub>μ</sub>-RD3.

#### *Cleavage from solid-support, isolation & purification*

Following complete assembly of the peptide and deprotection of the final Fmoc group, the peptides were subsequently cleaved from the resin and the side-chain protecting groups simultaneously globally deprotected by treatment of the resin with 92.5/2.5/2.5/2.5 TFA:TIPS:DODT:H<sub>2</sub>O (5 mL) for 2 hours. The cleavage mixture was pipetted from the resin and concentrated under a stream of nitrogen to 0.5-1 mL. The peptide was then precipitated by addition of diethyl ether (10 mL) and the mixture cooled to -20°C. The precipitate was pelleted by centrifugation (7600 rpm, 10 min), and the supernatant decanted. The pellet was dried under a nitrogen stream, and then dissolved in 1:1 ACN/H<sub>2</sub>O, before being syringe filtered (0.2 μm) and lyophilised to yield the crude peptide as a fluffy white powder. The peptides were purified by semi-preparative RP-HPLC on a Gilson GX-Prep system using a Phenomenex Luna C18(2) column (10 × 250 mm), over a linear ACN/H<sub>2</sub>O gradient optimised for each peptide sample. RP-HPLC solvents were (A) H<sub>2</sub>O with 0.1% TFA and (B) ACN with 0.1% TFA. The product containing fractions were pooled and lyophilised. The identity of the final compounds was confirmed by High Resolution Mass Spectrometry on an Agilent 6230 ESI-TOF LCMS. Purity of the peptides was confirmed by analytical RP-HPLC on an Agilent 1260 HPLC equipped with a Phenomenex Luna C18(2) column (250 × 4.6 mm) over a gradient of 5-50% B (15 min) and visualised at 220 nm. Characterisation data for all peptides is recorded in Table S1.

**Table S1:** Peptide characterisation data. All peptides are C-terminally amidated.

Name	Sequence	Mw	MF	M+ Calc	[M+4] <sup>++</sup> Calc	ESI+ [M+4] <sup>+</sup> Found	Purity % (220 nm)
p21 <sub>μ</sub> (141-155)	<sup>141</sup> KRRQTSMTDFYHSKR	1940.20	C <sub>82</sub> H <sub>134</sub> N <sub>30</sub> O <sub>23</sub> S <sub>1</sub>	1938.9959	485.7570	485.7566	96.3
p21 (139-160)	<sup>139</sup> GRKRRQTSMTDFYHSKRRLIFS	2770.20	C <sub>120</sub> H <sub>197</sub> N <sub>43</sub> O <sub>31</sub> S <sub>1</sub>	2769.4722	693.3760	693.3793	97.4
p21 <sub>μ</sub> -Q144D	KRRDTSMTDFYHSKR	1927.16	C <sub>81</sub> H <sub>131</sub> N <sub>29</sub> O <sub>24</sub> S <sub>1</sub>	1925.9642	482.4991	482.5010	98.8
p21 <sub>μ</sub> -Q144M	KRRMTSMTDFYHSKR	1943.27	C <sub>82</sub> H <sub>135</sub> N <sub>29</sub> O <sub>22</sub> S <sub>2</sub>	1941.9778	486.5024	486.5020	98.9
p21 <sub>μ</sub> -Q144S	KRRSTSMTDFYHSKR	1899.15	C <sub>80</sub> H <sub>131</sub> N <sub>29</sub> O <sub>23</sub> S <sub>1</sub>	1897.9693	475.5003	475.4999	97.3
p21 <sub>μ</sub> -Q144K	KRRKTSMTDFYHSKR	1940.24	C <sub>83</sub> H <sub>138</sub> N <sub>30</sub> O <sub>22</sub> S <sub>1</sub>	1939.0323	485.7661	485.7658	93.4
p21 <sub>μ</sub> -Q144N	KRRNTSMTDFYHSKR	1926.18	C <sub>81</sub> H <sub>132</sub> N <sub>30</sub> O <sub>23</sub> S <sub>1</sub>	1924.9802	482.2531	482.2527	90.7
p21 <sub>μ</sub> -T145K	KRRQKSMTDFYHSKR	1967.27	C <sub>84</sub> H <sub>139</sub> N <sub>31</sub> O <sub>22</sub> S <sub>1</sub>	1966.0432	492.5188	492.5206	90.2
p21 <sub>μ</sub> -T145R	KRRQRSMTDFYHSKR	1995.28	C <sub>84</sub> H <sub>139</sub> N <sub>33</sub> O <sub>22</sub> S <sub>1</sub>	1994.0493	499.5203	499.5215	91.0
p21 <sub>μ</sub> -S146K	KRRQTKMTDFYHSKR	1981.30	C <sub>85</sub> H <sub>141</sub> N <sub>31</sub> O <sub>22</sub> S <sub>1</sub>	1980.0588	496.0227	496.0235	87.0
p21 <sub>μ</sub> -S146R	KRRQTRMTDFYHSKR	2009.31	C <sub>85</sub> H <sub>141</sub> N <sub>33</sub> O <sub>22</sub> S <sub>1</sub>	2008.0650	503.0243	503.0250	94.0
p21 <sub>μ</sub> -TS145/146KK	KRRQKKMTDFYHSKR	2008.37	C <sub>87</sub> H <sub>146</sub> N <sub>32</sub> O <sub>21</sub> S <sub>1</sub>	2007.1061	3+: 670.0492	670.3799	91.3
p21 <sub>μ</sub> -TS145/146RK	KRRQKRMDFYHSKR	2036.38	C <sub>87</sub> H <sub>146</sub> N <sub>34</sub> O <sub>21</sub> S <sub>1</sub>	2035.1122	509.7861	509.7864	93.0
p21 <sub>μ</sub> -TS145/146KR	KRRQKRMDFYHSKR	2036.38	C <sub>87</sub> H <sub>146</sub> N <sub>34</sub> O <sub>21</sub> S <sub>1</sub>	2035.1122	509.7861	509.7859	91.9
p21 <sub>μ</sub> -TS145/146RR	KRRQRRMTDFYHSKR	2064.39	C <sub>87</sub> H <sub>146</sub> N <sub>36</sub> O <sub>21</sub> S <sub>1</sub>	2063.1184	516.7876	516.7877	90.9
p21 <sub>μ</sub> -M147I	KRRQTSITDFYHSK	1765.98	C <sub>77</sub> H <sub>124</sub> N <sub>26</sub> O <sub>22</sub>	1764.9383	442.2426	442.2419	99.1
p21 <sub>μ</sub> -M147L	KRRQTSITDFYHSKR	1922.16	C <sub>83</sub> H <sub>136</sub> N <sub>30</sub> O <sub>23</sub>	1921.0395	481.2679	481.2672	99.8
p21 <sub>μ</sub> -M147A	KRRQTSATDFYHSKR	1880.08	C <sub>80</sub> H <sub>130</sub> N <sub>30</sub> O <sub>23</sub>	1878.9925	470.7561	470.7556	99.2
p21 <sub>μ</sub> -M147W	KRRQTSWTFYHSKR	1995.22	C <sub>88</sub> H <sub>135</sub> N <sub>31</sub> O <sub>23</sub>	1994.0347	499.5167	499.5161	99.5
p21 <sub>μ</sub> -M147V	KRRQTSVTFYHSKR	1908.14	C <sub>82</sub> H <sub>134</sub> N <sub>30</sub> O <sub>23</sub>	1907.0238	477.7640	477.7632	98.5
p21 <sub>μ</sub> -T148D	KRRQTSMTDFYHSKR	1954.19	C <sub>82</sub> H <sub>132</sub> N <sub>30</sub> O <sub>24</sub> S <sub>1</sub>	1952.9751	489.2518	489.2514	91.5
p21 <sub>μ</sub> -T148E	KRRQTSMTDFYHSKR	1968.21	C <sub>83</sub> H <sub>134</sub> N <sub>30</sub> O <sub>24</sub> S <sub>1</sub>	1966.9908	492.7557	492.7562	90.4
p21 <sub>μ</sub> -D149E	KRRQTSMTDFYHSKR	1954.23	C <sub>83</sub> H <sub>136</sub> N <sub>30</sub> O <sub>23</sub> S <sub>1</sub>	1953.0115	489.2609	489.2620	88.5
p21 <sub>μ</sub> -TD148/149EE	KRRQTSMTDFYHSKR	1982.24	C <sub>84</sub> H <sub>136</sub> N <sub>30</sub> O <sub>24</sub> S <sub>1</sub>	1981.0065	496.2596	496.2595	89.4
p21 <sub>μ</sub> -TS148/149DE	KRRQTSMTDFYHSKR	1968.21	C <sub>83</sub> H <sub>134</sub> N <sub>30</sub> O <sub>24</sub> S <sub>1</sub>	1966.9908	492.7557	492.7555	90.1
p21 <sub>μ</sub> -F150H	KRRQTSMTDFYHSKR	1930.16	C <sub>79</sub> H <sub>132</sub> N <sub>32</sub> O <sub>23</sub> S <sub>1</sub>	1928.9864	483.2546	483.2542	94.4
p21 <sub>μ</sub> -F150Y	KRRQTSMTDFYHSKR	1956.20	C <sub>82</sub> H <sub>134</sub> N <sub>30</sub> O <sub>24</sub> S <sub>1</sub>	1954.9908	489.7557	489.7552	93.8
p21 <sub>μ</sub> -Y151F	KRRQTSMTDFYHSKR	1924.20	C <sub>82</sub> H <sub>134</sub> N <sub>30</sub> O <sub>22</sub> S <sub>1</sub>	1923.0010	481.7582	481.7576	94.2
p21 <sub>μ</sub> -FY150/151YF	KRRQTSMTDFYHSKR	1940.20	C <sub>82</sub> H <sub>134</sub> N <sub>30</sub> O <sub>23</sub> S <sub>1</sub>	1938.9959	485.7570	485.7589	98.1
p21 <sub>μ</sub> -DNALig1	KRRQRSIMSFFHFSKR	1963.32	C <sub>85</sub> H <sub>143</sub> N <sub>33</sub> O <sub>19</sub> S <sub>1</sub>	1962.0959	491.5320	491.5311	98.2
p21 <sub>μ</sub> -FEN1	KRRQGRLLDFFHFSKR	1945.20	C <sub>84</sub> H <sub>137</sub> N <sub>33</sub> O <sub>21</sub>	1944.0667	487.0247	487.0241	99.2
p21 <sub>μ</sub> -Pogo	KRRQKKITDYFFHFSKR	1990.32	C <sub>88</sub> H <sub>148</sub> N <sub>32</sub> O <sub>21</sub>	1989.1497	498.2954	498.2965	98.0
p21 <sub>μ</sub> -XPG	KRRQLRIDSFFHFSKR	1973.29	C <sub>87</sub> H <sub>145</sub> N <sub>33</sub> O <sub>20</sub>	1972.1344	494.0416	494.0409	99.1
p21 <sub>μ</sub> -MCMT	KRRQTITSHFFHFSKR	1882.14	C <sub>80</sub> H <sub>136</sub> N <sub>32</sub> O <sub>21</sub>	1881.0558	471.2719	471.2714	93.9
p21 <sub>μ</sub> -p15	KRRQKGIGEFFHFSKR	1873.18	C <sub>83</sub> H <sub>137</sub> N <sub>31</sub> O <sub>19</sub>	1872.0707	469.0257	469.0265	96.5
p21 <sub>μ</sub> -Cdt1	KRRQRRVTDFFHFSKR	2016.33	C <sub>87</sub> H <sub>146</sub> N <sub>36</sub> O <sub>20</sub>	2015.1514	504.7959	504.7954	94.8
p21 <sub>μ</sub> -pol δ <sub>p66</sub>	KRRQVSITGFFHFSKR	1846.15	C <sub>82</sub> H <sub>136</sub> N <sub>30</sub> O <sub>19</sub>	1845.0598	462.2729	462.2727	99.2
p21 <sub>μ</sub> -RecQ5	KRRQNLIRHFFHFSKR	2022.37	C <sub>90</sub> H <sub>148</sub> N <sub>36</sub> O <sub>18</sub>	2021.1772	506.3024	506.3019	99.2
p21 <sub>μ</sub> -WRN	KRRQWKLRFHFSKR	2053.42	C <sub>92</sub> H <sub>153</sub> N <sub>35</sub> O <sub>19</sub>	2052.2082	514.0600	514.0593	99.3
p21 <sub>μ</sub> -PARG	KRRDSKITDHFHFSKR	1910.16	C <sub>81</sub> H <sub>136</sub> N <sub>32</sub> O <sub>22</sub>	1909.0507	478.2707	478.2705	98.4
p21 <sub>μ</sub> -pol β	KRRQLQKVHFFHFSKR	1847.18	C <sub>81</sub> H <sub>139</sub> N <sub>33</sub> O <sub>17</sub>	1846.1027	462.5337	462.5330	96.5
p21 <sub>μ</sub> -pol λ	KRRSVPVLELFFHFSKR	1851.21	C <sub>83</sub> H <sub>143</sub> N <sub>29</sub> O <sub>19</sub>	1850.1115	463.5359	463.5353	92.4
p21 <sub>μ</sub> -pol ι	KRRKGLIDYLLHFSKR	1932.29	C <sub>87</sub> H <sub>146</sub> N <sub>30</sub> O <sub>20</sub>	1931.1330	483.7913	483.7919	87.6
p21 <sub>μ</sub> -pol κ	KRRKHTLDIFFHFSKR	1968.32	C <sub>89</sub> H <sub>146</sub> N <sub>32</sub> O <sub>19</sub>	1967.1442	492.7941	492.7939	93.5
p21 <sub>μ</sub> -RFC <sub>p14</sub>	KRRMDIRKFFHFSKR	1904.30	C <sub>84</sub> H <sub>142</sub> N <sub>32</sub> O <sub>17</sub> S <sub>1</sub>	1903.0951	476.7818	476.7813	93.4
p21 <sub>μ</sub> -RNaseH2B	KRRMKSIDTFFHFSKR	1936.29	C <sub>85</sub> H <sub>142</sub> N <sub>30</sub> O <sub>20</sub> S <sub>1</sub>	1935.0737	484.7764	484.7761	97.7
p21 <sub>μ</sub> -Cdt2	KRRMRKICTYFFHFSKR	2009.46	C <sub>87</sub> H <sub>149</sub> N <sub>33</sub> O <sub>18</sub> S <sub>2</sub>	2008.1198	503.0380	503.0390	93.8
p21 <sub>μ</sub> -pol n	KRRMQTLESFFHFSKR	1950.28	C <sub>85</sub> H <sub>140</sub> N <sub>30</sub> O <sub>21</sub> S <sub>1</sub>	1949.0530	488.2713	488.2719	96.6
p21 <sub>μ</sub> -RD1	KRRQKRMEDYHFSKR	2080.39	C <sub>88</sub> H <sub>146</sub> N <sub>34</sub> O <sub>23</sub> S <sub>1</sub>	2079.1021	520.7835	520.7836	91.0
p21 <sub>μ</sub> -RD2	KRRQTRITEYHFSKR	2005.30	C <sub>87</sub> H <sub>145</sub> N <sub>33</sub> O <sub>22</sub>	2004.1242	502.0389	502.0396	97.1
p21 <sub>μ</sub> -RD3	KRRQKRITEYHFSKR	2048.39	C <sub>89</sub> H <sub>150</sub> N <sub>34</sub> O <sub>22</sub>	2047.1664	512.7994	512.8001	92.3



