

# Supporting Information

## A Modular Synthetic Route Involving *N*-Aryl-2-Nitrosoaniline Intermediates Leads to a New Series of 3-Substituted Halogenated Phenazine Antibacterial Agents

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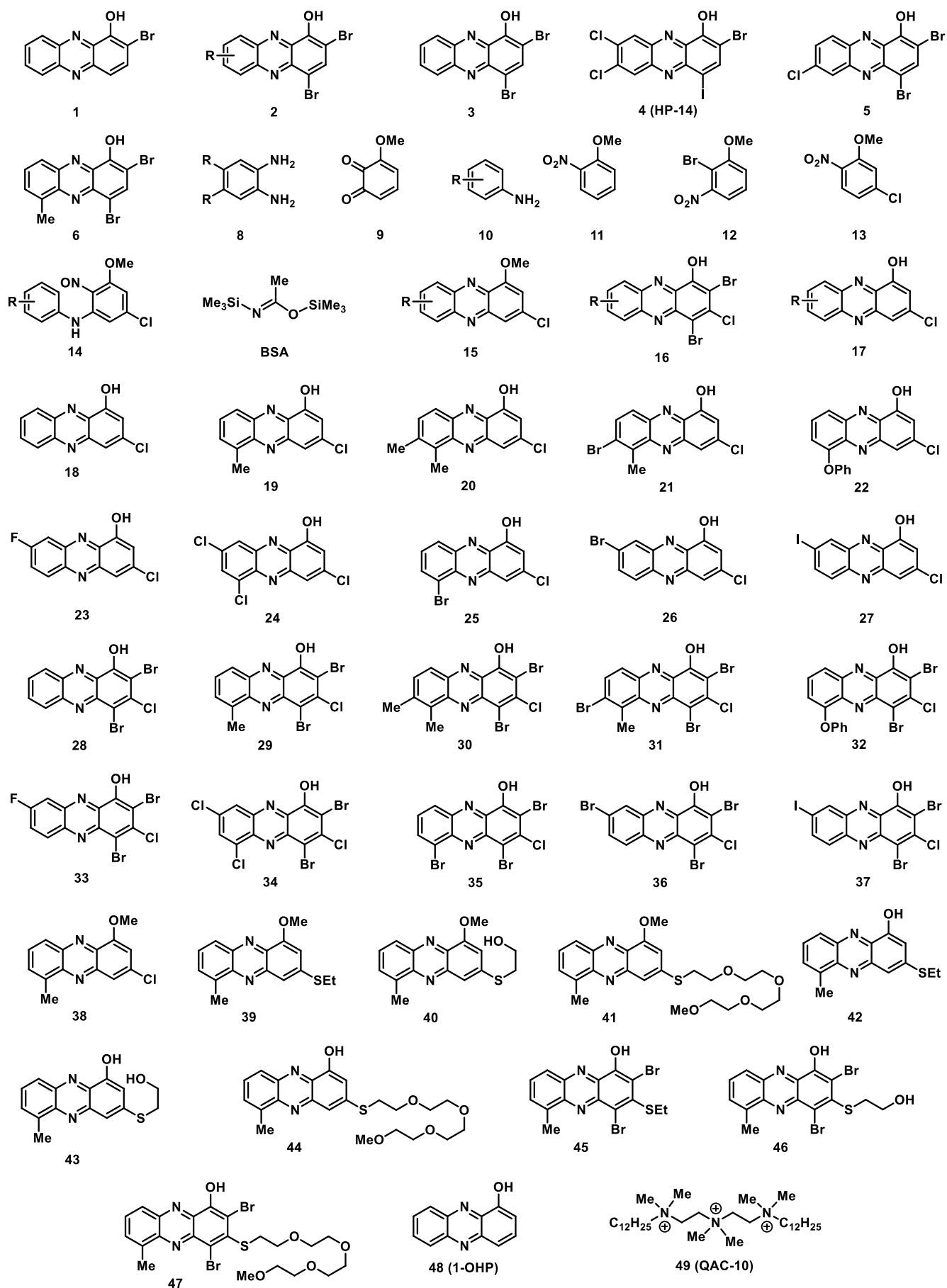
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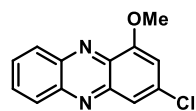
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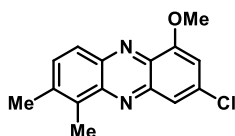
1.) **Figure S1.** Compounds reported in the manuscript.



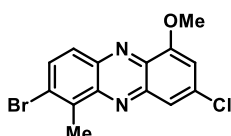
2.) **Figure S2.** Compounds reported in the supporting information.



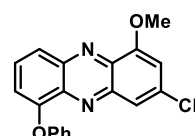
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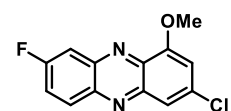
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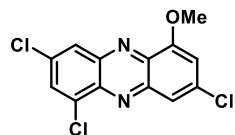
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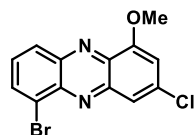
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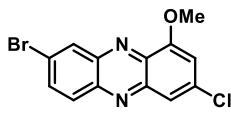
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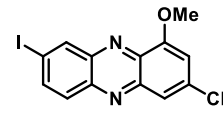
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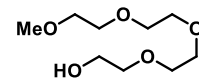
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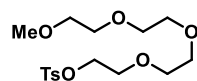
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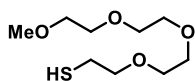
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59

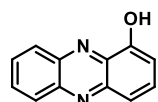


60



61

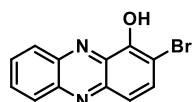
### 3.) Figure S3. Activity profiles with pKa and CLogP values for select HP analogues.



48 (1-OHP)

CLogP: 3.17  
pKa: 9.37

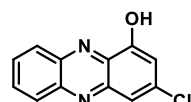
	MIC	MBEC (μM)
MRSA:	50	--
MRSE:	37.5	--
VRE:	> 50	--



1

CLogP: 3.80  
pKa: 7.61

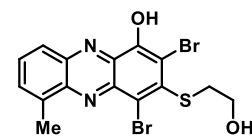
	MIC	MBEC (μM)
MRSA:	3.13	--
MRSE:	3.13	--
VRE:	6.25	--



18

CLogP: 3.93  
pKa: 8.04

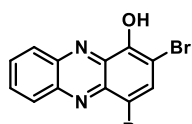
	MIC	MBEC (μM)
MRSA:	25	--
MRSE:	18.8	--
VRE:	> 50	--



46

CLogP: 4.61  
pKa: 8.07

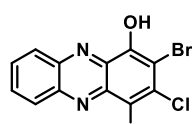
	MIC	MBEC (μM)
MRSA:	6.25	--
MRSE:	4.69	--
VRE:	12.5	--



3

CLogP: 4.68  
pKa: 7.12

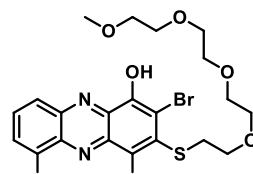
	MIC	MBEC (μM)
MRSA:	1.56	100
MRSE:	2.35	50
VRE:	4.69	12.5



28

CLogP: 5.12  
pKa: 5.93

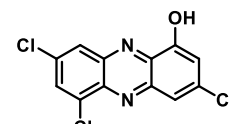
	MIC	MBEC (μM)
MRSA:	0.15	50
MRSE:	0.59	25
VRE:	1.17	2.35



47

CLogP: 5.12  
pKa: 7.93

	MIC	MBEC (μM)
MRSA:	2.35	--
MRSE:	2.35	--
VRE:	3.13	--



24

CLogP: 5.37  
pKa: 6.89

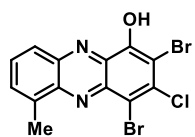
	MIC	MBEC (μM)
MRSA:	0.30	75
MRSE:	1.17	--
VRE:	2.35	--

#### KEY:

Increased Potency ↑	MIC (μM)	MBEC (μM)
	< 0.10	< 1
	0.10 - 0.59	1 - 4.69
	1.17 - 3.13	6.25 - 25
	4.69 - 6.25	37.5 - 75
	12.5 - 50	100 - 200
> 50	> 200	

Most Potent Antibacterial Activities  
CLogP > 5.6

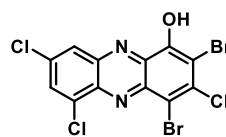
pKa does not dramatically impact activities



29

CLogP: 5.62  
pKa: 6.79

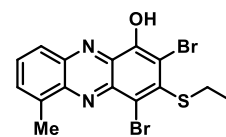
	MIC	MBEC (μM)
MRSA:	0.075	2.35
MRSE:	0.075	0.59
VRE:	0.10	0.59



34

CLogP: 6.55  
pKa: 6.07

	MIC	MBEC (μM)
MRSA:	0.59	3.13
MRSE:	0.05	0.30
VRE:	0.075	0.20



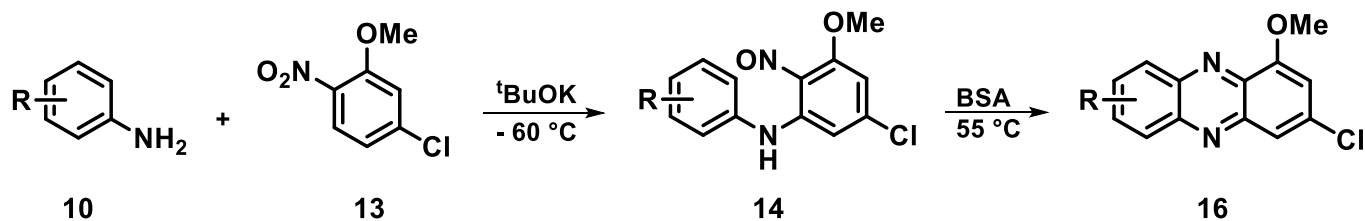
45

CLogP: 6.45  
pKa: 7.88

	MIC	MBEC (μM)
MRSA:	0.30	200
MRSE:	0.30	4.69
VRE:	0.39	3.13

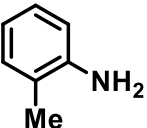
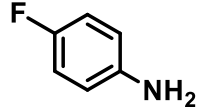
#### 4.) Table S1. Select nitroso intermediates characterized.

Select nitroso intermediates en route to 1-methoxyphenazines were identified using low resolution MS. Comparative yields of 1-methoxyphenazine from cyclization with purified nitroso intermediates and cyclization with crude intermediates are included below.



Nitroso Intermediate

Confirmed by Low Resolution MS

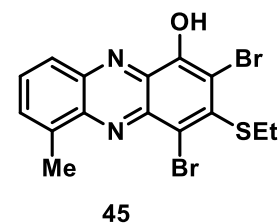
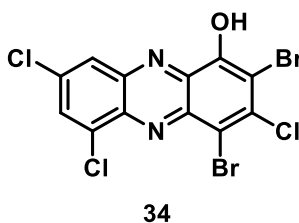
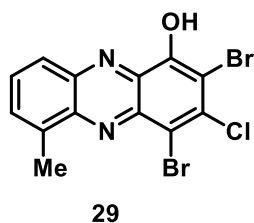
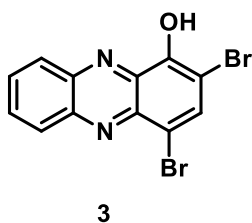
Aniline	Nitroso 14 %Yield	1-OMe HPs (16) Yield Over 2 Steps	
		Purified Nitroso	Crude Nitroso
	84%	73%	80%
	77%	70%	82%

**Note:** Crude nitroso intermediates were synthesized and divided into two equal portions. One portion was purified by flash chromatography to get the pure nitroso intermediate (confirmed by low resolution MS) before cyclization to the corresponding 1-methoxyphenazine. The other nitroso portion was directly subjected to cyclization crude, similar to the synthetic approach detailed in the manuscript. Comparison of the final yield of 1-methoxyphenazines showed that cyclization of the crude nitroso gave slightly higher yields than with pure nitroso intermediates in our hands.

**5.) Table S2. Summary of MIC assays against MRSA clinical isolates.** All MIC values are reported in micromolar ( $\mu\text{M}$ ) concentrations.

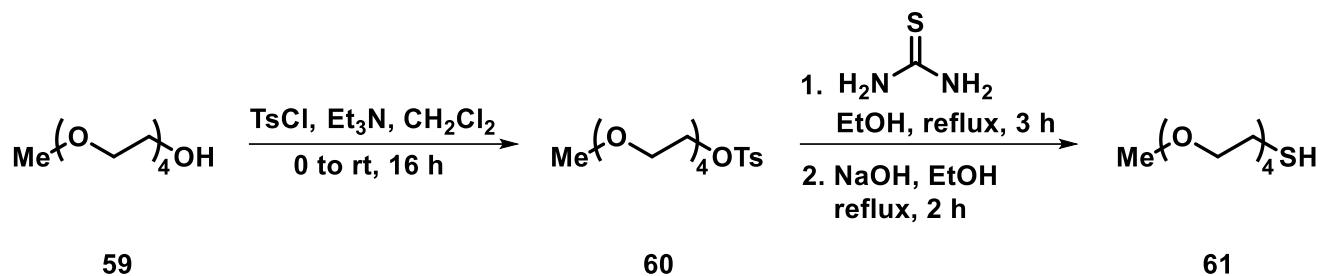
Compound	MRSA-1707	MRSA 1	MRSA 2	<i>S. aureus</i> 129	<i>S. aureus</i> 147	<i>S. aureus</i> 138	<i>S. aureus</i> 156
<b>3</b>	1.56 <sup>b</sup>	1.25	1.88 <sup>a</sup>	1.88 <sup>a</sup>	1.88 <sup>a</sup>	2.5	1.25
<b>29</b>	0.08 <sup>a</sup>	0.06 <sup>a</sup>	0.12 <sup>a</sup>	0.12 <sup>a</sup>	0.12 <sup>a</sup>	0.08	0.08
<b>34</b>	0.59 <sup>a</sup>	1.25	0.94	0.63	0.47 <sup>a</sup>	0.94 <sup>a</sup>	0.47 <sup>a</sup>
<b>45</b>	0.30 <sup>a</sup>	0.31	0.63	0.47 <sup>a</sup>	0.47 <sup>a</sup>	0.47 <sup>a</sup>	0.47 <sup>a</sup>
<b>Vancomycin</b>	0.39	0.39	0.39	0.39	0.59 <sup>a</sup>	0.39	0.39
<b>Methicillin</b>	18.8 <sup>a</sup>	37.5 <sup>a</sup>	>100	37.5 <sup>a</sup>	6.25	37.5 <sup>a</sup>	37.5 <sup>a</sup>
<b>Ciprofloxacin</b>	0.59 <sup>a</sup>	0.78	>100	>100	1.17 <sup>a</sup>	>100	0.78 <sup>b</sup>

Notes: All MIC values were recorded from a minimum of three independent biological assays. <sup>a</sup> = midpoint of a 2-fold range in observed MIC values. <sup>b</sup> = midpoint of a 4-fold range in observed MIC values. For these MIC assays, HPs were tested at a range of 0.01 - 10  $\mu\text{M}$ , while antibiotics were tested at a range of 0.1 - 100  $\mu\text{M}$ .



Clinical isolates MRSA 1, MRSA 2, *S. aureus* 129, *S. aureus* 147, *S. aureus* 138 and *S. aureus* 156 were obtained from the Emerging Pathogens Institute (EPI) at the University of Florida and isolated from patients treated at UF Health Shands Hospital (Gainesville, FL). Methicillin has an MIC = 0.78  $\mu\text{M}$  against *S. aureus* strain ATCC 29213, which is susceptible to this antibiotic. MRSA-1707 was obtained from ATCC (MRSA strain BAA-1707) and included in this table as a comparator.

## 6.) Synthetic Procedures.



**Procedures.** The title compound was synthesized according to modified literature procedures.<sup>1,2</sup>

### First step:

Tetraethylene glycol monomethyl ether (2.18 g, 13.4 mmol) was dissolved in dichloromethane (12 mL) before triethylamine (2.25 mL, 16.02 mmol) was added. Then, a solution of 4-toluenesulfonyl chloride (2.55 g, 13.4 mmol) in dichloromethane (2 mL) was added at 0 °C and the resulting reaction mixture was allowed to stir at room temperature for 3 days before being quenched with brine. The mixture was then transferred to a separatory funnel and extracted with dichloromethane (3 x 30 mL). The organic layers were collected and dried with anhydrous sodium sulfate, filtered, and concentrated *in vacuo* to afford **60** as a colorless oil (100%, 5.5 g). Note: Our <sup>1</sup>H NMR align with the previously reported spectra for **60** (CAS: 62921-76-0).<sup>1</sup>

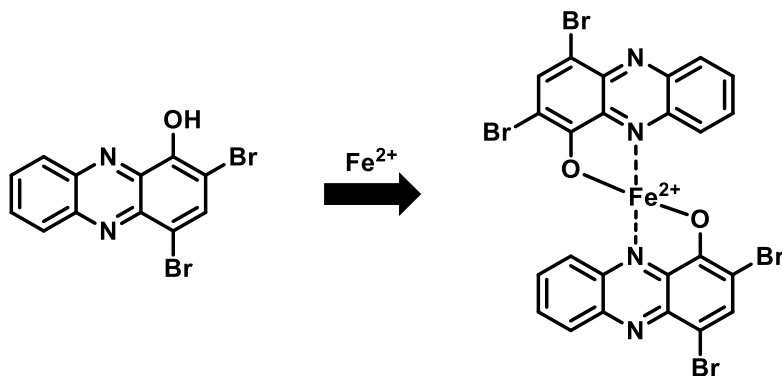
### Second step:

Intermediate **60** (2.90 g, 8.00 mmol) was dissolved in ethanol (5 mL). Then, thiourea (609 mg, 8.00 mmol) was added to the solution along with deionized water (3.5 mL). The resulting reaction mixture was then heated to reflux and allowed to stir for 3 hours. After that time, a solution of sodium hydroxide (384 mg, 9.60 mmol) in water (2.5 mL) was added to the mixture, which was heated to reflux and allowed to stir for an additional 2 hours. When the reaction was complete, the mixture was cooled to room temperature and concentrated to 6 mL *in vacuo*, neutralized with 1 M hydrochloric acid until pH 7, and extracted with dichloromethane. The organic layers were combined and dried with anhydrous sodium sulfate, filtered, and concentrated *in vacuo*. The resulting crude oil was purified using flash column chromatography with 4:1 to 1:1 hexanes:ethyl acetate to yield **61** as a colorless oil (82%, 1.38 g). Note: Our <sup>1</sup>H NMR align with the previously reported spectra for **61** (CAS: 52190-55-3).<sup>2</sup>

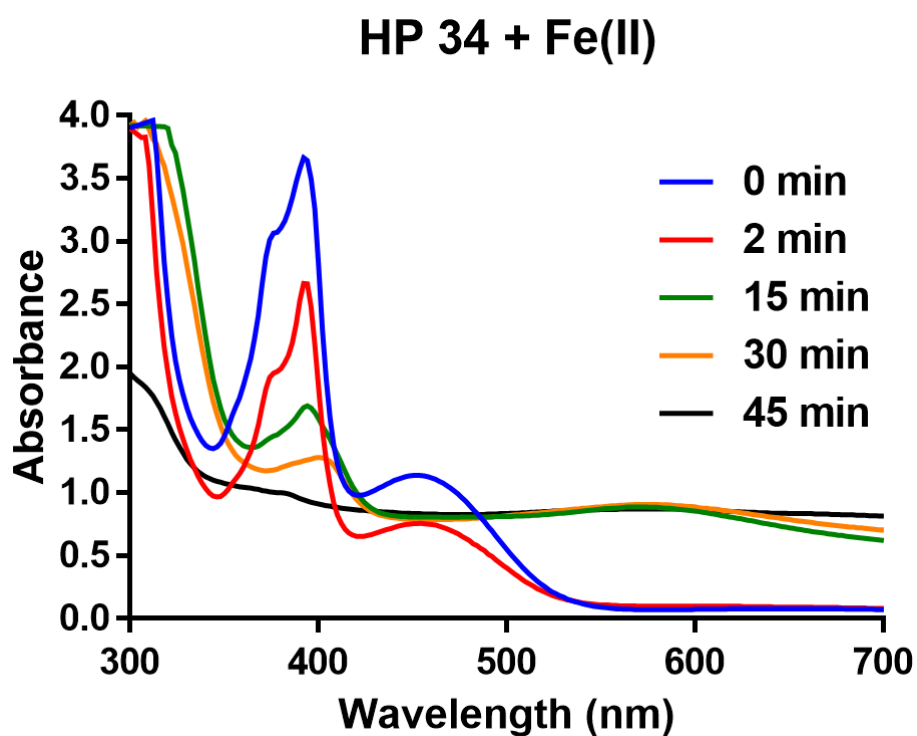


## 7.) UV-Vis Spectroscopy.

We use UV-Vis to show iron(II) binding, which form a 2:1 HP:iron(II) complex as shown below.<sup>3-5</sup>



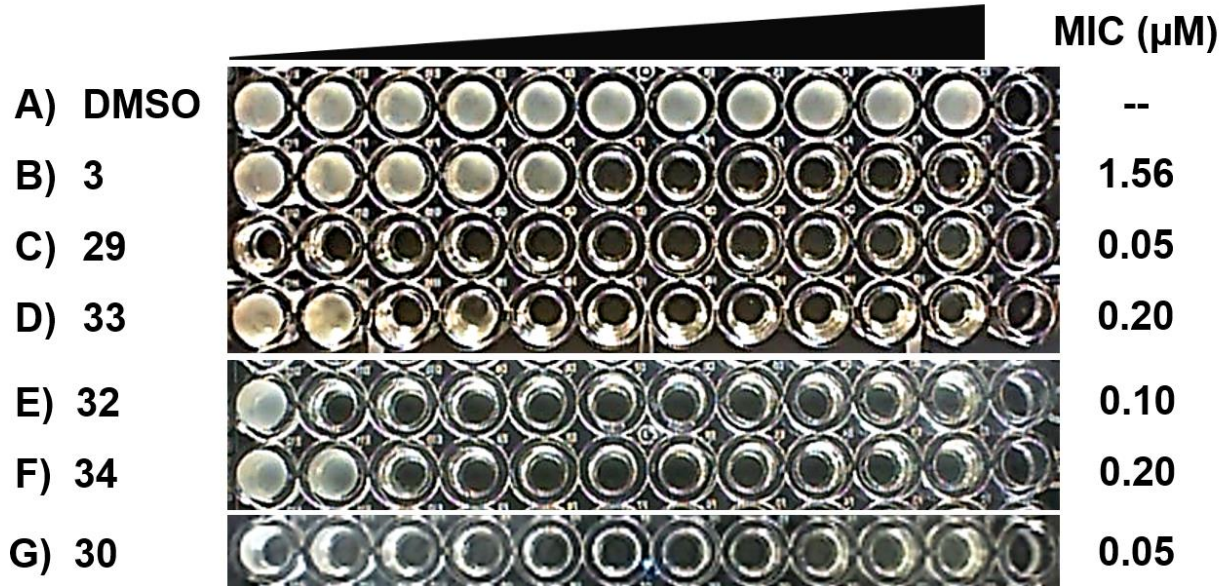
UV-Vis analysis of iron(II) binding with halogenated phenazine **34** (other HPs are presented in the manuscript).



## 8.) MIC Assays.

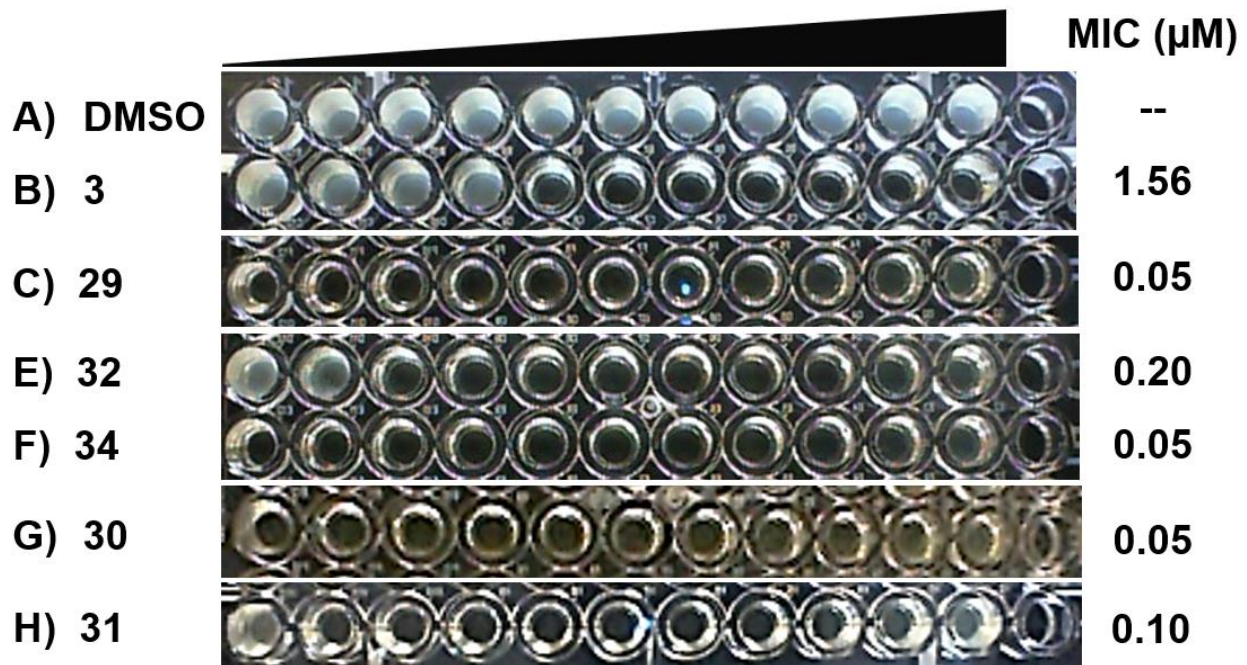
Images of representative MIC assays performed during these studies (2-fold serial dilution concentrations).

### MRSA-44



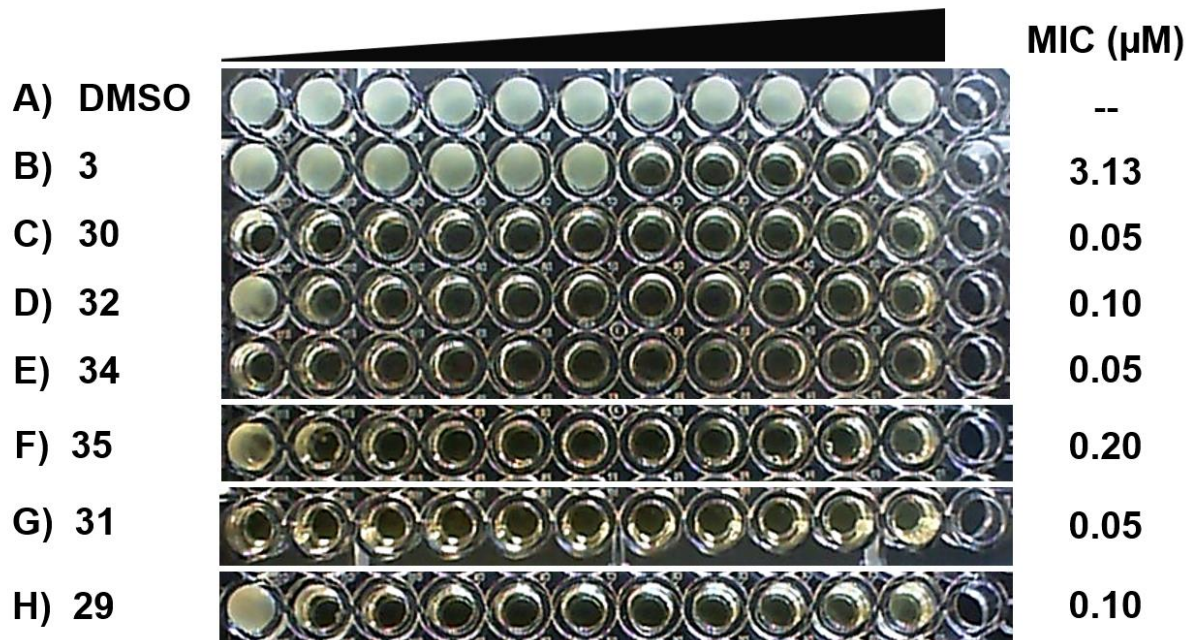
Test concentration: 0.05 - 50 µM

### MRSE 35984



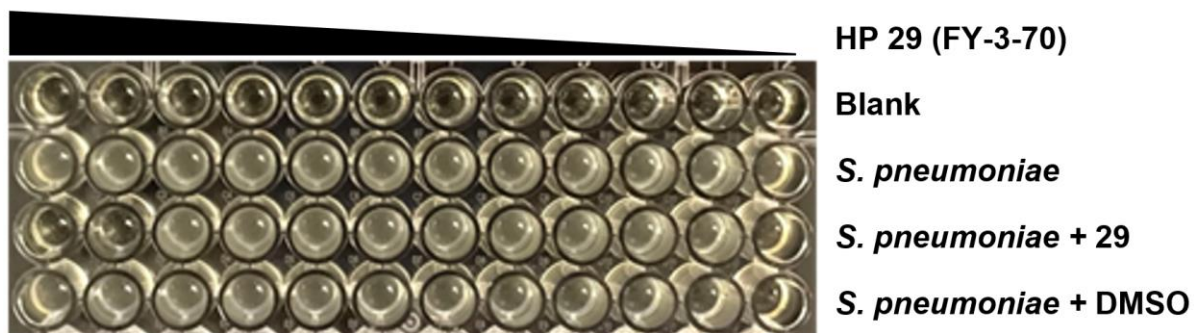
Test concentration: 0.05 - 50 µM

# VRE 700221



Test concentration: 0.05 - 50  $\mu\text{M}$

## HP 29 (FY-3-70) against *S. pneumoniae* 6303

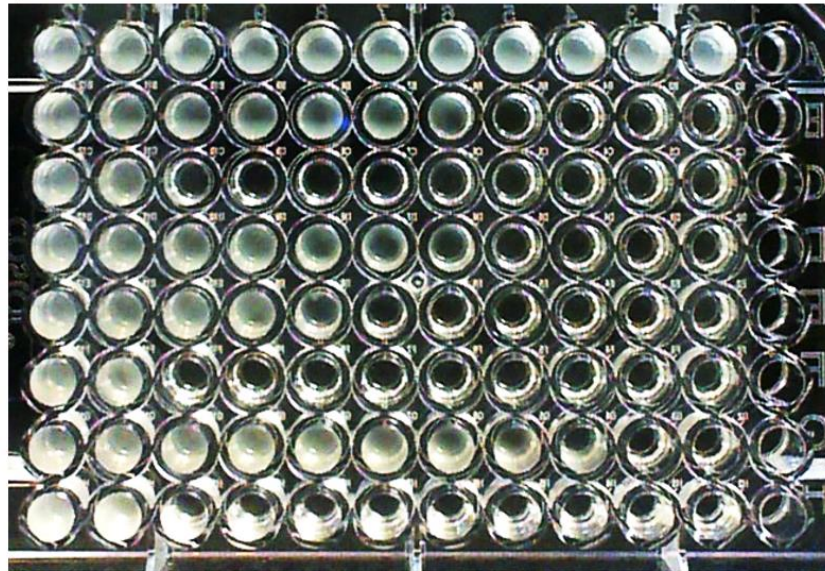


Test concentration: 0.5 - 1000 nM

Vanco. = vancomycin, Cipro. = ciprofloxacin

## MRSA 1

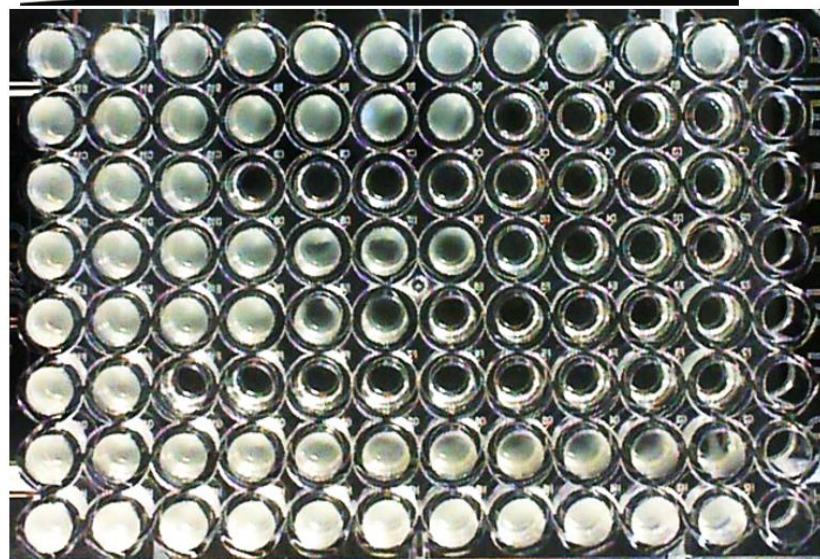
	MIC ( $\mu\text{M}$ )
A) DMSO	--
B) 3	1.25
C) 29	0.04
D) 34	1.25
E) 45	0.31
F) Vanco.*	0.39
G) Methicillin*	25
H) Cipro.*	0.78



Test concentration: 0.01 - 10  $\mu\text{M}$ ; 0.1 - 100  $\mu\text{M}$ \*









## MRSA 2

	MIC ( $\mu\text{M}$ )
A) DMSO	--
B) 3	1.25
C) 29	0.08
D) 34	1.25
E) 45	0.63
F) Vanco.*	0.39
G) Methicillin*	> 100
H) Cipro.*	> 100











Test concentration: 0.01 - 10  $\mu\text{M}$ ; 0.1 - 100  $\mu\text{M}$ \*

# *S. aureus* 129

		MIC ( $\mu\text{M}$ )
A) DMSO		--
B) 3		2.5
C) 29		0.16
D) 34		0.63
E) 45		0.63
F) Vanco.*		0.39
G) Methicillin*		50
H) Cipro.*		> 100

Test concentration: 0.01 - 10  $\mu\text{M}$ ; 0.1 - 100  $\mu\text{M}$ \*

# *S. aureus* 147

		MIC ( $\mu\text{M}$ )
A) DMSO		--
B) 3		2.5
C) 29		0.08
D) 34		0.63
E) 45		0.63
F) Vanco.*		0.78
G) Methicillin*		6.25
H) Cipro.*		0.78

Test concentration: 0.01 - 10  $\mu\text{M}$ ; 0.1 - 100  $\mu\text{M}$ \*

# *S. aureus* 138

- A) DMSO
- B) 3
- C) 29
- D) 34
- E) 45
- F) Vanco.\*
- G) Methicillin\*
- H) Cipro.\*

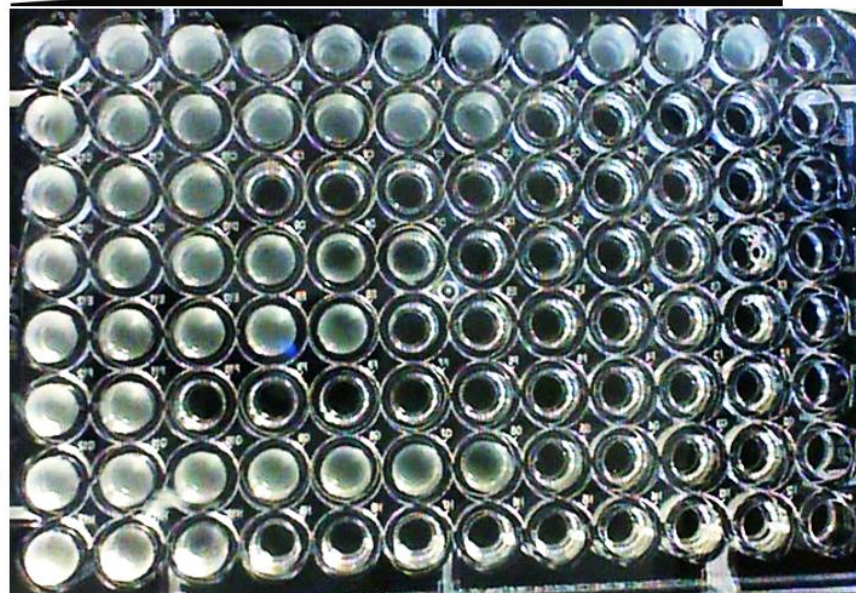


MIC ( $\mu\text{M}$ )
--
2.5
0.08
0.63
0.31
0.39
25
> 100

Test concentration: 0.01 - 10  $\mu\text{M}$ ; 0.1 - 100  $\mu\text{M}$ \*

# *S. aureus* 156

- A) DMSO
- B) 3
- C) 29
- D) 34
- E) 45
- F) Vanco.\*
- G) Methicillin\*
- H) Cipro.\*

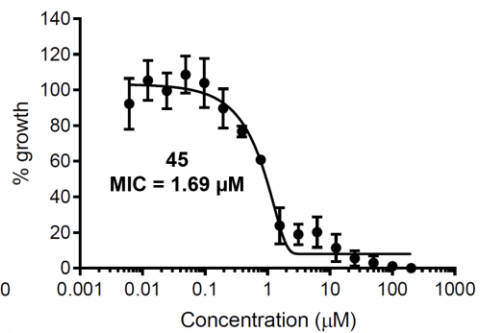
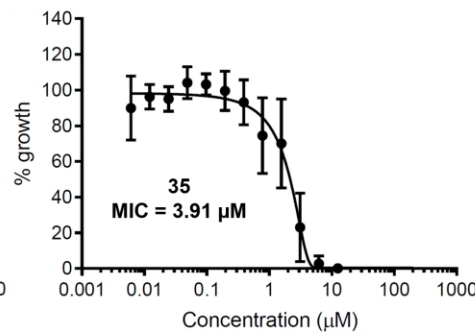
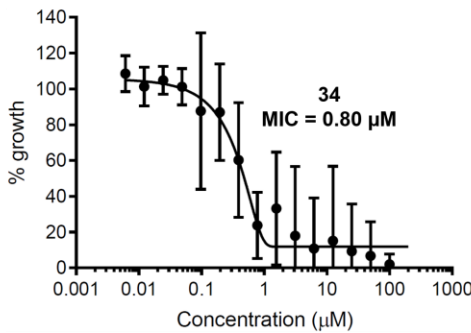
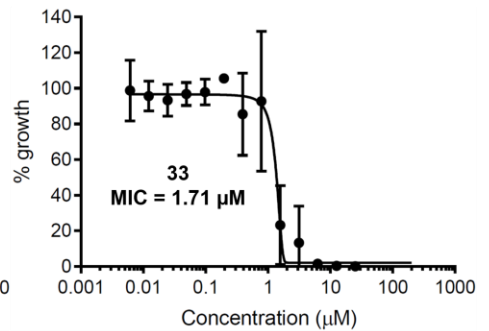
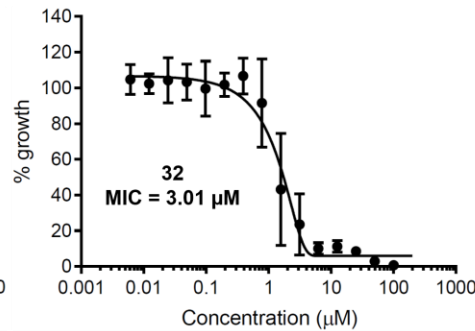
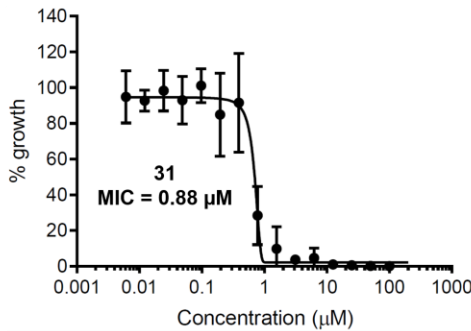
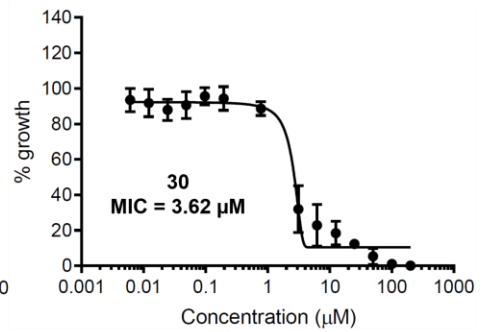
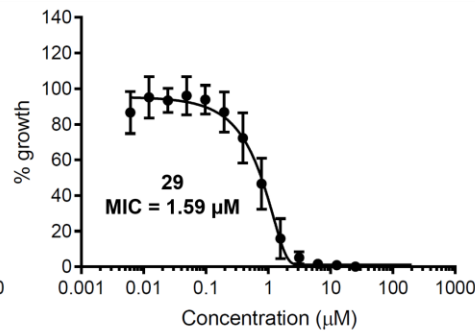
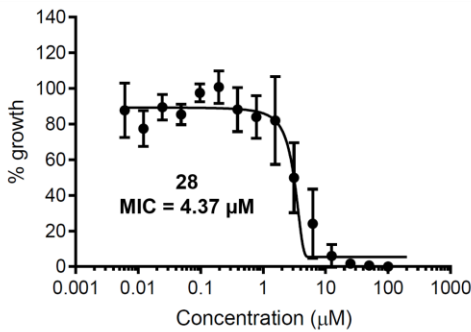
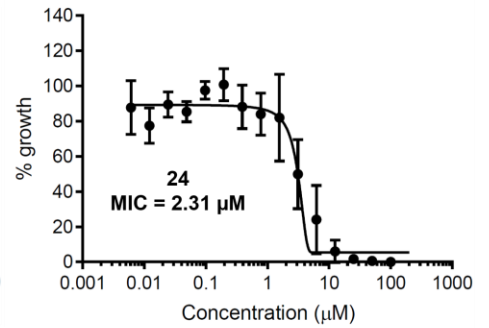
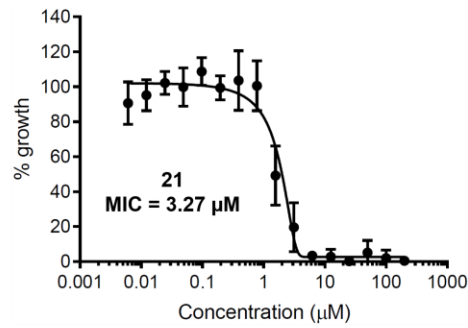
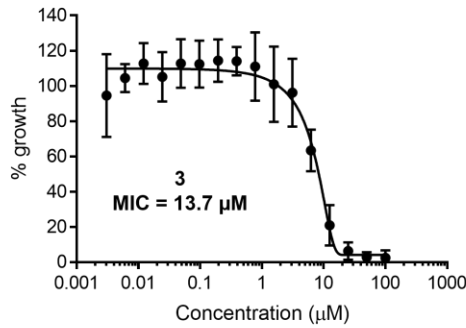


MIC ( $\mu\text{M}$ )
--
1.25
0.08
0.63
0.31
0.39
25
0.78

Test concentration: 0.01 - 10  $\mu\text{M}$ ; 0.1 - 100  $\mu\text{M}$ \*

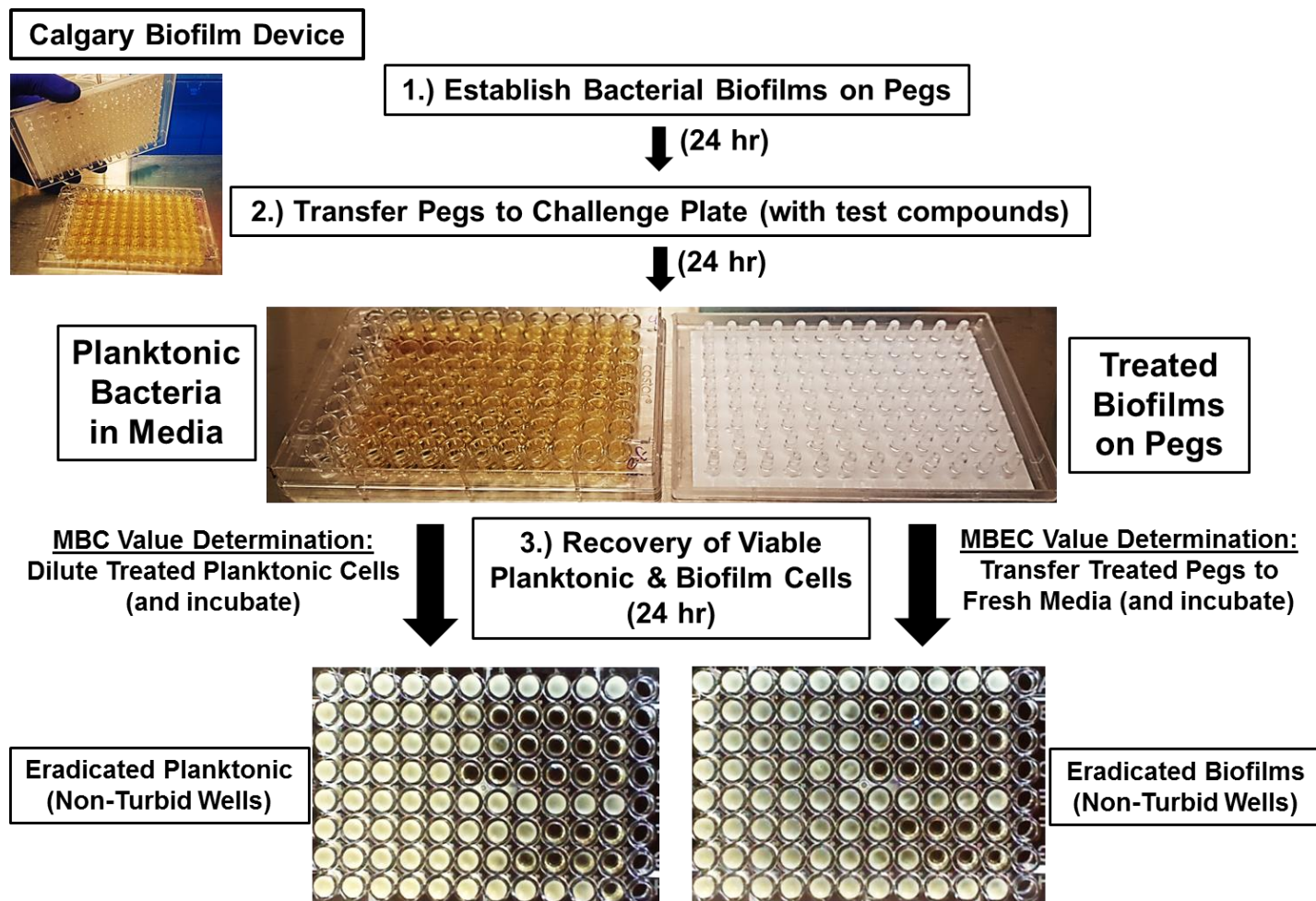
## 9.) Dose-Response Curves of HPs against *Mycobacterium tuberculosis* CDC1551.