**SPR:**

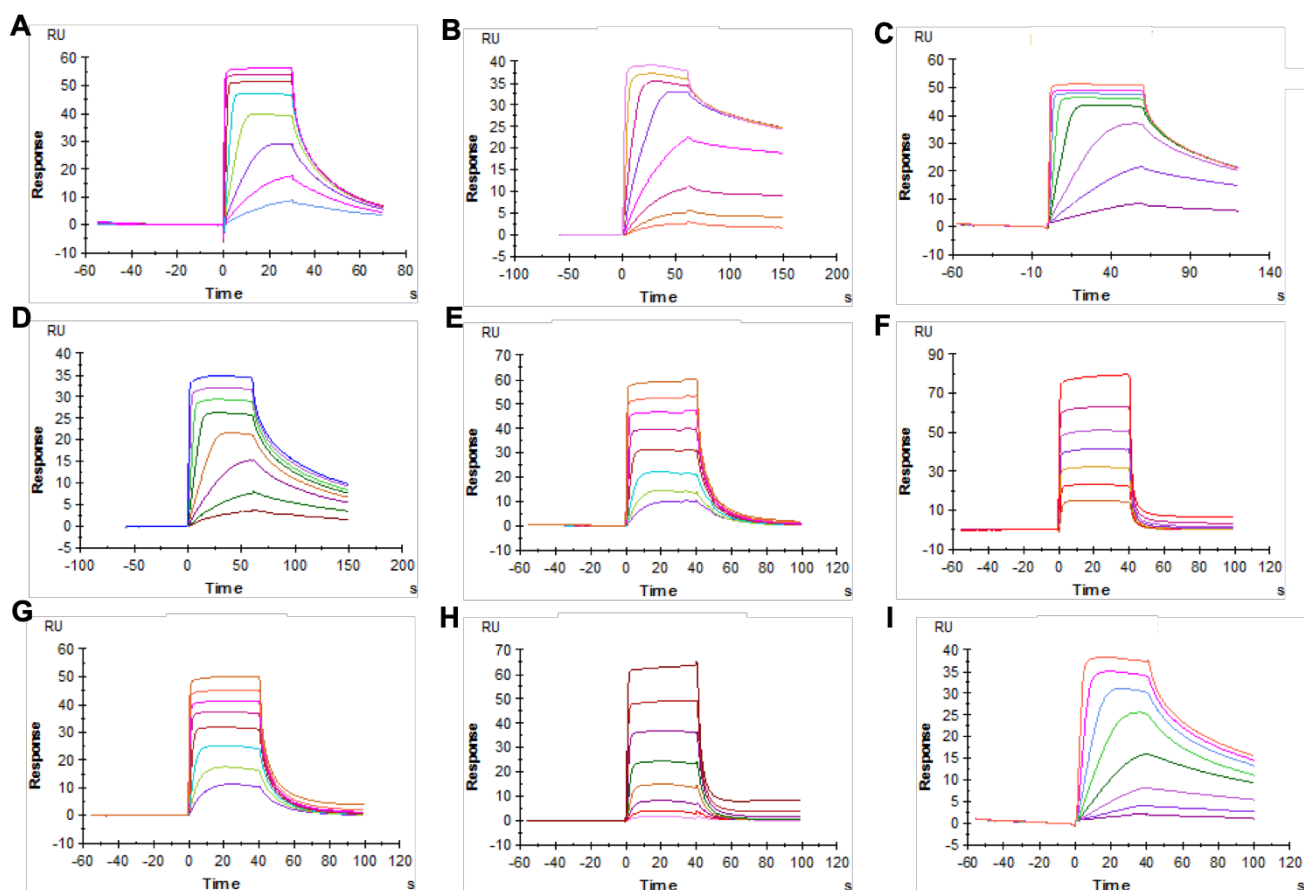
**Table S2: Peptide SPR data against hPCNA.** Top Conc is the highest concentration of 8x 1 in 2 dilutions used to calculate the steady state affinity.  $K_D$  is the affinity constant. SE, standard fitting error; On/Off, indicate times for contact and dissociation phases of each run. All peptides are C-terminally amidated.

Name	Sequence	$\epsilon_{205}^*$	Top Conc (nM)	Affinity $K_D$ (nM)	$K_D$ SE (nM)	$\chi^2$	On/Off (s)
p21 <sub>μ</sub> (141-155)	<sup>141</sup> KRRQTSMTDFYHSKR	67860	500	12.3	0.598	0.196	30/60
p21 (139-160)	<sup>139</sup> GRKRRQTSMTDFYHSKRRLIFS	98620	50	4.319	1.31	10.9	60/90
p21 <sub>μ</sub> -Q144D	KRRDTSMTDFYHSKR	67460	2000	714	30.4	0.126	40/60
p21 <sub>μ</sub> -Q144M	KRRMTSMTDFYHSKR	69290	5000	1544	159	1.57	40/60
p21 <sub>μ</sub> -Q144S	KRRSTSMTDFYHSKR	67460	5000	1032	116	2.07	40/60
p21 <sub>μ</sub> -Q144K	KRRKTSMTDFYHSKR	67460	5000	1123	148	4.19	40/60
p21 <sub>μ</sub> -Q144N	KRRNTSMTDFYHSKR	67860	5000	772	117	3.07	40/60
p21 <sub>μ</sub> -T145K	KRRQKSMTDFYHSKR	67860	2000	98.05	10.8	2.58	40/60
p21 <sub>μ</sub> -T145R	KRRQRSMTDFYHSKR	69210	1000	88.51	13.2	2.97	40/60
p21 <sub>μ</sub> -S146K	KRRQTKMTDFYHSKR	67860	300	12.97	1.55	0.520	60/90
p21 <sub>μ</sub> -S146R	KRRQTRMTDFYHSKR	69210	100	4.297	1.35	6.26	60/90
p21 <sub>μ</sub> -TS145/146KK	KRRQKKMTDFYHSKR	67860	300	20.18	4.80	6.70	60/90
p21 <sub>μ</sub> -TS145/146RK	KRRQRKMTDFYHSKR	69210	300	17.30	4.66	7.94	60/90
p21 <sub>μ</sub> -TS145/146KR	KRRQKRMTDFYHSKR	69210	300	37.00	11.0	9.63	60/90
p21 <sub>μ</sub> -TS145/146RR	KRRQRRMTDFYHSKR	70560	30	1.833	0.453	5.80	60/90
p21 <sub>μ</sub> -M147I	KRRQTSITDFYHSK	66030	200	11.1	0.258	0.0251	40/60
p21 <sub>μ</sub> -M147L	KRRQTSITDFYHSKR	66030	200	20.49	1.75	0.784	40/60
p21 <sub>μ</sub> -M147A	KRRQTSATDFYHSKR	66030	20000	7591	626	1.43	40/60
p21 <sub>μ</sub> -M147W	KRRQTSWTFYHSKR	86430	5000	3566	165	0.274	40/60
p21 <sub>μ</sub> -M147V	KRRQTSVTFYHSKR	66030	500	29.29	1.83	0.326	40/60
p21 <sub>μ</sub> -T148D	KRRQTSMDDFYHSKR	67860	2000	96.47	7.32	0.477	40/60
p21 <sub>μ</sub> -T148E	KRRQTSMEDFYHSKR	67860	200	77.40	2.62	0.0533	60/90
p21 <sub>μ</sub> -D149E	KRRQTSMEFYHSKR	67860	500	12.67	1.4	0.387	60/90
p21 <sub>μ</sub> -TD148/149EE	KRRQTSMEEFYHSKR	67860	1000	48.52	1.16	0.0475	60/90
p21 <sub>μ</sub> -TD148/149DE	KRRQTSMEDEFYHSKR	67860	2000	345.8	2.67	0.273	60/90
p21 <sub>μ</sub> -F150H	KRRQTSMDHYHSKR	64460	1000	159.0	62.8	0.119	40/60
p21 <sub>μ</sub> -F150Y	KRRQTSMDYHSKR	65340	300	20.2	0.424	0.0286	40/60
p21 <sub>μ</sub> -Y151F	KRRQTSMTDFHHSKR	70380	300	10.61	1.48	0.721	60/90
p21 <sub>μ</sub> -FY150/151YF	KRRQTSMTDYFHSKR	67860	30	2.175	0.531	3.01	60/90
p21 <sub>μ</sub> -DNALig1	KRRQRSIMSFHHSKR	71730	250	40.74	9.20	3.42	60/90
p21 <sub>μ</sub> -FEN1	KRRQGRLDDFFHHSKR	69900	2000	165.6	2.07	1.67	40/60
p21 <sub>μ</sub> -Pogo	KRRQKKITDYFHSKR	66030	100	8.816	3.30	6.73	60/90
p21 <sub>μ</sub> -XPG	KRRQLRIDSFHHSKR	69900	200	16.39	4.81	11.2	60/90
p21 <sub>μ</sub> -MCMT	KRRQTITSHFHSKR	65150	2000	106.0	24.4	5.32	40/60
p21 <sub>μ</sub> -p15	KRRQKGIGEFFHHSKR	68550	2000	234.0	42.4	5.21	40/60
p21 <sub>μ</sub> -Cdt1	KRRQRRVTDFFHHSKR	71250	150	8.760	2.22	10.1	40/60
p21 <sub>μ</sub> -pol δ <sub>p66</sub>	KRRQVSITGFFHHSKR	68550	1000	123.7	4.24	3.21	40/60
p21 <sub>μ</sub> -RecQ5	KRRQNLIRHFFHHSKR	75500	20000	>40000**			
p21 <sub>μ</sub> -WRN	KRRQWKLLRDFHHSKR	81700	2000	1155	67.3	0.952	40/60
p21 <sub>μ</sub> -PARG	KRRDSKITDHFHHSKR	64750	2000	90.03	18.4	2.38	40/60
p21 <sub>μ</sub> -pol β	KRRQLQKVHFHHSKR	62770	20000	5732	822	18.6	40/60
p21 <sub>μ</sub> -pol λ	KRRSVPVLELFHHSKR	59550	20000	8144	766	4.81	40/60
p21 <sub>μ</sub> -pol ι	KRRKGLIDYLFHHSKR	63110	1500	111.2	5.41	0.0710	60/90
p21 <sub>μ</sub> -pol κ	KRRKHTLIDFFHHSKR	73350	10000	3296	353	6.62	40/60
p21 <sub>μ</sub> -RFC <sub>p14</sub>	KRRMDIRKFFHHSKR	68550	1000	145.2	34.5	6.42	40/60
p21 <sub>μ</sub> -Cdt2	KRRMRKICTYFHSKR	69500		NS***			
p21 <sub>μ</sub> -pol η	KRRMQTLESFFHHSKR	70380	10000	2537	745	10.2	40/60
p21 <sub>μ</sub> -RNAseH2B	KRRMKSIDTFHHSKR	69980	2000	640.4	51.1	0.851	40/60
p21 <sub>μ</sub> -RD1	KRRQRKMEDYYHSKR	66690	100	8.152	1.27	1.51	40/60
p21 <sub>μ</sub> -RD2	KRRQRTRITEYFHSKR	67380	50	1.118	0.330	2.40	60/90
p21 <sub>μ</sub> -RD3	KRRQRTRITEYYHSKR	64860	200	6.70	1.58	3.96	40/60

\* Determined using online calculator, from Anthis 2013 (1)

\*\* Peptide bound very non-specifically to protein where the RU significantly exceeded the expected maximum and a  $K_D$  could not be determined.