Dose-response curves determined from OD readings of MIC assays performed in 96-well plates.



## 10.) Biofilm Eradication Assays with Calgary Biofilm Devices.

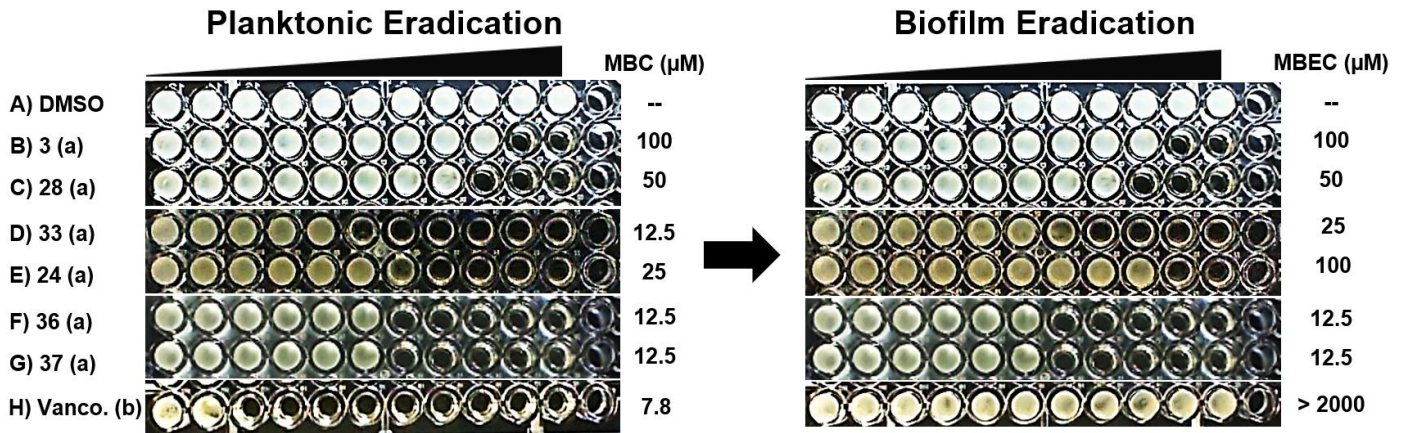
Assay workflow to determine planktonic and biofilm eradication activities.





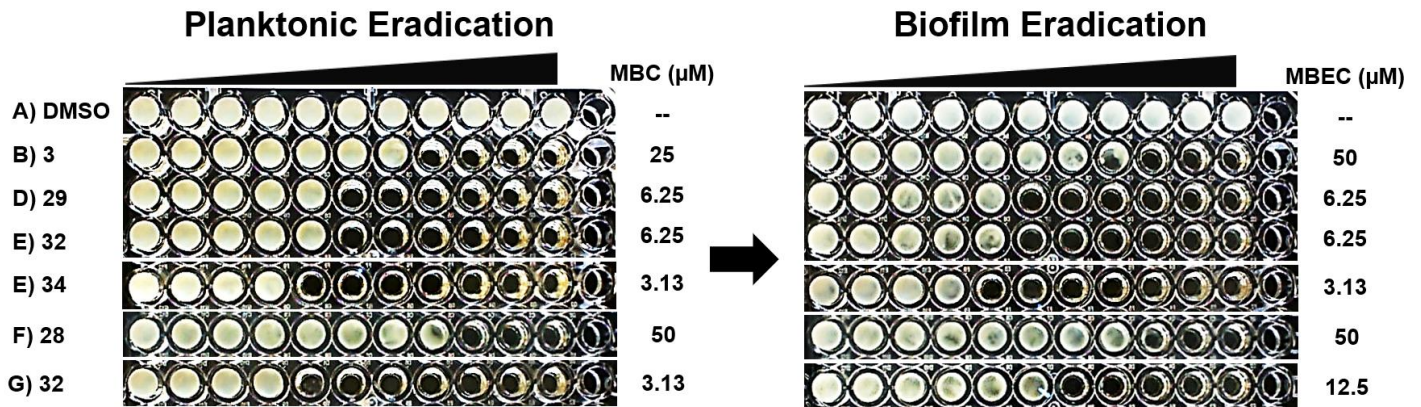
Images of representative CBD assays performed during these studies.

### MRSA-1707



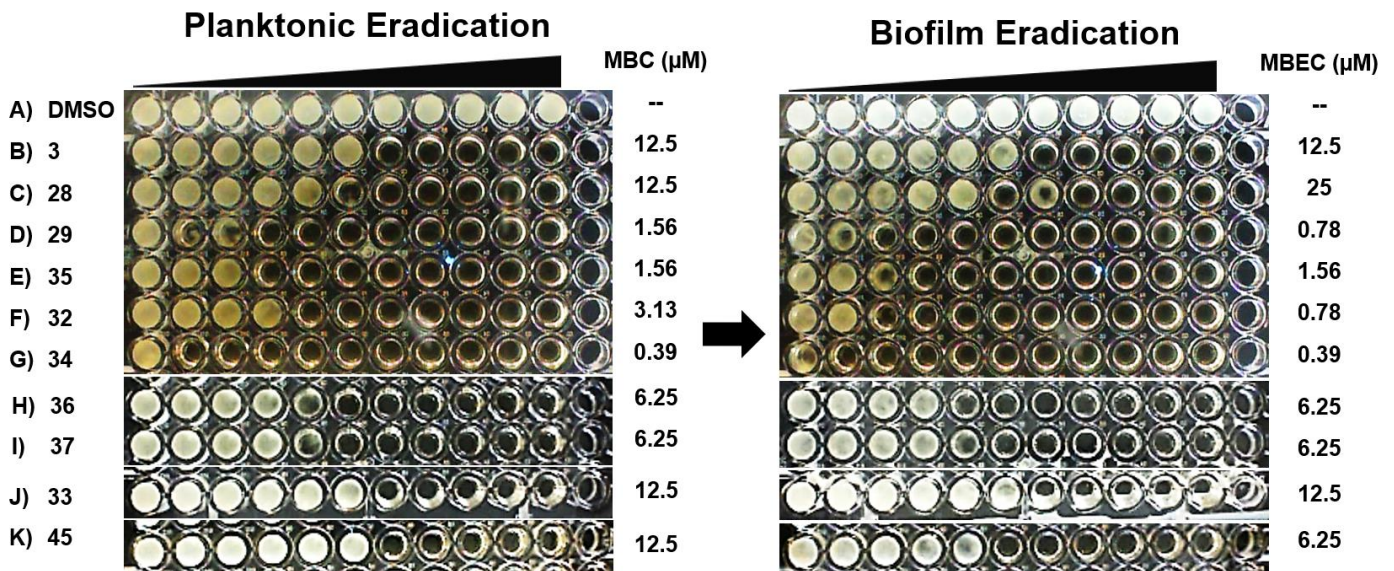
(a) Test concentration: 0.2 – 200  $\mu\text{M}$ ; (b) Test concentration: 2-2000  $\mu\text{M}$ ; Vanco. (Vancomycin).

### MRSA-44



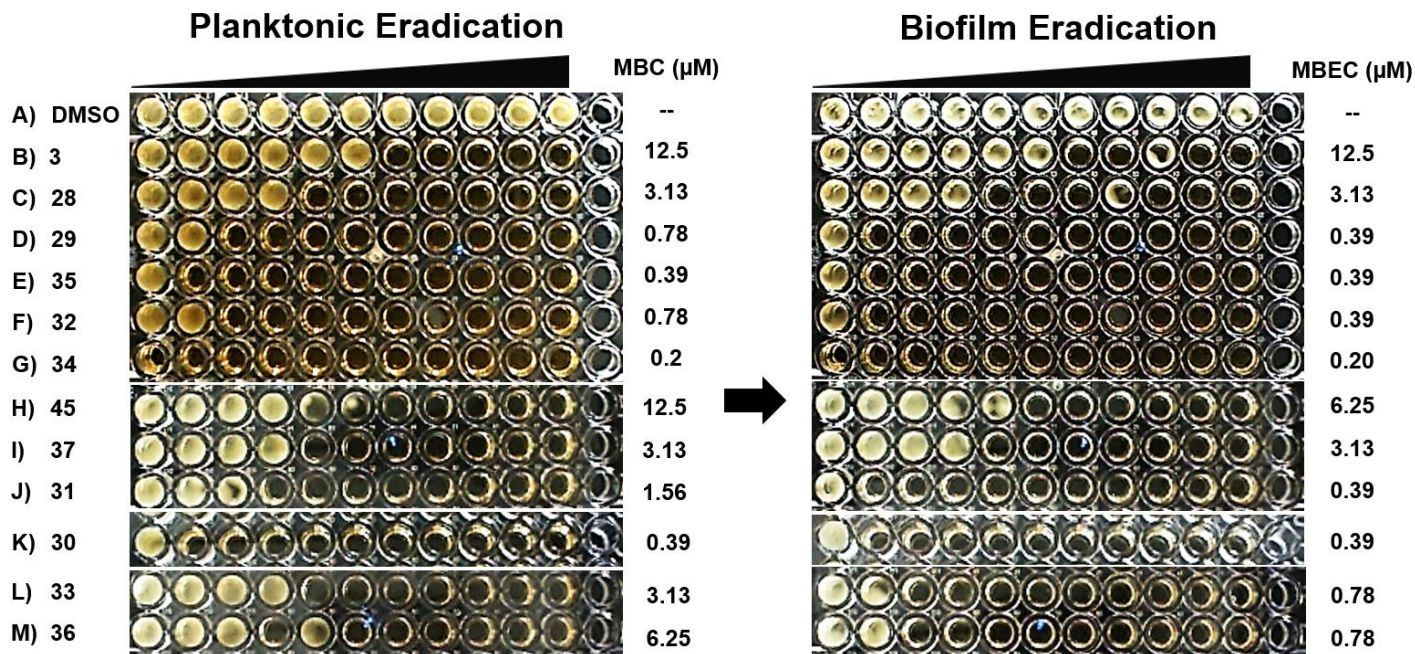
Test concentration: 0.2 – 200  $\mu\text{M}$

### MRSE 35984



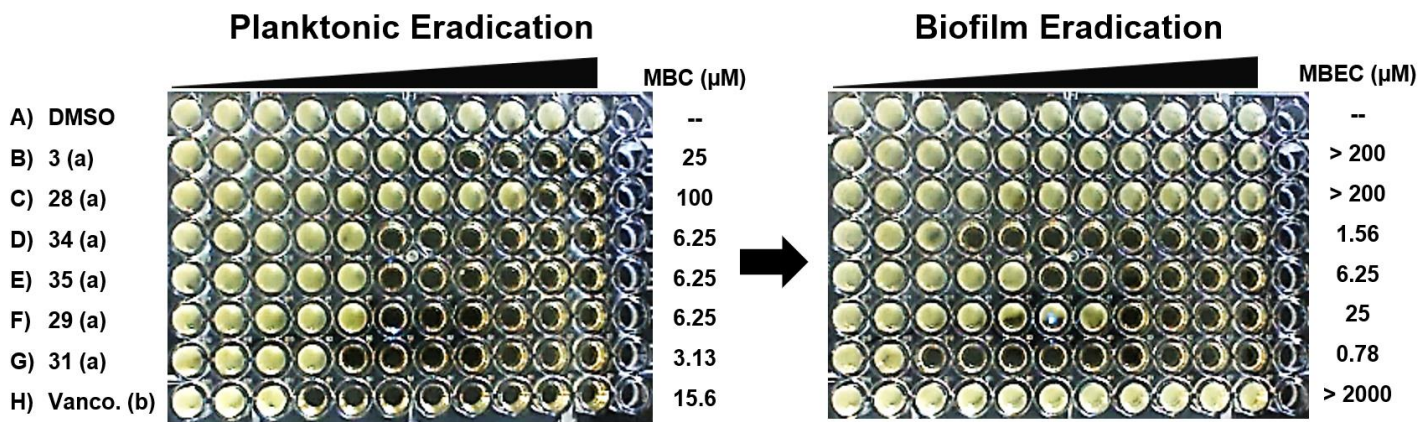
Test concentration: 0.2 – 200  $\mu\text{M}$

# VRE 700221



Test concentration: 0.2 – 200  $\mu\text{M}$

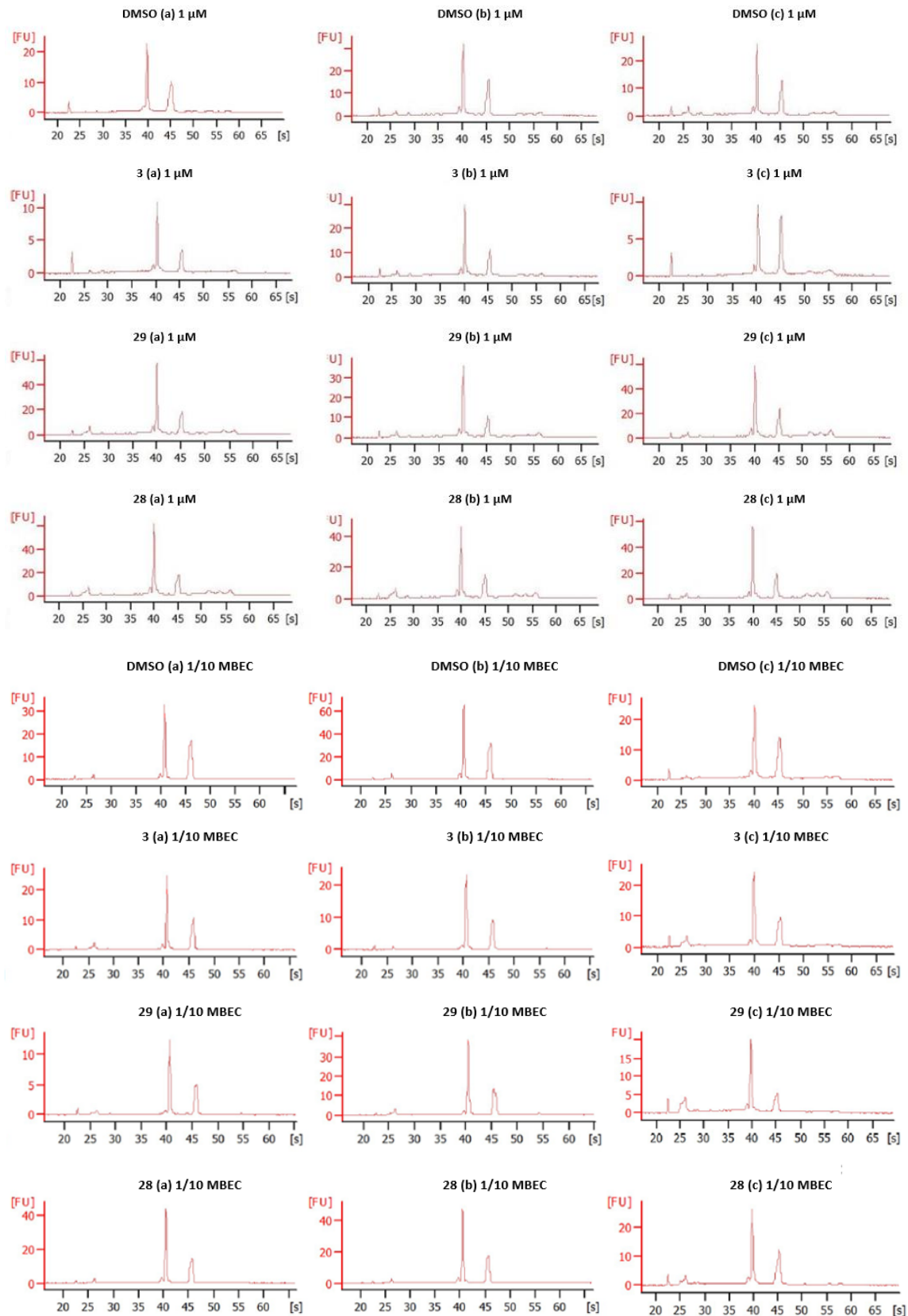
# *E. faecalis* OG1RF



(a) Test concentration: 0.2 – 200  $\mu\text{M}$ ; (b) Test concentration: 2-2000  $\mu\text{M}$ ; Vanco. (Vancomycin).

## 11.) Iron Starvation in MRSA Biofilms via RT-qPCR.

Time course quality control data for biofilm experiments obtained from the Interdisciplinary Center for Biotechnology Research (ICBR) at the University of Florida.



Time course RT-qPCR validation of MRSA-1707 biofilm upon treatment with **3**, **28**, and **29** at (A) 1  $\mu$ M, (B) 1/10 MBEC for 4 hours.

(A) MRSA-1707 biofilms treated with HPs at 1  $\mu$ M.

Gene	DMSO	3	28	29	Gene Information / Function
	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	
<i>isdB</i>	1.0 $\pm$ 0.02	5.3 $\pm$ 4.00	7.6 $\pm$ 2.13	7.0 $\pm$ 3.31	hemoglobin receptor required for heme iron utilization
<i>sbnC</i>	1.0 $\pm$ 0.02	5.4 $\pm$ 1.85	10.8 $\pm$ 2.16	12.7 $\pm$ 2.98	staphyloferrin B (siderophore) biosynthesis; iron acquisition
MW0695	1.0 $\pm$ 0.05	4.4 $\pm$ 3.10	6.1 $\pm$ 4.08	10.2 $\pm$ 4.99	hypo. protein, similar to ferrichrome ABC transporters
<i>sfaA</i>	1.0 $\pm$ 0.02	3.4 $\pm$ 0.50	4.4 $\pm$ 0.48	3.9 $\pm$ 1.47	staphyloferrin A (siderophore); iron acquisition

(B) MRSA-1707 biofilms treated with HPs at 1/10 MBEC.

Gene	DMSO	3	28	29	Gene Information / Function
	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	Mean $\pm$ SD	
<i>isdB</i>	1.0 $\pm$ 0.01	7.3 $\pm$ 1.70	14.4 $\pm$ 5.73	8.6 $\pm$ 2.04	hemoglobin receptor required for heme iron utilization
<i>sbnC</i>	1.1 $\pm$ 0.08	15.5 $\pm$ 4.35	14.1 $\pm$ 5.86	15.3 $\pm$ 4.82	staphyloferrin B (siderophore) biosynthesis; iron acquisition
MW0695	1.0 $\pm$ 0.04	1.9 $\pm$ 0.66	2.4 $\pm$ 1.84	1.9 $\pm$ 0.87	hypo. protein, similar to ferrichrome ABC transporters
<i>sfaA</i>	1.0 $\pm$ 0.02	3.2 $\pm$ 1.06	2.9 $\pm$ 1.45	3.5 $\pm$ 2.66	staphyloferrin A (siderophore); iron acquisition

Primers used for qPCR validation experiments.

Gene	Symbol	Forward primer	Reverse primer
MW0091	<i>sbnC</i>	TCCGGTACATCCTTGGCAAT	GACATGGTTATACGGTGCGC
MW1011	<i>isdB</i>	CCAGCAGCAAAGCCACTAA	CGAGAGTTTGGTGCGCTATG
MW0695	n.a.	CTGGAACGATGGAATGGGCT	CGTGCGCAATGAGATGAAGG
MW2106	<i>sfaA</i>	GACGATGTGTGCCATTGGTG	TTGTTCGTACGTGCCCATTTGA
MW1668	<i>ptaA</i>	TCCTAGCGAGTTCAGTTGCA	CCAATGGAATGTAGCTGCGA

**Notes:** Primers were designed using OligoPerfect Primer Designer (Thermofisher). During these studies, *ptaA* was used as the housekeeping gene for MRSA-1707 (*S. aureus* MW2) biofilms.

Materials used added to a 1.5 mL Eppendorf tube on ice.

Material	Amount / Reaction (20 $\mu$ L)	Triplicate + Excess (88 $\mu$ L)
SYBR	10 $\mu$ L	44 $\mu$ L
Primer (5 nM)_forward	1.5 $\mu$ L	6.5 $\mu$ L
Primer (5 nM)_reverse	1.5 $\mu$ L	6.5 $\mu$ L
Rt enzyme	0.16 $\mu$ L	0.704 $\mu$ L
RNA	30 ng	132 ng
Water	20 $\mu$ L	88 $\mu$ L

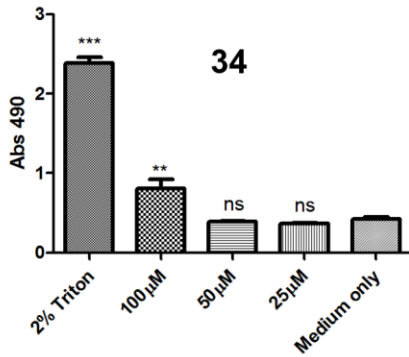
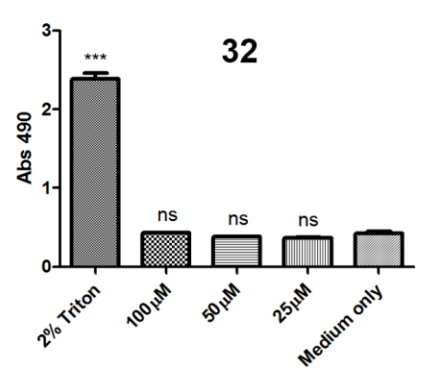
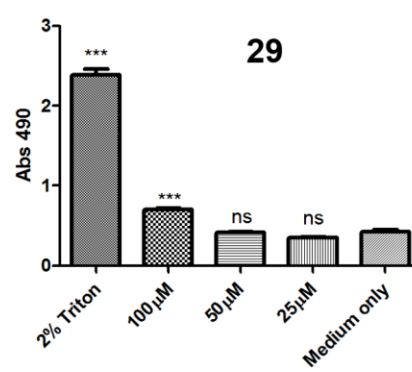
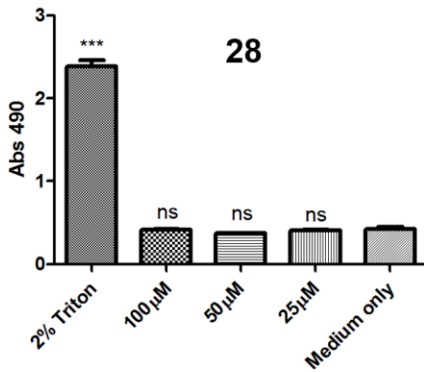
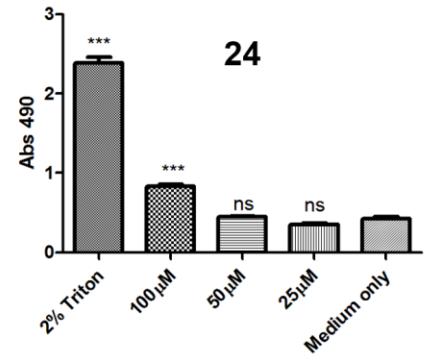
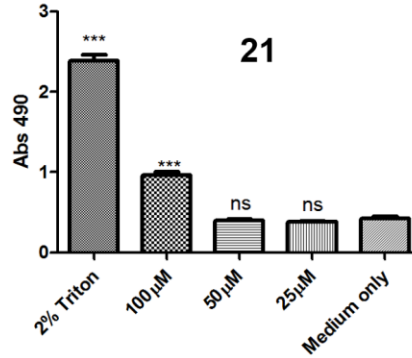
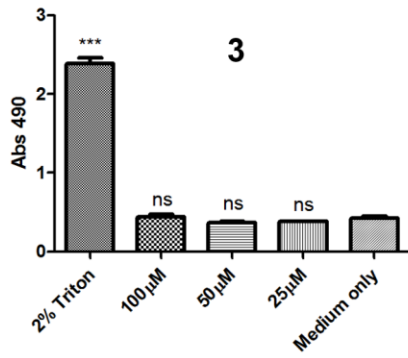
Summary of the RNA concentrations and the RIN (RNA Integrity Number) numbers obtained from the RNA samples (independent experiments) isolated from MRSA-1707 biofilms.

Sample	1 $\mu$ M		1/10 MBEC	
	[ng / $\mu$ L]	RIN	[ng / $\mu$ L]	RIN
DMSO (a)	133.8	8.6	141.467	9.0
DMSO (b)	135.64	8.3	193.387	10.0
DMSO (c)	97.64	8.3	135	8.2
<b>3</b> (a)	52.84	7.7	112.507	9.0
<b>3</b> (b)	186.52	7.9	93.467	9.0
<b>3</b> (c)	78.2	8.8	116.68	8.1
<b>29</b> (a)	231.24	8.3	40.587	9.0
<b>29</b> (b)	214.68	8.0	121.707	10.0
<b>29</b> (c)	319.08	8.5	102.76	7.4
<b>28</b> (a)	324.92	8.3	146.667	9.0
<b>28</b> (b)	260.52	8.2	149.467	9.0
<b>28</b> (c)	236.2	8.7	111.32	8.2

Data obtained from the Interdisciplinary Center for Biotechnology Research (ICBR) at the University of Florida.

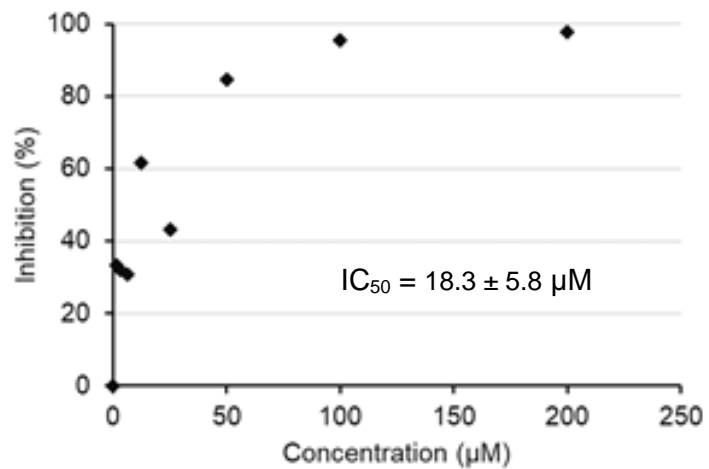
## 12.) HeLa Cytotoxicity: LDH Release Results.

HP analogues demonstrate low levels of cytotoxicity against HeLa cells at 100  $\mu$ M.



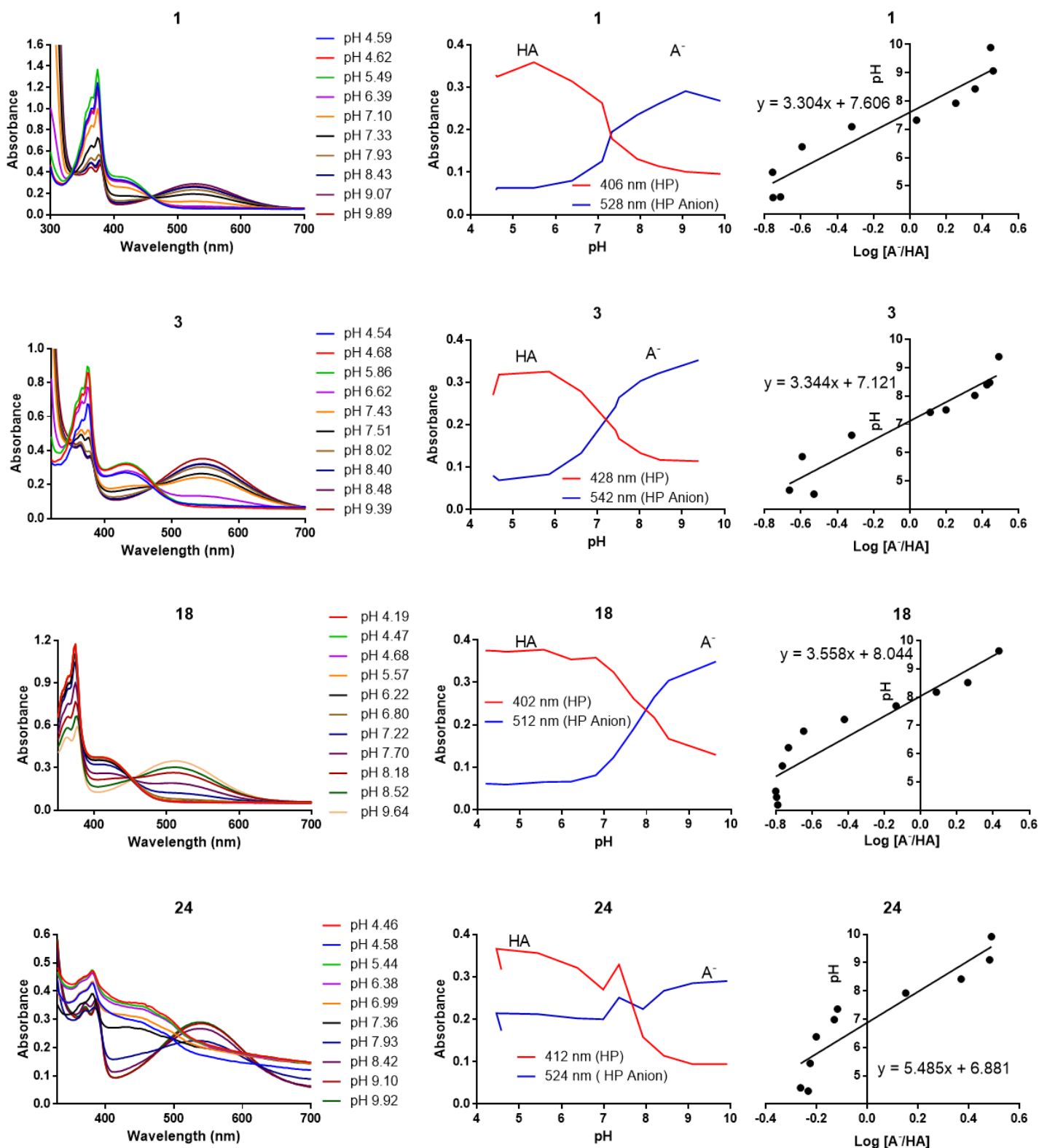
Key: Triton-X = 100% cell death; Medium Only = 0% cell death.

13.) HEK-293 Cytotoxicity of HP 29.

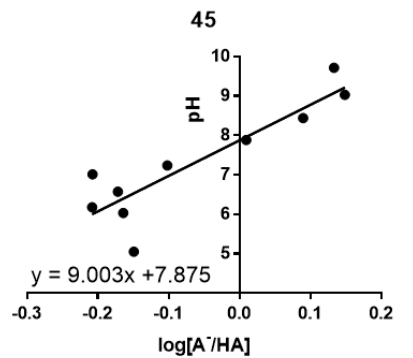
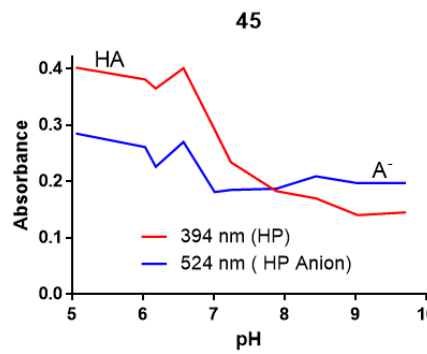
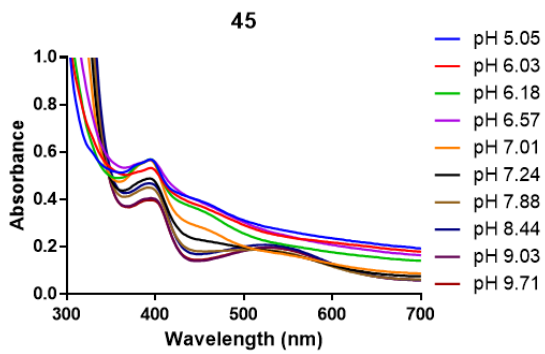
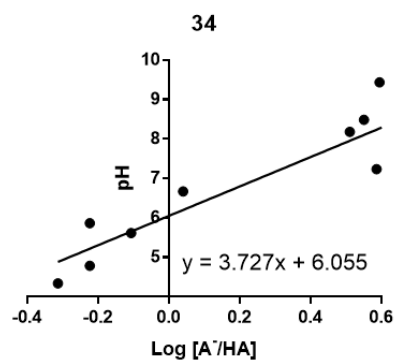
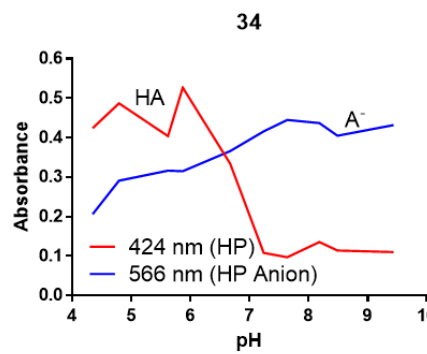
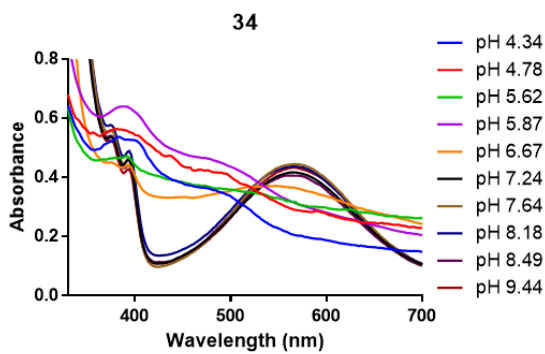
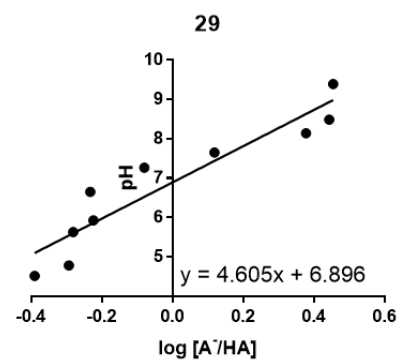
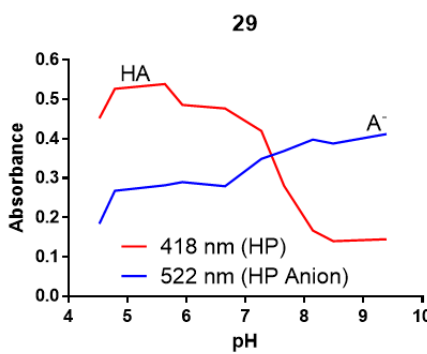
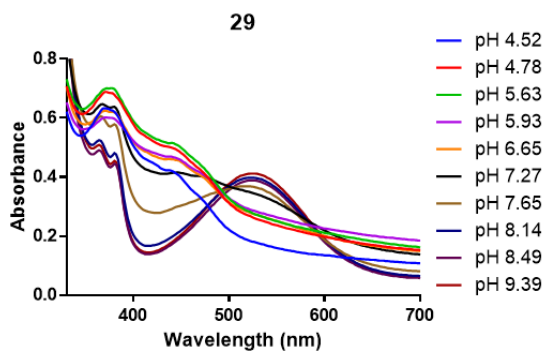
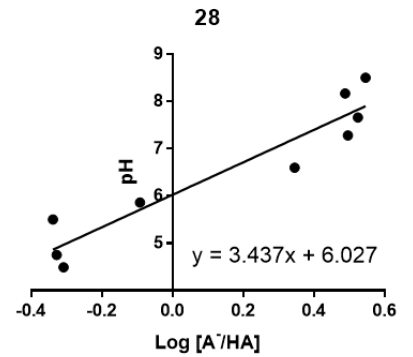
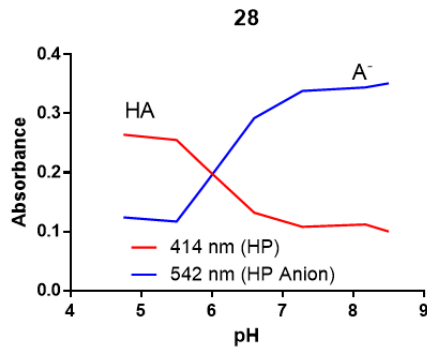
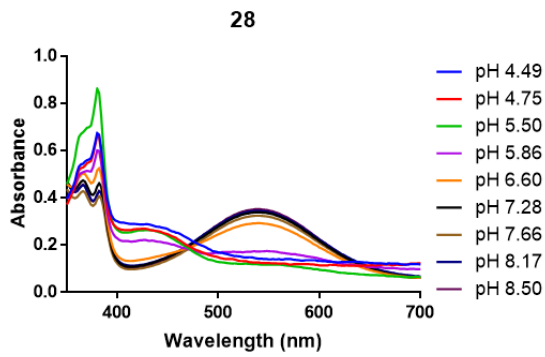


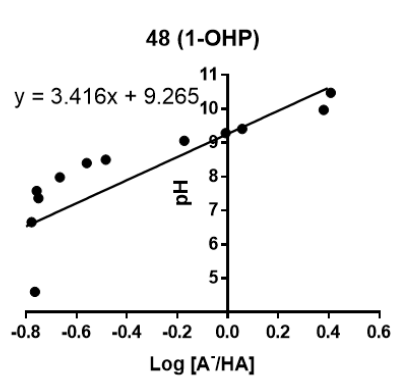
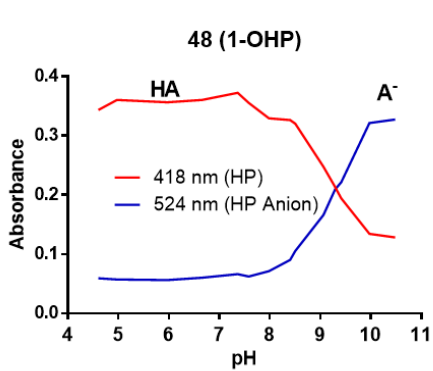
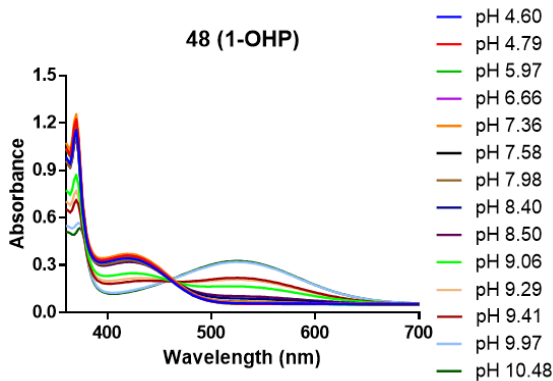
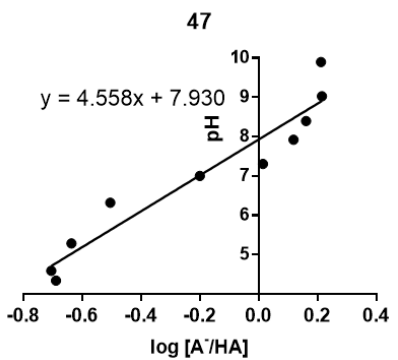
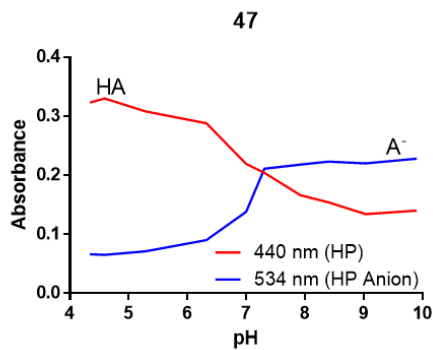
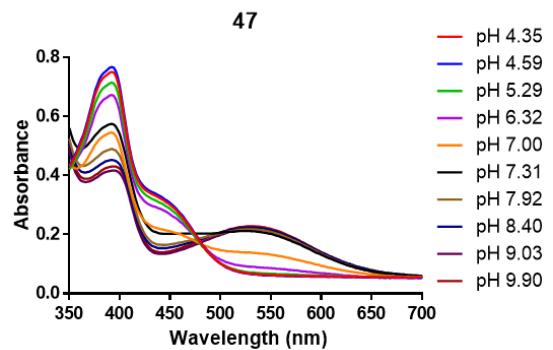
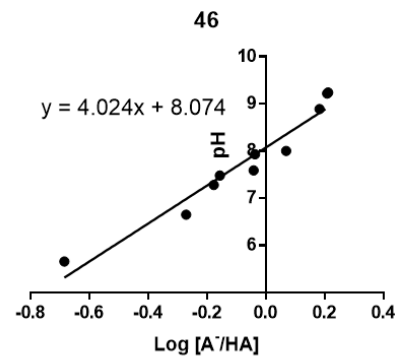
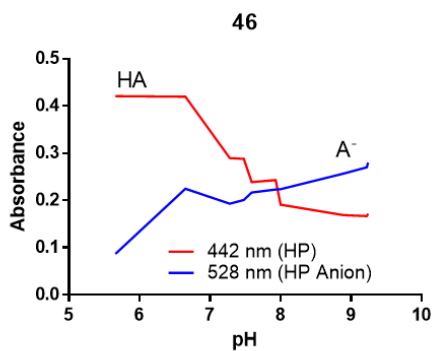
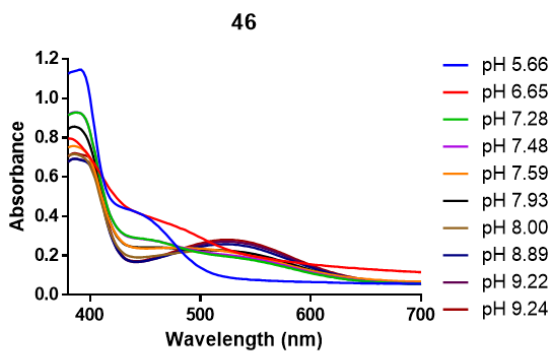
## 14.) pKa Value Determination for Select HPs.

pH-Dependent spectral scan of HPs in buffer; normalized plot of absorbance vs. pH for protonated (HA) and deprotonated (A<sup>-</sup>) HPs and plot of pH versus log[A<sup>-</sup>/HA] for HPs. A linear regression line is shown with a Y-intercept corresponding to a calculated pKa value.



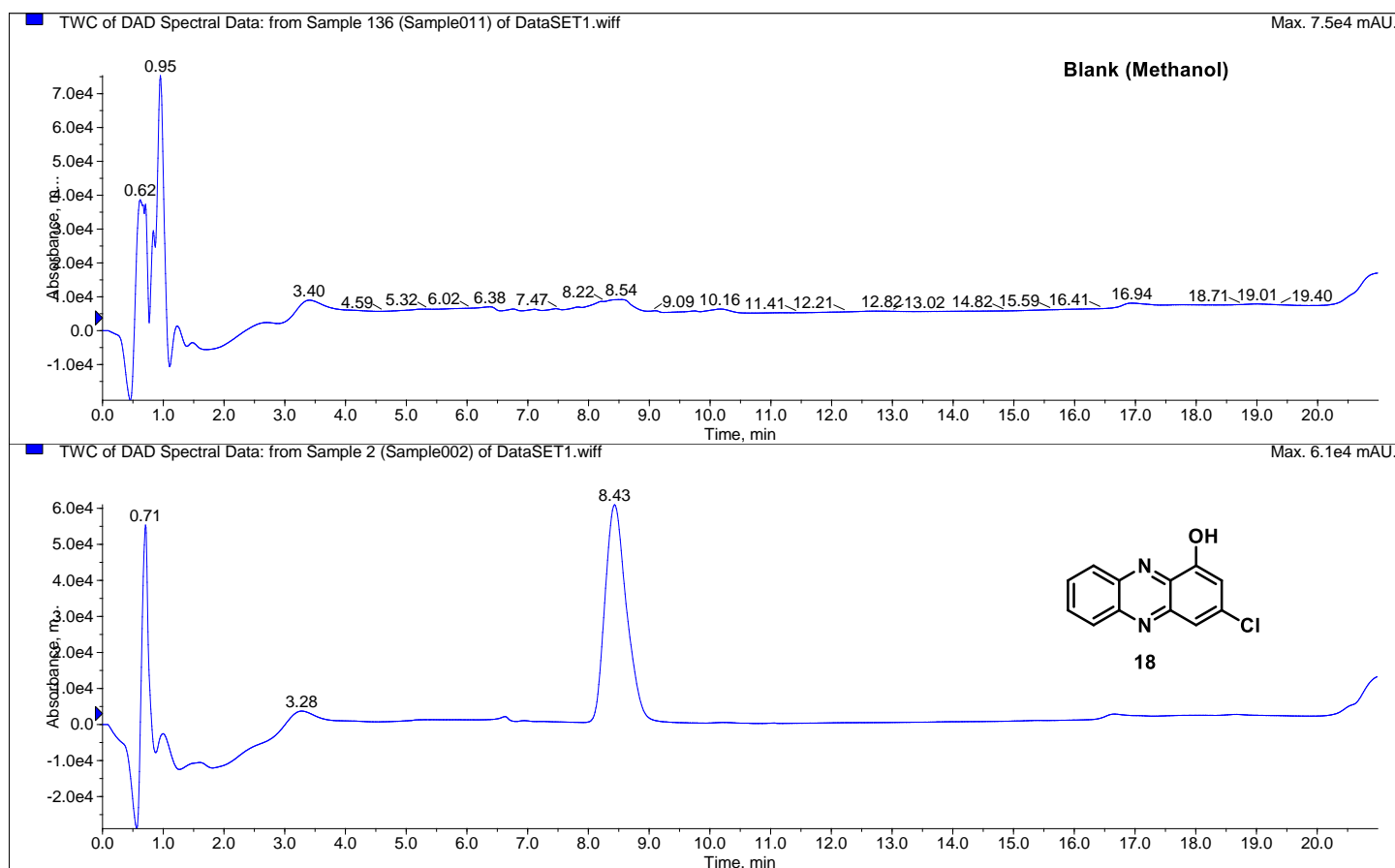




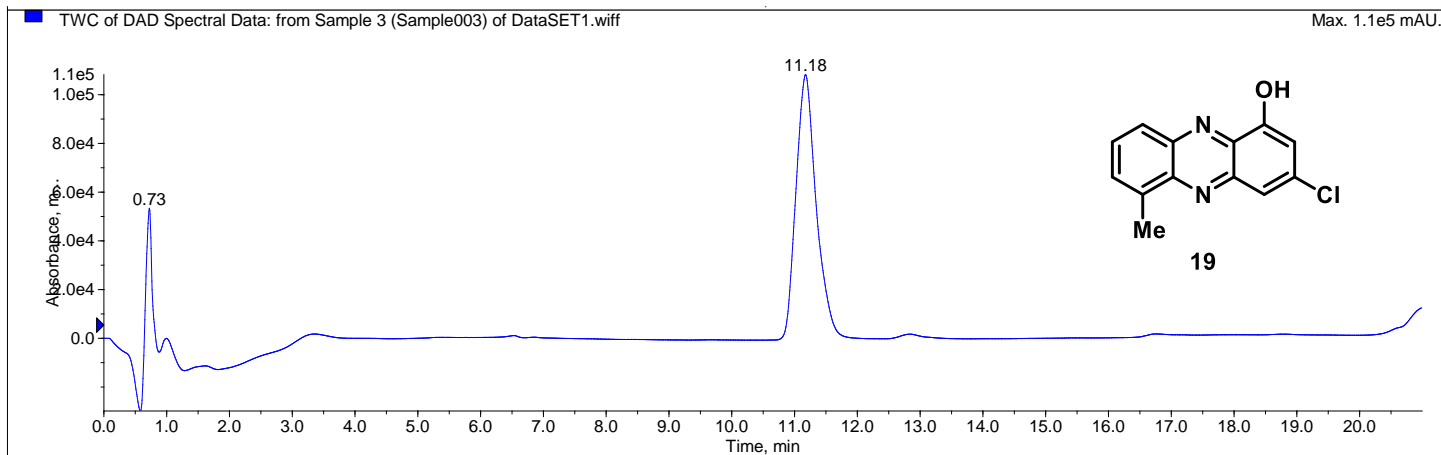


## 15.) Compound Purity Analysis.

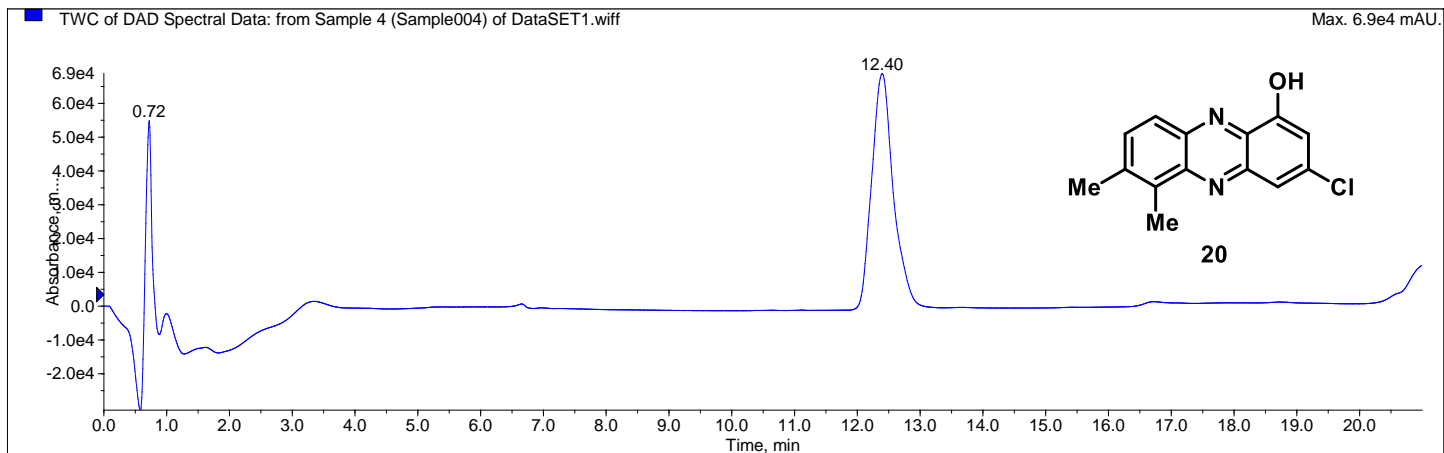
Each HPLC experiment to determine the purity of halogenated phenazine analogues was carried out over 20 minutes. The purity of each compound was determined following the characterization via  $^1\text{H}/^{13}\text{C}$  NMR, HRMS and melting point analysis. A blank methanol run is shown below and used to remove background noise from the purity assessment of each compound. Overall, the purity range from these experiments was found to be 95.5 to >99.9% pure based on LC traces.



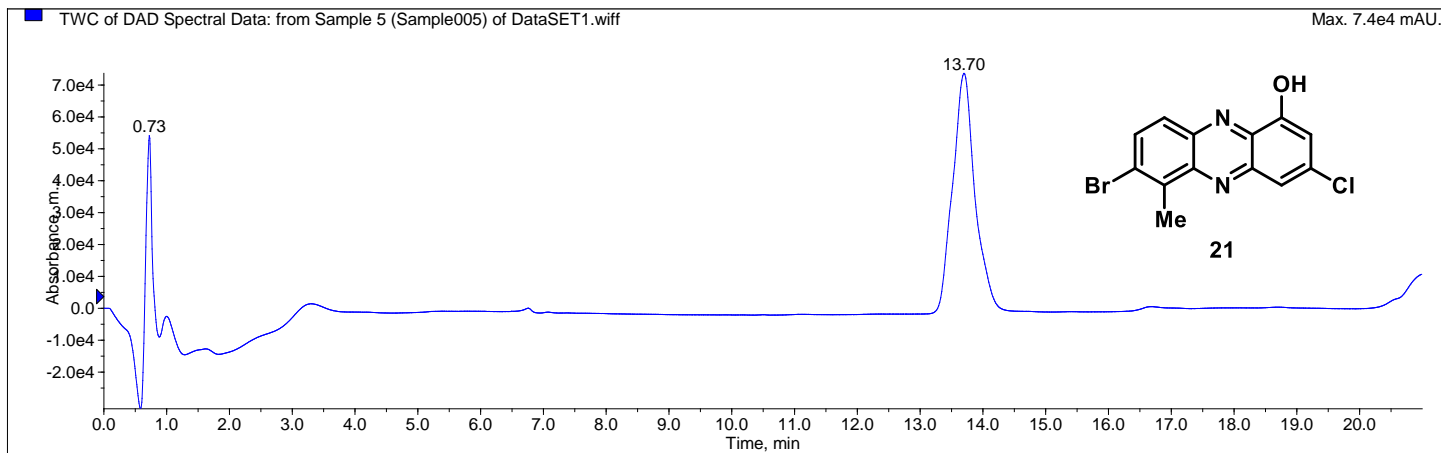
Retention Time (Min)	Peak Area
6.63	17500
6.95	2034
8.43 (18)	1480000
<b>Sum of Area</b>	<b>1499534</b>
<b>% Purity</b>	<b>98.7%</b>



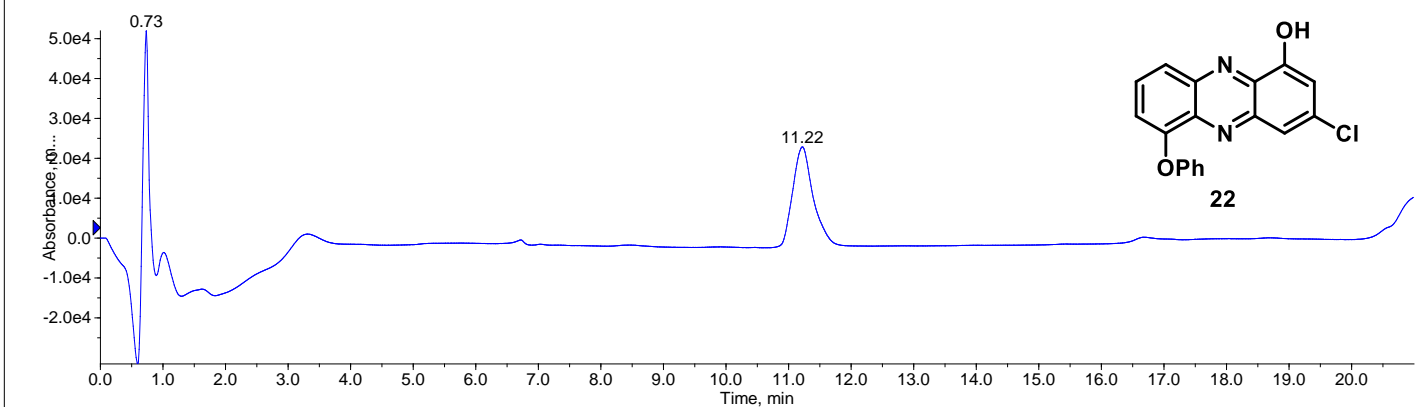
Retention Time (Min)	Peak Area
6.53	11800
11.18 (19)	2440000
12.84	44300
<b>Sum of Area</b>	<b>2496100</b>
<b>% Purity</b>	<b>97.8%</b>



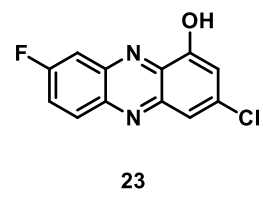
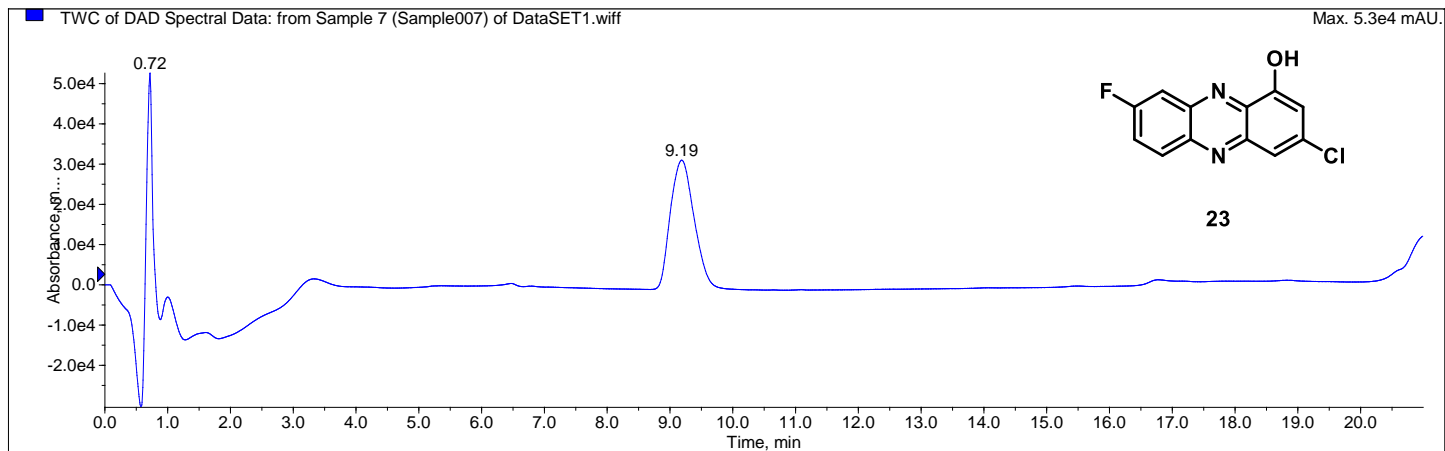
Retention Time (Min)	Peak Area
6.65	15800
12.40 ( <b>20</b> )	1670000
<b>Sum of Area</b>	1685800
<b>% Purity</b>	<b>99.1%</b>



Retention Time (Min)	Peak Area
6.76	13800
13.70 (21)	1850000
<b>Sum of Area</b>	<b>1863800</b>
<b>% Purity</b>	<b>99.3%</b>

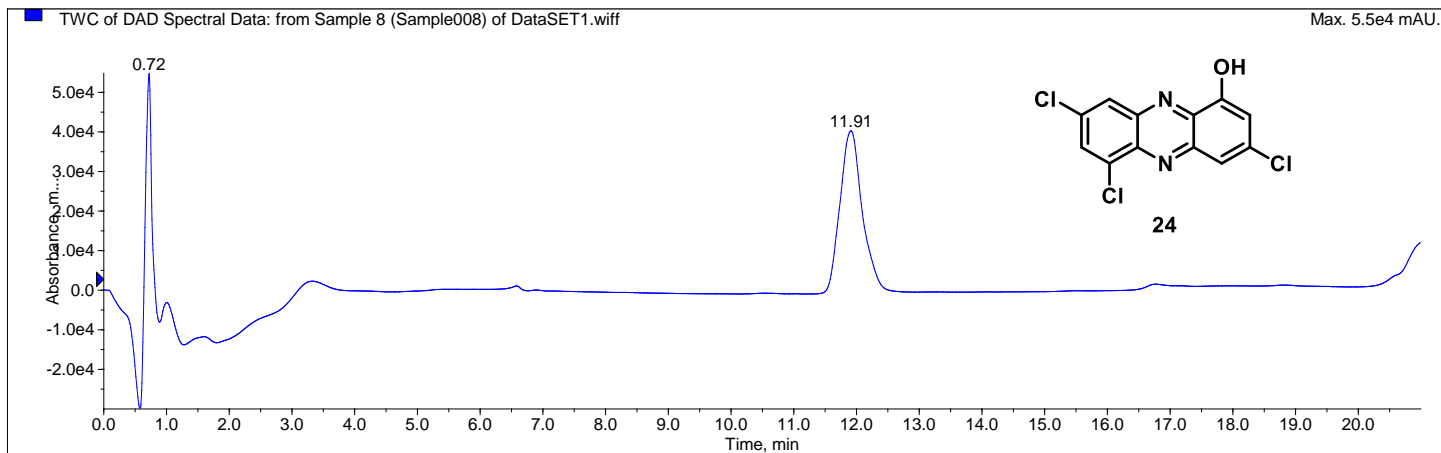


Retention Time (Min)	Peak Area
6.71	13200
7.03	1806
11.22 (22)	571000
<b>Sum of Area</b>	<b>586006</b>
<b>% Purity</b>	<b>97.4%</b>

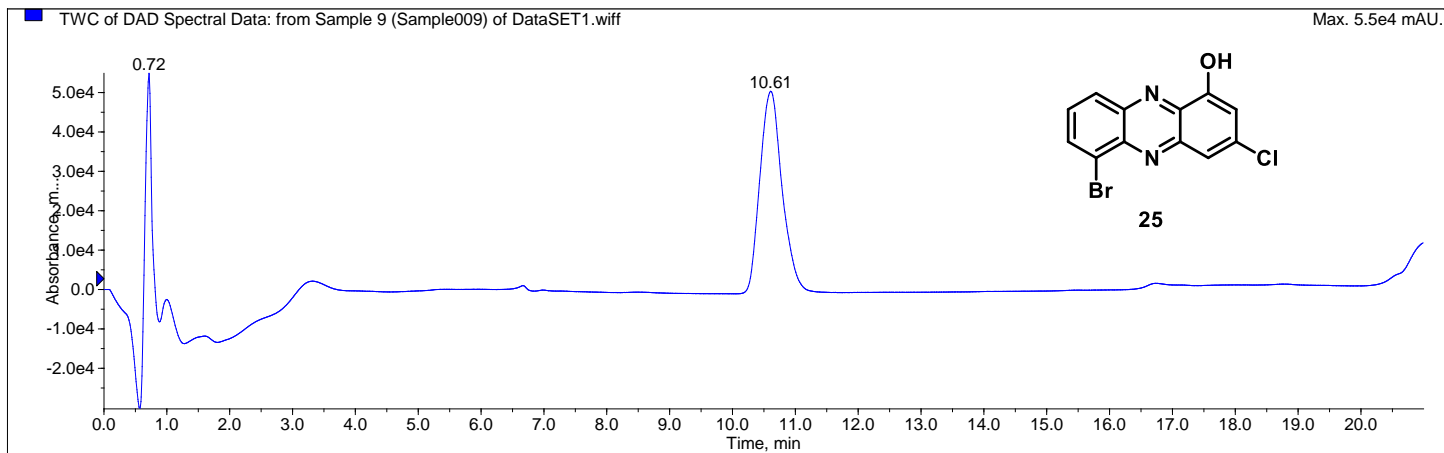


Retention Time (Min)	Peak Area
9.19 (23)	842000
10.31	147
<b>Sum of Area</b>	<b>842147</b>
<b>% Purity</b>	<b>&gt;99.9%</b>

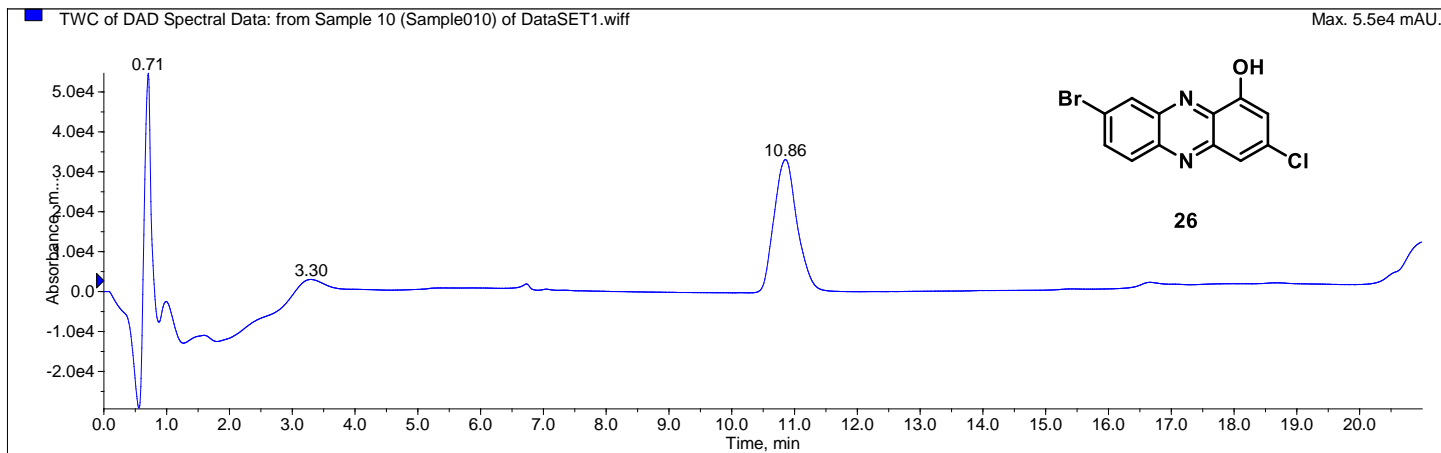




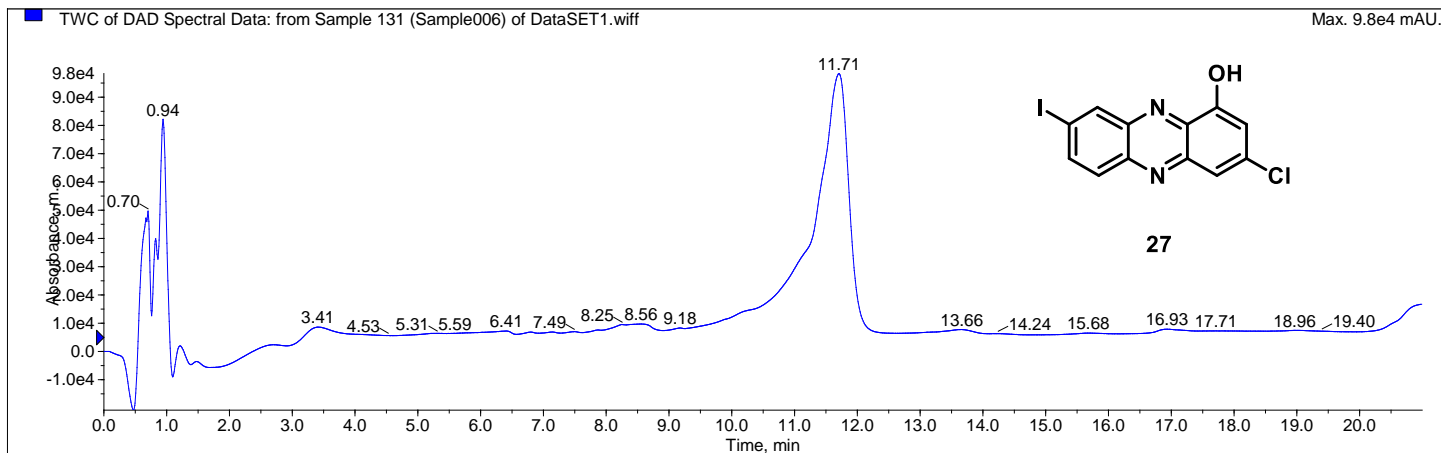
Retention Time (Min)	Peak Area
6.58	17000
6.90	1888
10.62	5083
11.91 (24)	1010000
15.52	367
<b>Sum of Area</b>	<b>1034338</b>
<b>% Purity</b>	<b>97.6%</b>



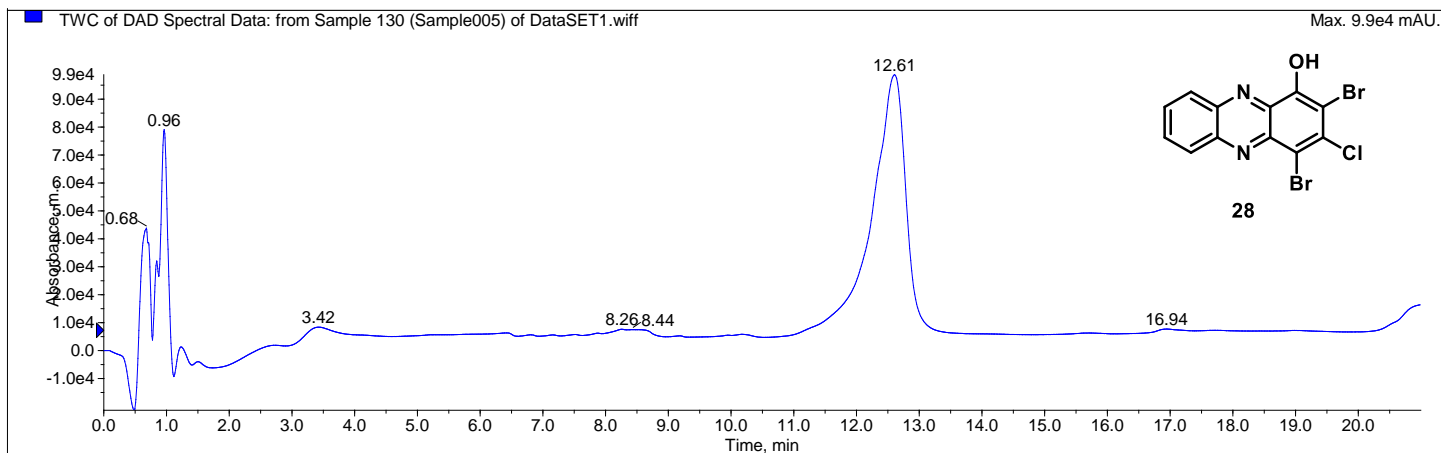
Retention Time (Min)	Peak Area
6.67	20400
6.99	2430
10.61 (25)	1250000
17.98	256
<b>Sum of Area</b>	<b>1273086</b>
<b>% Purity</b>	<b>98.2%</b>



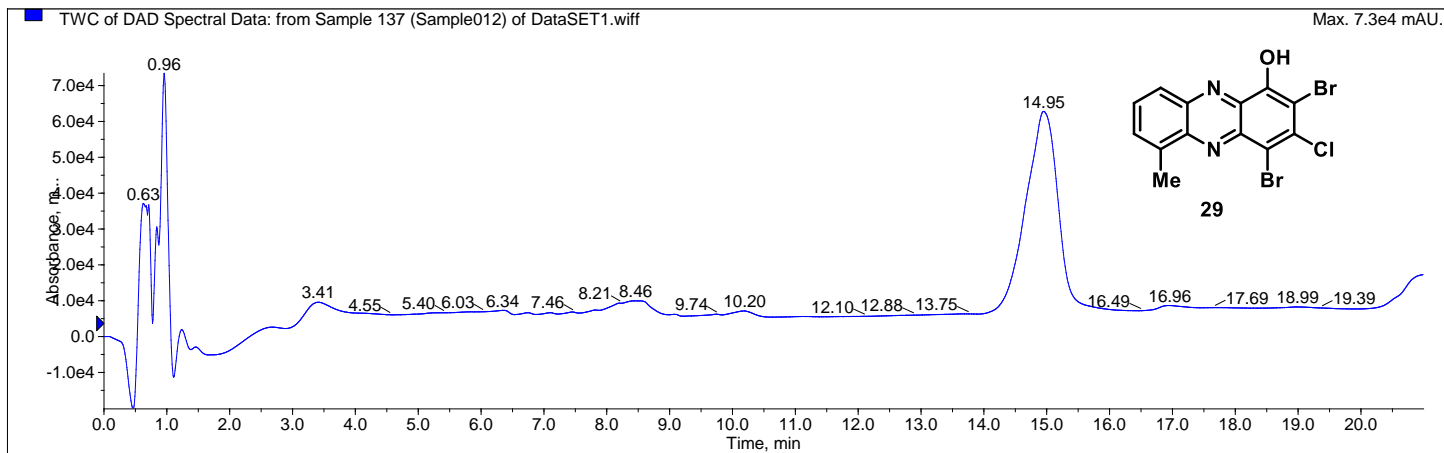
Retention Time (Min)	Peak Area
6.73	14000
7.04	2179
10.86 ( <b>26</b> )	847000
<b>Sum of Area</b>	863179
<b>% Purity</b>	<b>98.1%</b>



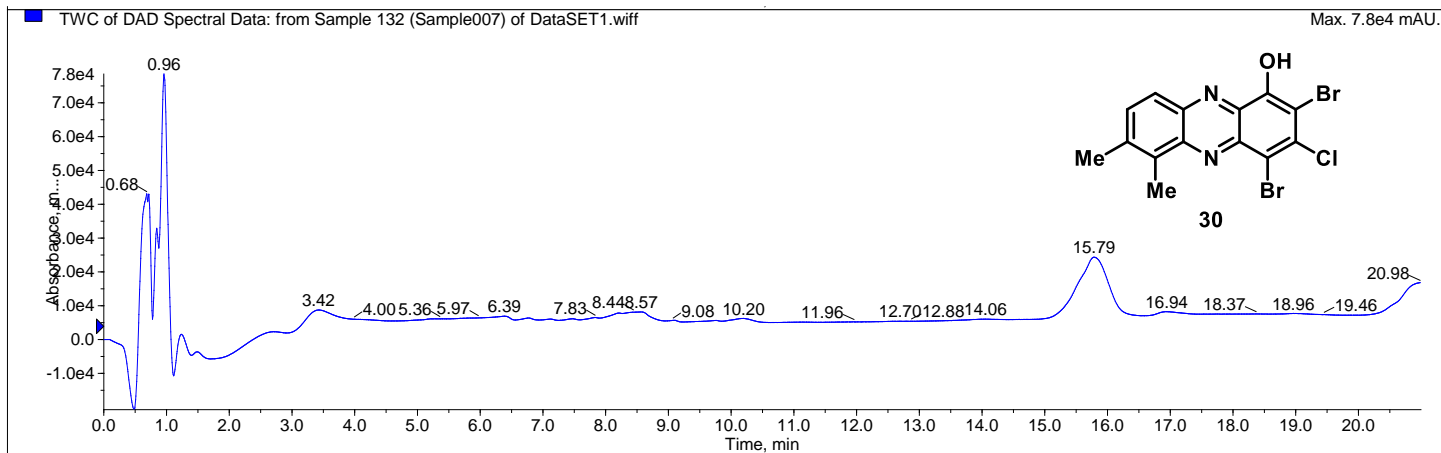
Retention Time (Min)	Peak Area
6.41	32600
6.80	4841
9.18	3346
11.71 ( <b>27</b> )	3800000
13.67	40300
<b>Sum of Area</b>	<b>3881087</b>
<b>% Purity</b>	<b>97.9%</b>



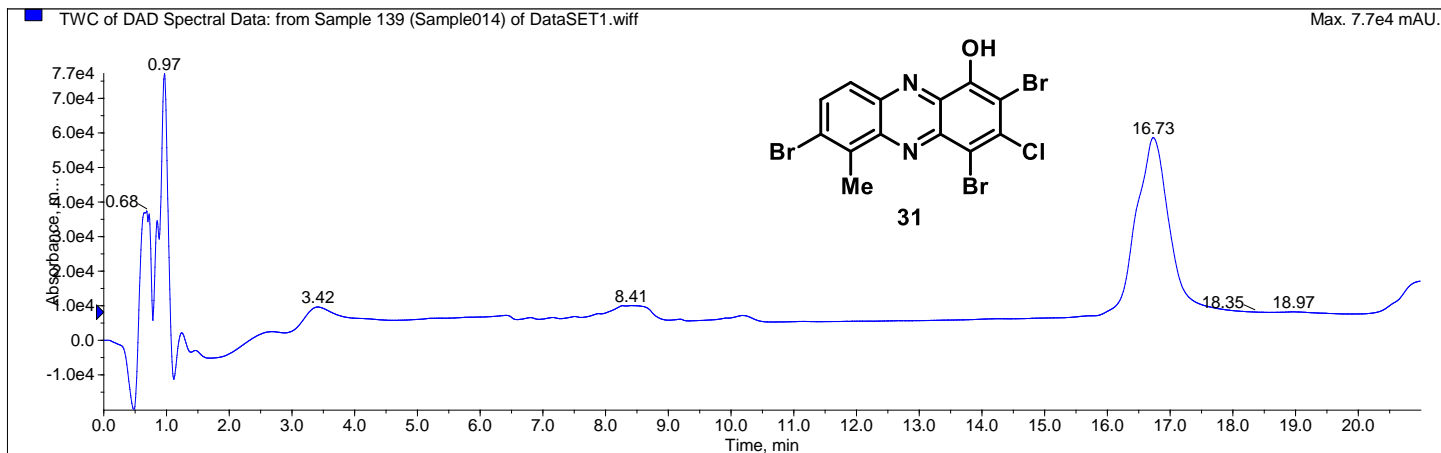
Retention Time (Min)	Peak Area
12.61( <b>28</b> )	3530000
15.67	6526
17.72	10003
<b>Sum of Area</b>	<b>3546529</b>
<b>% Purity</b>	<b>99.5%</b>



Retention Time (Min)	Peak Area
6.28	24300
11.19	337
14.95 (29)	1960000
<b>Sum of Area</b>	<b>1984637</b>
<b>% Purity</b>	<b>98.8%</b>

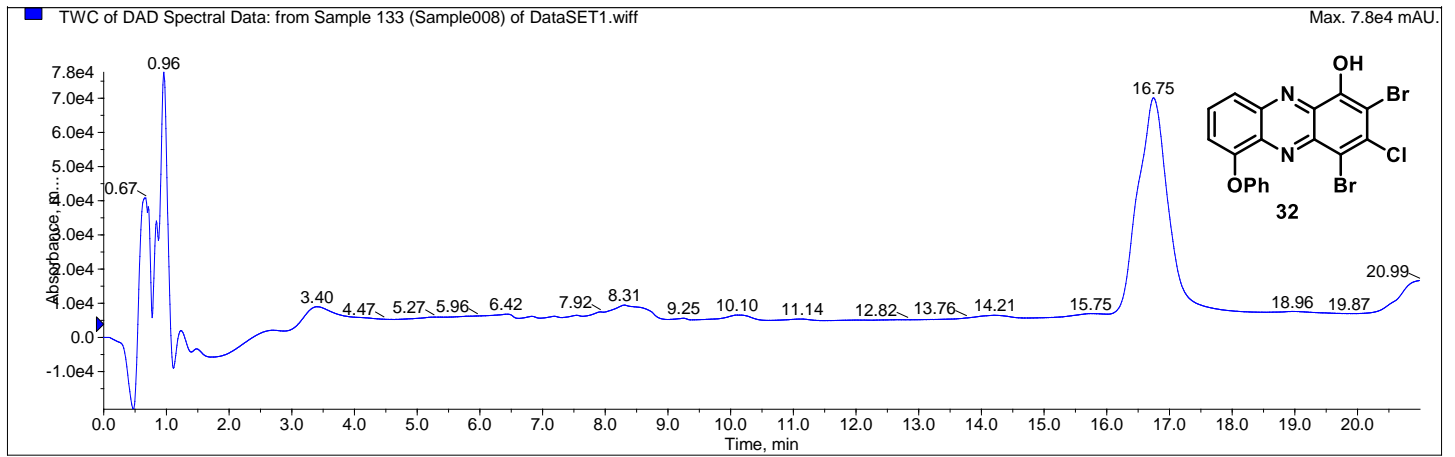


Retention Time (Min)	Peak Area
6.39	22700
6.76	5874
15.79 (30)	601000
<b>Sum of Area</b>	<b>629574</b>
<b>% Purity</b>	<b>95.5%</b>

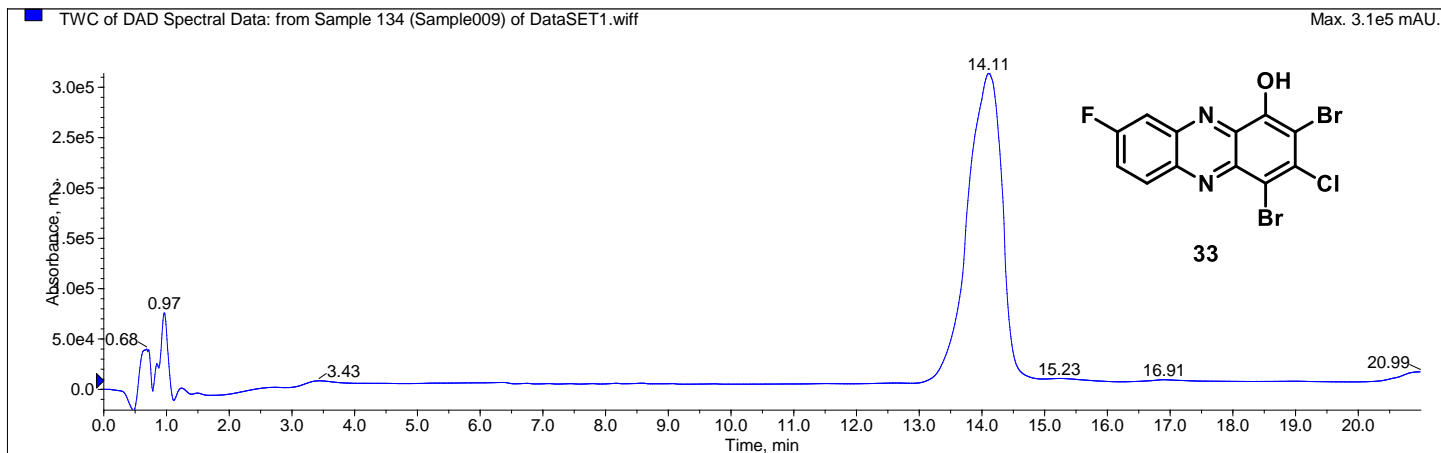


Retention Time (Min)	Peak Area
12.24	727
14.25	488
16.73 (31)	1810000
18.35	6
<b>Sum of Area</b>	<b>1811221</b>
<b>% Purity</b>	<b>&gt;99.9%</b>

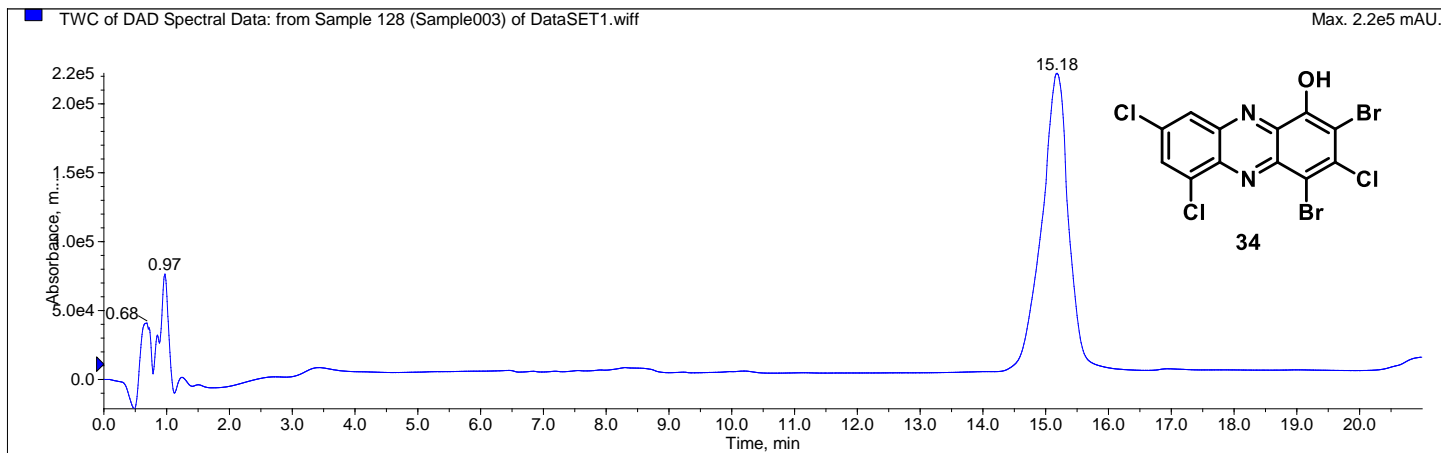




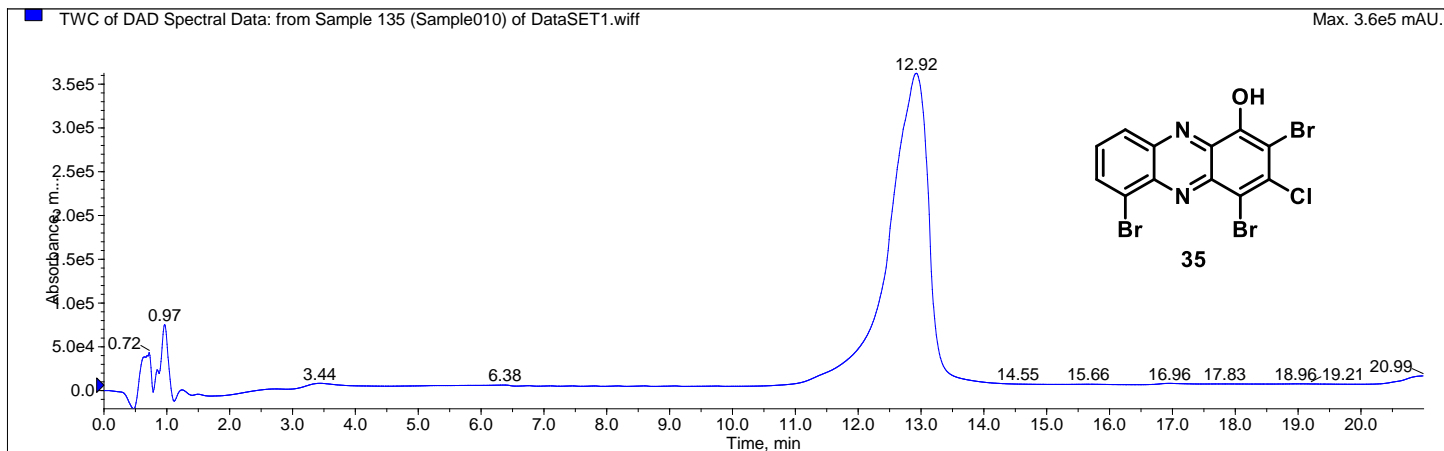
Retention Time (Min)	Peak Area
11.14	8699
13.76	499
14.21	27200
15.75	2104
16.75 (32)	2030000
<b>Sum of Area</b>	<b>2068502</b>
<b>% Purity</b>	<b>98.1%</b>



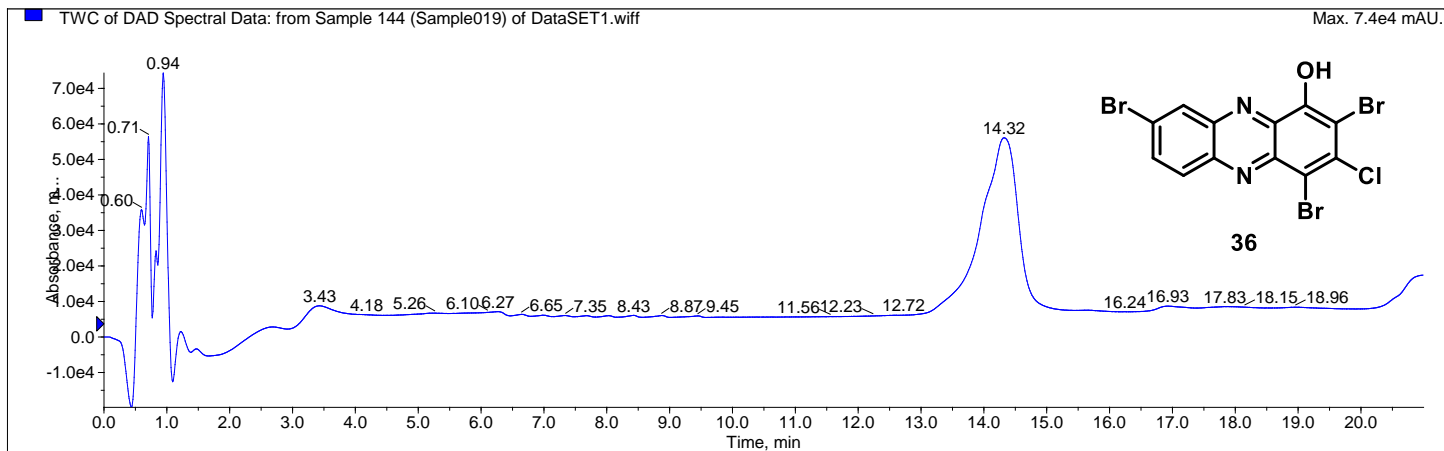
Retention Time (Min)	Peak Area
11.51	4725
12.64	4539
14.11 ( <b>33</b> )	11600000
15.23	32900
<b>Sum of Area</b>	11642164
<b>% Purity</b>	<b>99.6%</b>



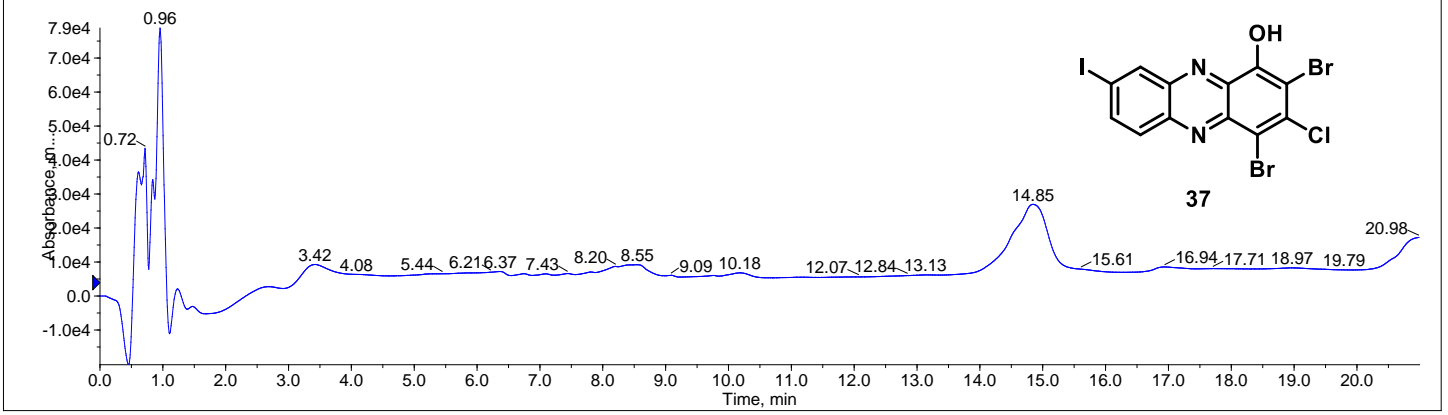
Retention Time (Min)	Peak Area
11.27	5207
15.18 (34)	6400000
17.73	2988
<b>Sum of Area</b>	<b>6408195</b>
<b>% Purity</b>	<b>99.9%</b>