\*\*\* Peptide bound significantly to the reference cell of sensor chip such that a  $K_D$  could not be determined



**Figure S1:** Representative sample of SPR sensorgrams. **A** p21 $\mu$  **B** p21 $\mu$ -S146R **C** p21 $\mu$ -FY150/151YF **D** p21 $\mu$ -Y151F **E** p21 $\mu$ -FEN1 **F** p21 $\mu$ -pol  $\eta$  **G** p21 $\mu$ -PARG **H** p21 $\mu$ -RNaseH2B **I** p21 $\mu$ -RD1

## COCRYSTAL EXPERIMENTS

**Table S3:** Data collection and refinement statistics of hPCNA bound with p21 $\mu$  (RCSB PDB ID: 7KQ1), and hPCNA bound with p21 $\mu$ -F150Y (RCSB PDB ID: 7KQ0). Statistics for the highest-resolution shell are shown in parentheses.

PDB ID	7KQ1	7KQ0
Wavelength	0.9537	0.9537
Resolution range	41.09 - 3.3 (3.418 - 3.3)	41.15 - 2.4 (2.486 - 2.4)
Space group	P 32 2 1	P 3
Unit cell	136.66 136.66 104.005 90 90 120	142.563 142.563 41.03 90 90 120
Unique reflections	17208 (1689)	32881 (3598)
Multiplicity	19.9 (18.2)	10.7 (11.1)
Completeness (%)	99.83 (99.64)	90.13 (99.94)
Mean I/sigma(I)	10.45 (0.91)	40.58 (13.50)
<sup>a</sup> R-merge	0.2823 (4.453)	0.04334 (0.1841)
<sup>b</sup> Rpim	0.065 (0.755)	0.014 (0.570)
CC1/2	0.999 (0.469)	1 (0.987)
Reflections used in refinement	17192 (1684)	32881 (3598)
Reflections used for R-free	1716 (171)	2031 (224)
<sup>c</sup> R-work	0.2419 (0.3591)	0.1537 (0.2023)
<sup>d</sup> R-free	0.2685 (0.3618)	0.2011 (0.2813)
Number of non-hydrogen atoms	6084	6086
macromolecules	6084	5976
Protein residues	816	807
RMS(bonds)	0.003	0.004
RMS(angles)	0.69	0.68
Ramachandran favoured (%)	94.78	98.10
Ramachandran allowed (%)	4.73	1.64
Ramachandran outliers (%)	0.50	0.25
Rotamer outliers (%)	0.00	8.80
Clashscore	8.80	3.15
Average B-factor	108.50	32.34

$$^a R_{\text{merge}} = \sum |I - \langle I \rangle| / \sum I.$$

$$^b R_{\text{pim}} = \sum h [1 / (I_h - 1)]^{1/2} \sum | \langle I_h \rangle - I_{h,i} | / \sum_h \sum_i I_{h,i} \quad (2)$$

$$^c R_{\text{work}} = \sum |F_o - F_c| / \sum |F_o| \text{ for all data excluding data used to calculate } R_{\text{free}}.$$

$$^d R_{\text{free}} = \sum |F_o - F_c| / \sum |F_o|, \text{ for all data.}$$

**Table S4:** Secondary Interaction Summary for co-crystal structure of p21<sub>μ</sub> with hPCNA (PDB ID: 7KQ1) calculated using the RING server. Only peptide chain interactions reported (chains B, D and F). Interactions reported are an average of the number of interactions observed for the three chains. RING session ID: [5ef50f2b0e9f94078ea226bd](#)

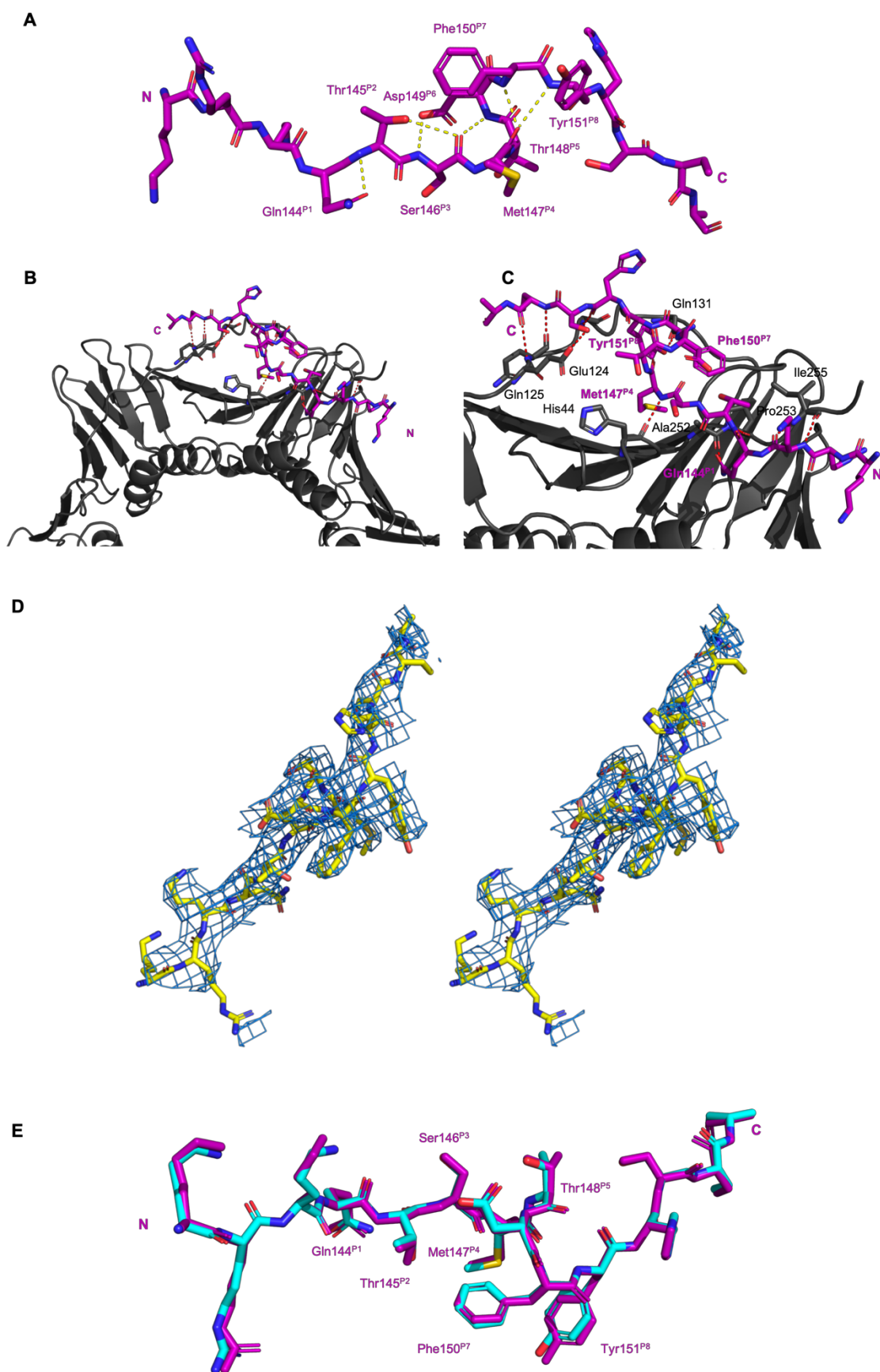
	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	0.67	0	0	0	
	143	0	0.67	0	0	
PIP-box	* 144	1.67	0	0	0	
	145	2.33	1.00	0.67	0.33	
	146	0.67	0.33	0.33	1.00	
	* 147	6.67	1.00	0.33	1.67	
	148	0.33	0	0	0	
	149	0	0	0	0	
	* 150	2.67	0.00	0	0	
	* 151	2.00	0.67	0	0	
	152	1.00	1.00	0	0	
FI	153	1.67	0.33	0	0	
	154	0.67	1.00	0	0	
	155	0	0	0	0	
	<b>Total</b>	20.3	6.0	1.3	3.0	30.67
<b>PIP-box</b>		16	3	1	3	23.67
<b>Flanking (FI)</b>		4	3	0	0	7
<b>Conserved (*) PIP-box residues</b>		11.00	1	0	2	14.00
<b>Non-conserved PIP-box residues</b>		3.33	1.33	1.00	1.33	7.00

**Other:** Intermolecular pi-stack between F-Tyr151 to E-Tyr133, and D-Tyr151 to C-Tyr250

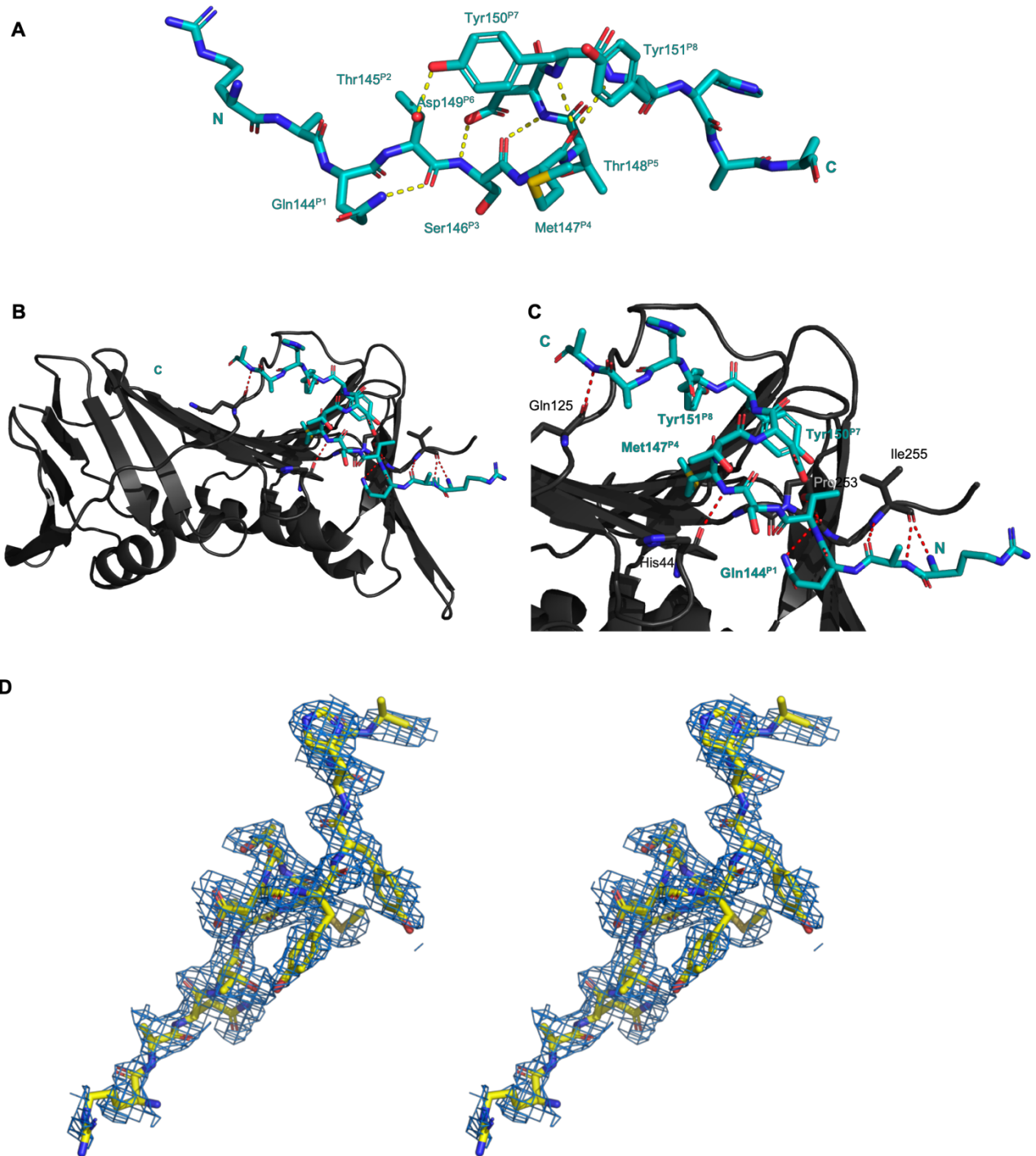
**Table S5:** Secondary Interaction Summary for co-crystal structure of p21<sub>μ</sub>-F150Y with hPCNA (PDB ID: 7KQ0) calculated using the RING server. Only peptide chain interactions reported (chains B, D and F). Interactions reported are an average of the number of interactions observed for the three chains. RING session ID: [5f595dab0e9f94078ea22f07](#)

	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	1	0	0	0	
	143	0	1	0	0	
PIP-box	* 144	1	0	0	0	
	145	1	1	0	1	
	146	1	0	1	0	
	* 147	3	1	2	1	
	148	1	0	0	0	
	149	0	0	0	0	
	* 150	2	0	0	0	
	* 151	2	0	0	0	
	152	0	1	0	0	
FI	153	0	0	0	0	
	154	0	0	0	0	
	155	1	0	0	0	
	<b>Total</b>	13.0	4.0	2.67	2.67	22.33
<b>PIP-box</b>		11	2	3	3	18.33
<b>Flanking (FI)</b>		2	2	0	0	4
<b>Conserved (*) PIP-box residues</b>		6.67	1	2	1	10.67
<b>Non-conserved PIP-box residues</b>		2.00	0.67	1.00	1.33	5.00