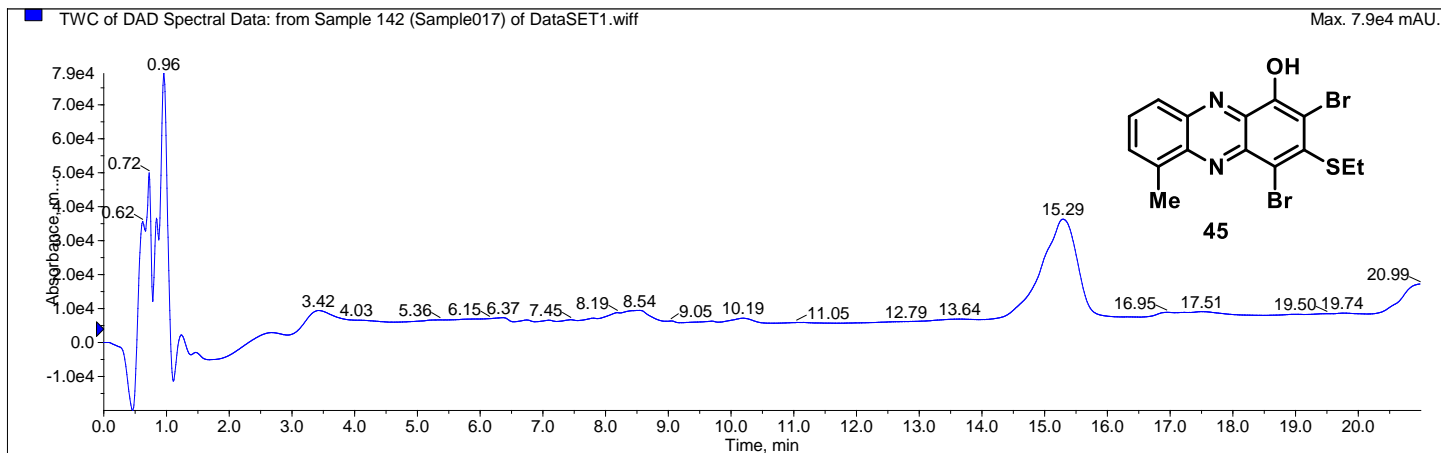
Retention Time (Min)	Peak Area
12.92 (35)	15700000
15.66	3507
17.66	5228
<b>Sum of Area</b>	<b>15708735</b>
<b>% Purity</b>	<b>&gt;99.9%</b>



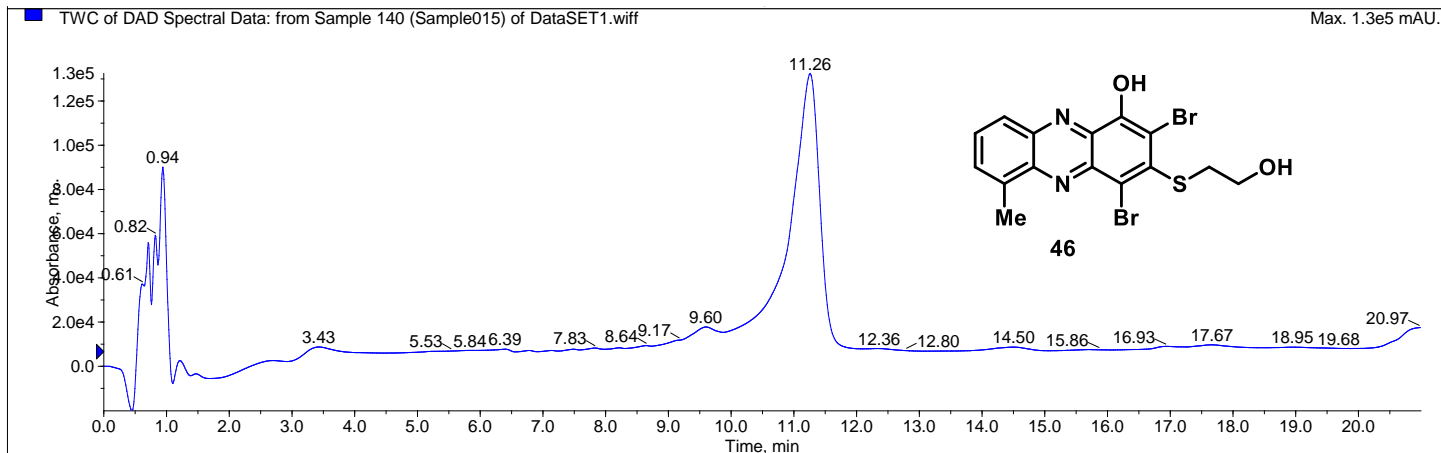
Retention Time (Min)	Peak Area
8.87	5904
9.45	5161
14.32 ( <b>36</b> )	1990000
17.87	11086
<b>Sum of Area</b>	2012151
<b>% Purity</b>	<b>98.9%</b>



Retention Time (Min)	Peak Area
11.18	1420
13.13	825
14.85 (37)	760000
<b>Sum of Area</b>	<b>762245</b>
<b>% Purity</b>	<b>99.7%</b>

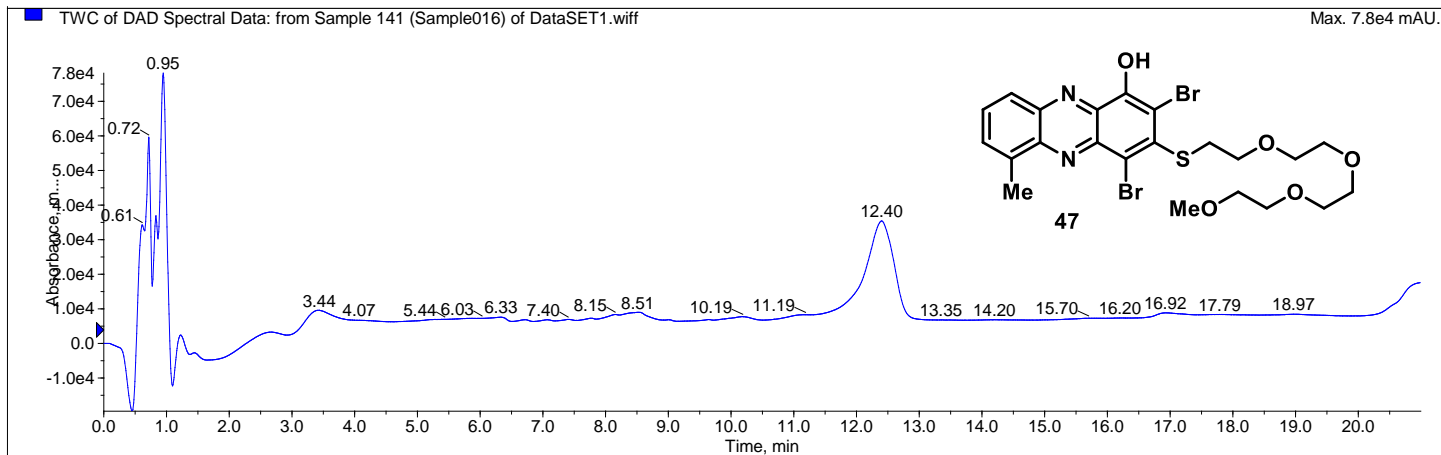


Retention Time (Min)	Peak Area
11.15	4579
13.64	3454
15.29 (45)	1140000
17.51	17500
<b>Sum of Area</b>	<b>1165533</b>
<b>% Purity</b>	<b>97.8%</b>



Retention Time (Min)	Peak Area
9.17	1184
9.60	76600
11.26 (46)	3860000
14.50	51900
17.67	45300
<b>Sum of Area</b>	<b>4034984</b>
<b>% Purity</b>	<b>95.7%</b>



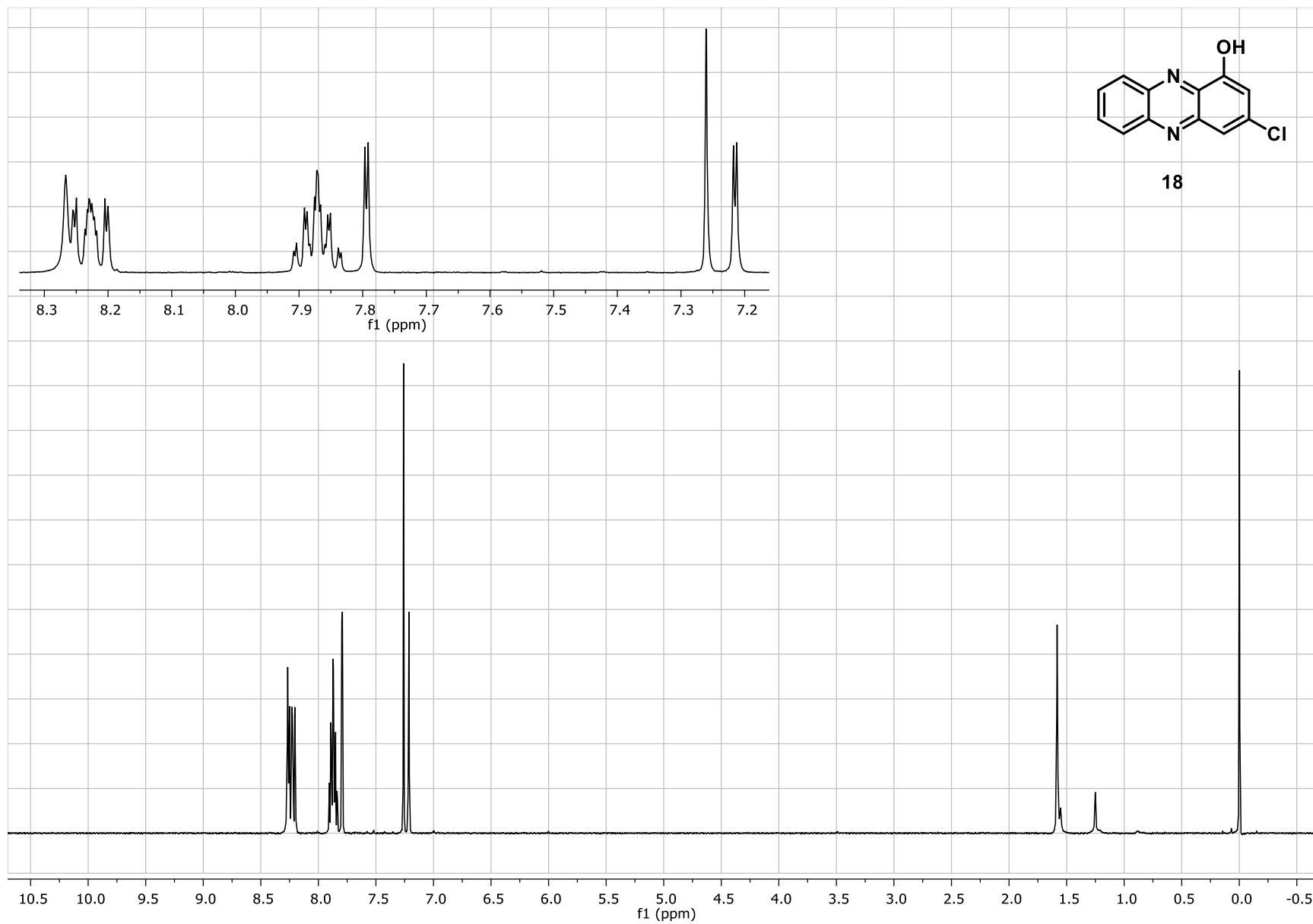


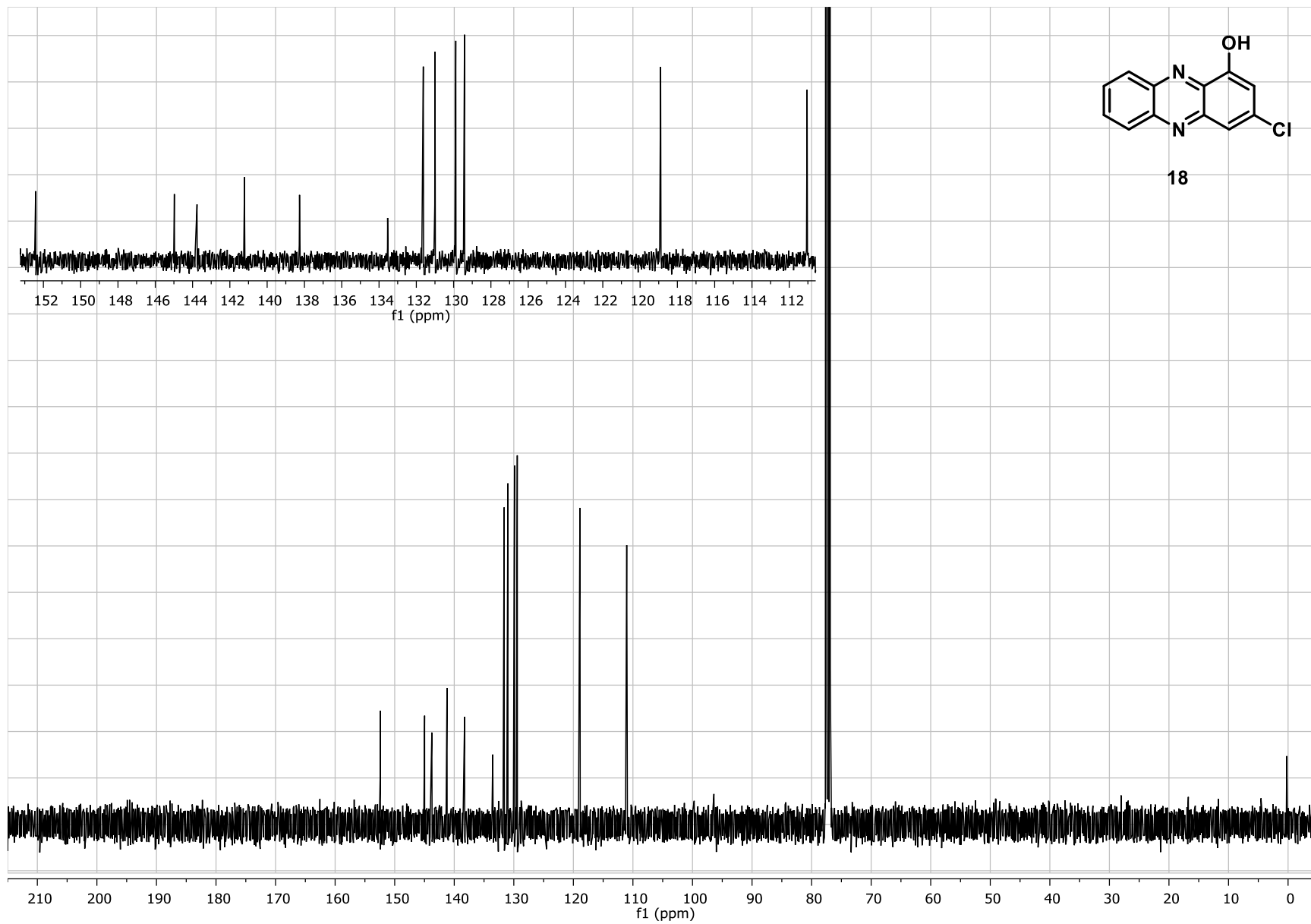
Retention Time (Min)	Peak Area
11.19	176
12.40 (47)	887000
13.35	67
<b>Sum of Area</b>	<b>887243</b>
<b>% Purity</b>	<b>&gt;99.9%</b>

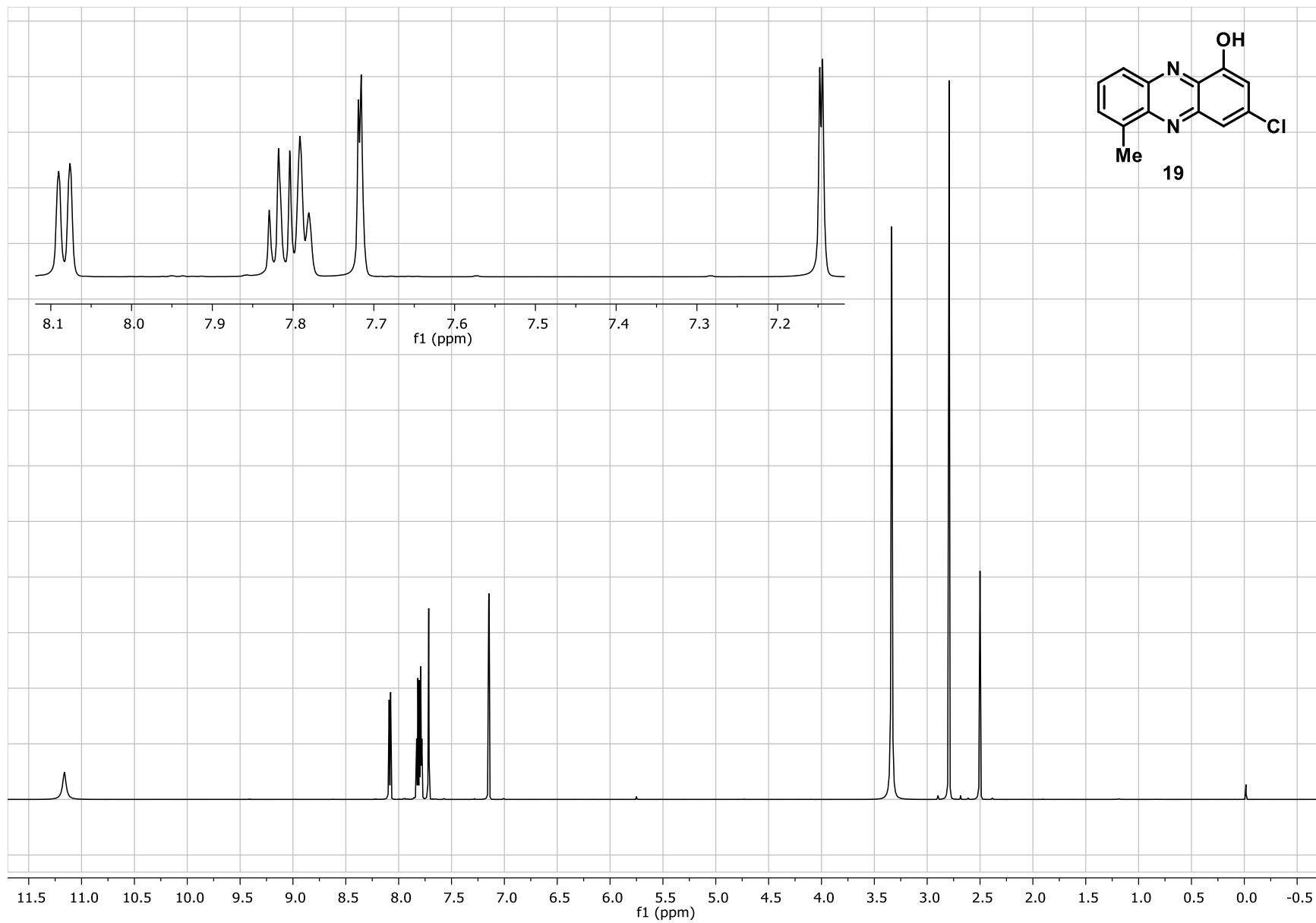
## 16.) Literature References.

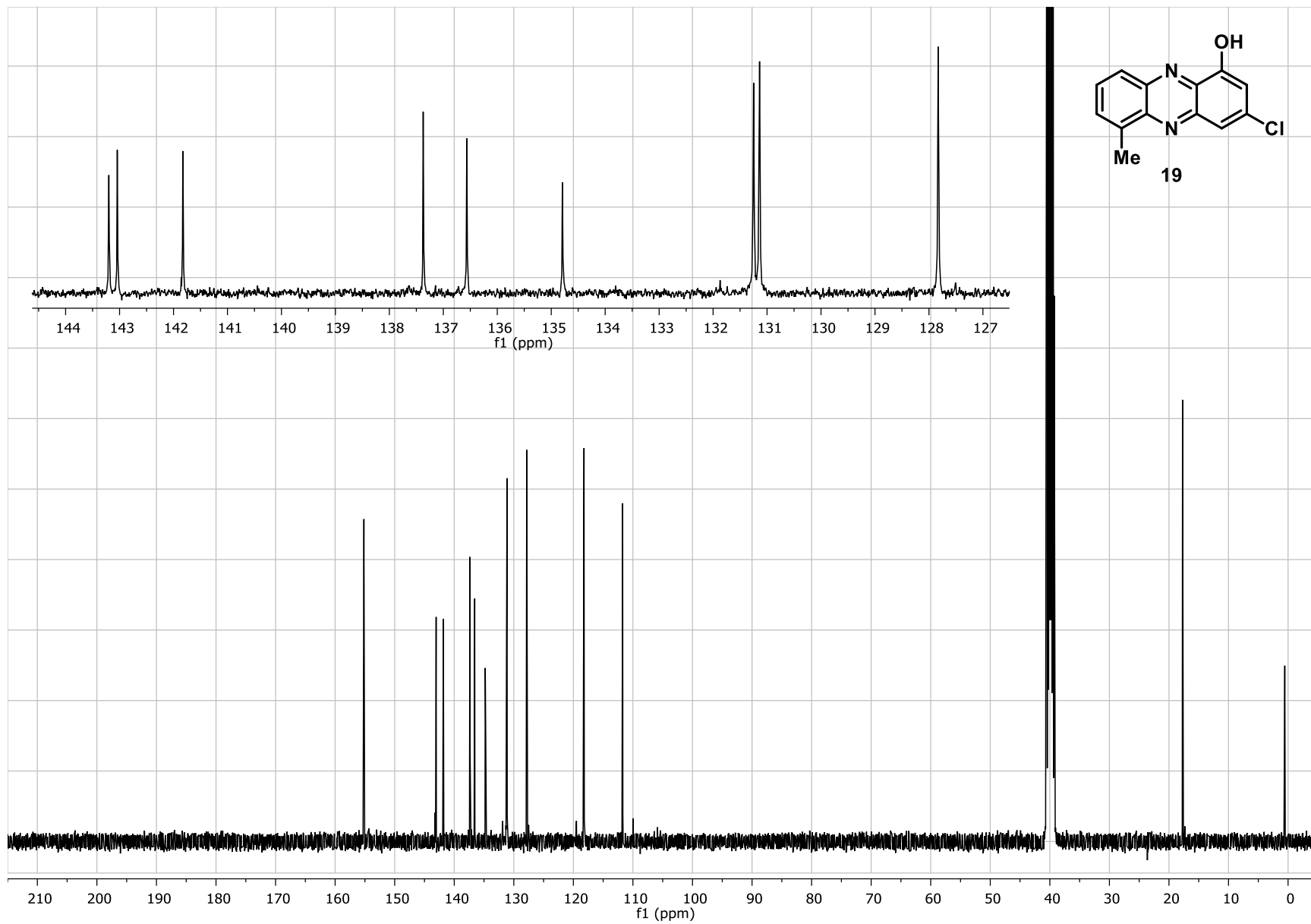
1. Sachin, K.; Jadhav, V.H.; Kim, E.M.; Kim, H.L.; Lee, S.B.; Jeong, H.J.; Lim, S.T.; Sohn, M.H.; Kim, D.W. *Bioconjug. Chem.* **2012**, *23*, 1680-1686.
2. Snow, A.W.; Foos, E.E. *Synthesis* **2003**, *14*, 0509-0512.
3. Garrison, A.T.; Abouelhassan, Y.; Norwood IV, V.M.; Kallifidas, D.; Bai, F.; Nguyen, M.; Rolfe, M.; Burch, G.M.; Jin, S.; Luesch, H.; Huigens III, R.W. *J. Med. Chem.* **2016**, *59*, 3808-3825.
4. Yang, H.; Abouelhassan, Y.; Burch, G.M.; Kallifidas, D.; Huang, G.; Yousaf, H.; Jin, S.; Luesch, H.; Huigens III, R.W. *Sci. Rep.* **2017**, *7*, 2003.
5. Garrison, A.T.; Abouelhassan, Y.; Kallifidas, D.; Tan, H.; Kim, Y.S.; Jin, S.; Luesch, H.; Huigens III, R.W. *J. Med. Chem.* **2018**, *61*, 3962-3983.

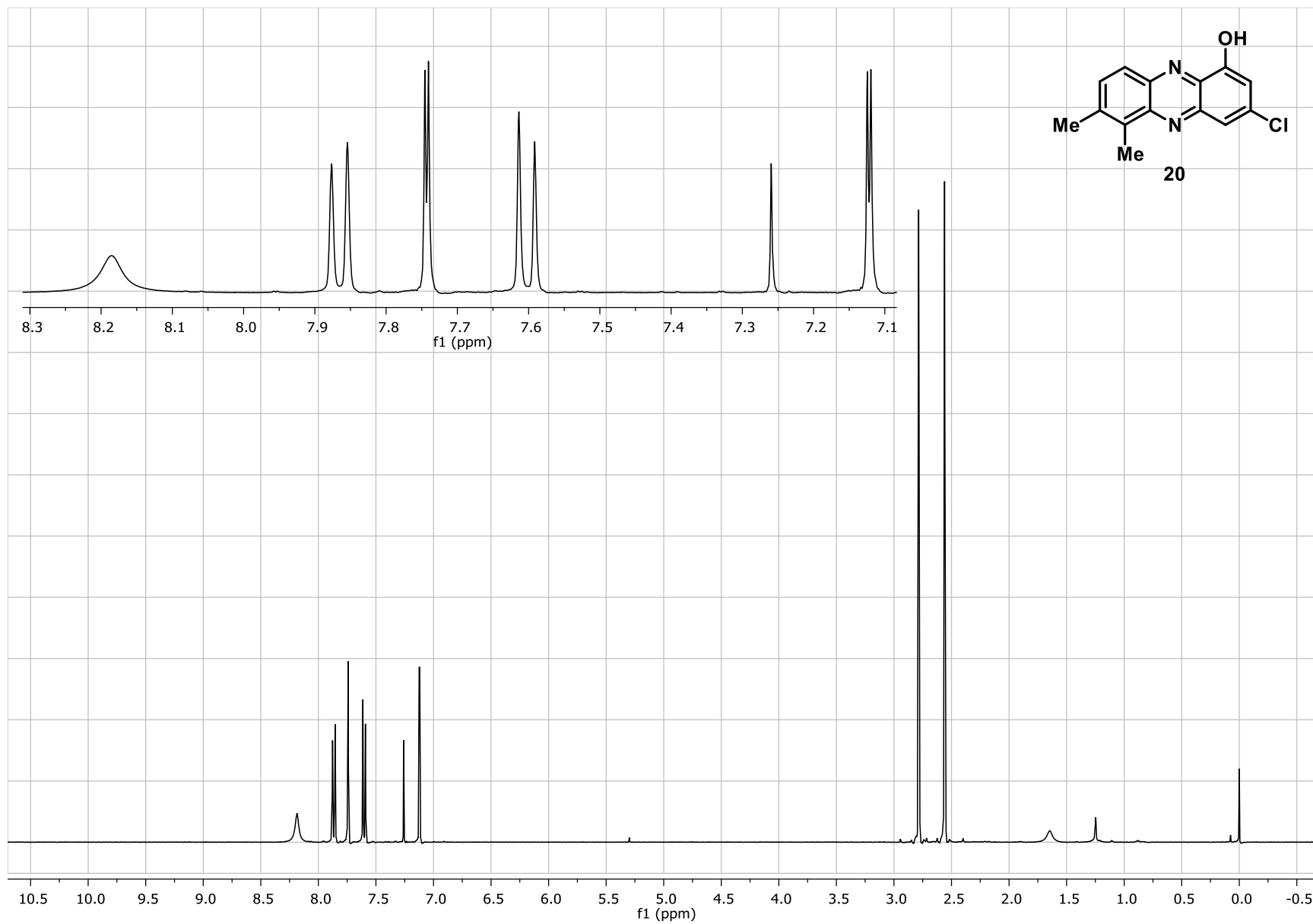
## 17. NMR Spectra.



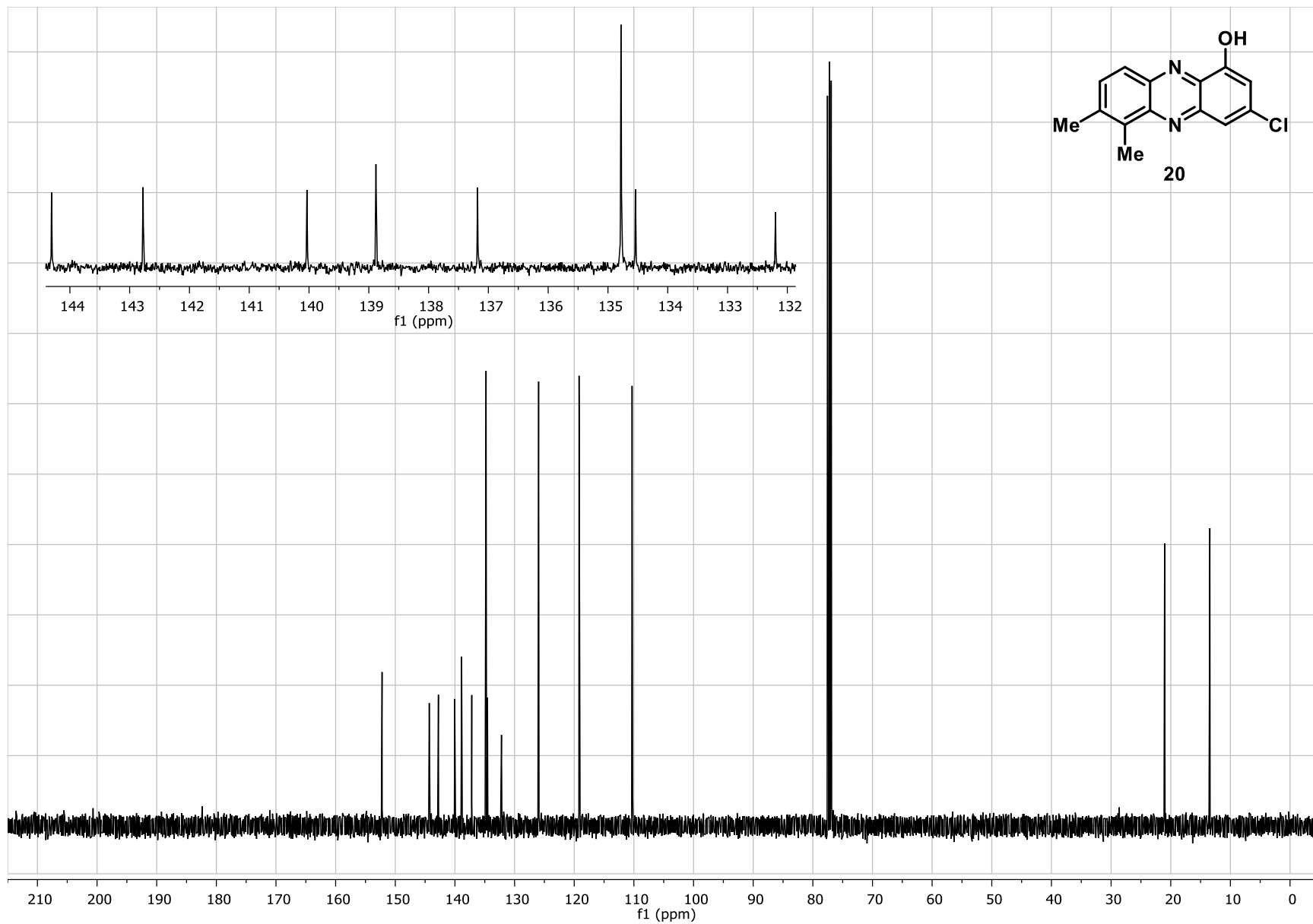


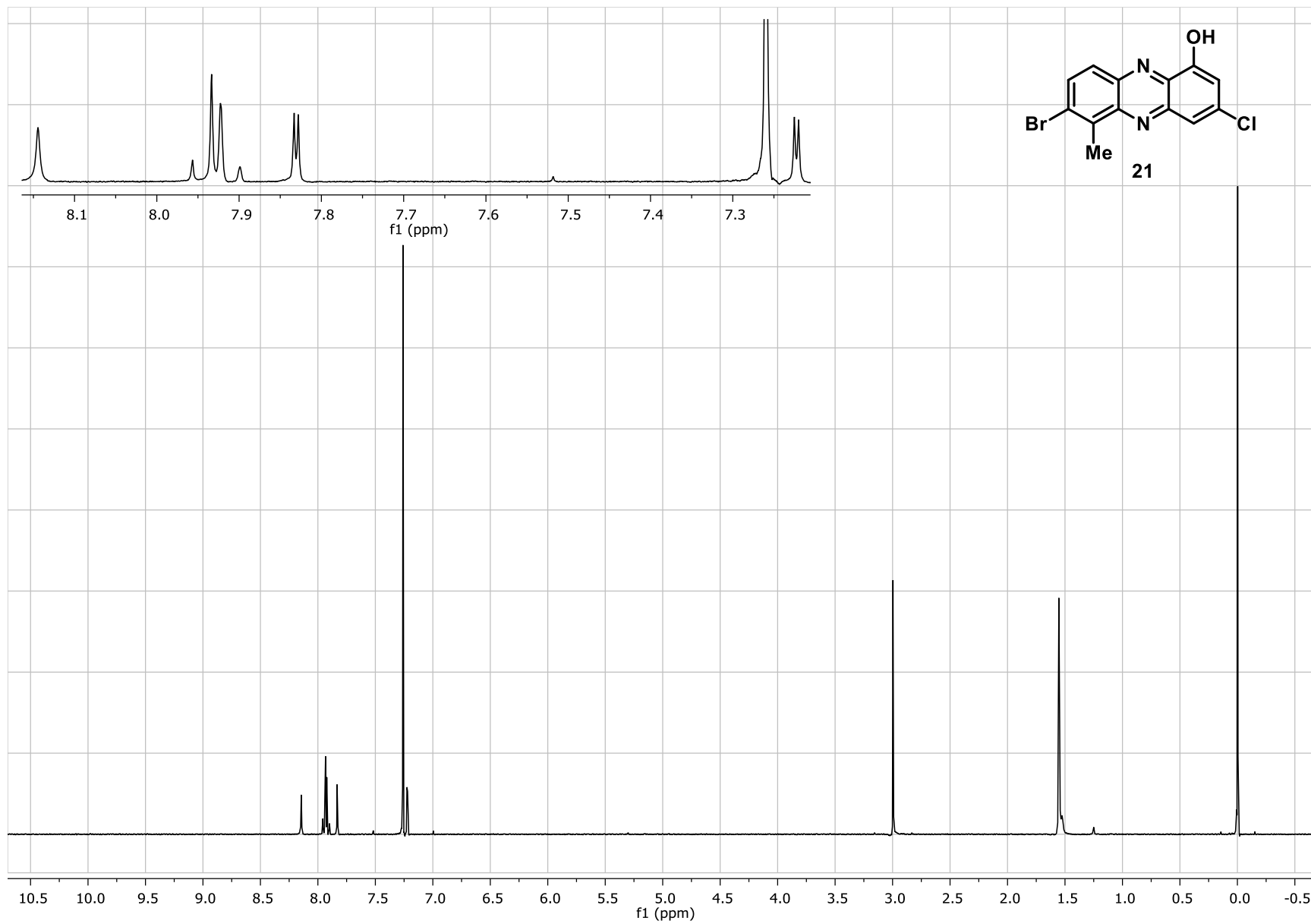


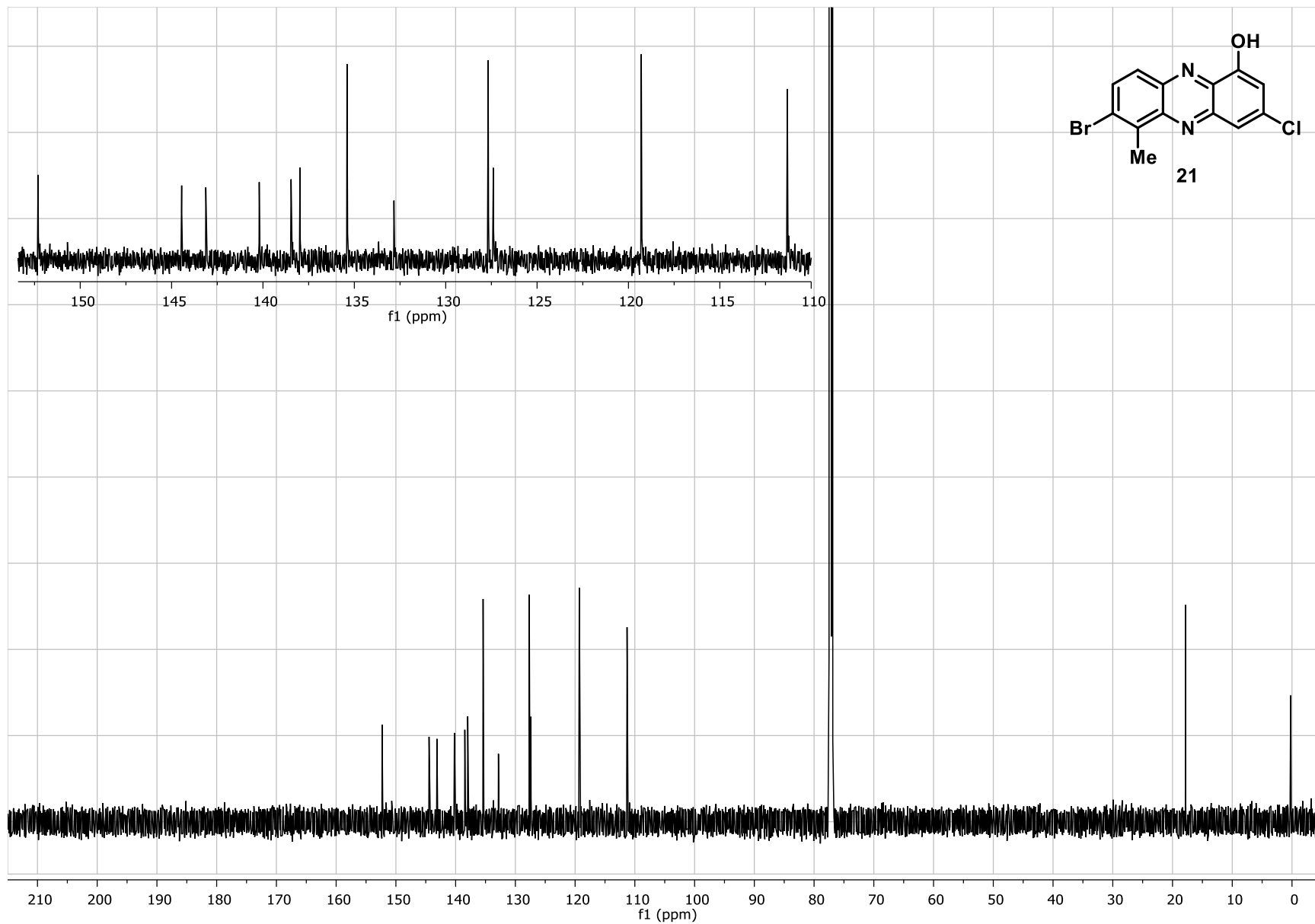


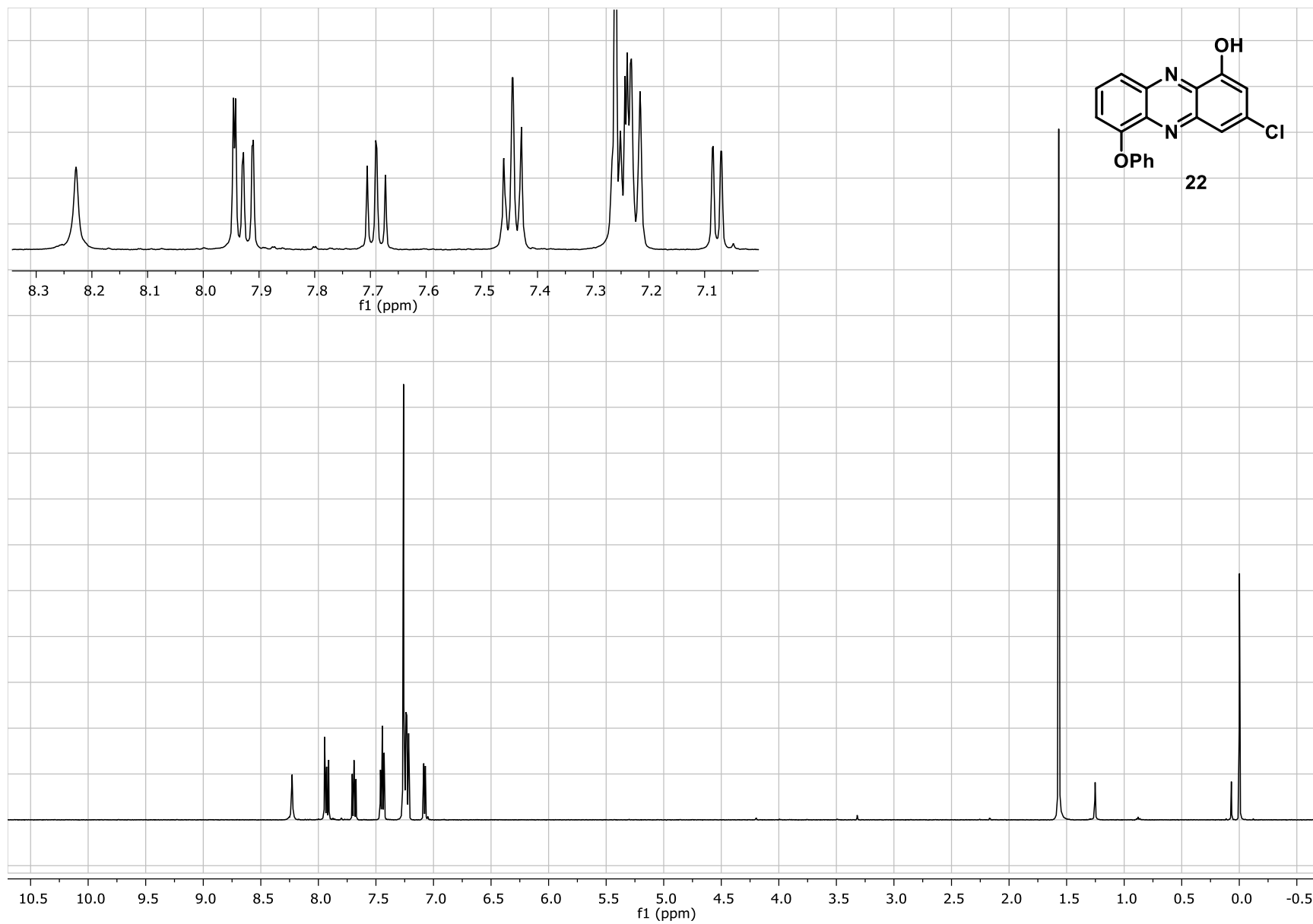


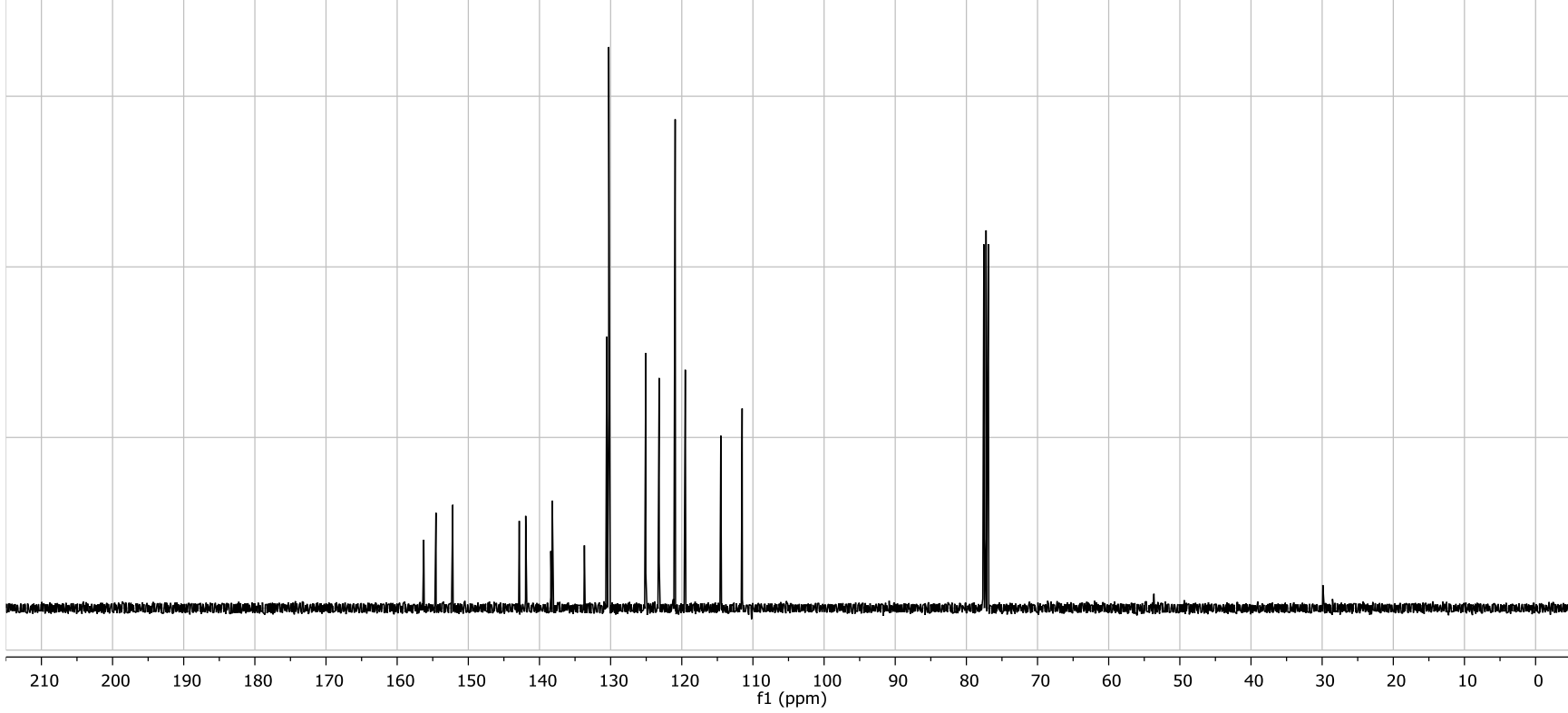
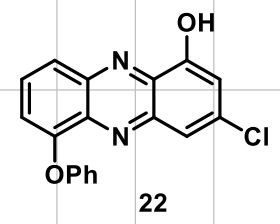
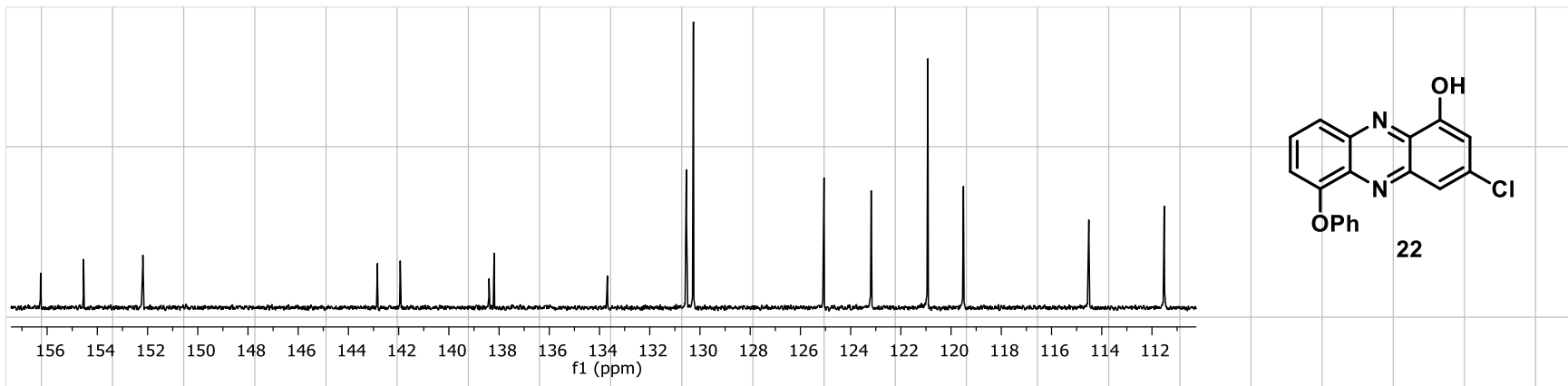


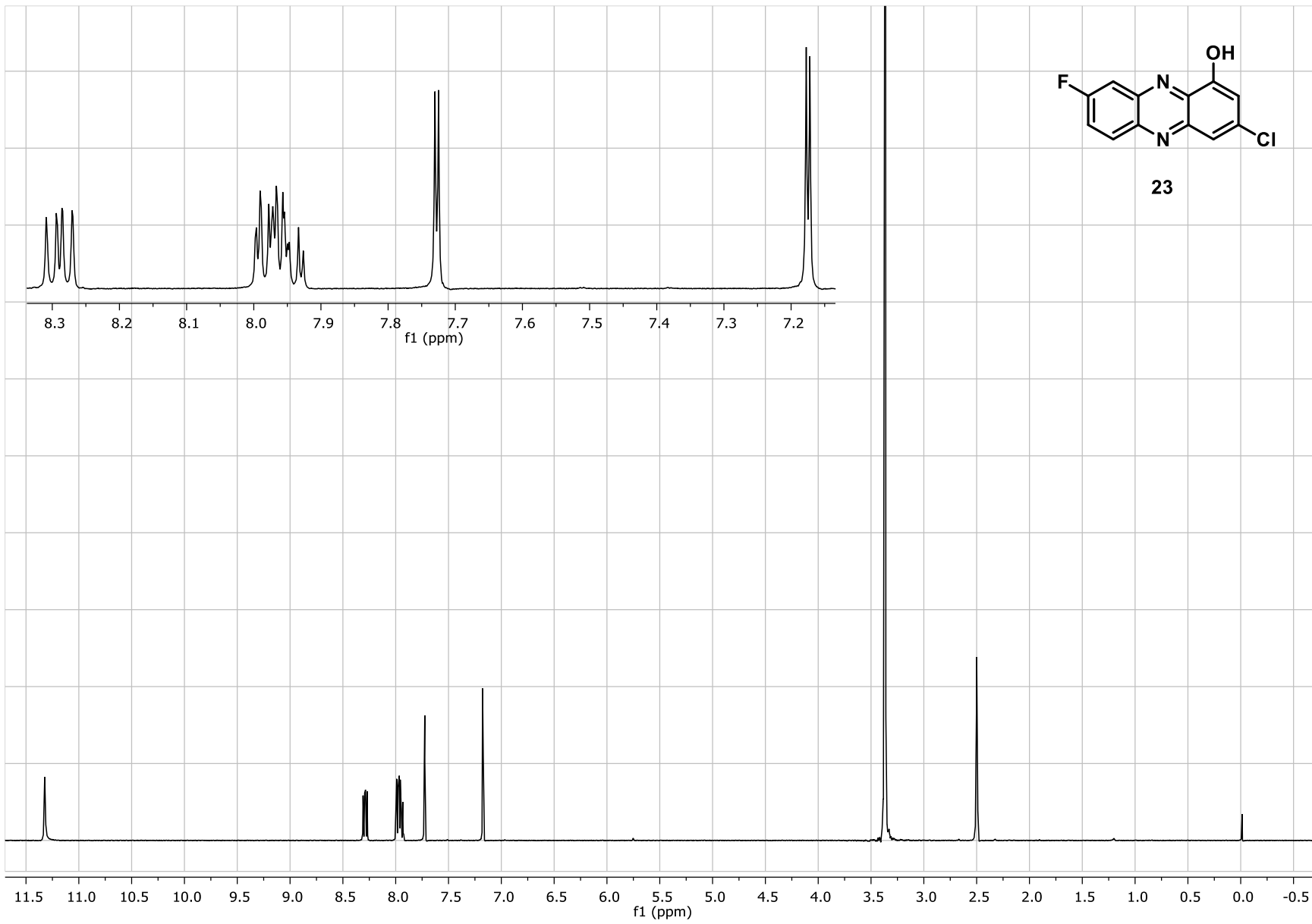


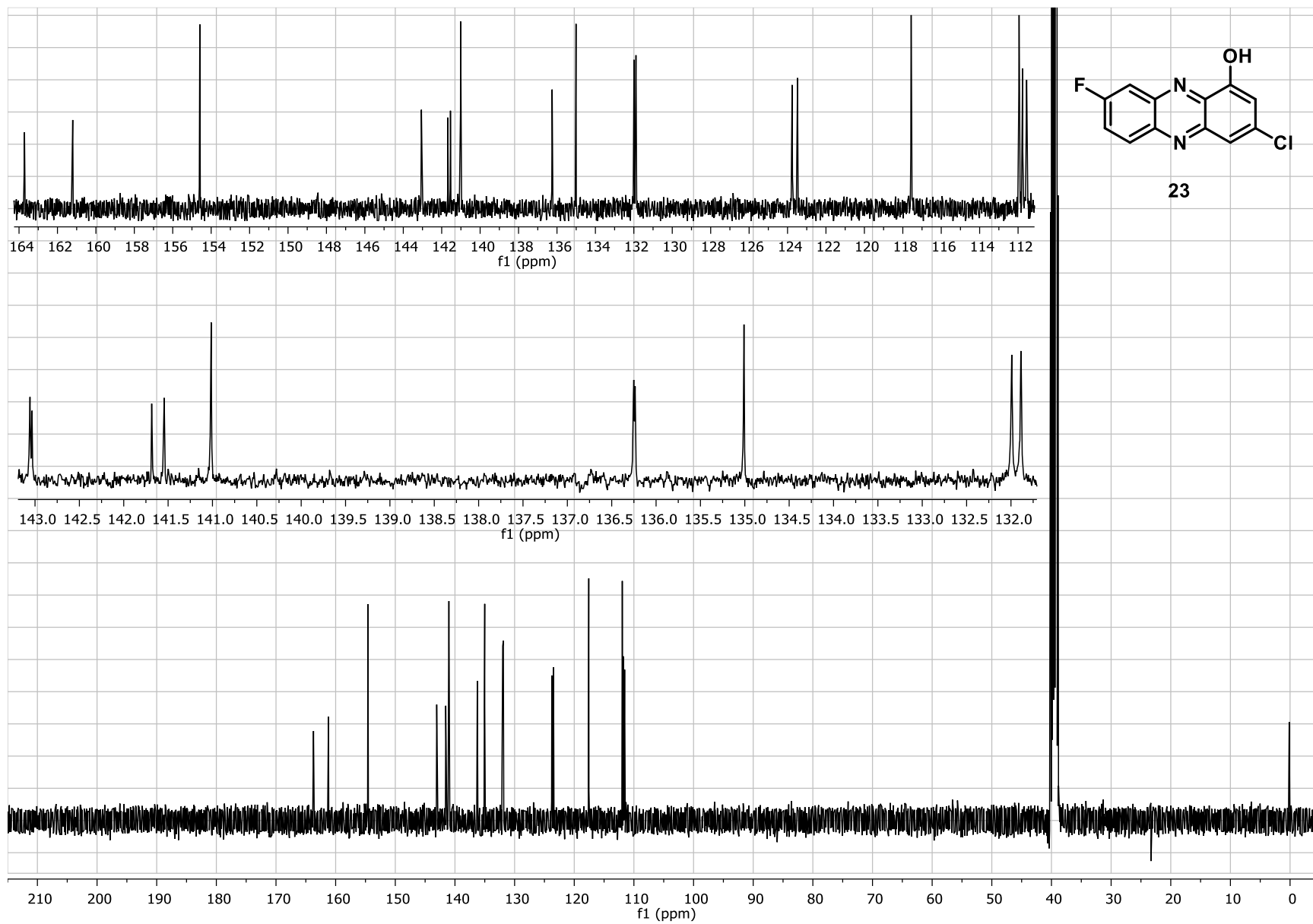


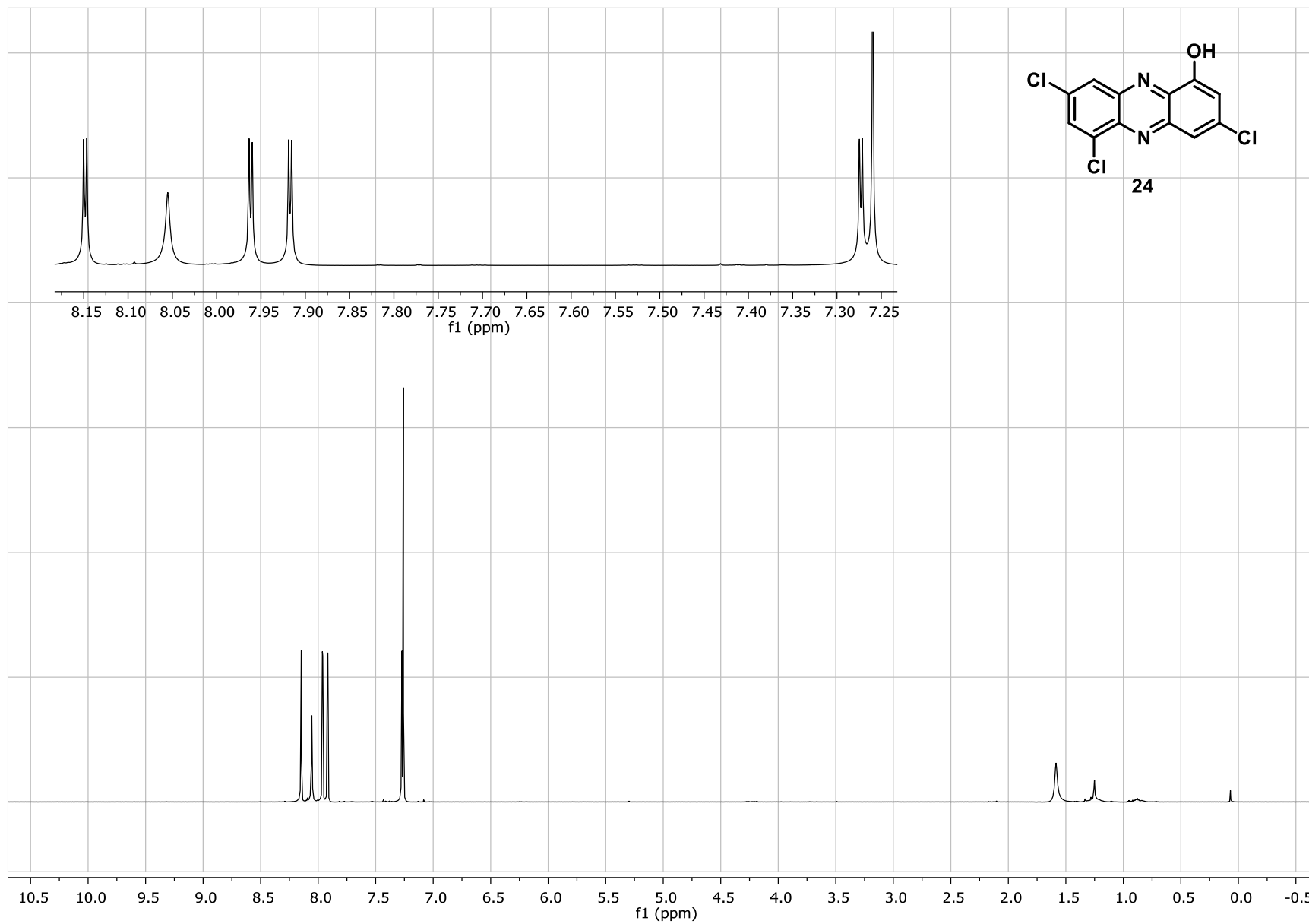




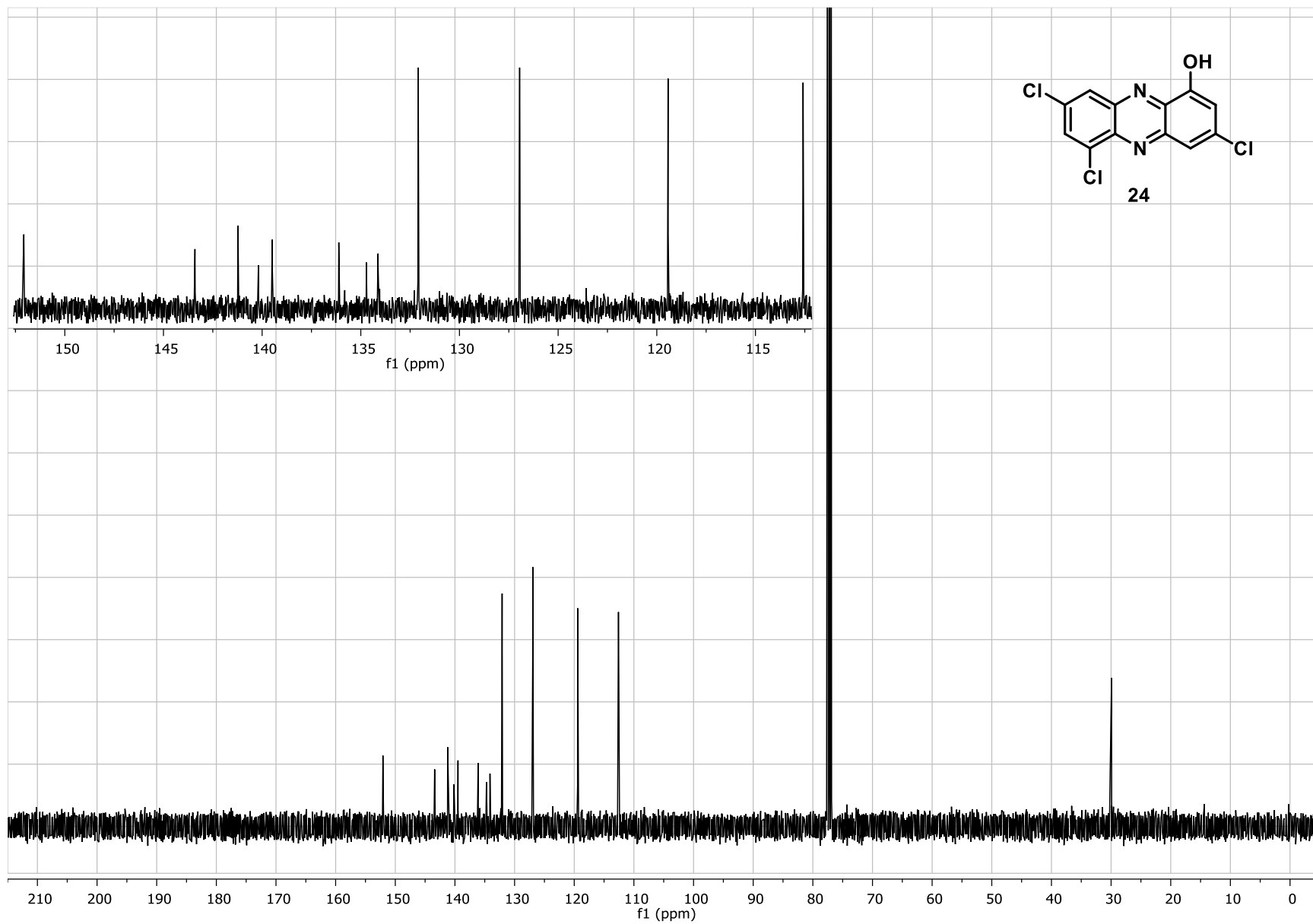


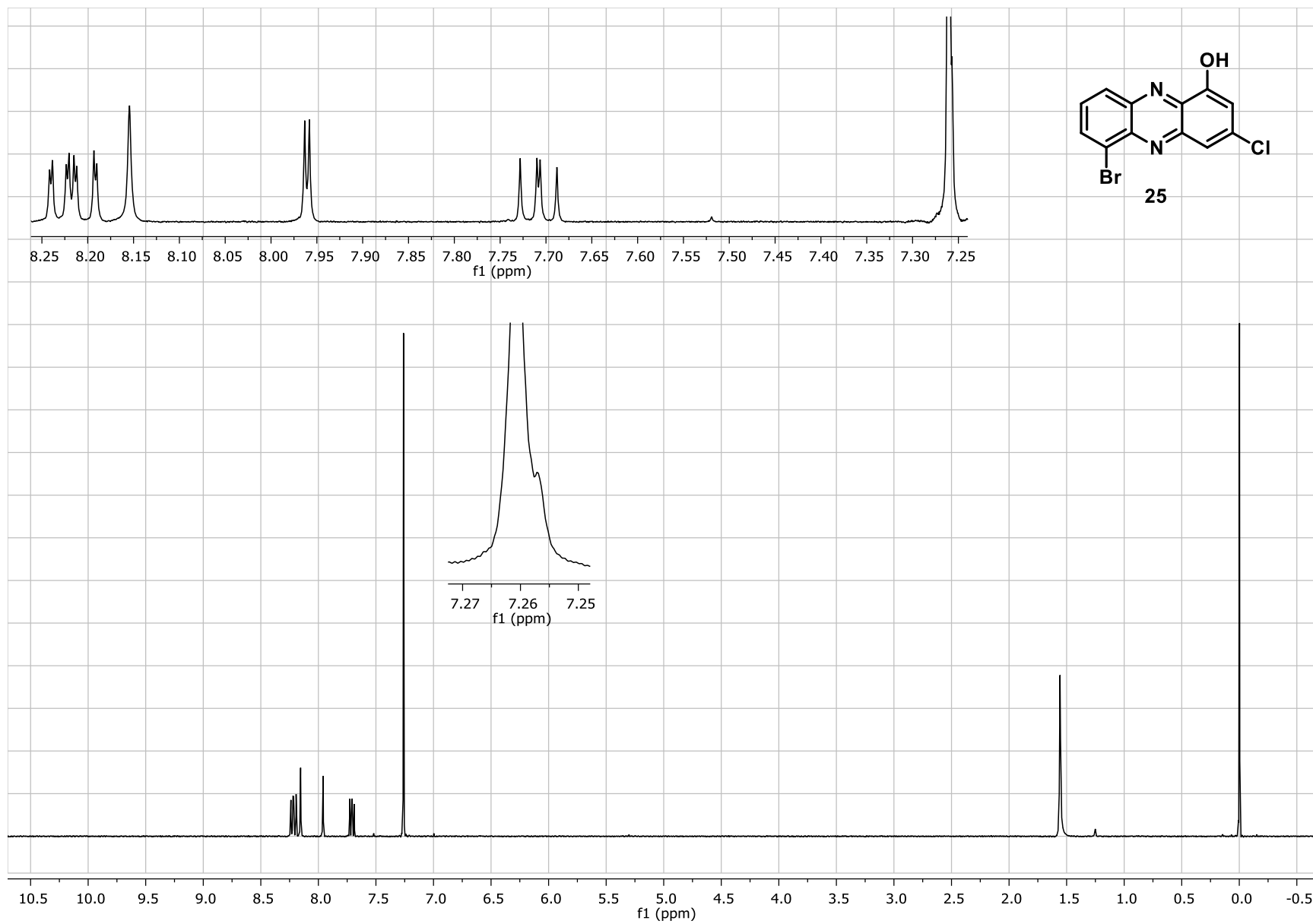


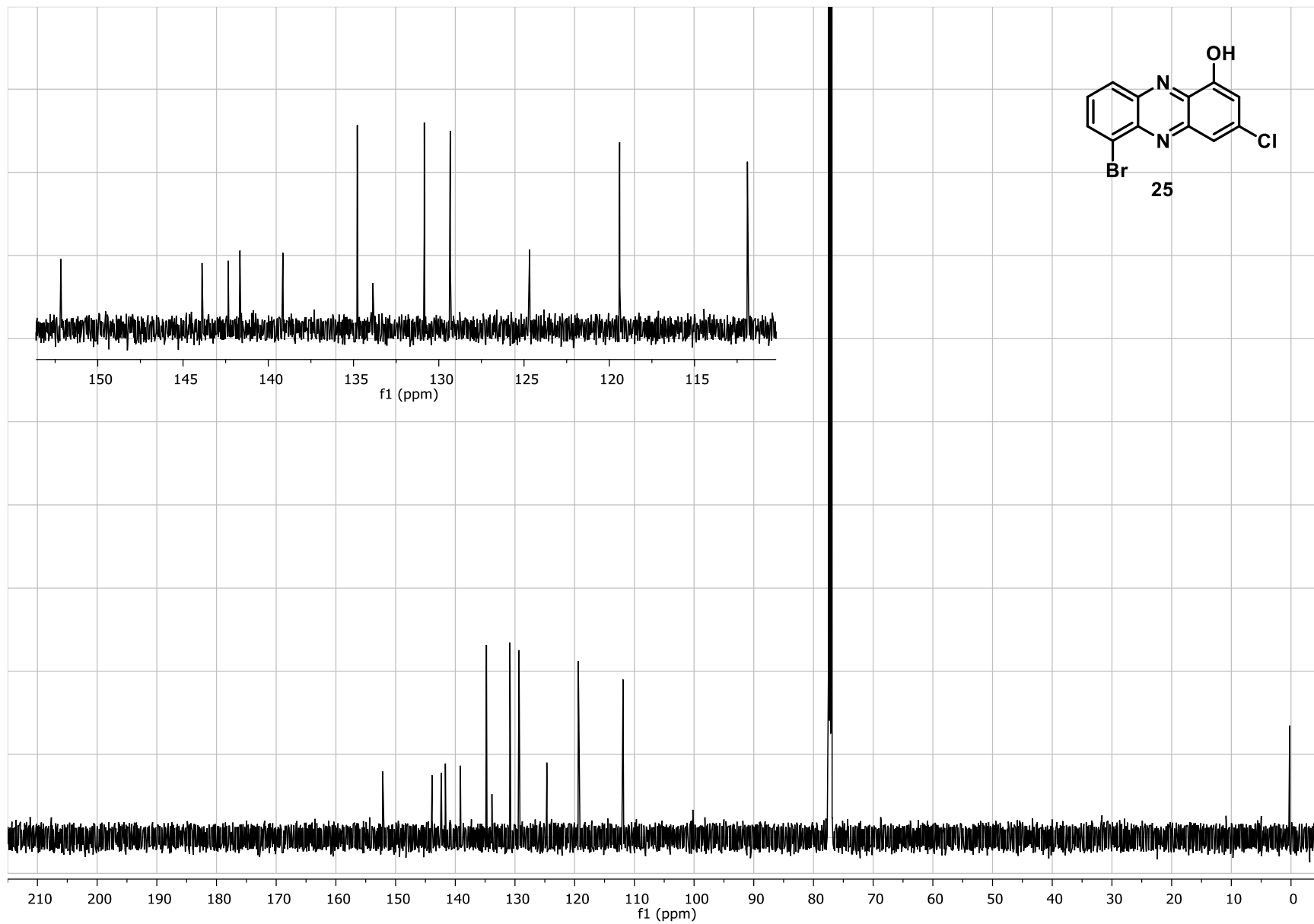


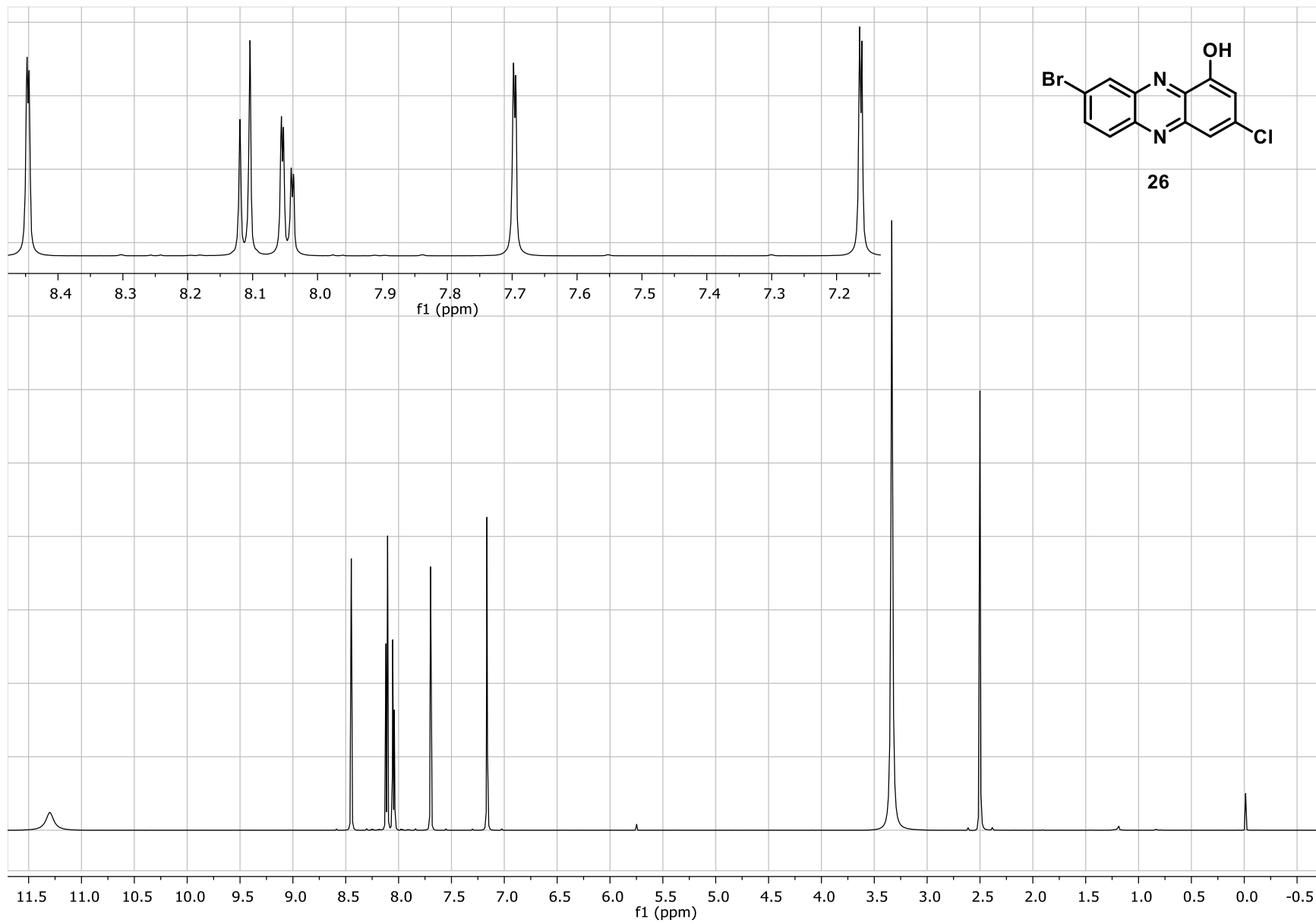


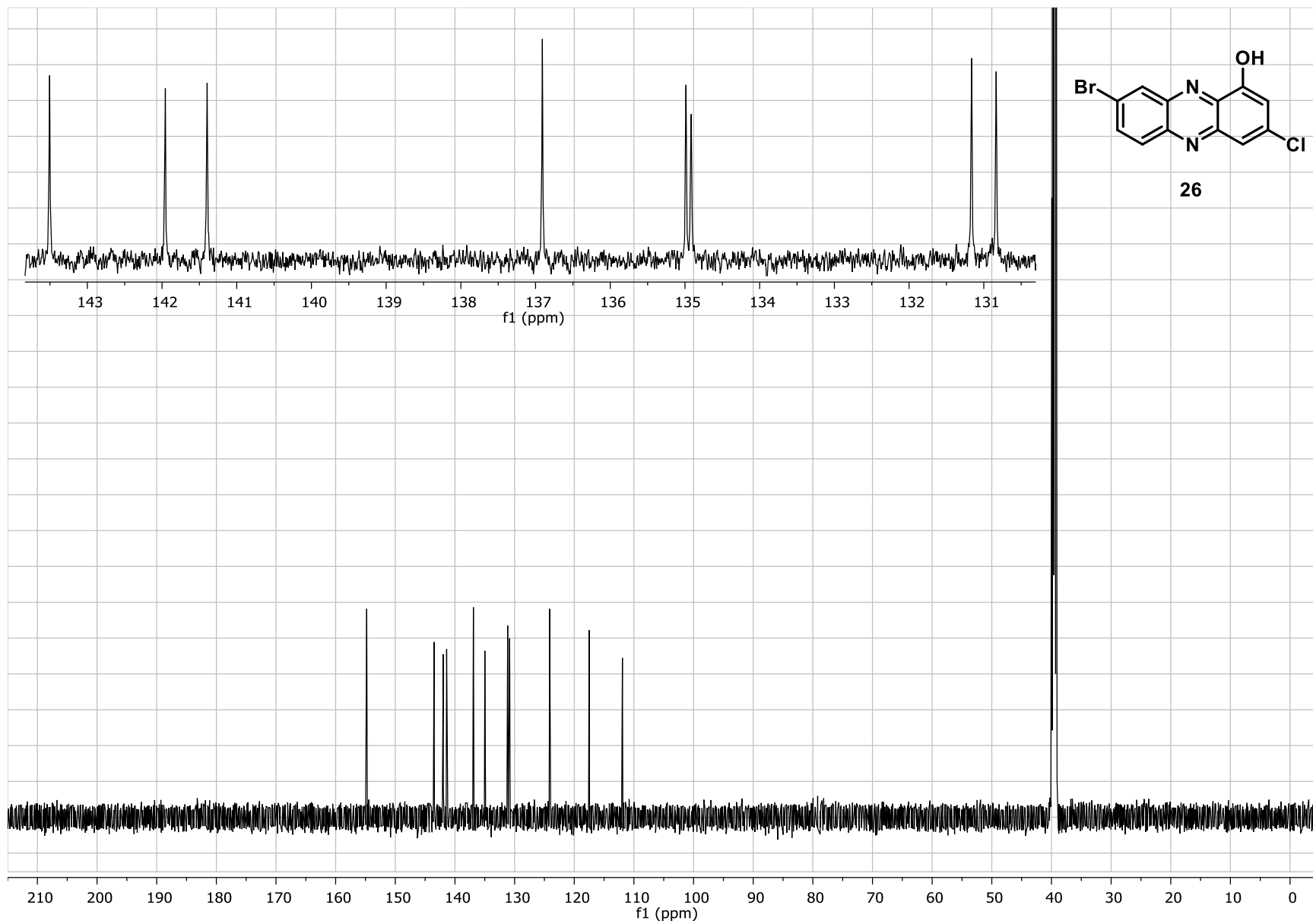


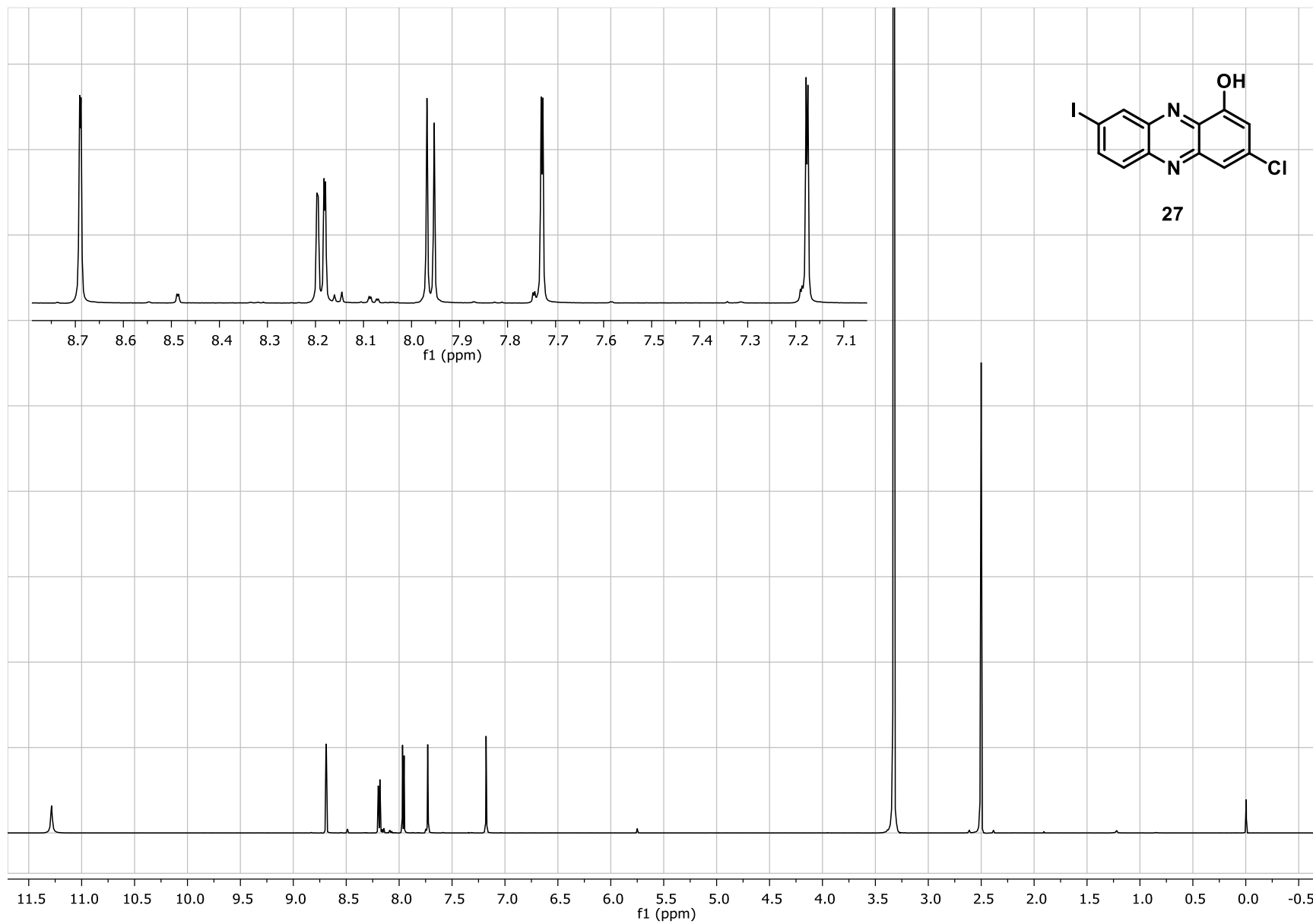


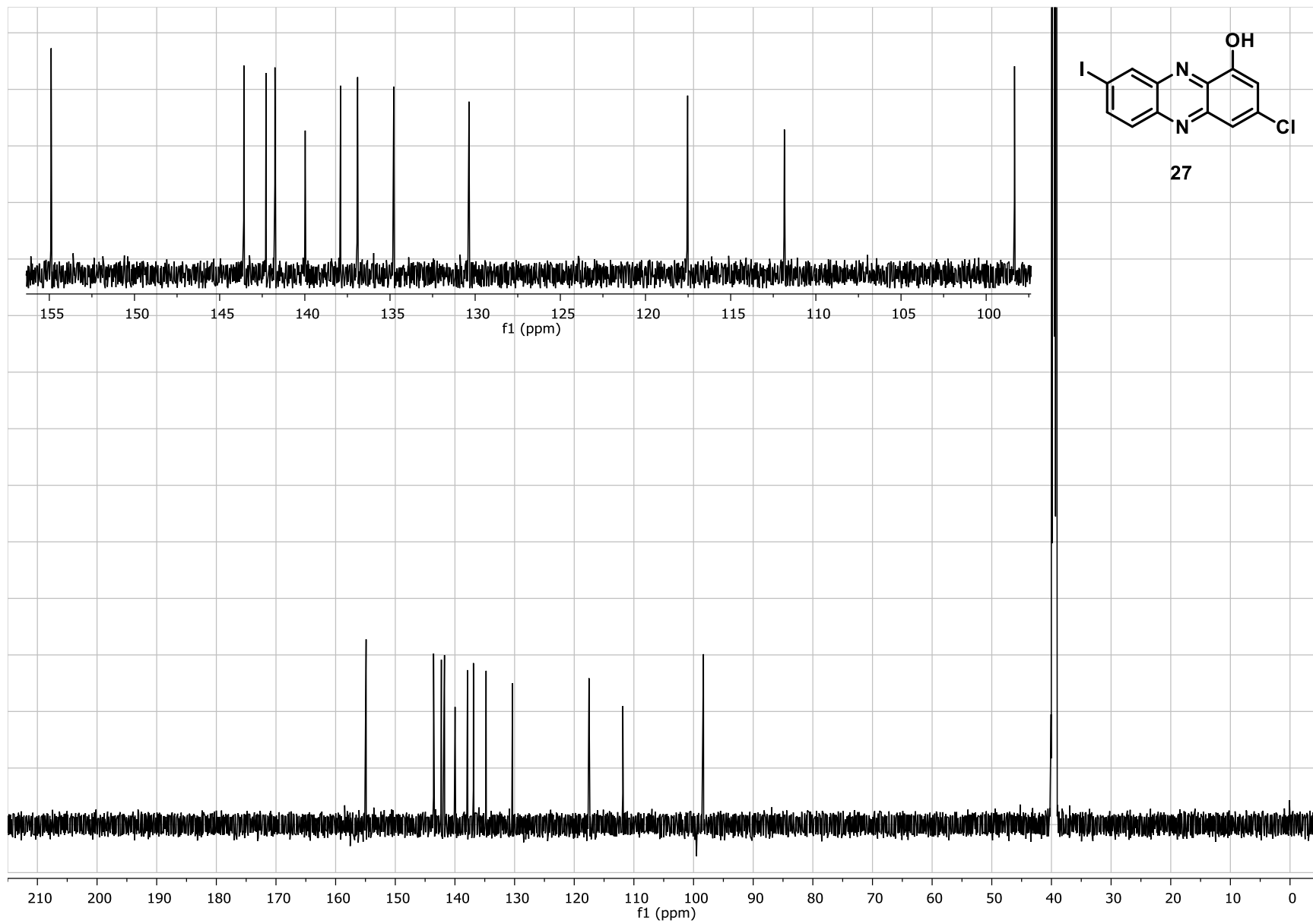


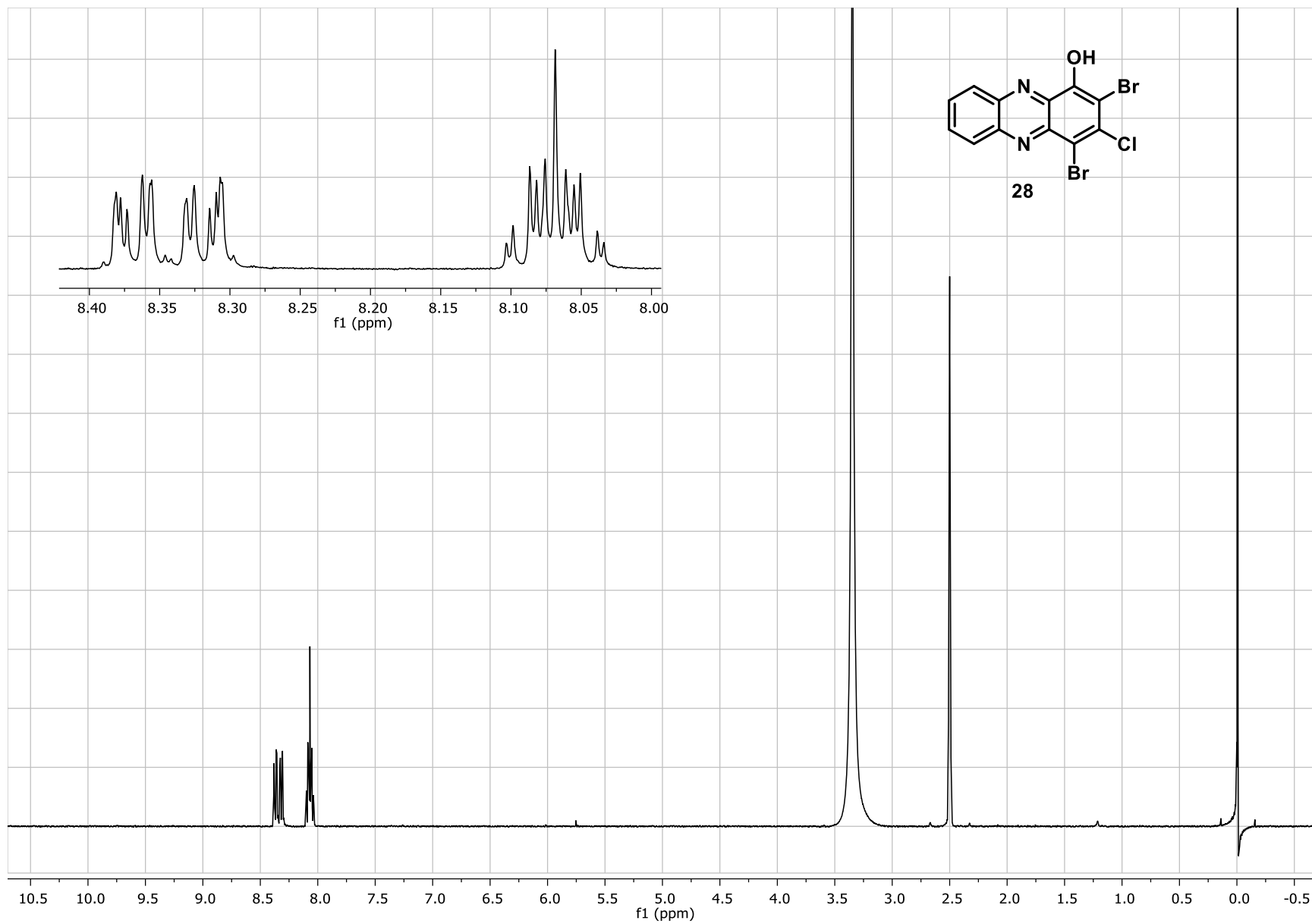




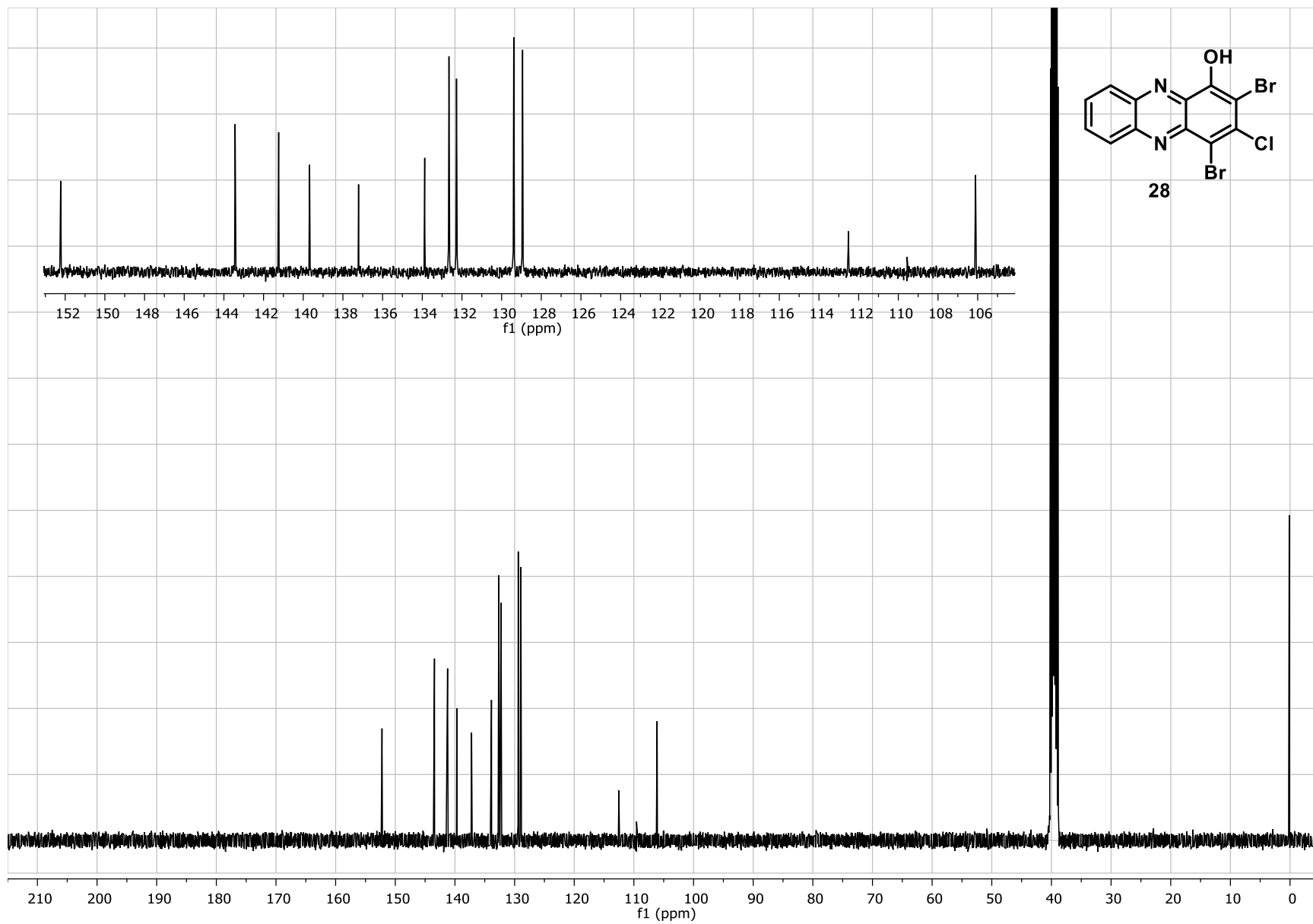


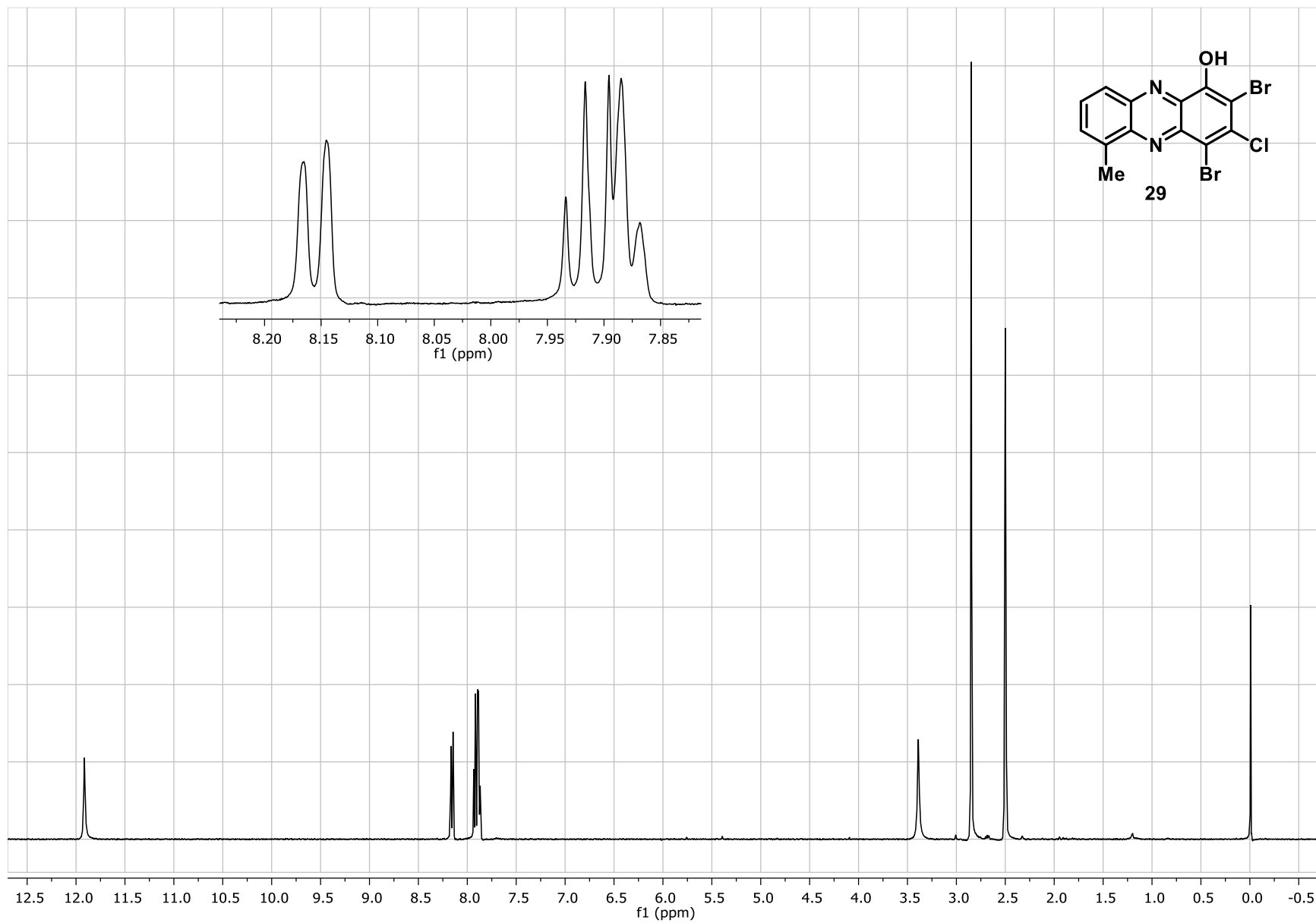


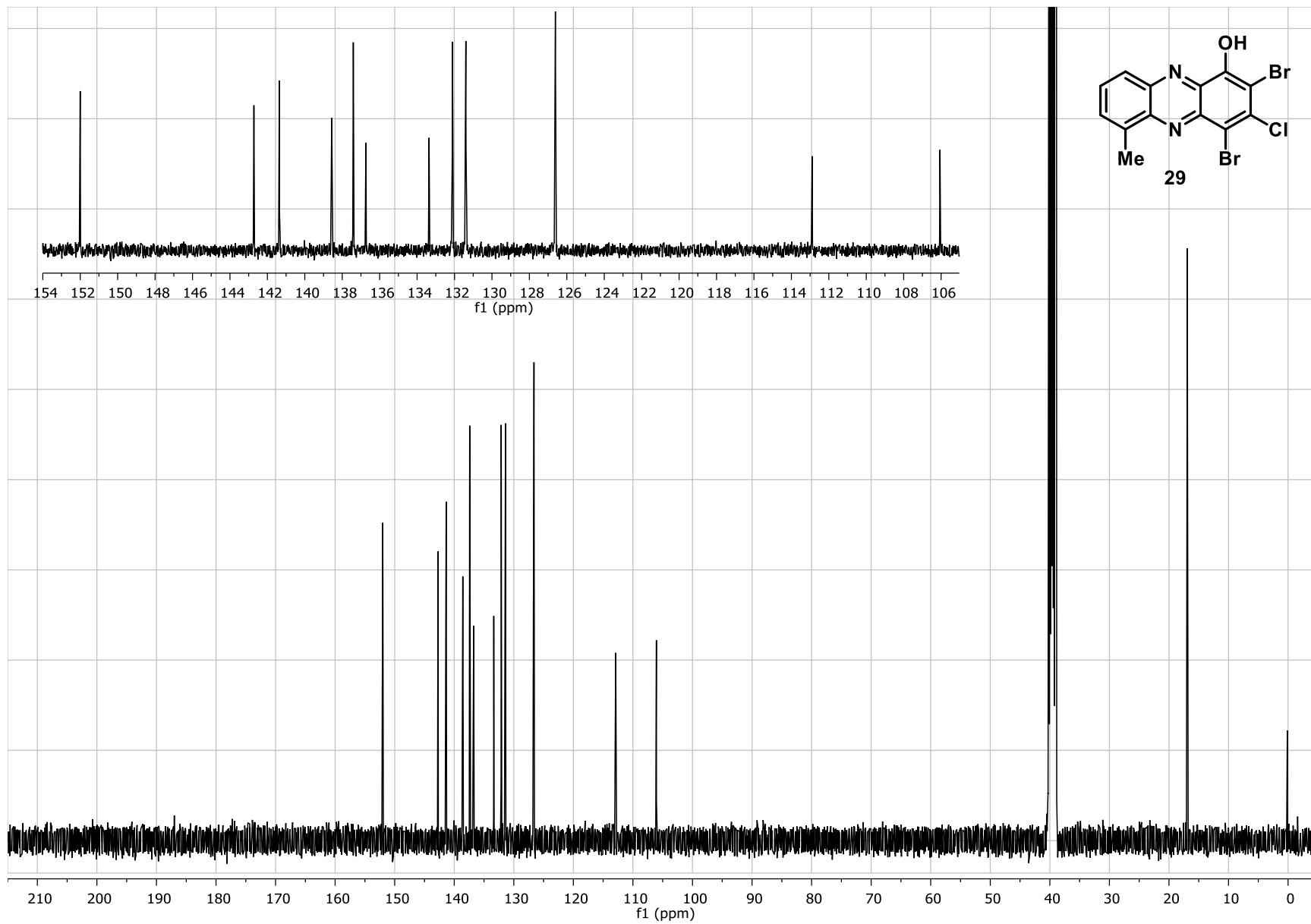


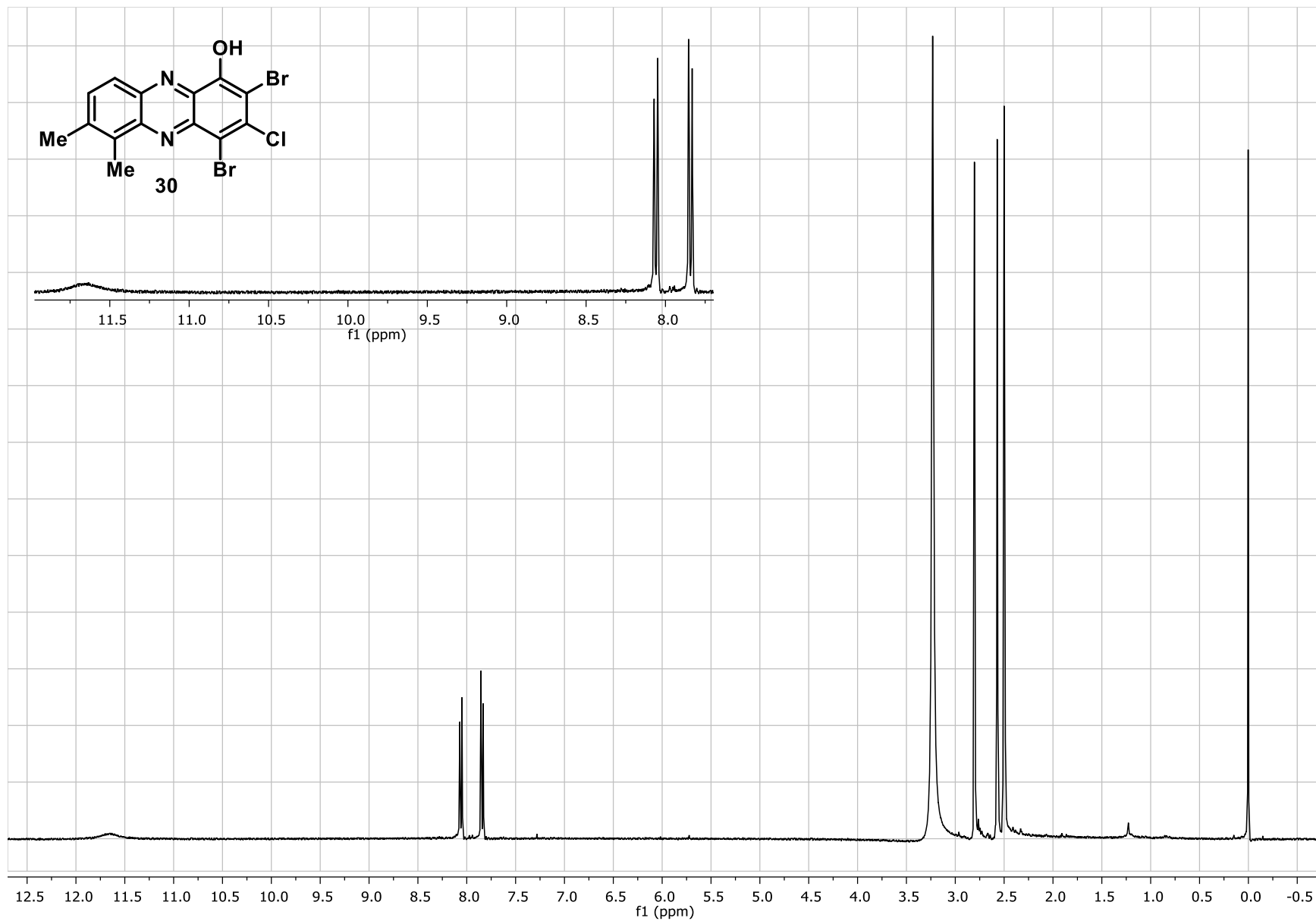
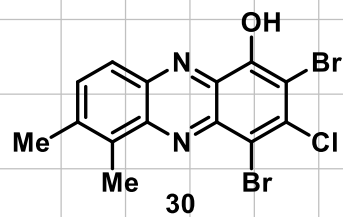