**Other:**

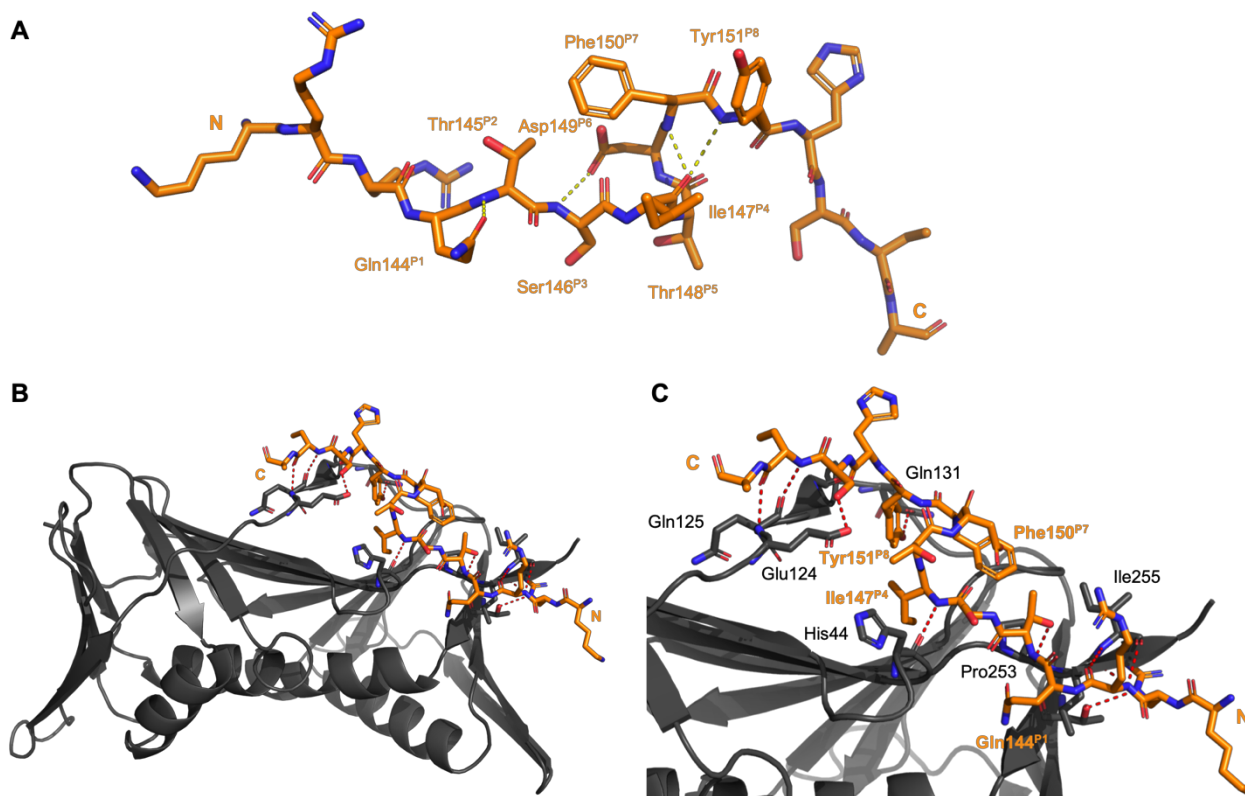


**Figure S2:** Co-crystal structure of p21<sub>μ</sub> (purple, sticks) bound to PCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in purple. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in purple. **D** Representative electron density of p21<sub>μ</sub> (yellow, sticks) shown as a wall-eye stereo image 2Fo-Fc composite omit map, view contoured at 1.5σ. **E** Overlay of p21<sub>μ</sub> (purple, sticks) bound to PCNA (not shown) co-crystal structure, with p21<sub>μ</sub> (cyan, sticks) that has been energy minimised on the PCNA surface which shows a high degree of structural similarity, and validates the computational modelling approach.



**Figure S3:** Co-crystal structure of p21<sub>μ</sub>-F150Y (blue, sticks) bound to PCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in blue. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in blue. **D** Representative electron density of p21<sub>μ</sub>-F150Y (yellow, sticks) shown as a wall-eye stereo image 2Fo-Fc composite omit map, view contoured at 1.5σ.

## COMPUTATIONAL MODELLING OF PCNA MONOMERS BOUND TO P21 $\mu$ PEPTIDES



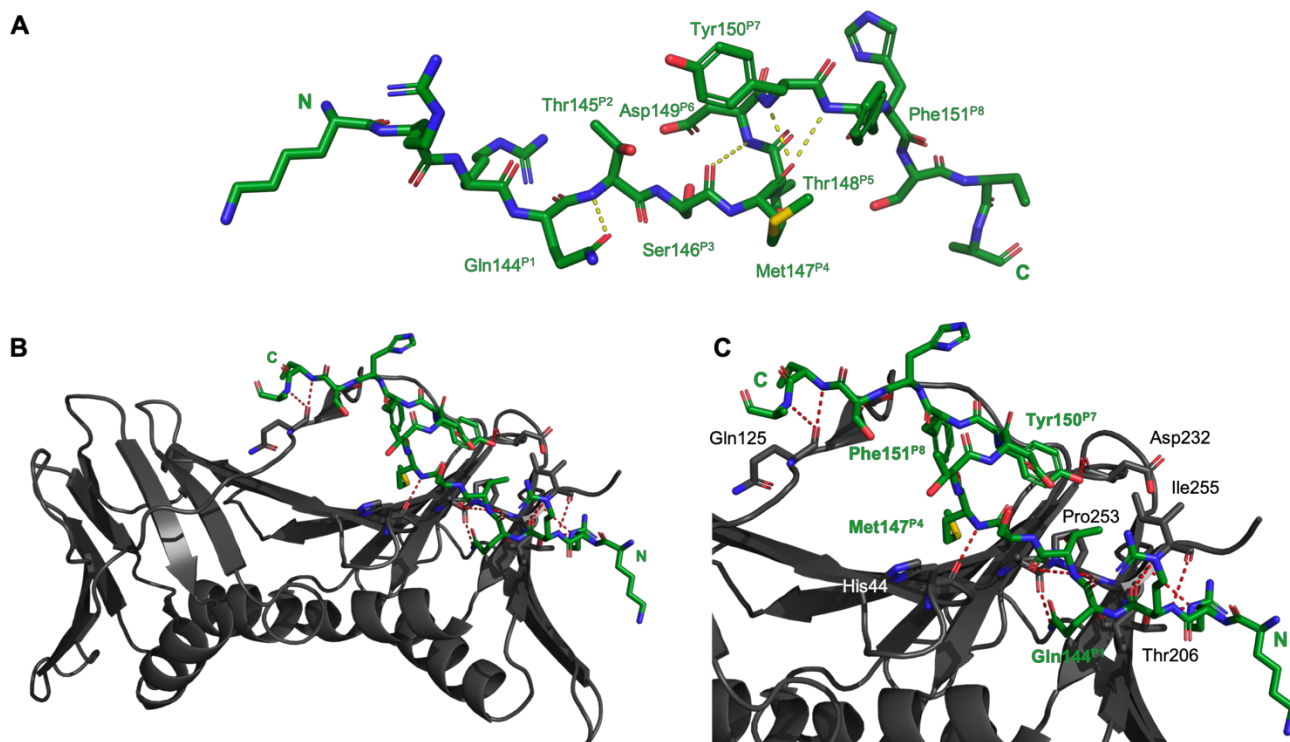
**Figure S4:** Computationally modelled structure of p21 $\mu$ -M147I (orange, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in orange. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in orange.

**Table S6:** Secondary Interaction Summary for computationally modelled structure of p21 $\mu$ -M147I with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b21280e9f94078ea22cfe](https://ring-server.org/session/5f3b21280e9f94078ea22cfe)

	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	3	1	0	0	
	143	1	2	0	0	
PIP-box	* 144	1	0	0	0	
	145	1	1	2	0	
	146	1	0	0	0	
	* 147	2	1	0	1	
	148	2	0	0	0	
	149	0	0	0	0	
	* 150	3	0	0	0	
	* 151	3	2	0	0	
FI	152	0	1	0	0	
	153	1	0	0	0	
	154	1	1	0	0	
	155	0	0	0	0	
<b>Total</b>		19	9	2	1	31
<b>PIP-box</b>		13	4	2	1	20
<b>Flanking (FI)</b>		6	5	0	0	11
<b>Conserved (*) PIP-box residues</b>		6	1	0	1	8
<b>Non-conserved PIP-box residues</b>		4	1	2	0	7

**Other:** Intermolecular pi-stack between B-Tyr151 and A-Tyr133



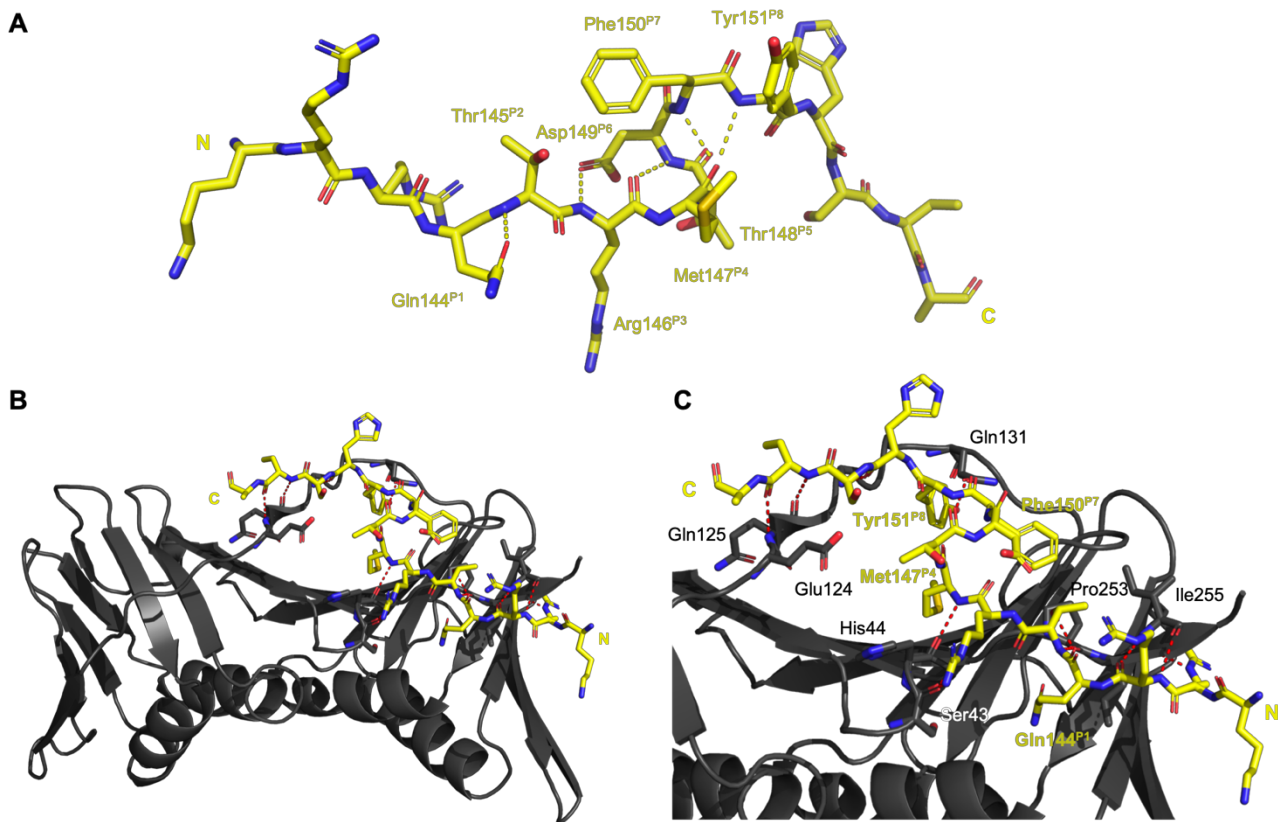


**Figure S5:** Computationally modelled structure of p21<sub>μ</sub>-FY150/151YF (dark green, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in dark green **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in dark green.

**Table S7:** Secondary Interaction Summary for computationally modelled structure of p21<sub>μ</sub>-FY150/151YF with hPCNA calculated using the RING server. Chain B interactions only. RING session ID: [5f3b22bc0e9f94078ea22d03](https://ring.rutgers.edu/session/5f3b22bc0e9f94078ea22d03)

	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	4	2	0	0	
	143	1	1	0	0	
PIP-box	* 144	2	0	0	0	
	145	1	1	0	0	
	146	1	0	0	1	
	* 147	6	1	1	2	
	148	1	0	0	0	
	149	0	0	0	0	
	* 150	3	1	0	0	
* 151	2	0	0	0		
FI	152	0	0	0	0	
	153	1	0	0	0	
	154	2	1	0	0	
	155	0	1	0	0	
	<b>Total</b>	24	8	1	3	36
	<b>PIP-box</b>	16	3	1	3	23
	<b>Flanking (FI)</b>	8	5	0	0	13
	<b>Conserved (*) PIP-box residues</b>	11	2	1	2	16
	<b>Non-conserved PIP-box residues</b>	3	1	0	1	5

**Other:** Intermolecular ionic interaction between B-Arg143 and A-Asp257; Intermolecular pi-stack between B-Tyr151 and A-Tyr250