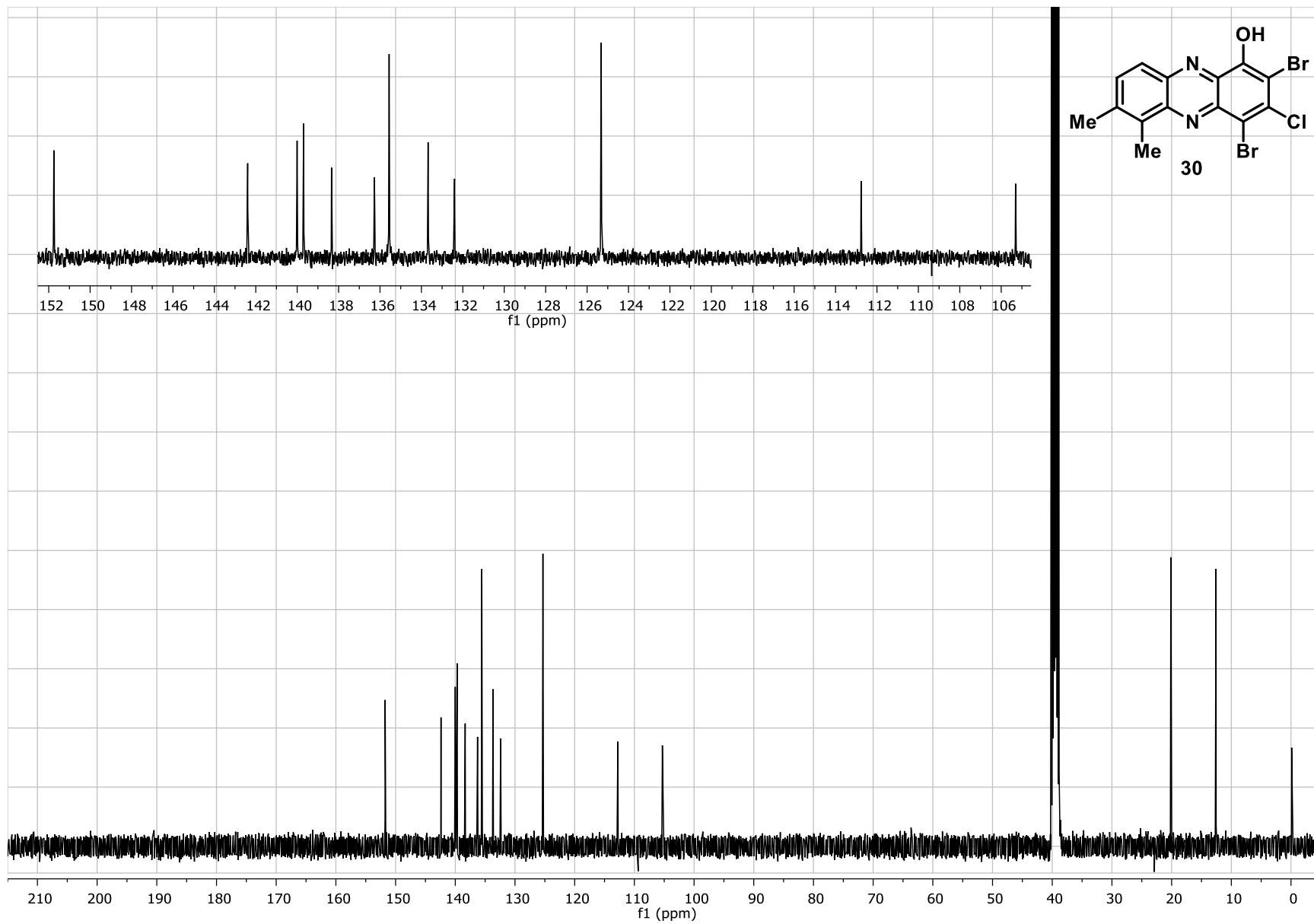


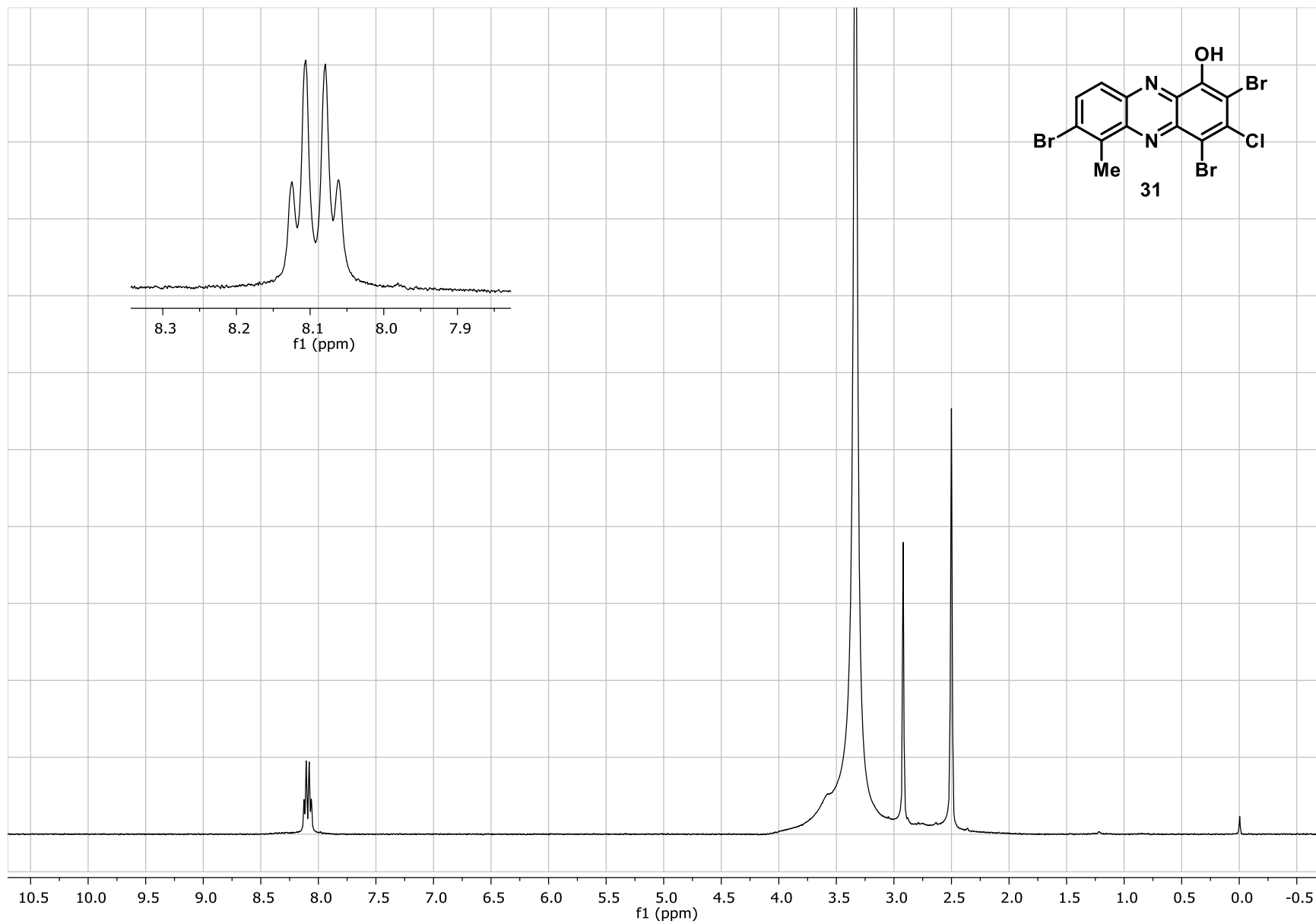


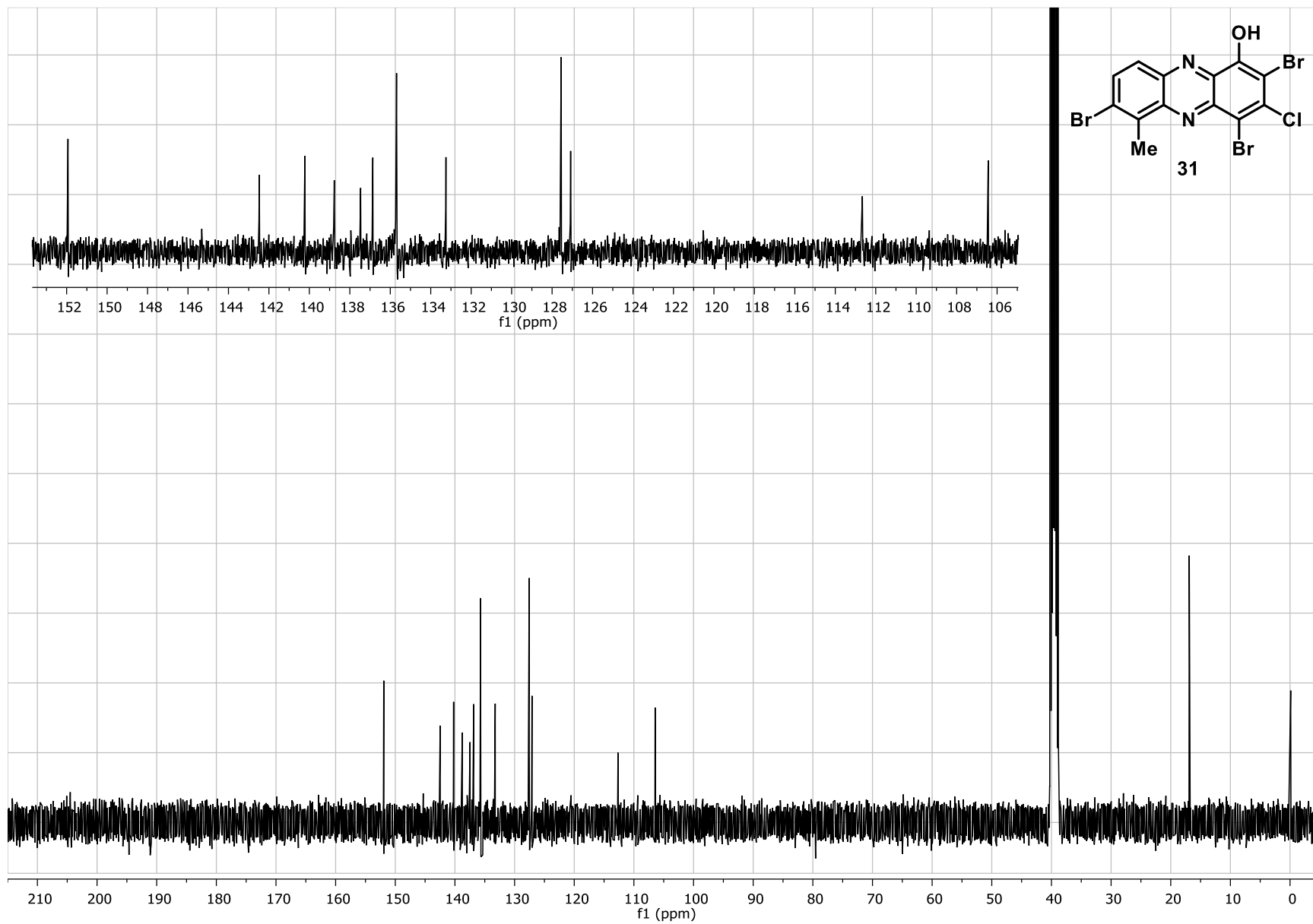


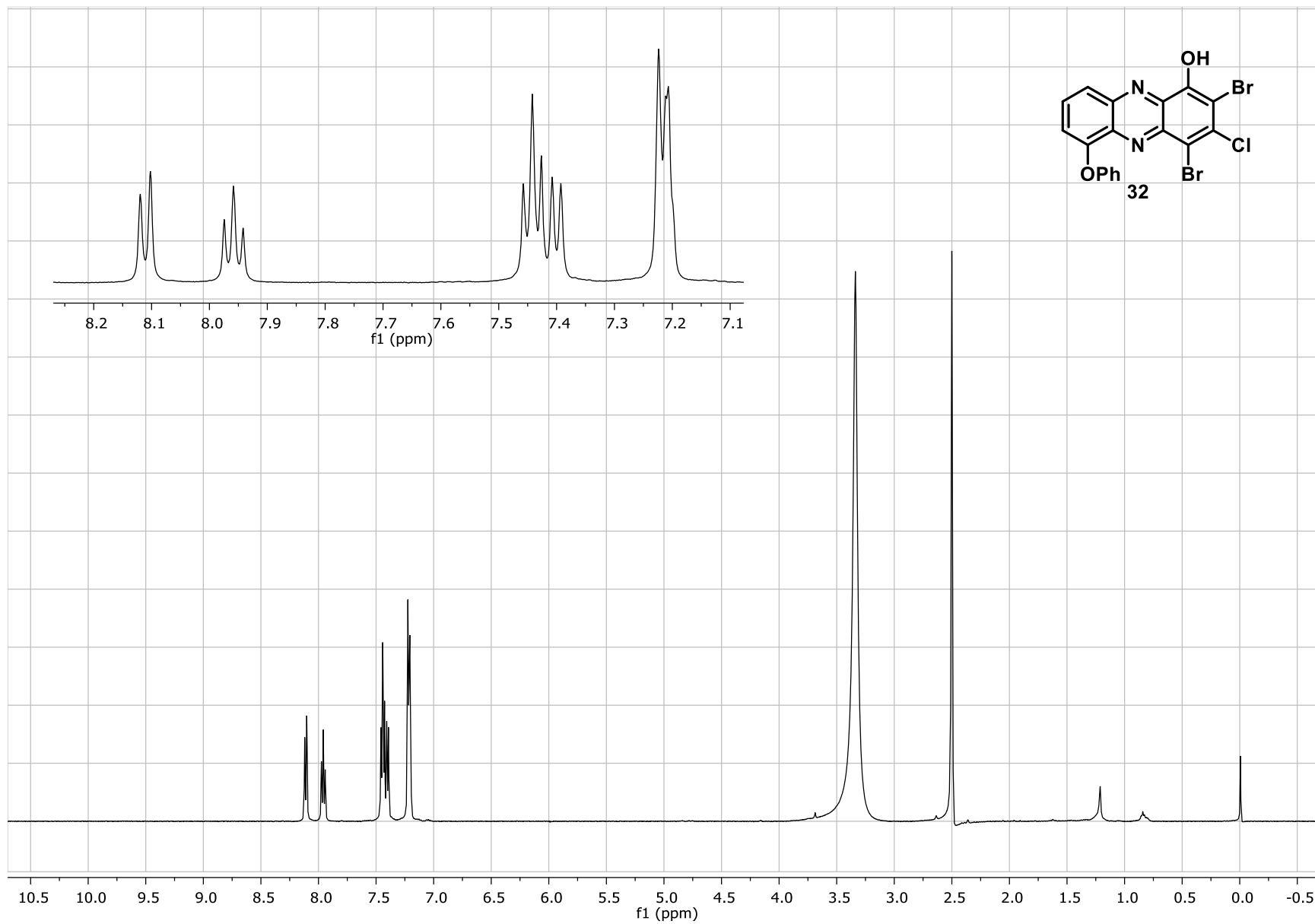




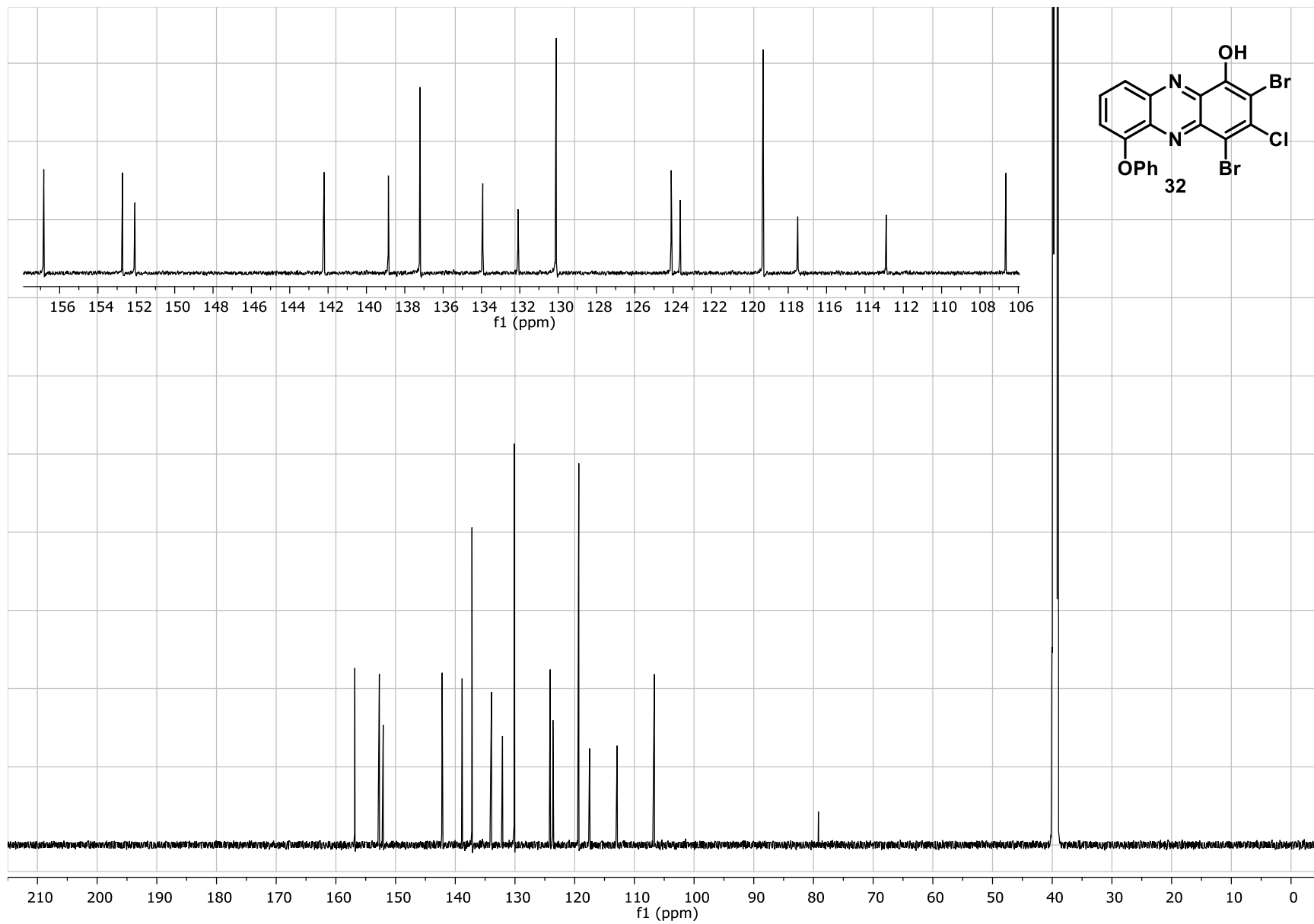


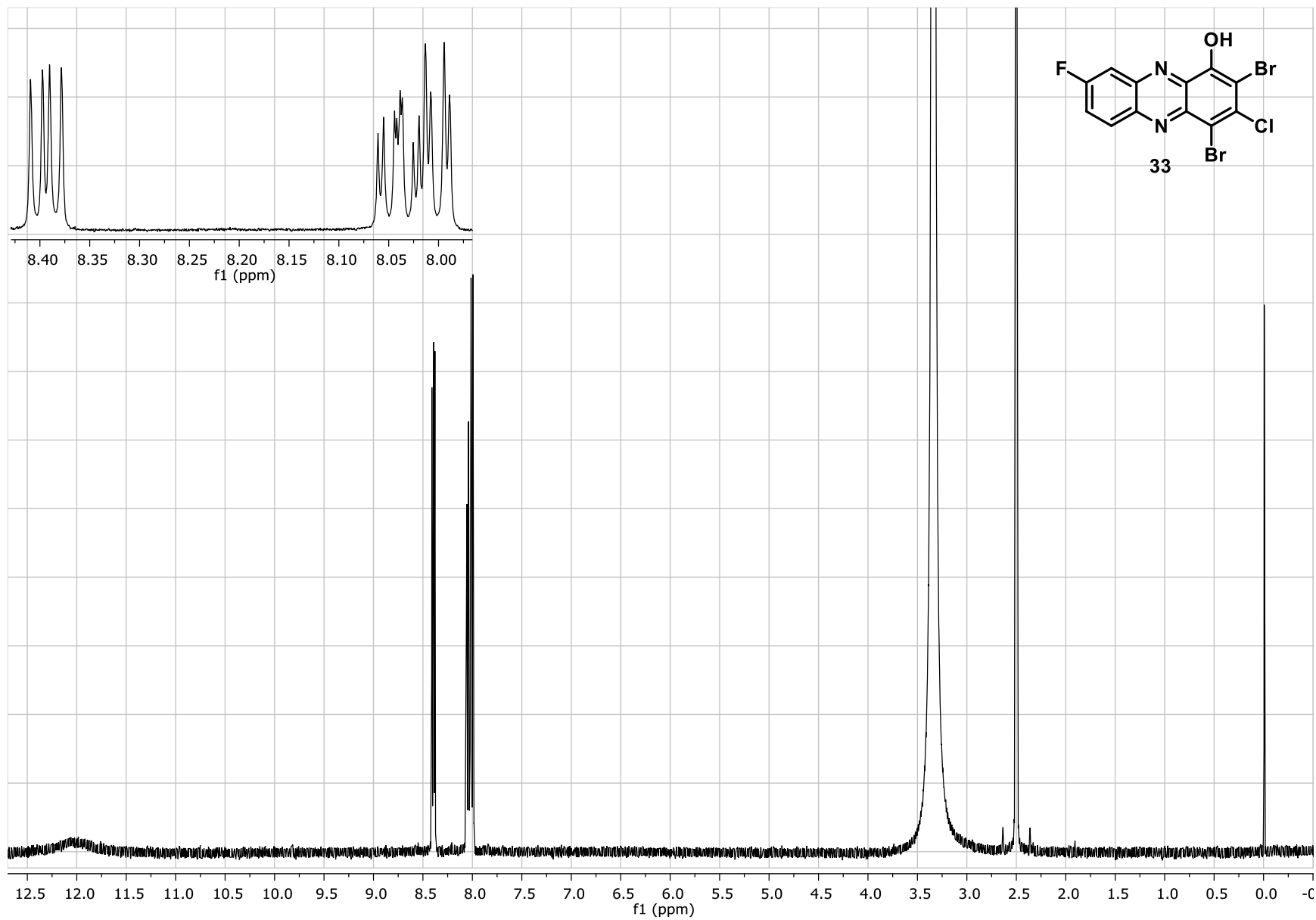


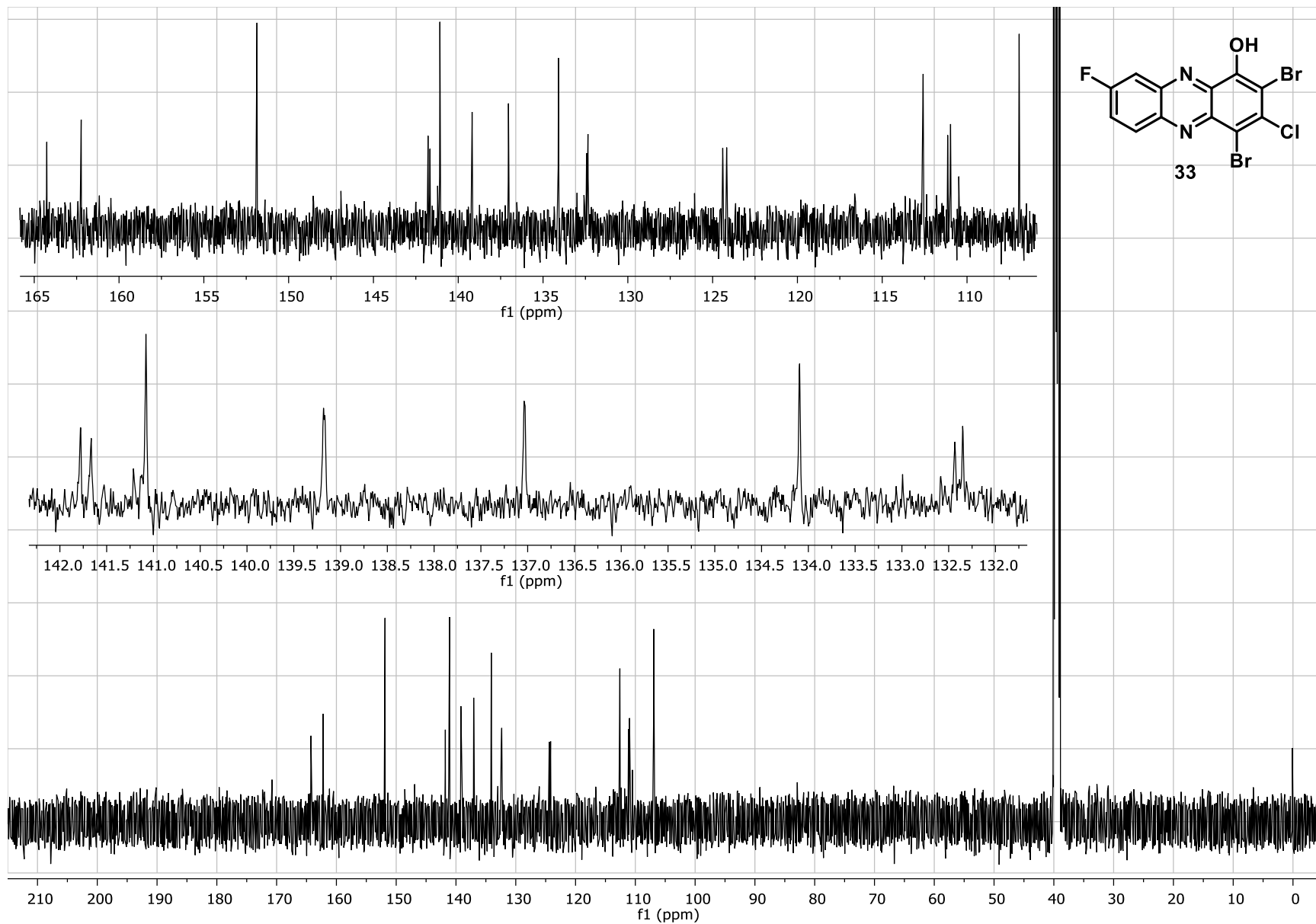


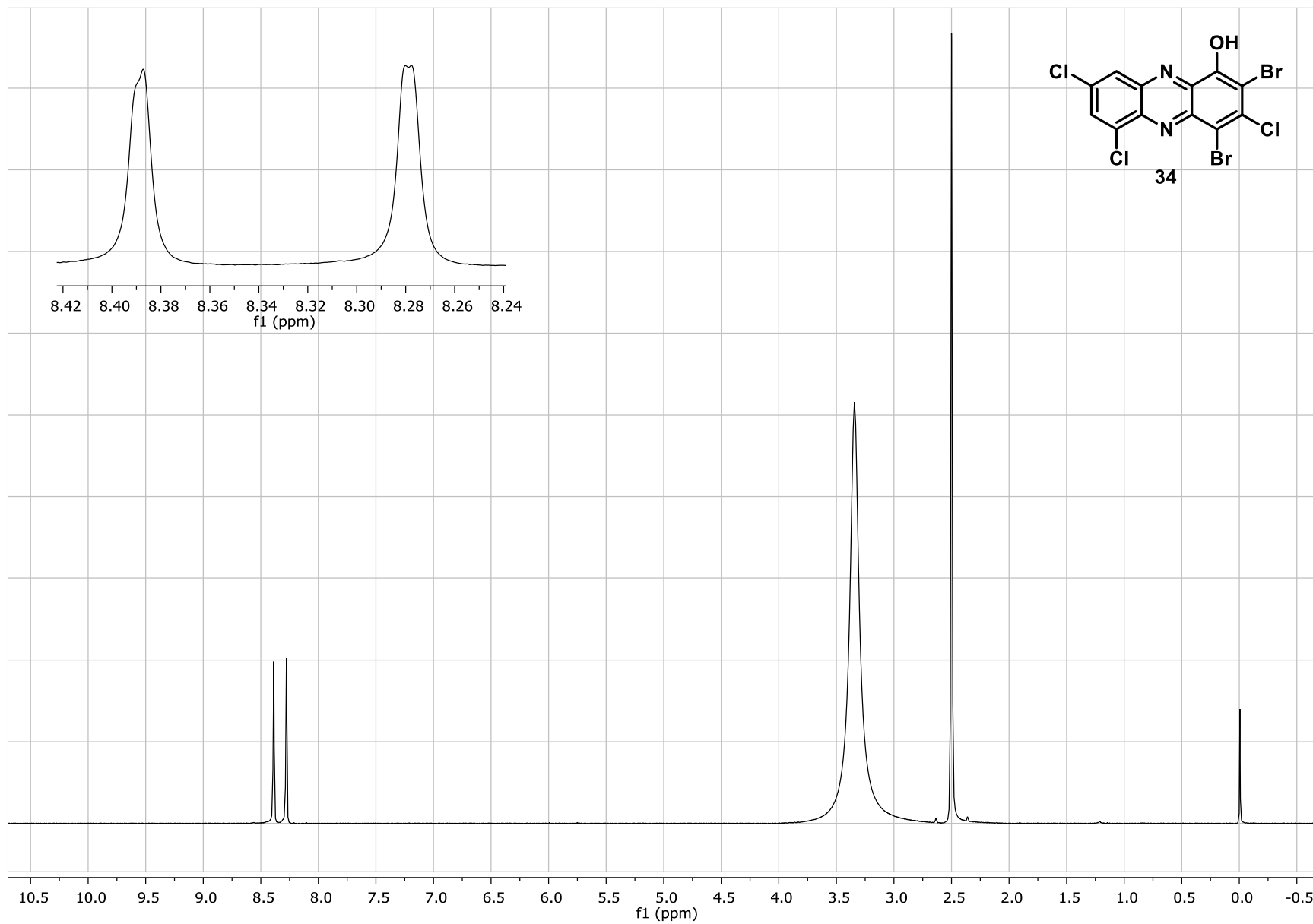


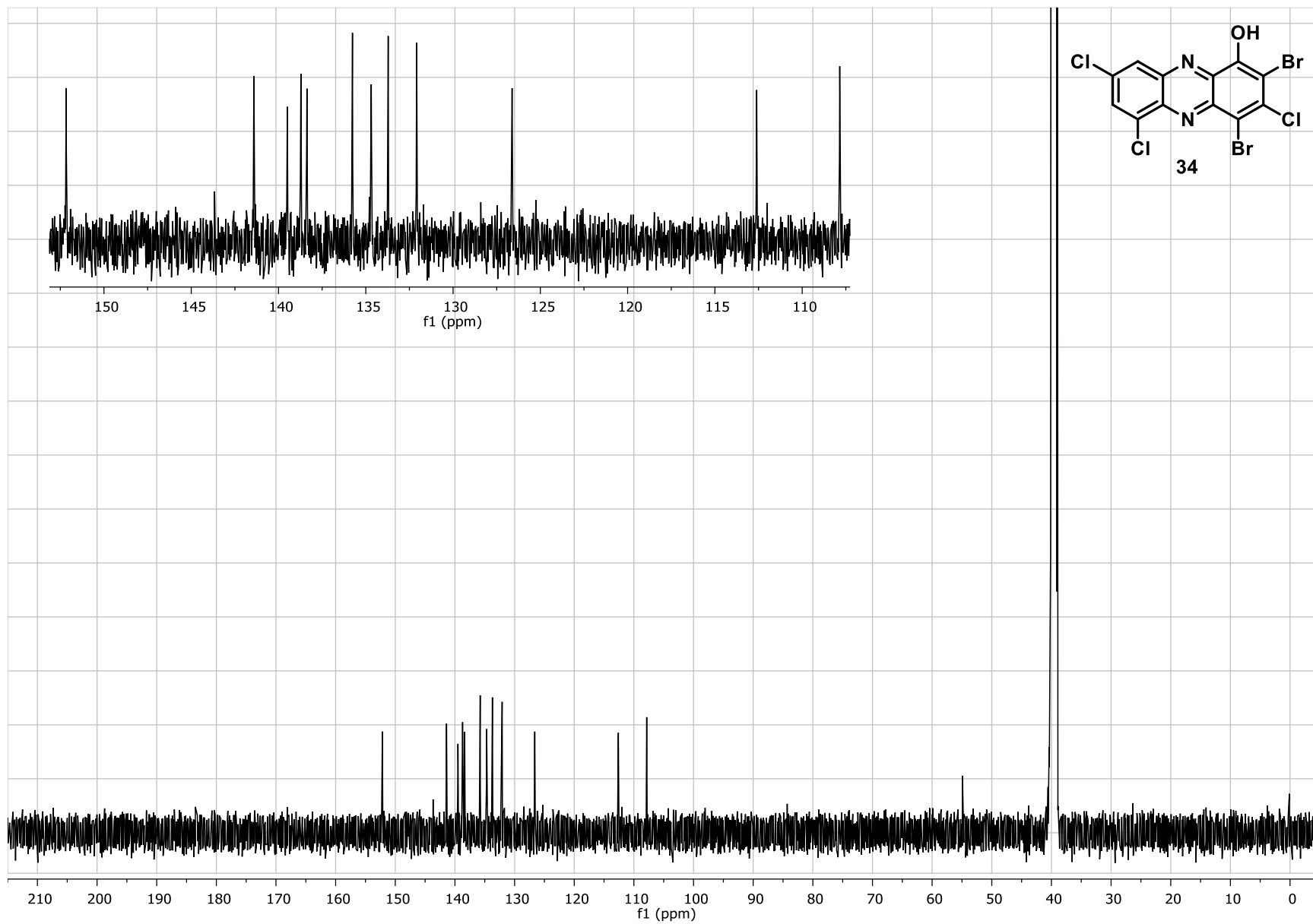


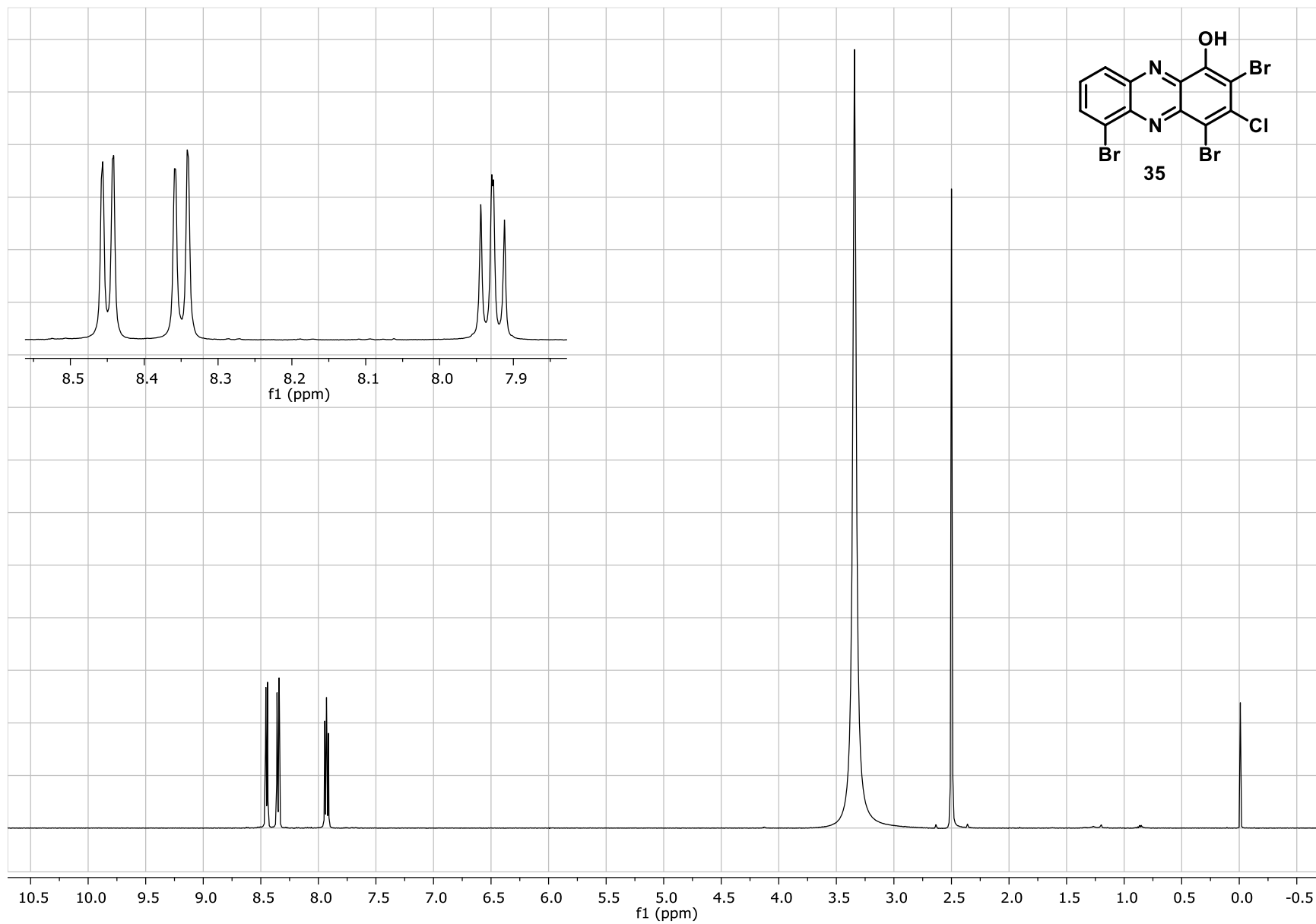


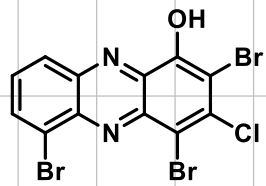




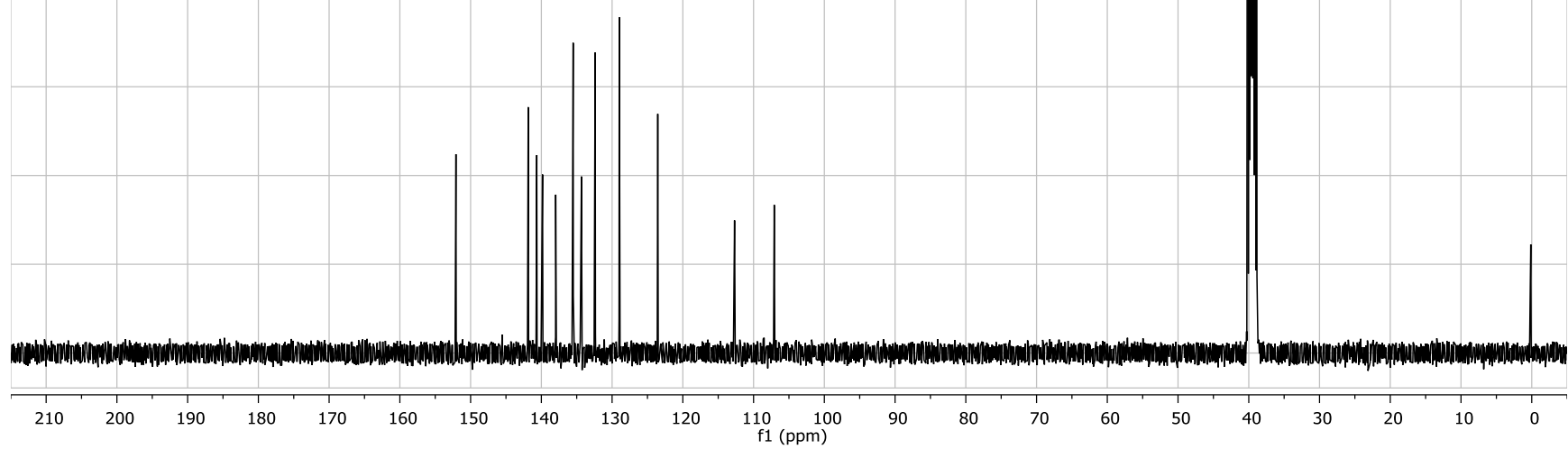
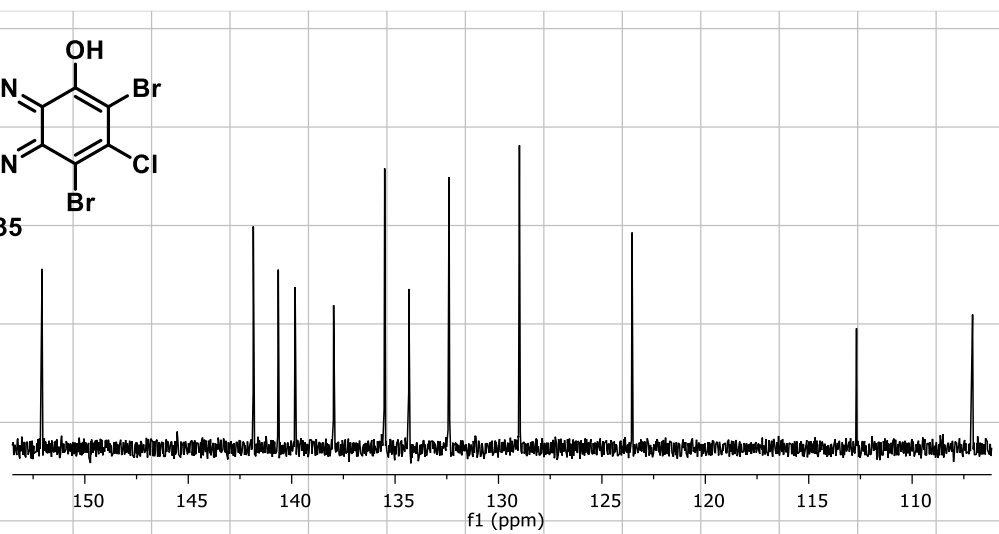


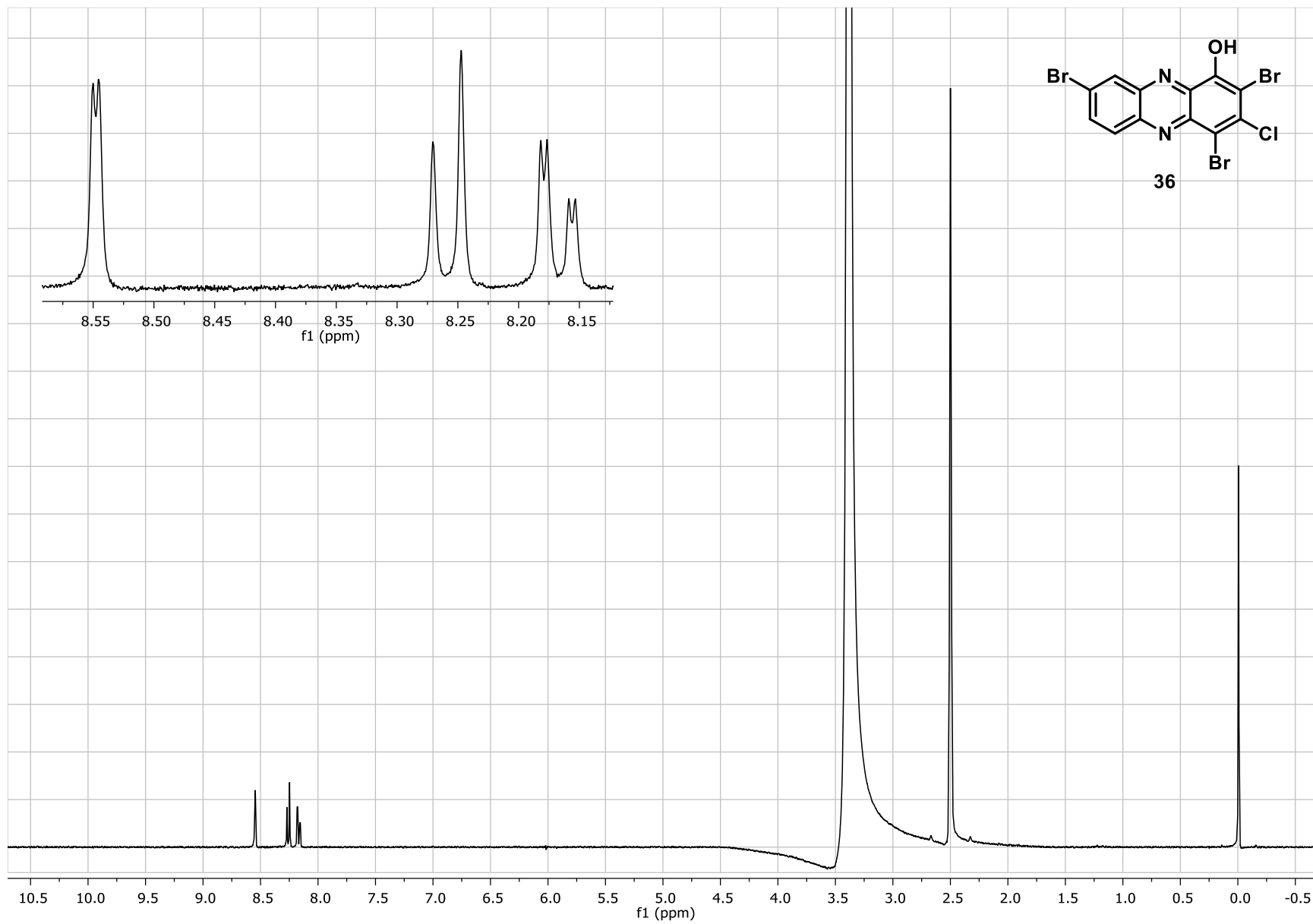




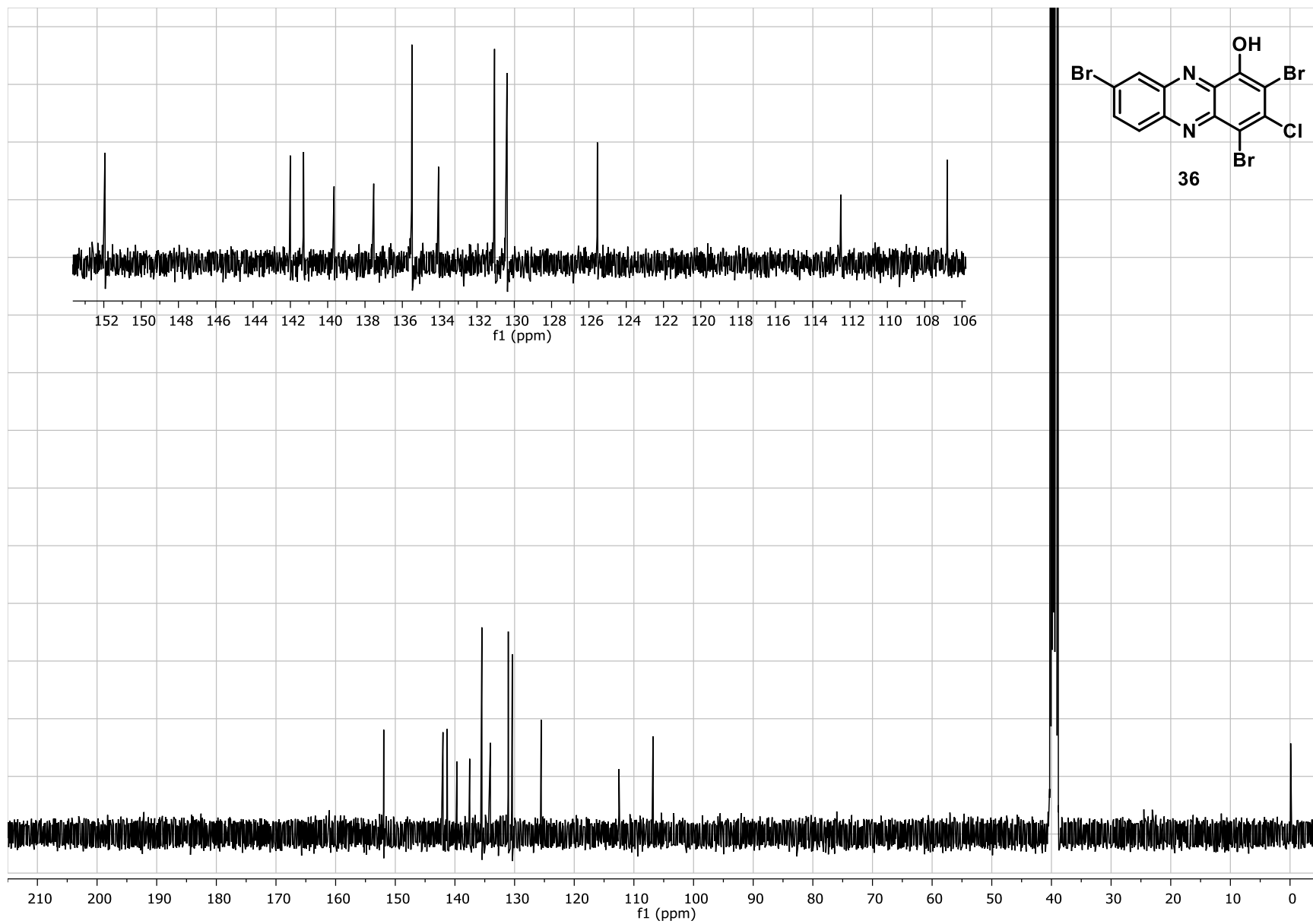


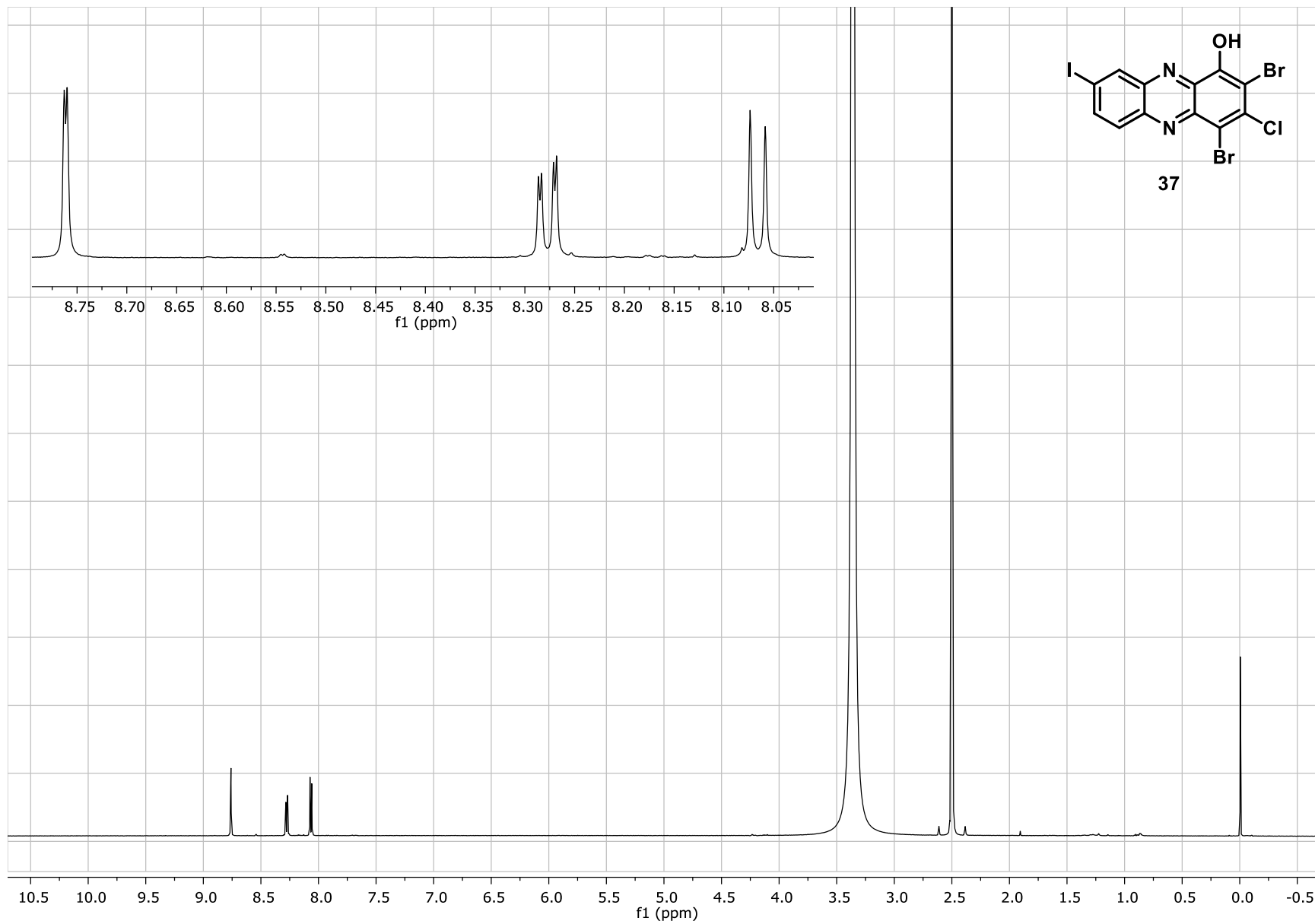
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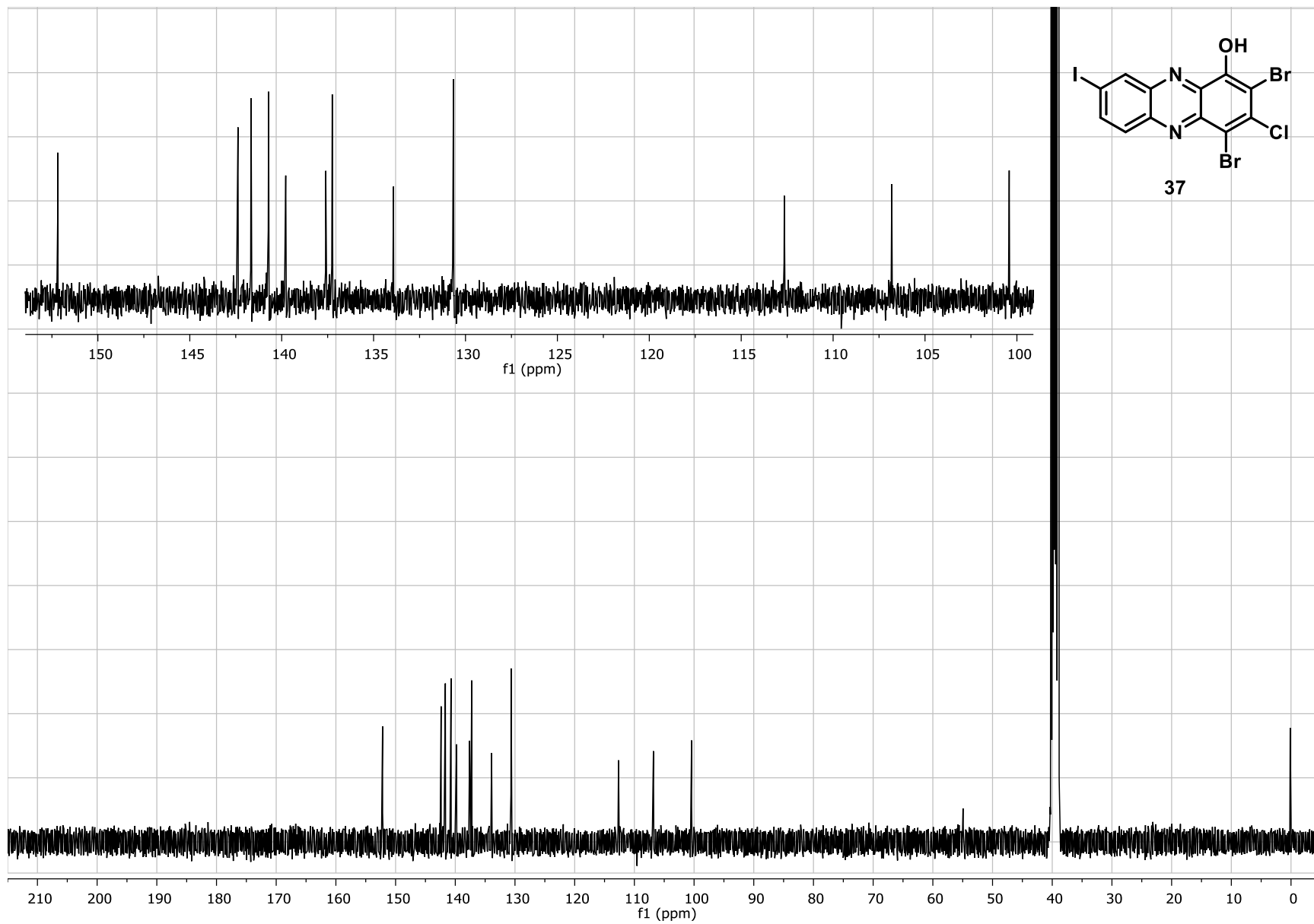


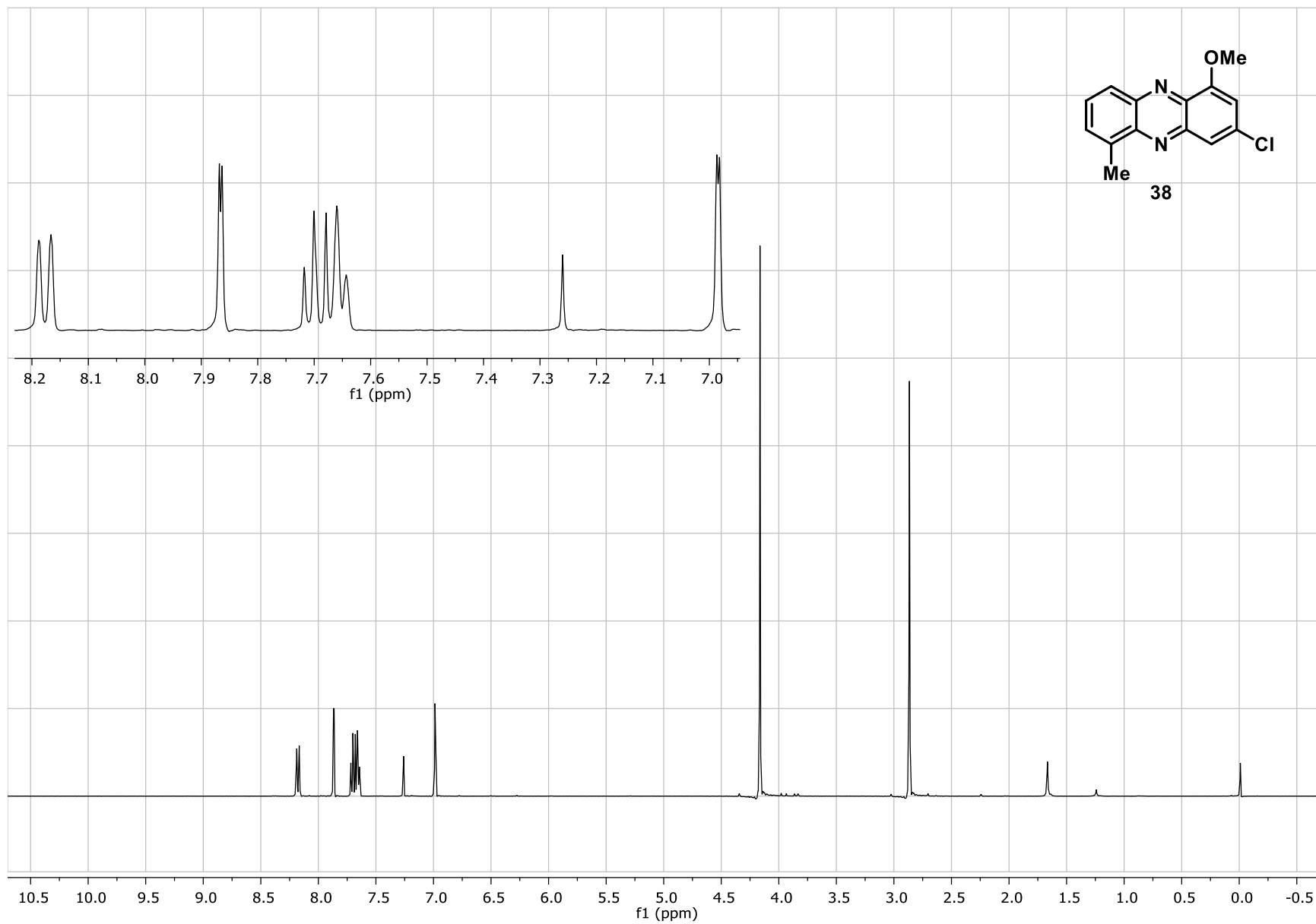


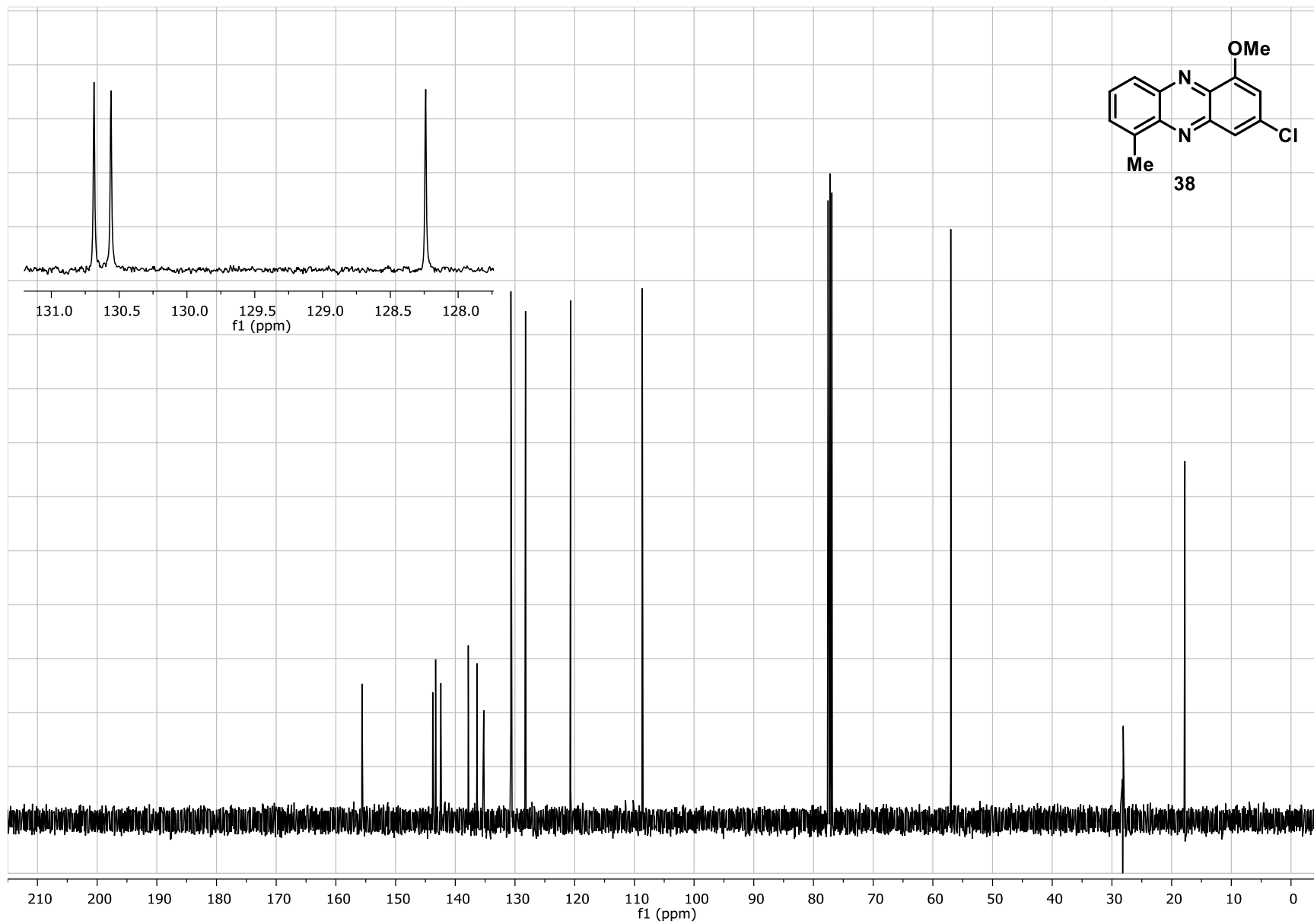


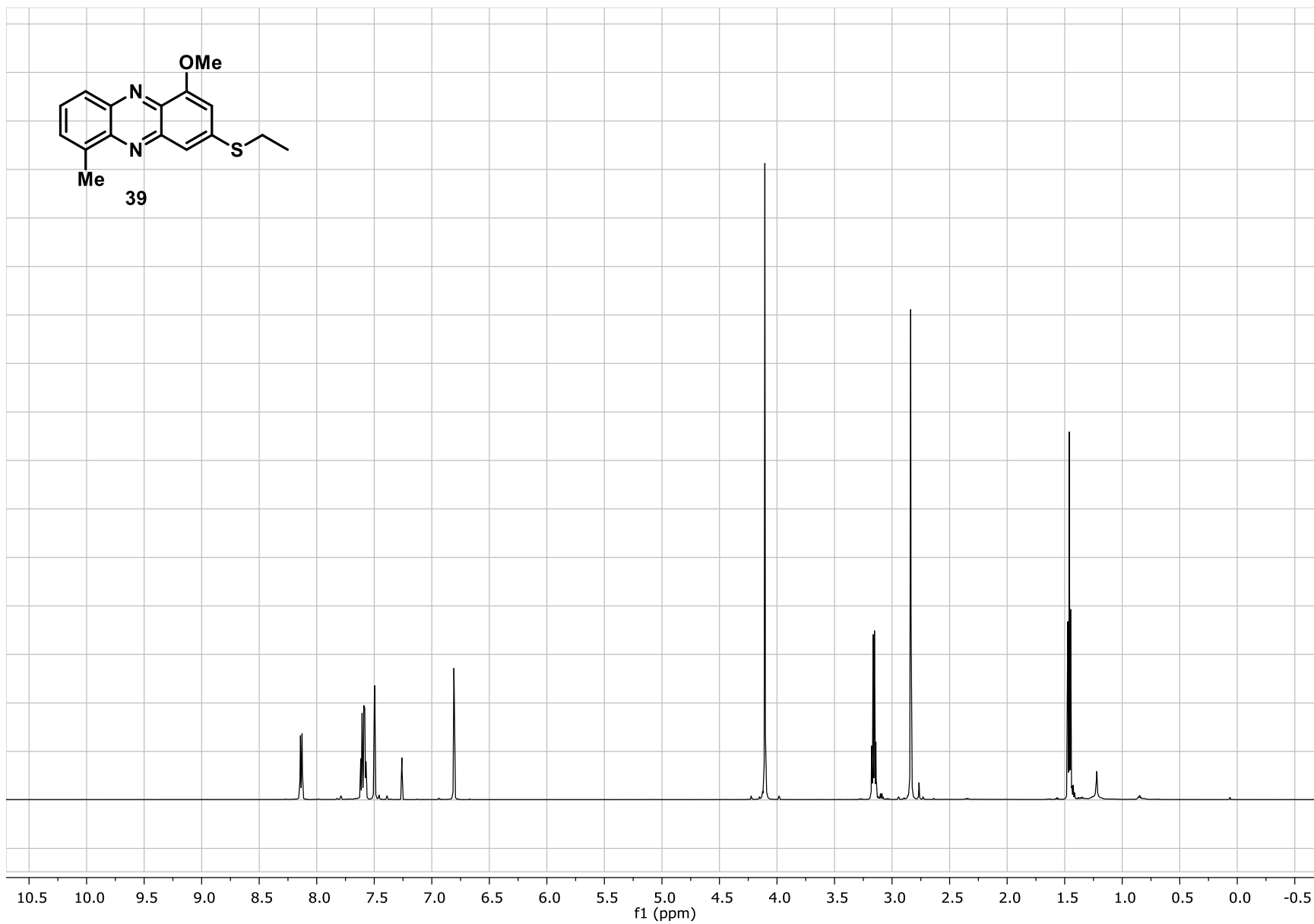
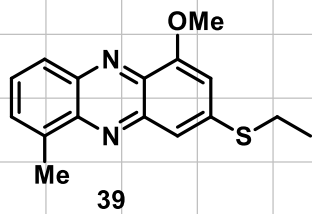


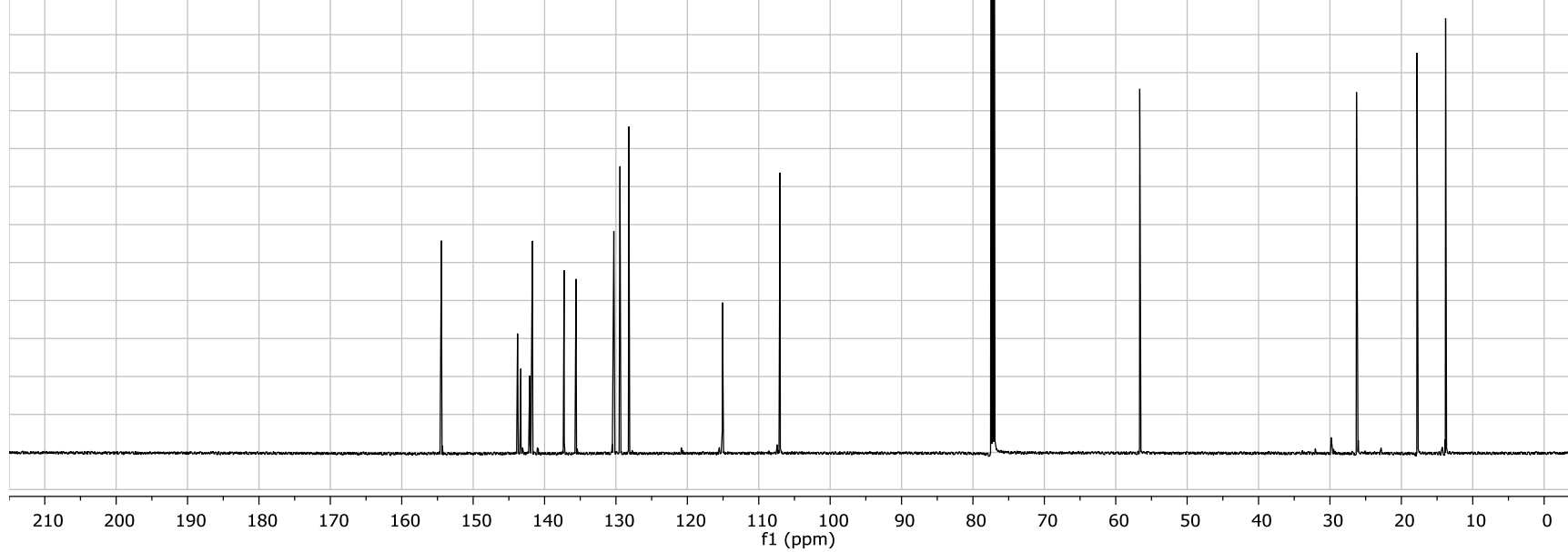
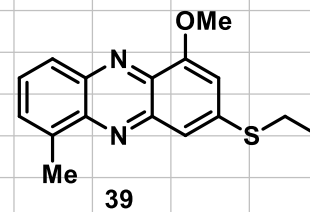
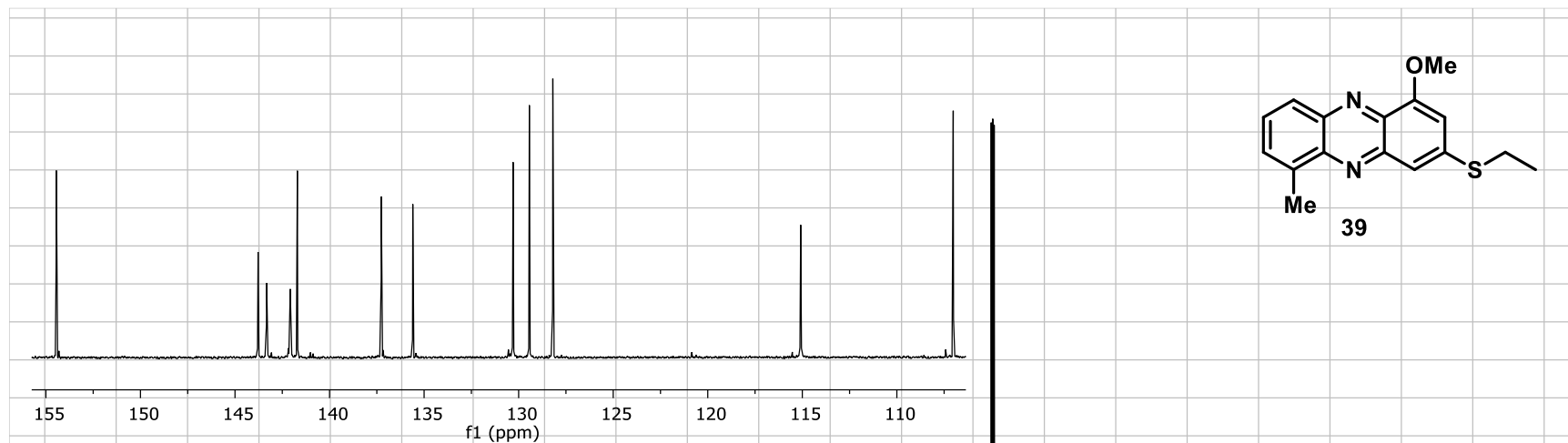