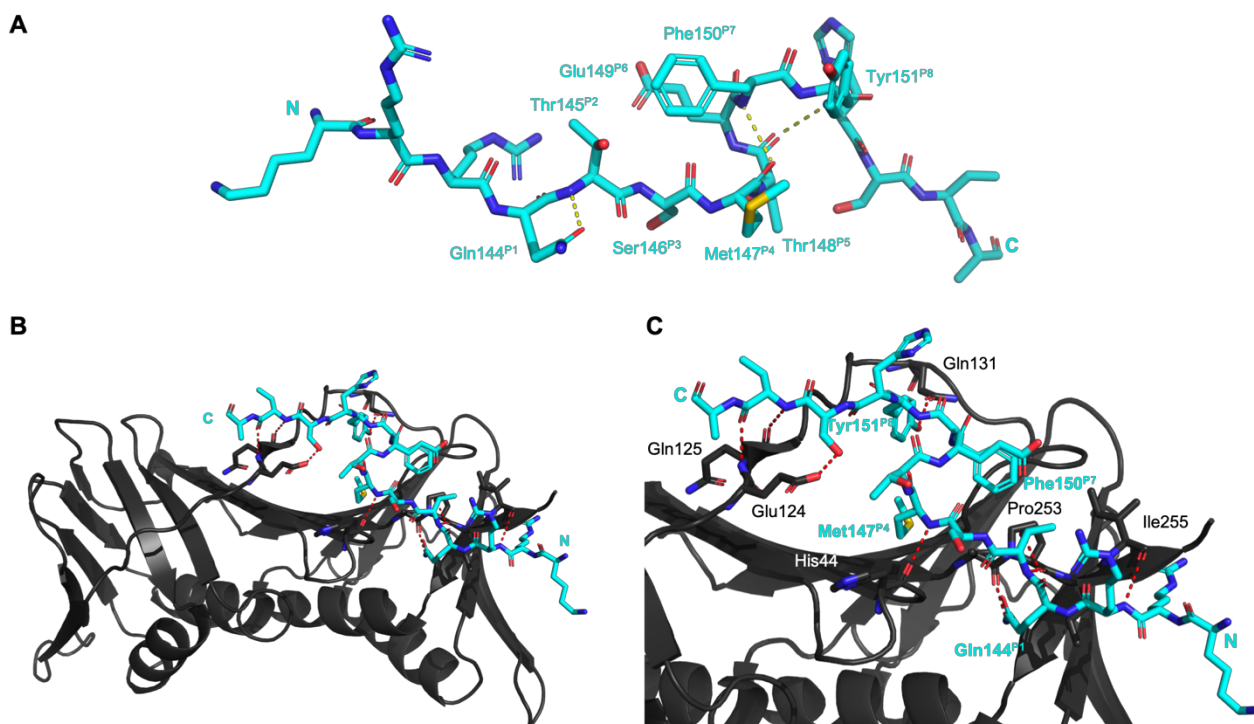
**Figure S6:** Computationally modelled structure of p21<sub>μ</sub>-S146R (yellow, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in yellow. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in yellow.

**Table S8:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-S146R with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b1f6b0e9f94078ea22cf9](https://ring.rdg.ac.uk/session/5f3b1f6b0e9f94078ea22cf9)

	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	3	1	0	0	
	143	2	2	0	0	
	* 144	1	0	0	0	
PIP-box	145	1	1	1	0	
	146	2	1	0	1	
	* 147	4	1	1	2	
	148	0	0	0	0	
	149	0	0	0	0	
	* 150	3	0	0	0	
	* 151	2	1	0	0	
	152	0	1	0	0	
FI	153	2	0	0	0	
	154	0	1	0	0	
	155	0	1	0	0	
	<b>Total</b>	20	10	2	3	35
	<b>PIP-box</b>	13	4	2	3	22
	<b>Flanking (FI)</b>	7	6	0	0	13
	<b>Conserved (*) PIP-box residues</b>	8	1	1	2	12
	<b>Non-conserved PIP-box residues</b>	3	2	1	1	7

**Other:** Intermolecular ionic interaction between B-Arg143 and A-Asp257



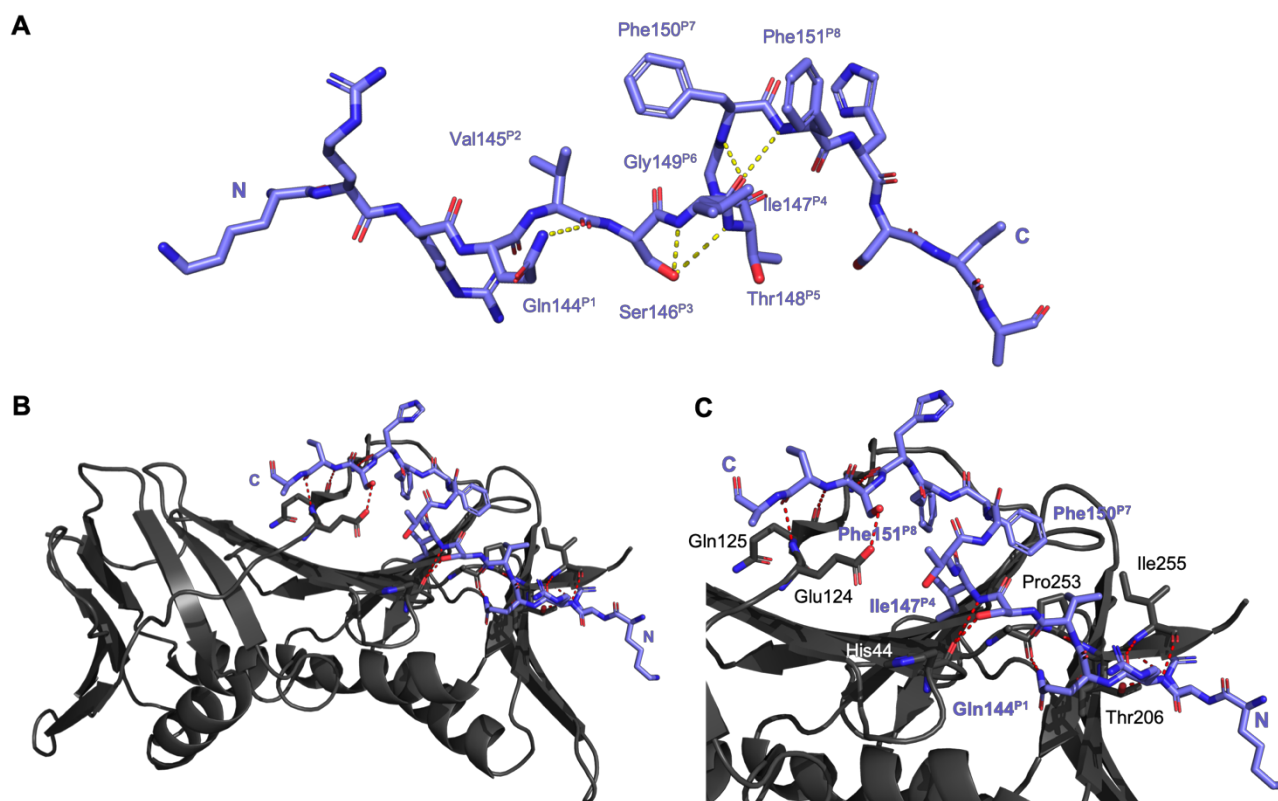


**Figure S7:** Computationally modelled structure of p21<sub>μ</sub>-D149E (light blue, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in light blue. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in light blue.

**Table S9:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-D149E with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b21fe0e9f94078ea22d00](https://ring-server.org/session/5f3b21fe0e9f94078ea22d00)

	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	2	2	0	0	
	143	0	2	0	0	
PIP-box	* 144	2	0	0	0	
	145	1	1	1	0	
	146	1	0	0	1	
	* 147	4	1	1	2	
	148	1	0	0	0	
	149	0	0	1	0	
	* 150	3	0	0	0	
	* 151	4	2	0	0	
FI	152	1	0	0	0	
	153	1	0	0	0	
	154	0	1	0	0	
	155	0	0	0	0	
	<b>Total</b>	20	9	3	3	35
	<b>PIP-box</b>	16	4	3	3	26
	<b>Flanking (FI)</b>	4	5	0	0	9
	<b>Conserved (*) PIP-box residues</b>	9	1	2	2	14
	<b>Non-conserved PIP-box residues</b>	3	1	2	1	7

**Other:** Intermolecular ionic interaction between B-Arg143 and A-Asp257; Intermolecular pi-stack between B-Tyr151 and A-Tyr250

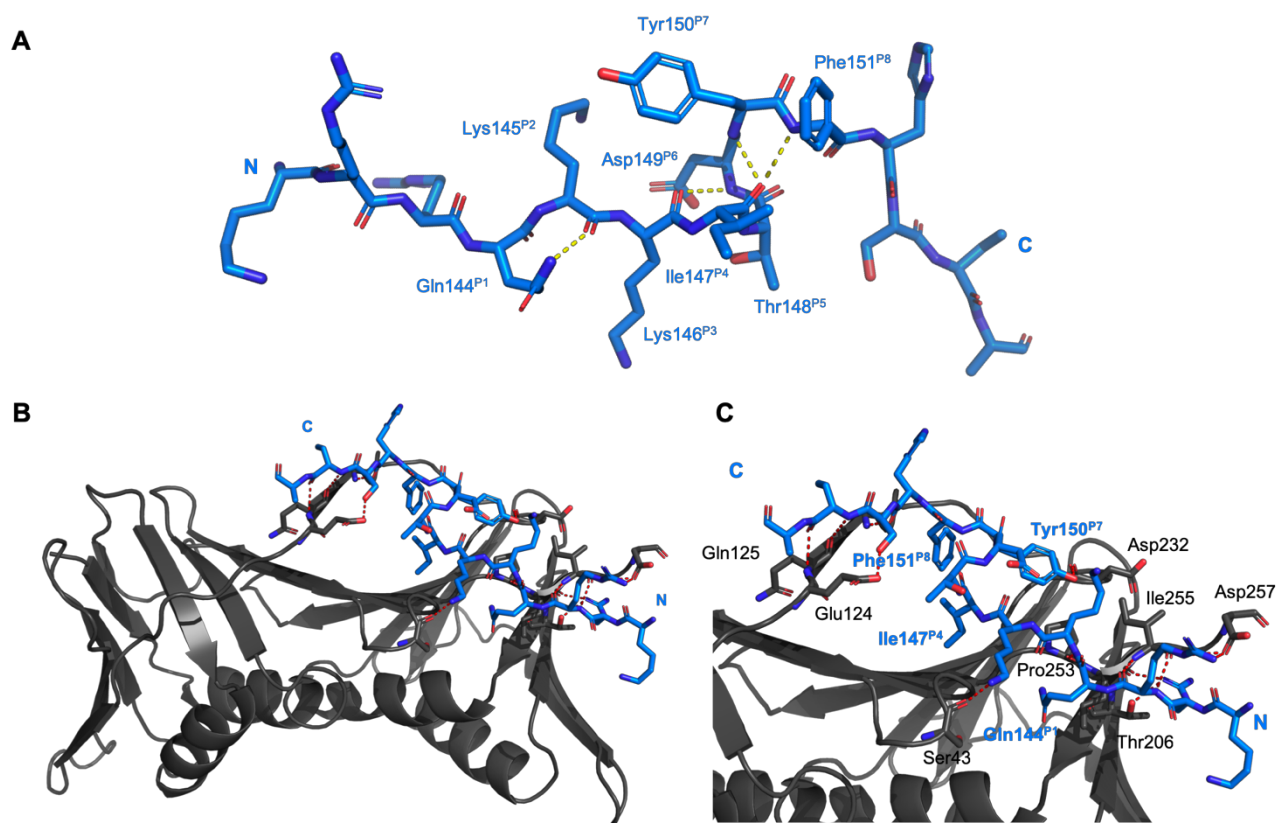


**Figure S8:** Computationally modelled structure of p21<sub>μ</sub>- pol δ<sub>p66</sub> (light purple, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in light purple. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in light purple.

**Table S10:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-pol δ<sub>p66</sub> with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b202a0e9f94078ea22cfb](https://ring-server.org/session/5f3b202a0e9f94078ea22cfb)

Residue	Intermolecular		Intramolecular		Total
	VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	
	142	3	0	2	
	143	1	0	2	
PIP-box	* 144	2	0	0	
	145	1	0	1	
	146	0	0	0	
	* 147	5	0	1	2
	148	0	0	0	0
	149	0	0	0	0
	* 150	1	0	0	0
	* 151	2	0	0	0
FI	152	0	0	1	
	153	2	0	0	
	154	0	0	1	
	155	1	0	0	
<b>Total</b>	<b>18</b>	<b>7</b>	<b>2</b>	<b>2</b>	<b>29</b>
<b>PIP-box</b>	<b>11</b>	<b>1</b>	<b>2</b>	<b>2</b>	<b>16</b>
<b>Flanking (FI)</b>	<b>7</b>	<b>6</b>	<b>0</b>	<b>0</b>	<b>13</b>
<b>Conserved (*) PIP-box residues</b>	<b>8</b>	<b>0</b>	<b>1</b>	<b>2</b>	<b>11</b>
<b>Non-conserved PIP-box residues</b>	<b>1</b>	<b>1</b>	<b>1</b>	<b>0</b>	<b>3</b>

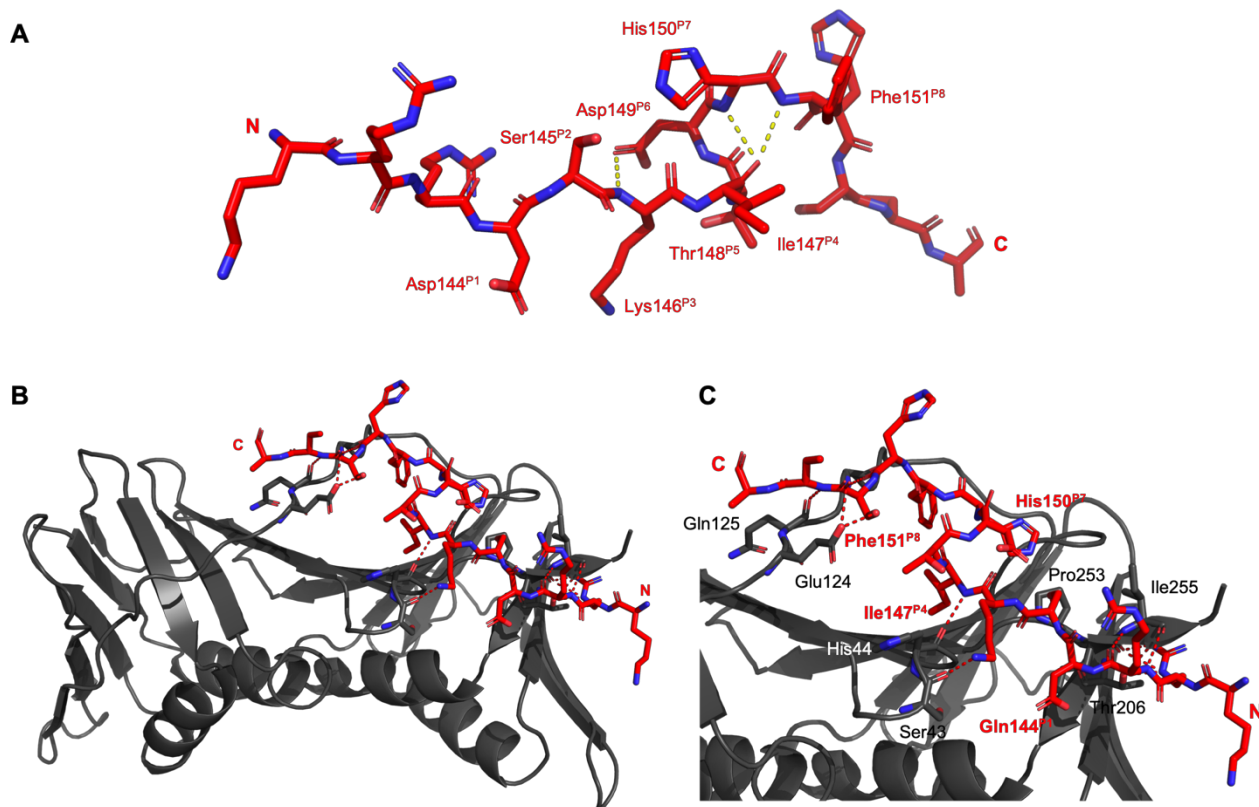
**Other:** Intermolecular pi-stack between B-Phe151 and A-Tyr250



**Figure S9:** Computationally modelled structure of p21<sub>μ</sub>-Pogo (dark blue, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in blue. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in blue.

**Table S11:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-Pogo with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b224d0e9f94078ea22d01](https://ring-server.org/session/5f3b224d0e9f94078ea22d01)

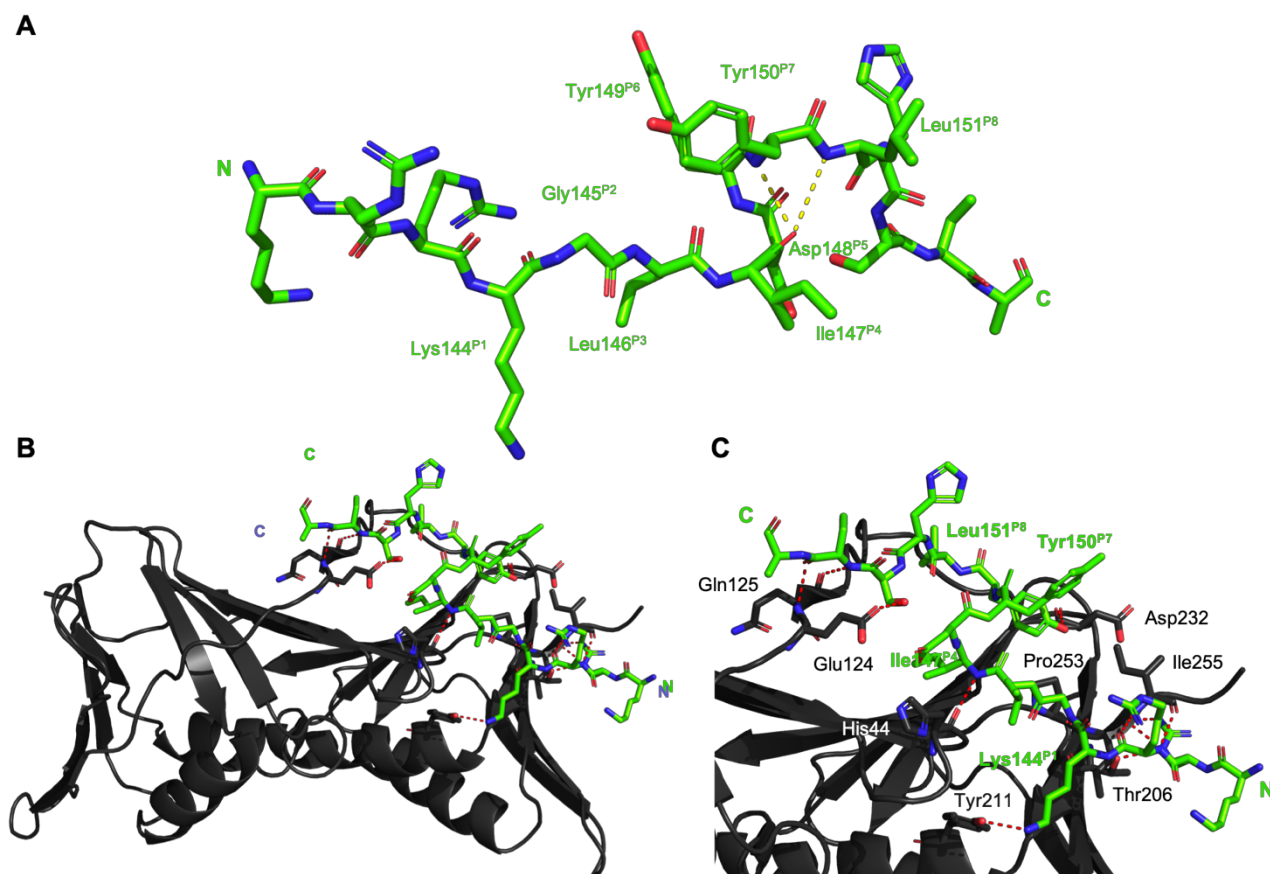
Residue	Intermolecular		Intramolecular		Total
	VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	
	142	3	2	0	
	143	1	2	0	
PIP-box	* 144	2	0	0	
	145	2	0	0	
	146	1	0	1	
	* 147	7	0	1	2
	148	1	0	0	0
	149	0	0	0	0
	* 150	3	1	0	0
	* 151	3	0	0	0
	152	0	1	0	0
FI	153	0	0	0	
	154	0	1	0	
	155	0	0	0	
	<b>Total</b>	23	7	1	3
<b>PIP-box</b>	19	1	1	3	24
<b>Flanking (FI)</b>	4	6	0	0	10
<b>Conserved (*) PIP-box residues</b>	12	1	1	2	16
<b>Non-conserved PIP-box residues</b>	4	0	0	1	5
<b>Other:</b> Intermolecular ionic interaction between B-Arg143 and A-Asp257; Intermolecular pi-stack between B-Phe151 and A-Tyr250					



**Figure S10:** Computationally modelled structure of p21<sub>μ</sub>-PARG (red, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in red. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in red.

**Table S12:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-PARG with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b1de50e9f94078ea22cf7](https://ring.rutgers.edu/session/5f3b1de50e9f94078ea22cf7)

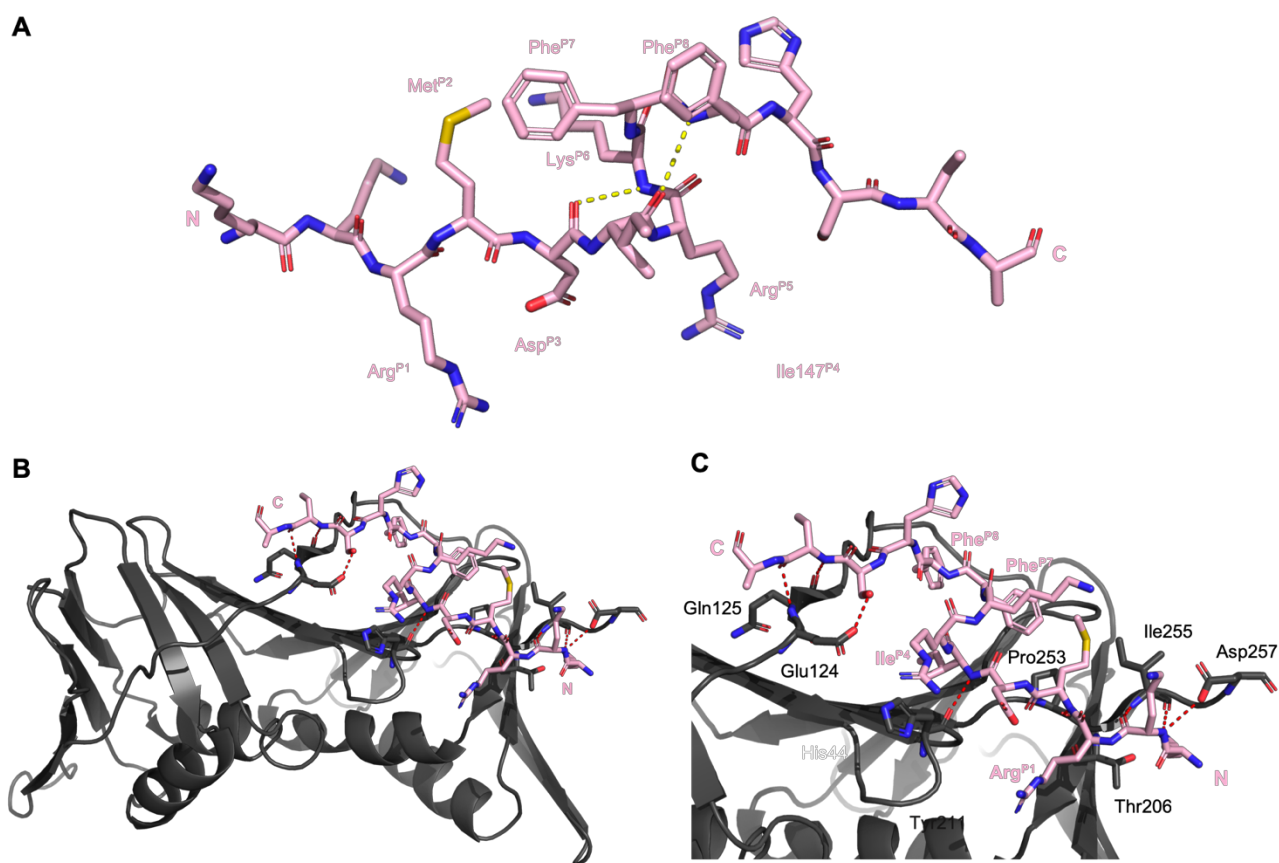
	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	2	2	0	0	
	143	0	2	0	0	
	* 144	1	0	0	0	
PIP-box	145	1	0	0	0	
	146	1	0	0	1	
	* 147	2	1	0	2	
	148	0	0	0	0	
	149	0	0	0	0	
	* 150	3	0	0	0	
	* 151	3	0	0	0	
	152	0	1	0	0	
FI	153	1	0	0	0	
	154	0	1	0	0	
	155	0	0	0	0	
	<b>Total</b>	14	7	0	3	24
	<b>PIP-box</b>	11	1	0	3	15
	<b>Flanking (FI)</b>	3	6	0	0	9
	<b>Conserved (*) PIP-box residues</b>	5	1	0	2	8
	<b>Non-conserved PIP-box residues</b>	3	0	0	1	4
<b>Other:</b> Intermolecular pi-stack between B-Tyr151 and A-Tyr250						



**Figure S11:** Computationally modelled structure of p21<sub>μ</sub>-pol I (green, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in green **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in green

**Table S13:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-pol I with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b1e580e9f94078ea22cf8](https://ring.rcsb.org/session/5f3b1e580e9f94078ea22cf8)

Residue	Intermolecular		Intramolecular		Total
	VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	
	142	2	2	0	
	143	0	2	0	
PIP-box	* 144	1	0	0	
	145	1	1	0	
	146	2	0	1	
	* 147	3	1	0	
	148	1	0	0	
	149	0	0	0	
	* 150	2	1	0	
	* 151	1	0	0	
FI	152	0	1	0	
	153	2	0	0	
	154	0	1	0	
	155	0	0	0	
<b>Total</b>	15	9	1	1	26
<b>PIP-box</b>	11	3	1	1	16
<b>Flanking (FI)</b>	4	6	0	0	10
<b>Conserved (*) PIP-box residues</b>	6	2	0	1	9
<b>Non-conserved PIP-box residues</b>	5	1	1	0	7
<b>Other:</b>					

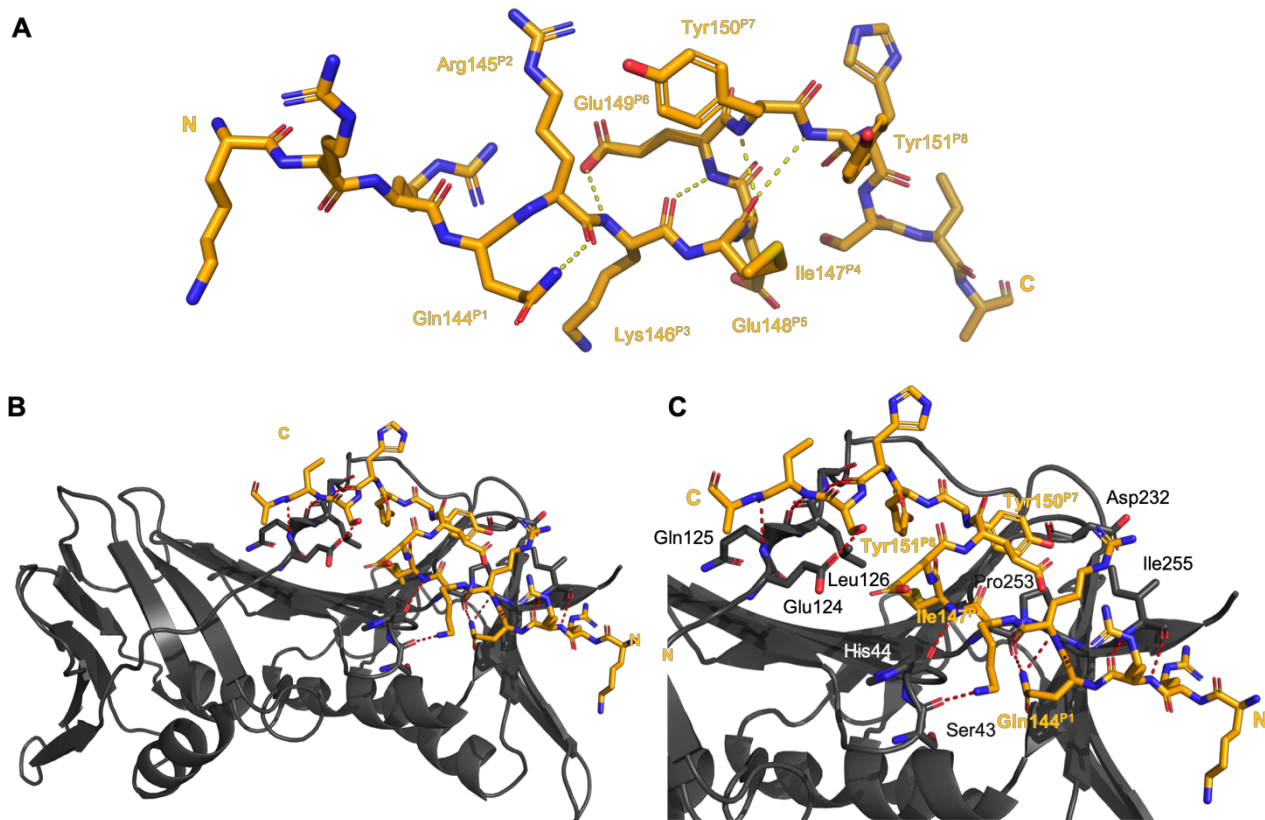


**Figure S12:** Computationally modelled structure of p21<sub>μ</sub>-RFC (light pink, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in light pink **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in light pink

**Table S14:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-RFC with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b20890e9f94078ea22cfd](https://ring-server.org/session/5f3b20890e9f94078ea22cfd)

	Residue	Intermolecular			Intramolecular		Total
		VDW	H-Bond		VDW	H-Bond	
FI	142	3	0		0	0	
	143	1	2		0	0	
PIP-box	144	1	0		0	0	
	145	2	1		1	0	
	146	1	0		0	2	
	* 147	2	1		0	2	
	148	1	0		0	0	
	149	0	0		0	0	
	* 150	2	0		0	0	
	* 151	2	0		0	0	
FI	152	0	0		0	0	
	153	2	0		0	0	
	154	0	1		0	0	
	155	0	0		0	0	
	<b>Total</b>	17	5		1	4	27
	<b>PIP-box</b>	11	2		1	4	18
	<b>Flanking (FI)</b>	6	3		0	0	9
	<b>Conserved (*) PIP-box residues</b>	6	1	0	0	2	9
	<b>Non-conserved PIP-box residues</b>	5	1	0	1	2	9
<b>Other:</b> Intermolecular ionic interaction between B-Arg143 and A-Asp257; Intermolecular pi-stack between B-Tyr151 and A-Tyr250							



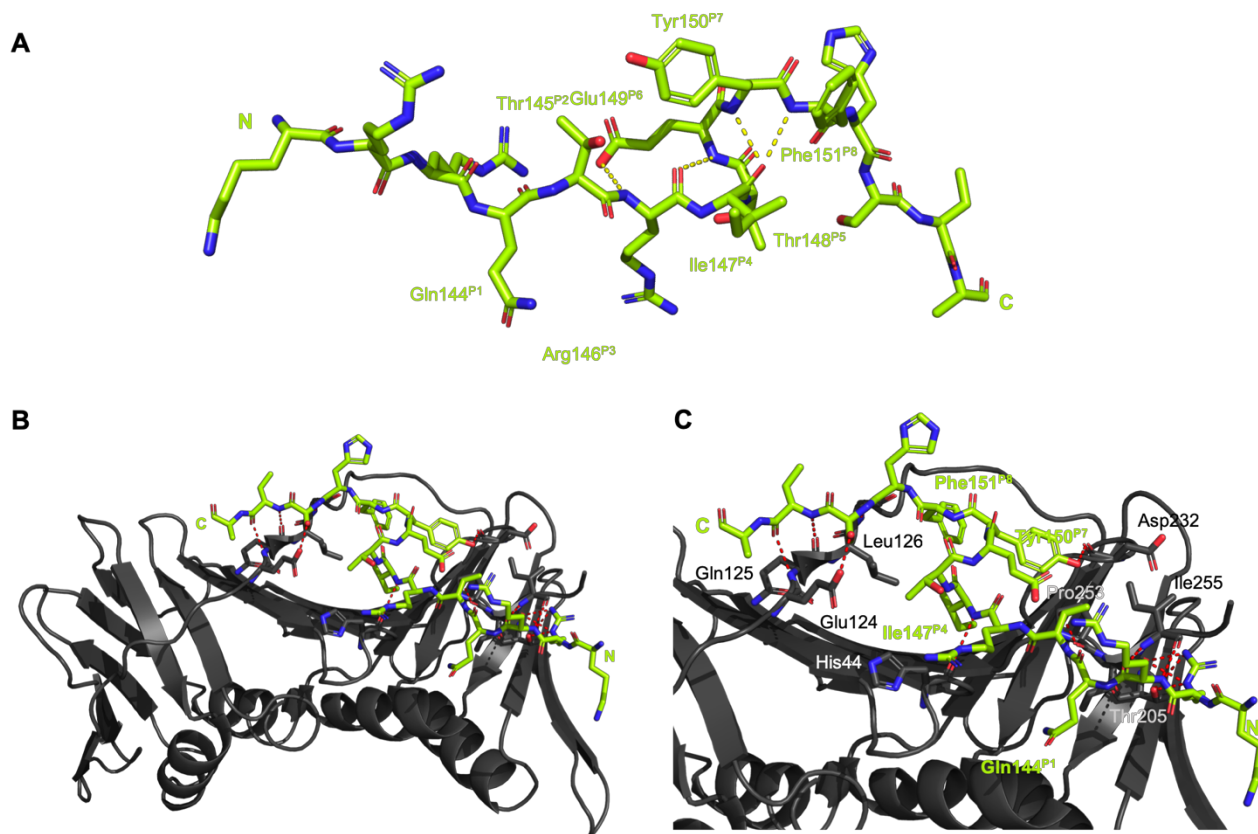


**Figure S13:** Computationally modelled structure of p21<sub>μ</sub>-RD1 (light orange, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in light orange. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in light orange.

**Table S15:** Secondary Interaction Summary for computationally modelled structured of p21<sub>μ</sub>-RD1 with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b21c10e9f94078ea22cff](https://ring-server.org/session/5f3b21c10e9f94078ea22cff)

Residue	Intermolecular		Intramolecular		Total
	VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	
	142	3	2	0	
	143	1	1	0	
	* 144	2	0	0	
	145	1	1	0	
	146	0	0	1	
	* 147	4	1	2	
	148	1	0	0	
	149	0	0	0	
	* 150	3	1	0	
	* 151	3	0	0	
	152	0	1	0	
	153	1	0	0	
	154	0	1	0	
	155	0	0	0	
<b>Total</b>	19	8	2	3	32
<b>PIP-box</b>	14	3	2	3	22
<b>Flanking (FI)</b>	5	5	0	0	10
<b>Conserved (*) PIP-box residues</b>	9	2	1	2	14
<b>Non-conserved PIP-box residues</b>	2	1	1	1	5

**Other:** Intermolecular ionic interaction between B-Arg143 and A-Asp257; Intermolecular pi-stack between B-Tyr151 and A-Tyr250

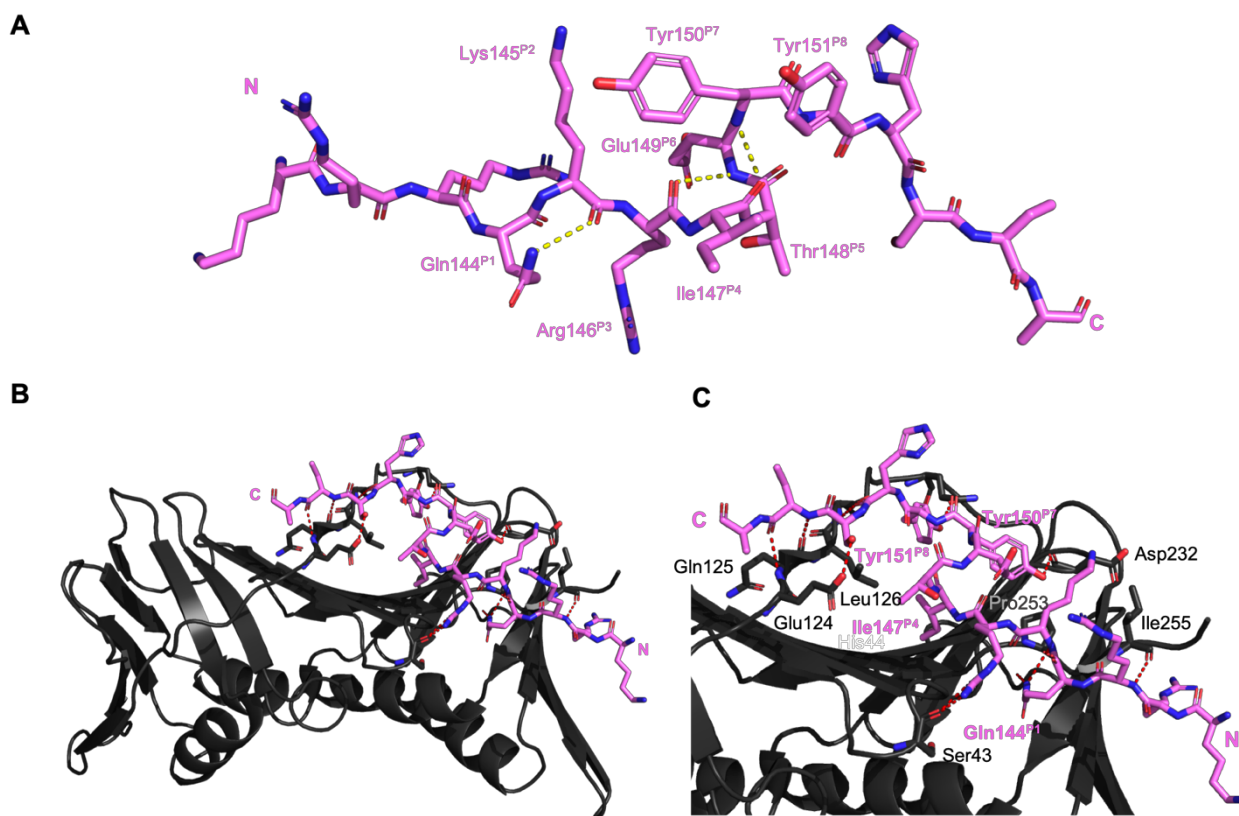


**Figure S14:** Computationally modelled structure of p21<sub>L</sub>-RD2 (green, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in green. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in green.

**Table S16:** Secondary Interaction Summary for computationally modelled structured of p21<sub>L</sub>-RD2 with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b1ff30e9f94078ea22cfa](https://ring-server.org/session/5f3b1ff30e9f94078ea22cfa)

Residue	Intermolecular		Intramolecular		Total
	VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	
	142	2	0	0	
	143	0	0	0	
PIP-box	* 144	1	0	0	
	145	1	0	0	
	146	2	1	0	
	* 147	7	1	1	
	148	1	0	0	
	149	0	0	0	
	* 150	2	1	0	
	* 151	2	0	0	
FI	152	0	0	0	
	153	0	0	0	
	154	2	1	0	
	155	0	1	0	
<b>Total</b>	20	9	1	3	33
<b>PIP-box</b>	16	4	1	3	24
<b>Flanking (FI)</b>	4	5	0	0	9
<b>Conserved (*) PIP-box residues</b>	10	2	1	2	15
<b>Non-conserved PIP-box residues</b>	4	2	0	1	7
<b>Other:</b>					





**Figure S15:** Computationally modelled structure of p21<sub>μ</sub>-RD3 (pink, sticks) on the PIP-box binding site of a hPCNA monomer (grey, cartoon). Heteroatoms indicated as blue, nitrogen; red, oxygen; yellow, sulfur. **A** Intramolecular interactions shown as yellow dashes, and PIP-box residues labelled in pink. **B & C** Intermolecular interactions shown as red dashes, PCNA residues labelled in grey/white and conserved PIP-box residues labelled in pink.

**Table S17:** Secondary Interaction Summary for computationally modelled structure of p21<sub>μ</sub>-RD3 with hPCNA calculated using the RING server. Chain B interactions only. RING Session ID: [5f3b22e70e9f94078ea22d04](https://www.rcsb.org/structure/5f3b22e70e9f94078ea22d04)

	Residue	Intermolecular		Intramolecular		Total
		VDW	H-Bond	VDW	H-Bond	
FI	141	0	0	0	0	
	142	2	1	0	0	
	143	2	2	0	0	
PIP-box	* 144	1	0	0	0	
	145	0	0	2	0	
	146	2	1	0	1	
	* 147	5	0	0	2	
	148	1	0	0	0	
	149	0	0	0	0	
	* 150	2	1	0	0	
	* 151	2	1	0	0	
FI	152	0	1	0	0	
	153	2	0	0	0	
	154	0	1	0	0	
	155	0	0	0	0	
	<b>Total</b>	19	8	2	3	32
	<b>PIP-box</b>	13	3	2	3	21
	<b>Flanking (FI)</b>	6	5	0	0	11
	<b>Conserved (*) PIP-box residues</b>	8	1	0	2	11
	<b>Non-conserved PIP-box residues</b>	3	1	2	1	7
	<b>Other:</b>					

## ANALYSIS OF STRUCTURES

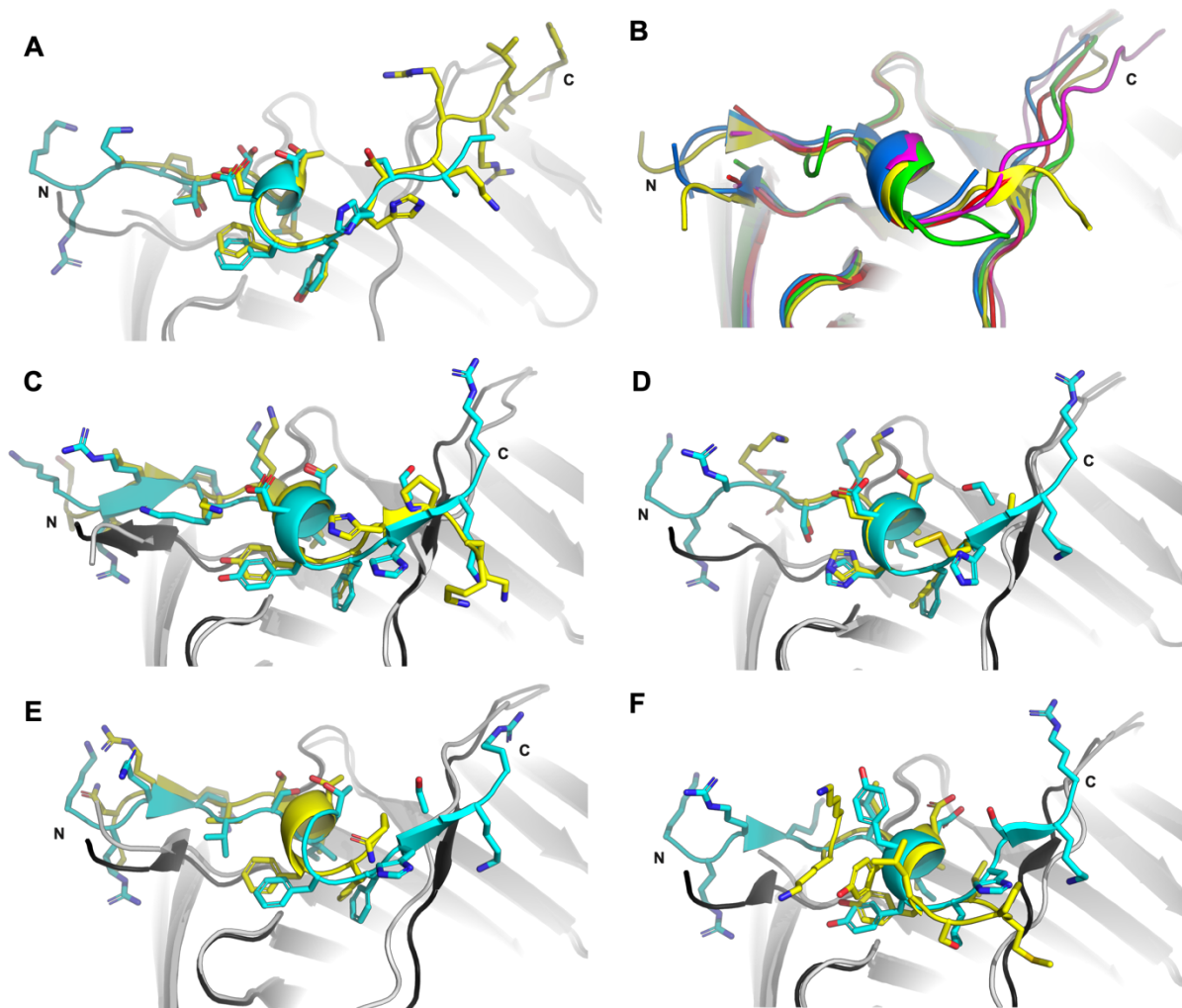
**Table S18:** Root-mean-squared deviation (RMSD) values of mutant peptides docked to the monomer of hPCNA compared to structures of p21<sub>μ</sub> bound to hPCNA and p21 bound to hPCNA.

Name	Structure Type	RMSD value against Wild Type p21 <sub>139-160</sub> (monomer of 1AXC)	RMSD value against monomer of p21 <sub>μ</sub> bound to hPCNA (PDB ID: 7KQ1)
p21 <sub>μ</sub>	Co-crystal – 7KQ1	0.511	-
p21 <sub>μ</sub>	Computational model	0.316	0.233
p21 <sub>μ</sub> -F150Y	Co-crystal – 7KQ0	0.342	0.451
p21 <sub>μ</sub> -S146R	Computational model	0.570	0.176
p21 <sub>μ</sub> -M147I	Computational model	0.571	0.191
p21 <sub>μ</sub> -D149E	Computational model	0.556	0.197
p21 <sub>μ</sub> -FY150/151YF	Computational model	0.559	0.183
p21 <sub>μ</sub> -PARG	Computational model	0.571	0.192
p21 <sub>μ</sub> -Pogo	Computational model	0.554	0.271
p21 <sub>μ</sub> -pol δ <sub>p66</sub>	Computational model	0.559	0.186
p21 <sub>μ</sub> -pol ι	Computational model	0.563	0.199
p21 <sub>μ</sub> -RFC	Computational model	0.567	0.209
p21 <sub>μ</sub> -RD1	Computational model	0.600	0.270
p21 <sub>μ</sub> -RD2	Computational model	0.558	0.181
p21 <sub>μ</sub> -RD3	Computational model	0.552	0.193

**Table S19:** Buried Surface Area (BSA) for PIP-box residues from the cocrystal structures and computationally modelled peptides calculated using the PDBePISA server v1.52 (<https://www.ebi.ac.uk/pdbe/pisa/>). Å<sup>2</sup> / Buried area percentage

PIP-box Residue	144 <sup>P1</sup>	145 <sup>P2</sup>	146 <sup>P3</sup>	147 <sup>P4</sup>	148 <sup>P5</sup>	149 <sup>P6</sup>	150 <sup>P7</sup>	151 <sup>P8</sup>
Peptide								
p21 (1AXC, Gulbis 1996)	107.86 / 70	48.31 / 70	30.39 / 60	135.10 / 100	47.60 / 60	0 / 0	59.31 / 50	132.20 / 90
p21 <sub>μ</sub> (7KQ1)	98.61 / 70	46.23 / 70	31.97 / 60	140.61 / 100	34.23 / 40	0 / 0	66.67 / 50	130.92 / 90
p21 <sub>μ</sub> -F150Y (7KQ0)	105.46 / 70	45.61 / 70	32.11 / 60	134.21 / 100	42.33 / 50	0 / 0	73.82 / 60	128.04 / 100
p21 <sub>μ</sub> -S146R	93.41 / 70	46.73 / 70	68.11 / 50	142.96 / 100	32.99 / 40	0 / 0	74.02 / 60	133.52 / 90
p21 <sub>μ</sub> -M147I	102.47 / 60	35.07 / 60	28.13 / 50	129.11 / 100	54.96 / 60	0 / 0	81.05 / 60	142.77 / 100
p21 <sub>μ</sub> -D149E	98.26 / 60	50.34 / 80	29.24 / 50	140.27 / 100	42.59 / 50	0 / 0	75.43 / 60	136.21 / 100
p21 <sub>μ</sub> -FY150/151YF	105.76 / 70	56.32 / 70	25.76 / 50	139.54 / 100	43.9 / 50	0 / 0	88.7 / 60	115.46 / 90
p21 <sub>μ</sub> -PARG	37.43 / 30	60.13 / 80	60.46 / 50	126.43 / 100	50.59 / 60	0 / 0	66.62 / 50	117.12 / 100
p21 <sub>μ</sub> -Pogo	73.05 / 60	71.12 / 60	53.08 / 50	131.07 / 100	62.18 / 60	0 / 0	83.77 / 60	120.27 / 90
p21 <sub>μ</sub> -pol δ <sub>p66</sub>	98.14 / 60	50.67 / 50	26.13 / 40	123.79 / 100	57.64 / 60	0 / 0	65.75 / 50	123.84 / 90
p21 <sub>μ</sub> -pol ι	88.44 / 50	33.75 / 70	46.18 / 50	142.62 / 100	59.19 / 70	0 / 0	97.25 / 60	74.44 / 60
p21 <sub>μ</sub> -RFC	76.16 / 40	86.28 / 60	23.01 / 40	138.71 / 100	85.43 / 50	0 / 0	62.59 / 60	96.61 / 70
p21 <sub>μ</sub> -RD1	99.56 / 60	56.85 / 40	55.85 / 50	143.05 / 100	77.77 / 60	0 / 0	81.89 / 60	127.73 / 90
p21 <sub>μ</sub> -RD2	72.88 / 50	57.05 / 90	65.92 / 50	136.97 / 100	52.52 / 50	0 / 0	76.75 / 50	104.72 / 100
p21 <sub>μ</sub> -RD3	88.02 / 80	22.38 / 20	63.49 / 50	137.11 / 100	60.31 / 80	0 / 0	75.71 / 50	123.64 / 90

## COMPARISON OF STRUCTURES TO NATIVE SEQUENCES



**Figure S16:** Overlaid structures of p21 $\mu$ :PIP-box hybrid (PCNA, white; peptide, blue) and native PIP-box peptide (PCNA, black; peptide, yellow) shown in cartoon representation and side-chains of peptide as sticks. **A** p21 $\mu$  (7KQ1) and p21<sup>139-160</sup> (1AXC) **B** Overlay of Native PIP-box peptides (p21<sup>139-160</sup> 1AXC, pink; PL 1VYJ, yellow; pol  $\delta_{p66}$  452-466 1U76, blue; PARG<sup>402-420</sup> 5MAV, red; pol I <sup>415-437</sup> 2ZVM, green **C** p21 $\mu$ -Pogo and PL (IVYJ) **D** p21 $\mu$ -PARG and PARG<sup>402-420</sup> (5MAV) **E** p21 $\mu$ -pol  $\delta$  and pol  $\delta_{p66}$  452-466 (1U76) **F** p21 $\mu$ - pol I and pol I <sup>415-437</sup> (2ZVM).

**Table S20:** Root-mean-squared deviation (RMSD) values of alternative PIP-box modified peptides docked to monomer of hPCNA compared to the structures of their respective native peptides bound to hPCNA

Alternative PIP box mutant peptide docked to hPCNA	Structure of native peptide bound to hPCNA	RMSD value
p21 $\mu$ -PARG	5MAV	0.496
p21 $\mu$ -Pogo	1VYJ	0.646
p21 $\mu$ -pol $\delta_{p66}$	1U76	0.696
p21 $\mu$ -pol I	2ZVM	1.136

**Table S21:** Number of interactions for p21 $\mu$ :hybrid peptides compared to native analogues determined from co-crystal or computationally modelled structures with RING server. For native structures the interactions are averaged over all subunit present in the pdb coordinate file.

Peptide	PDB ID:	Ref.	Affinity	# res	# Interactions					RING (3) session ID
					Total	PIP-box	Conserved	Non-conserved	Flanking	
p21 <sup>139-160</sup>	1AXC	(4)	5.96 nM	22	60.67	25	9.33	15.67	35.67	<a href="#">5ef515200e9f94078ea226cb</a>
p21 $\mu$	7KQ1		26.1 nM	15	30.67	23.67	16.67	7	7	<a href="#">5ef50f2b0e9f94078ea226bd</a>
PL <sup>1-16</sup> [mutant]	1VYJ	(5)	100 nM	16	22.33	17.67	13.33	4.33	4.67	<a href="#">5ef515730e9f94078ea226cc</a>
p21 $\mu$ -Pogo			9.12 nM	15	34	24	16	5	10	<a href="#">5f3b224d0e9f94078ea22d01</a>
pol $\delta_{p66}$ 452-466	1U76	(6)	15.6 $\mu$ M	15	30.33	24.33	17	7.33	6	<a href="#">5ef516b50e9f94078ea226cf</a>
p21 $\mu$ -pol $\delta_{p66}$			268 nM	15	29	16	11	3	13	<a href="#">5f3b202a0e9f94078ea226cfb</a>
PARG <sup>402-420</sup>	5MAV	(7)	3.3 $\mu$ M	19	26	19	4.83	14.17	7	<a href="#">5ef5164c0e9f94078ea226ce</a>
p21 $\mu$ -PARG			401 nM	15	24	15	8	4	9	<a href="#">5f3b1de50e9f94078ea226cf7</a>
pol I <sup>415-437</sup>	2ZVM	(8)	0.39 $\mu$ M	23	22.33	17.67	17.67	4.67	4.67	<a href="#">5ef515e00e9f94078ea226cd</a>
p21 $\mu$ -pol I			1.42 $\mu$ M	15	26	16	9	7	10	<a href="#">5f3b1e580e9f94078ea226cf8</a>

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