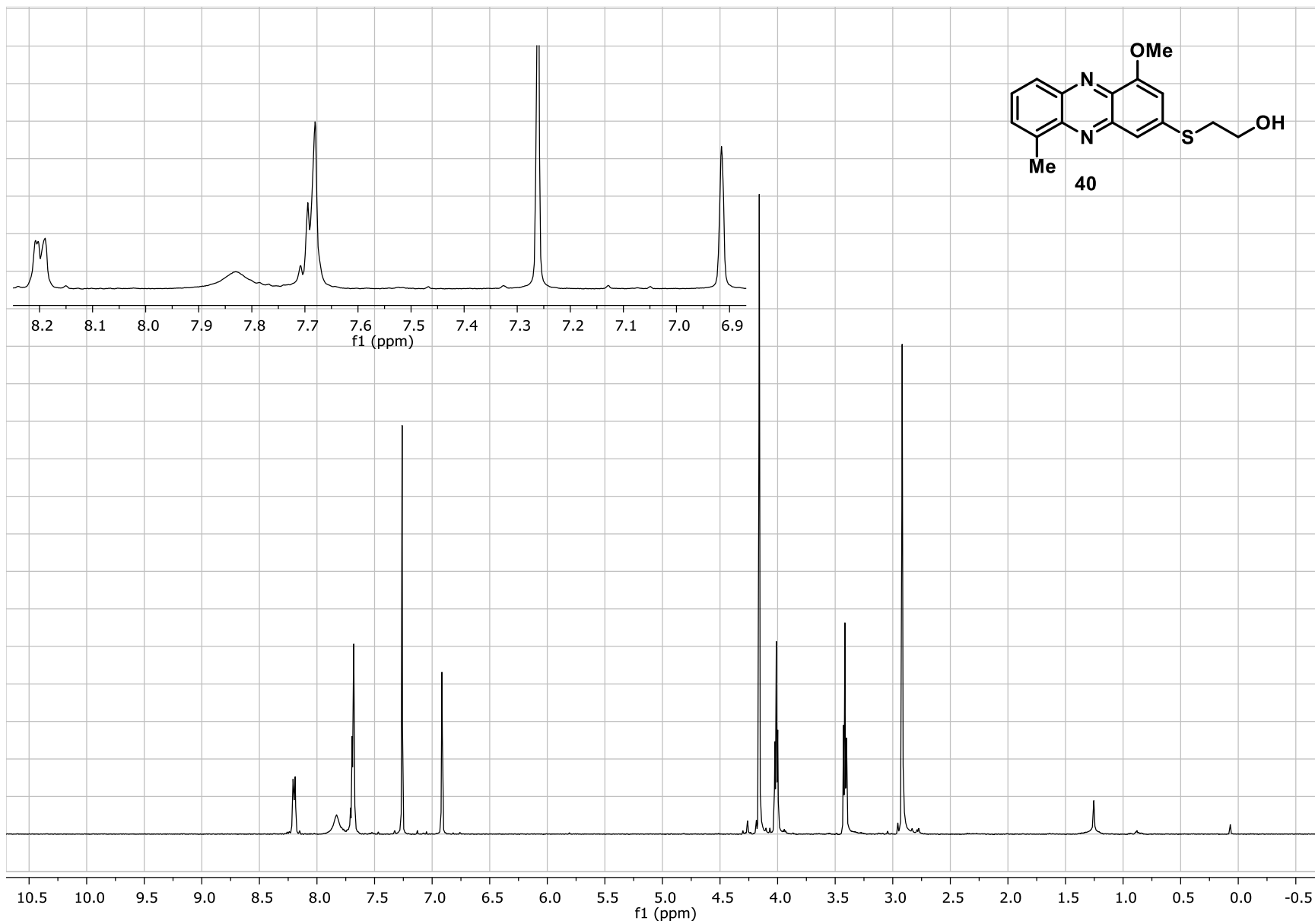




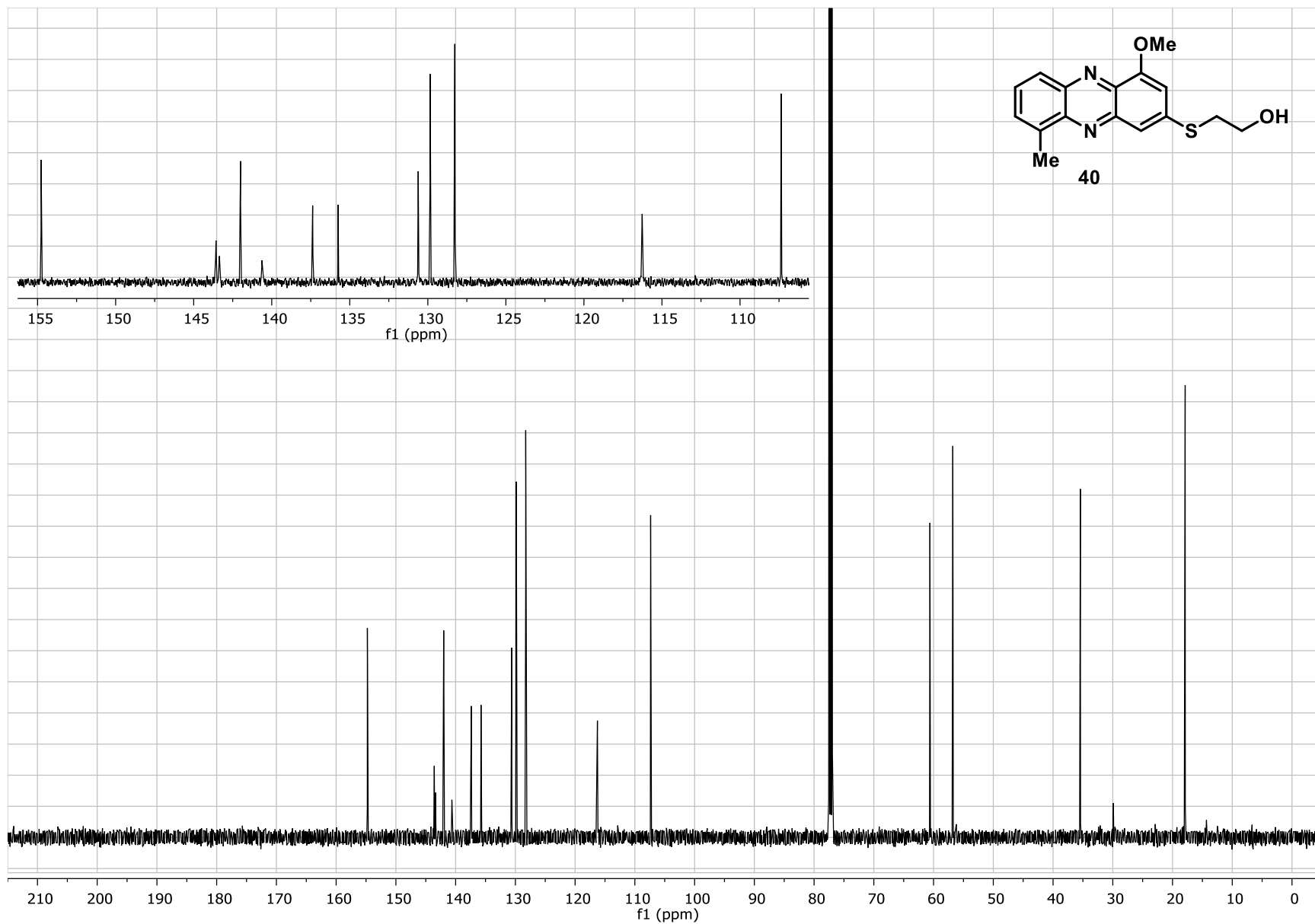


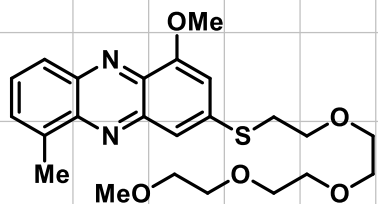




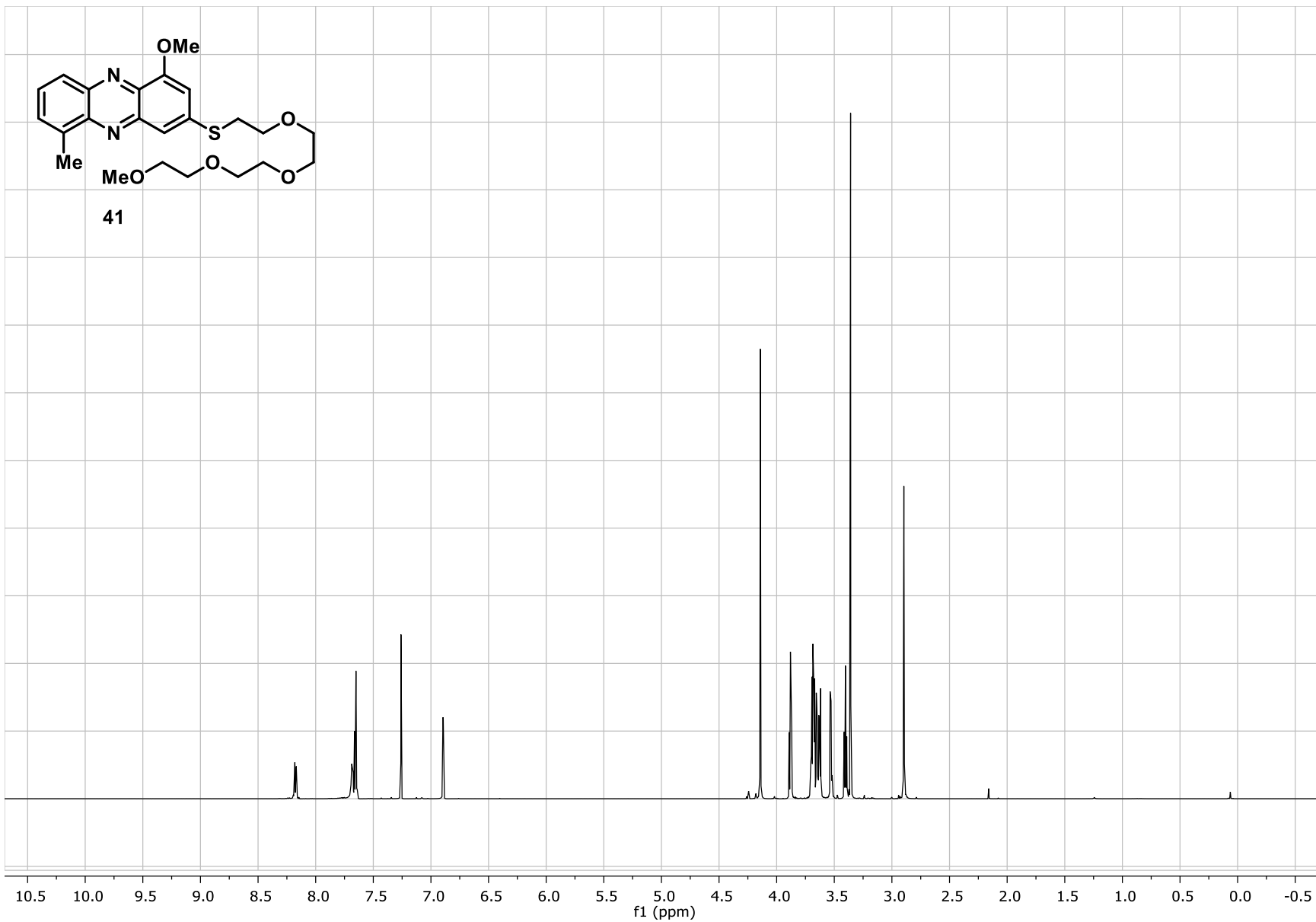




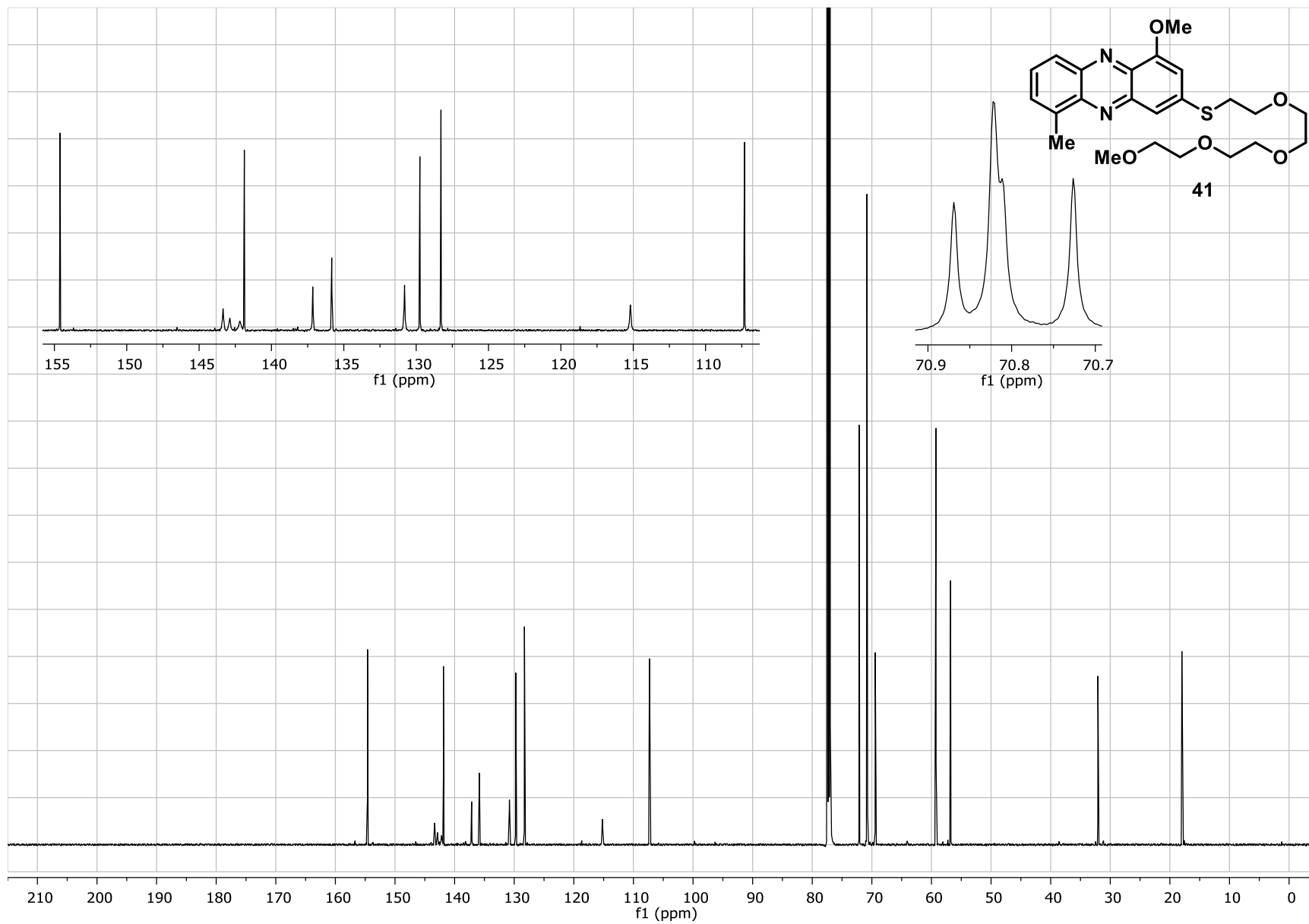


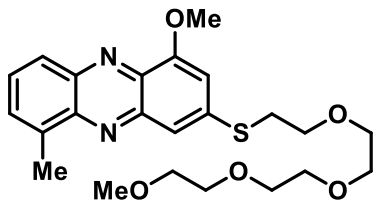


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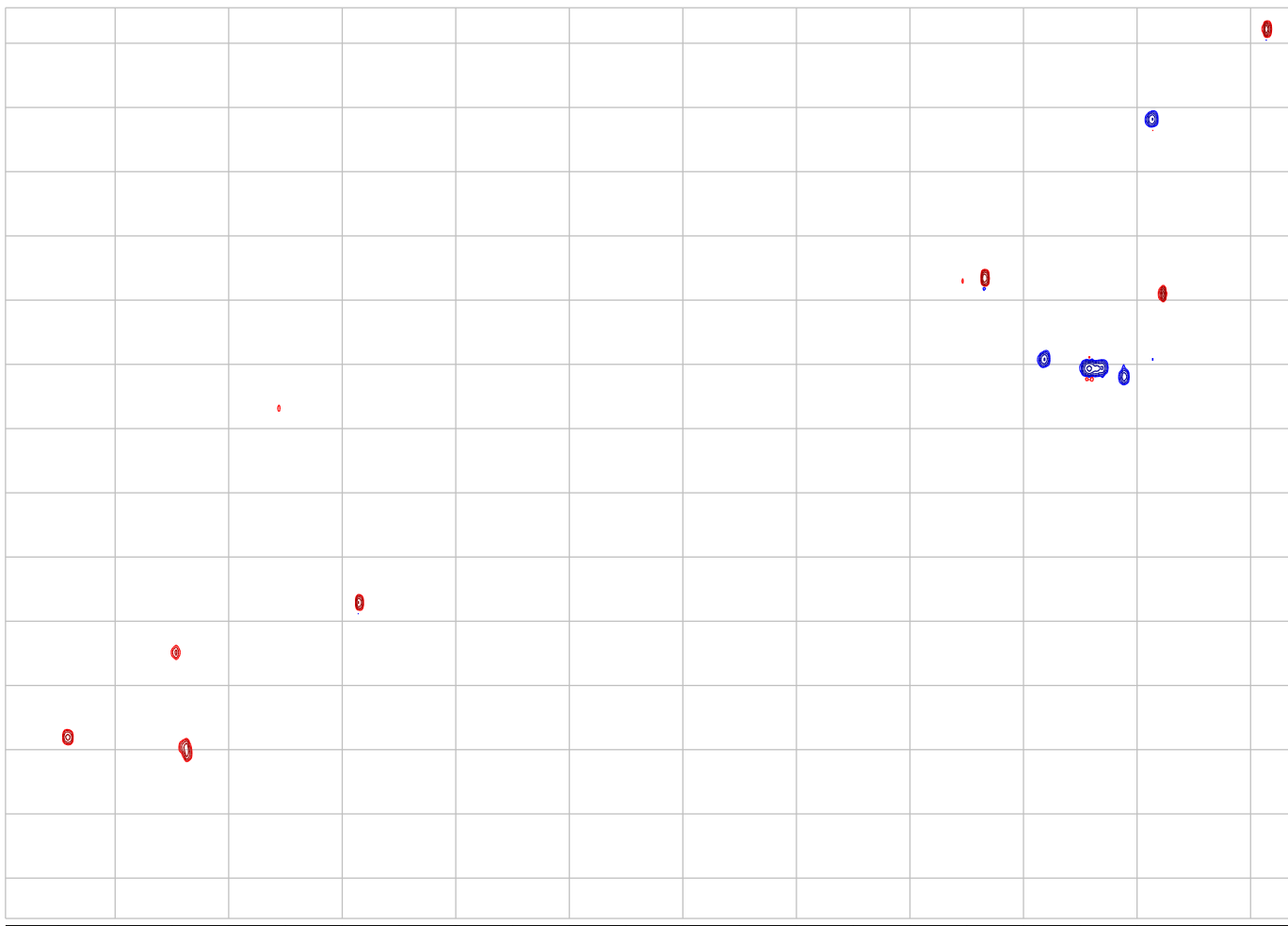
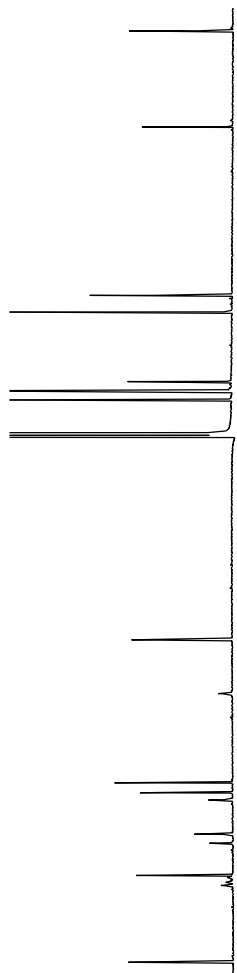


S98





41

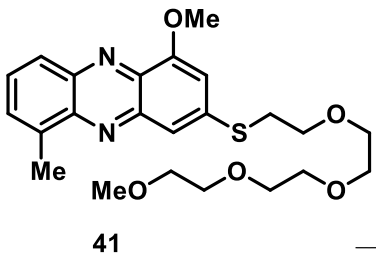


f1 (ppm)

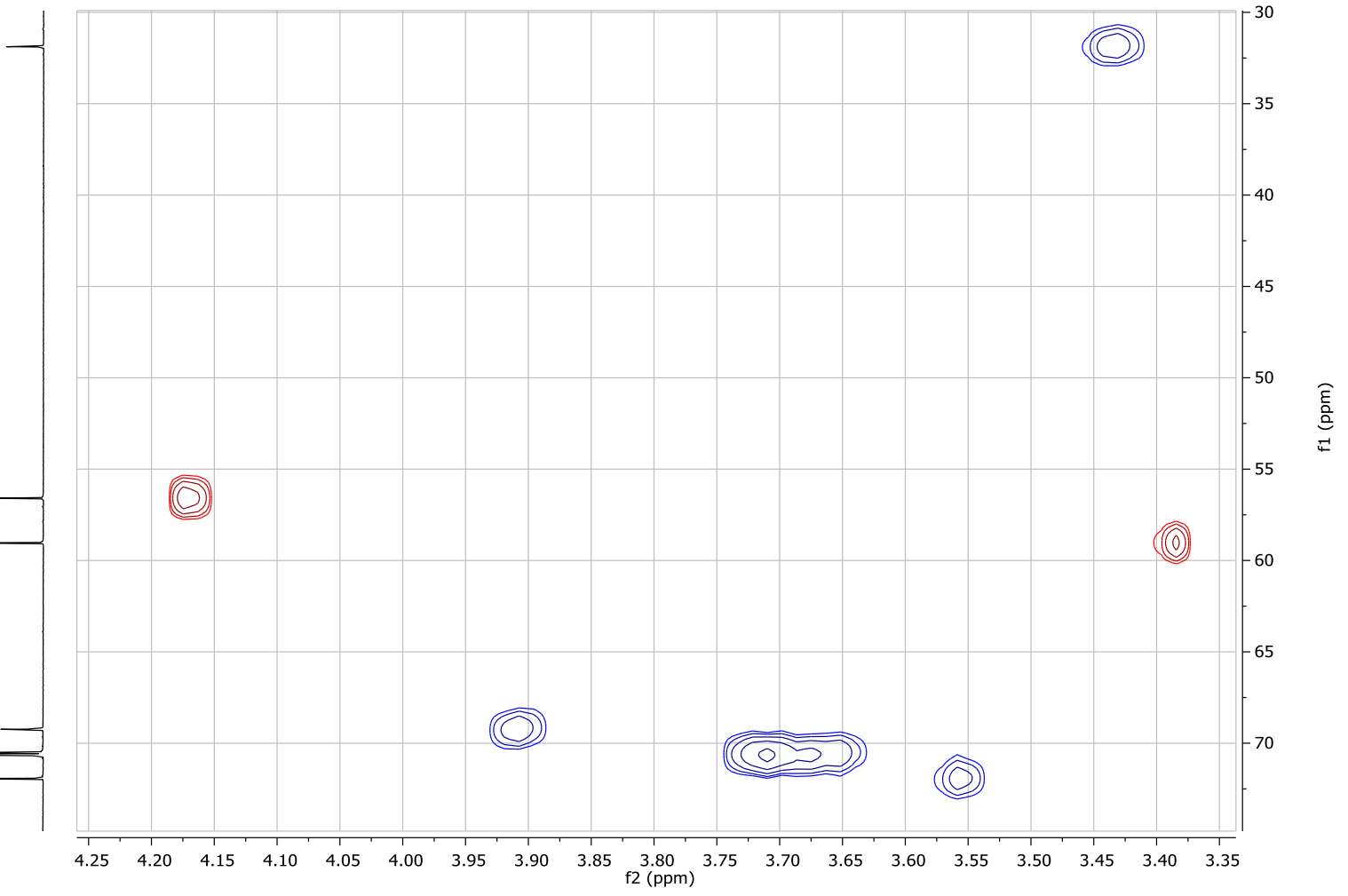
f2 (ppm)

HSQC (full)

S100

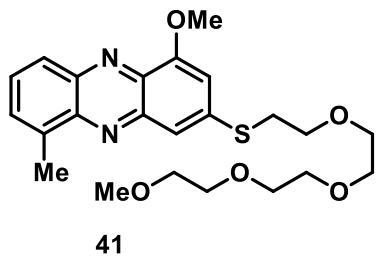


PEG protons indicated

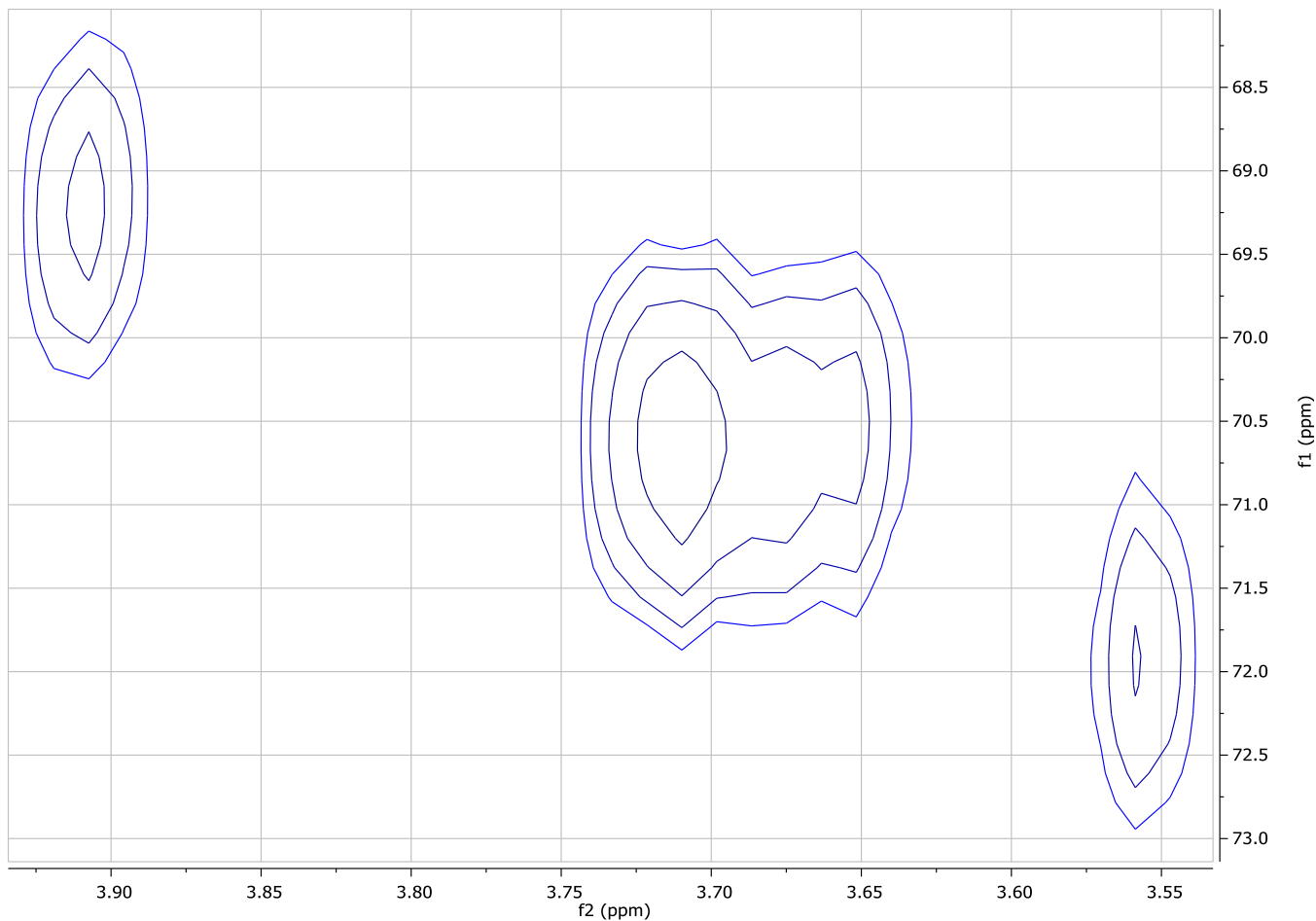
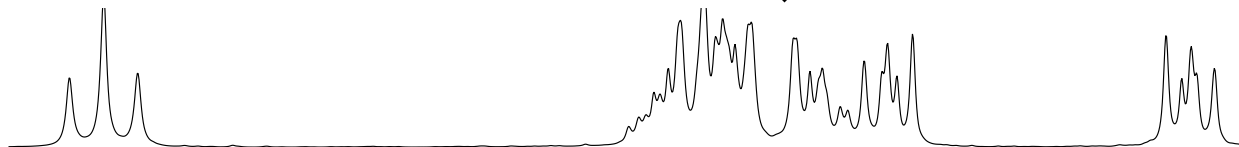


HSQC (zoomed in to relevant region bearing 19 PEG protons)

S101



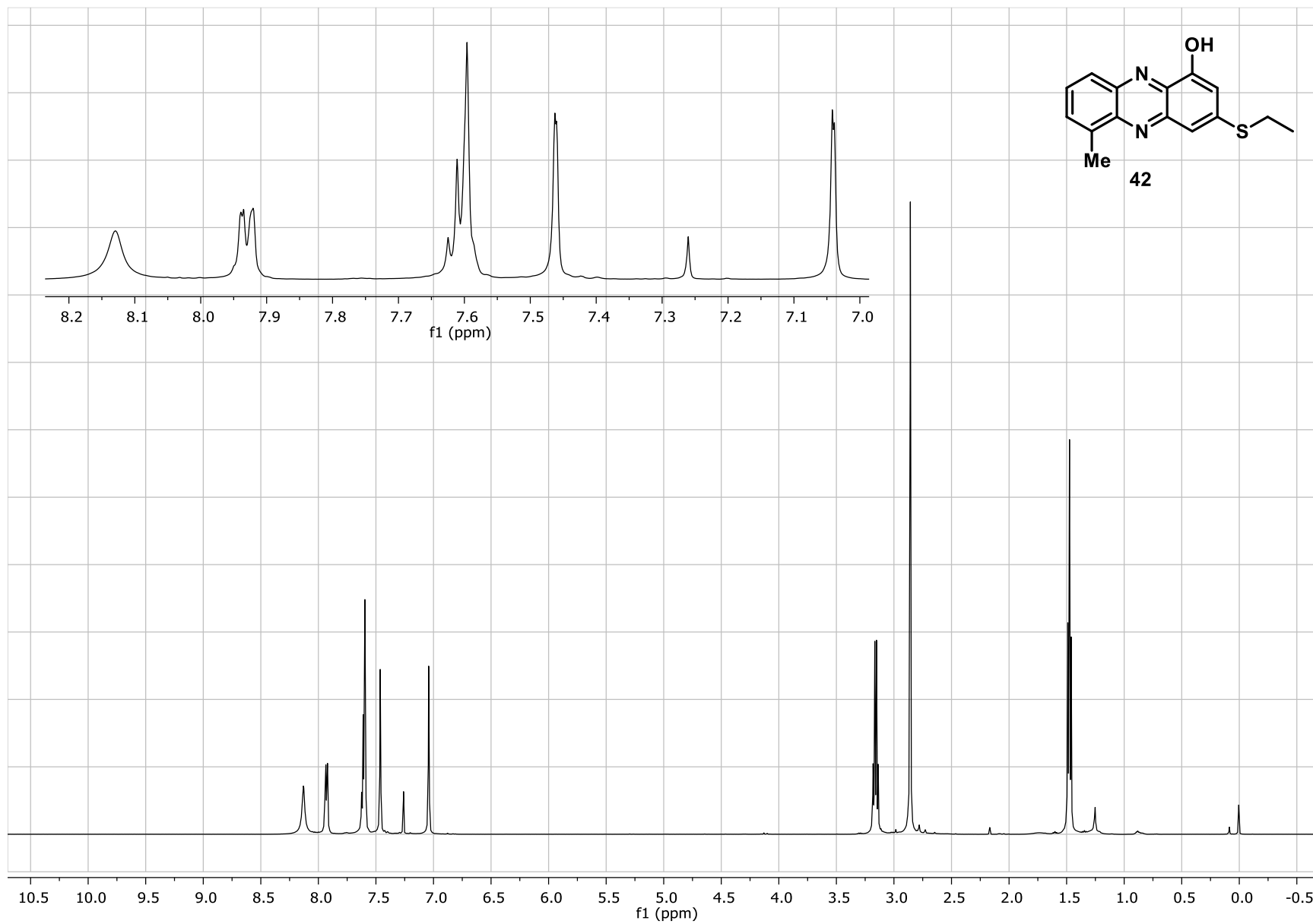
10 protons (3.72 - 3.60 ppm)



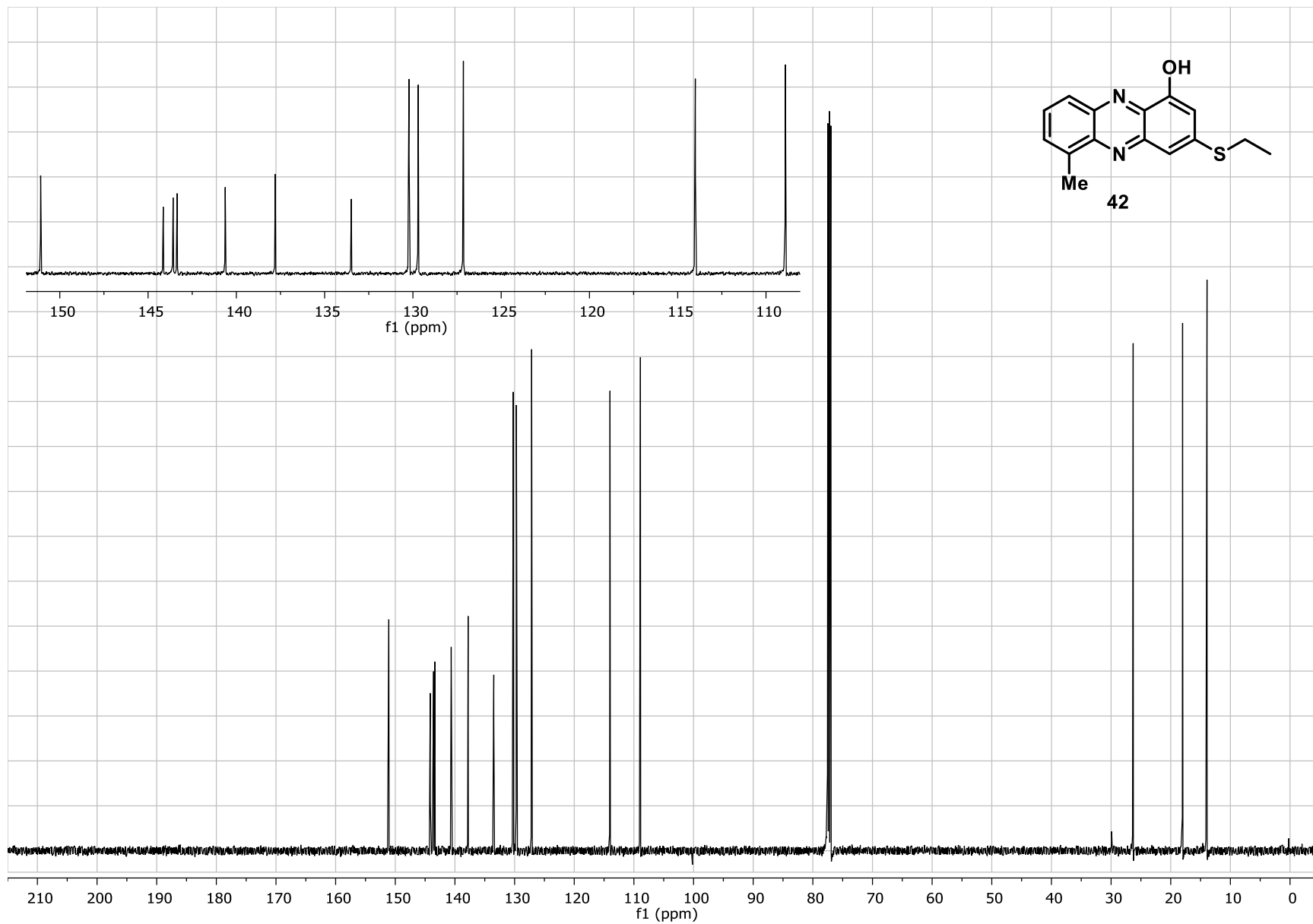
5 carbon atoms  
4 clear signals  
(1 overlap)



HSQC (zoomed in to show 10 protons on the PEG moiety on 5 carbon atoms in this region)

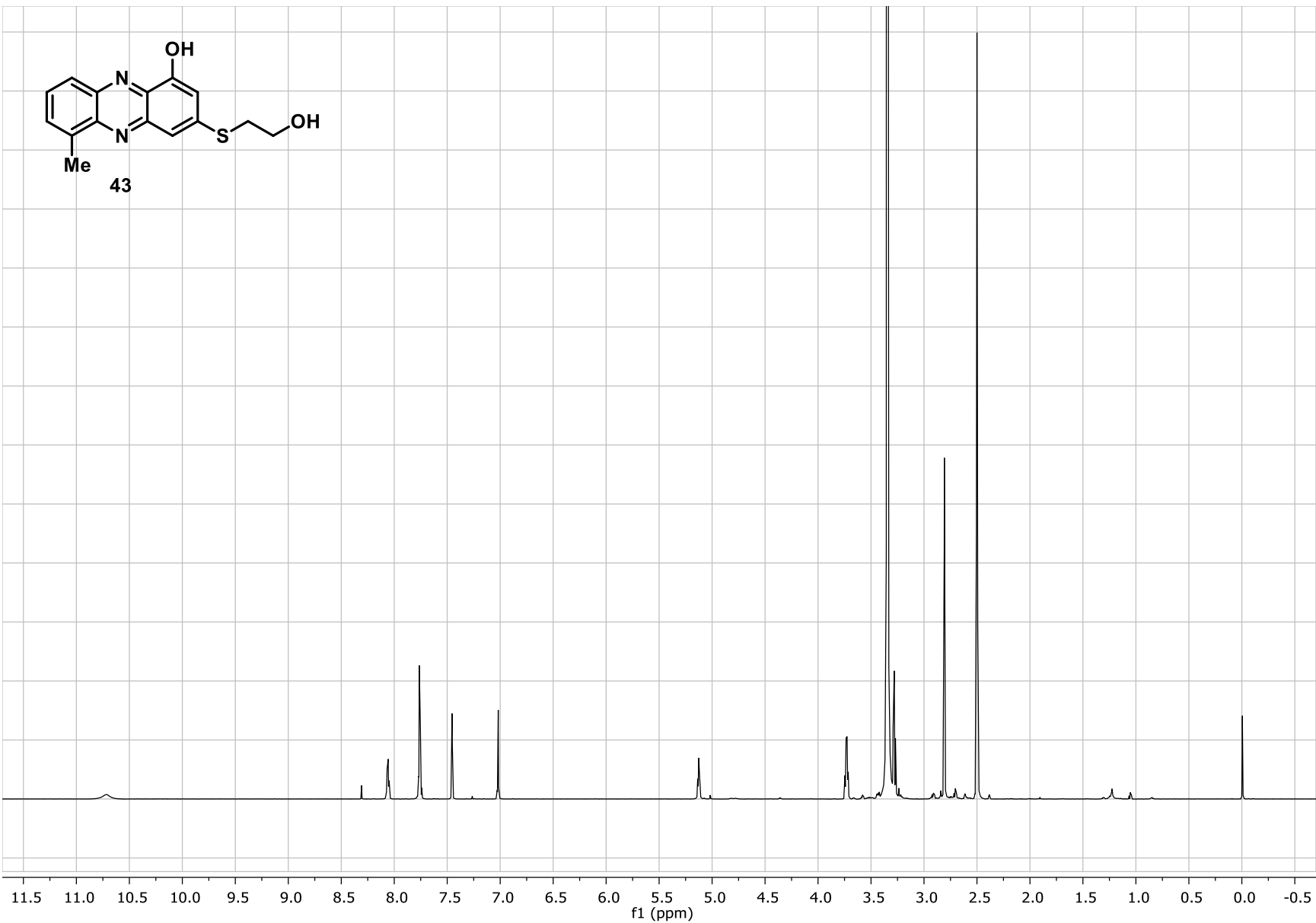
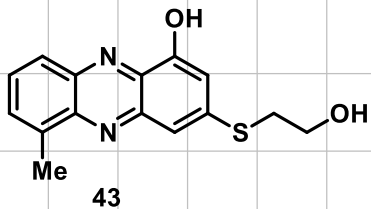


S103

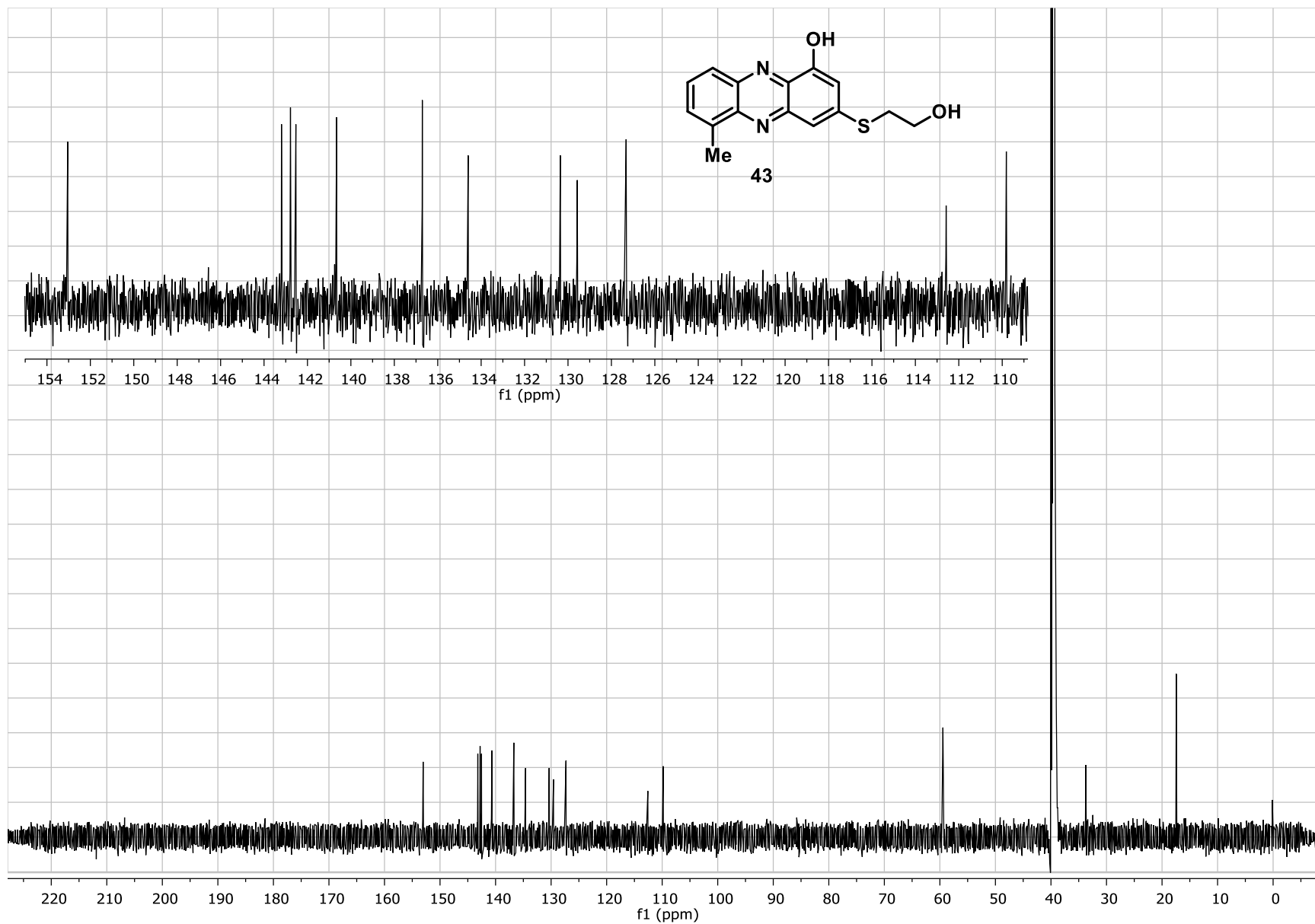


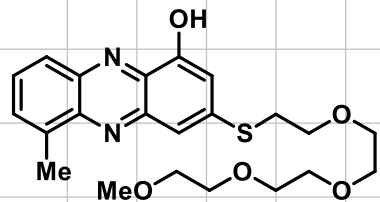
S104



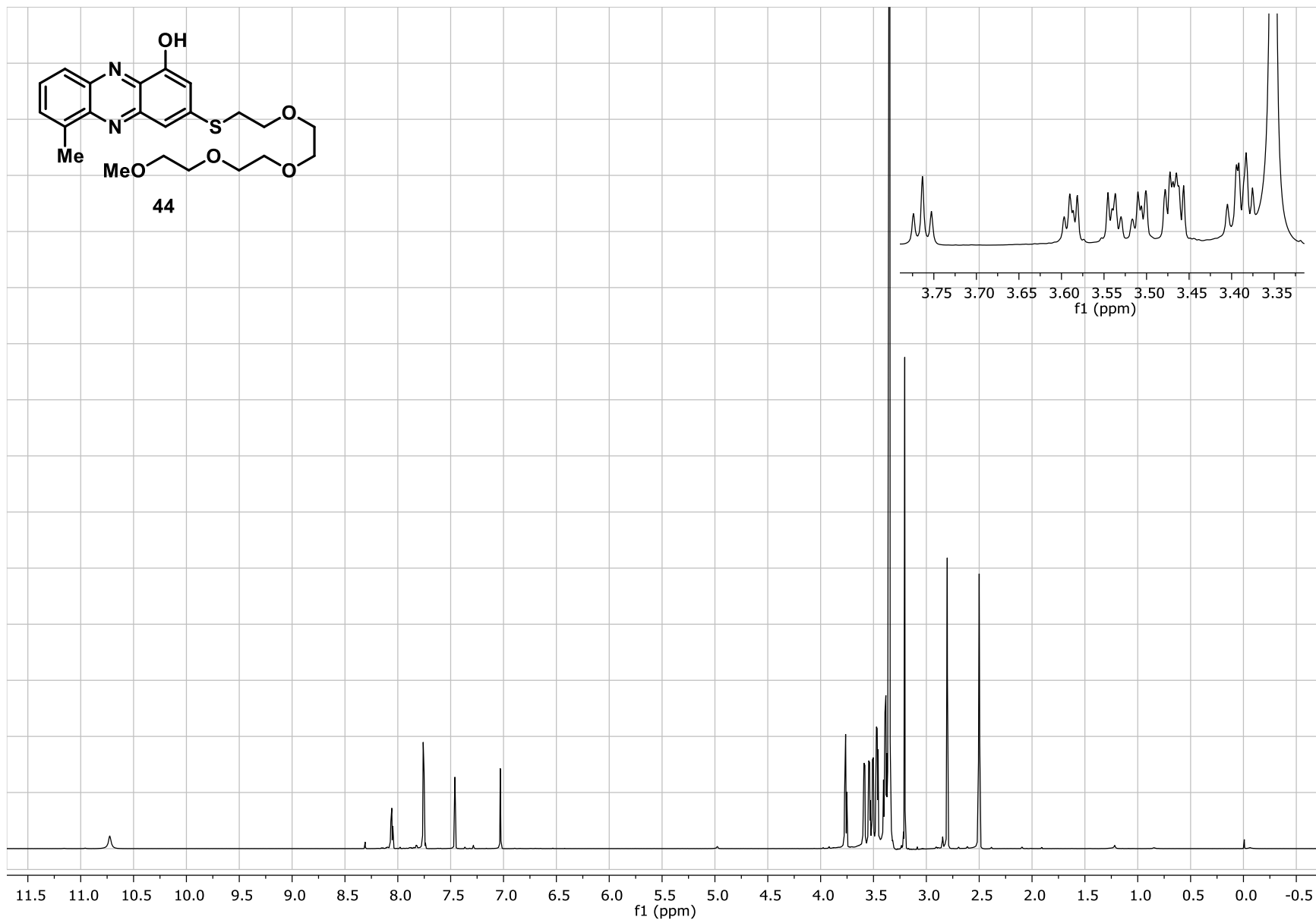


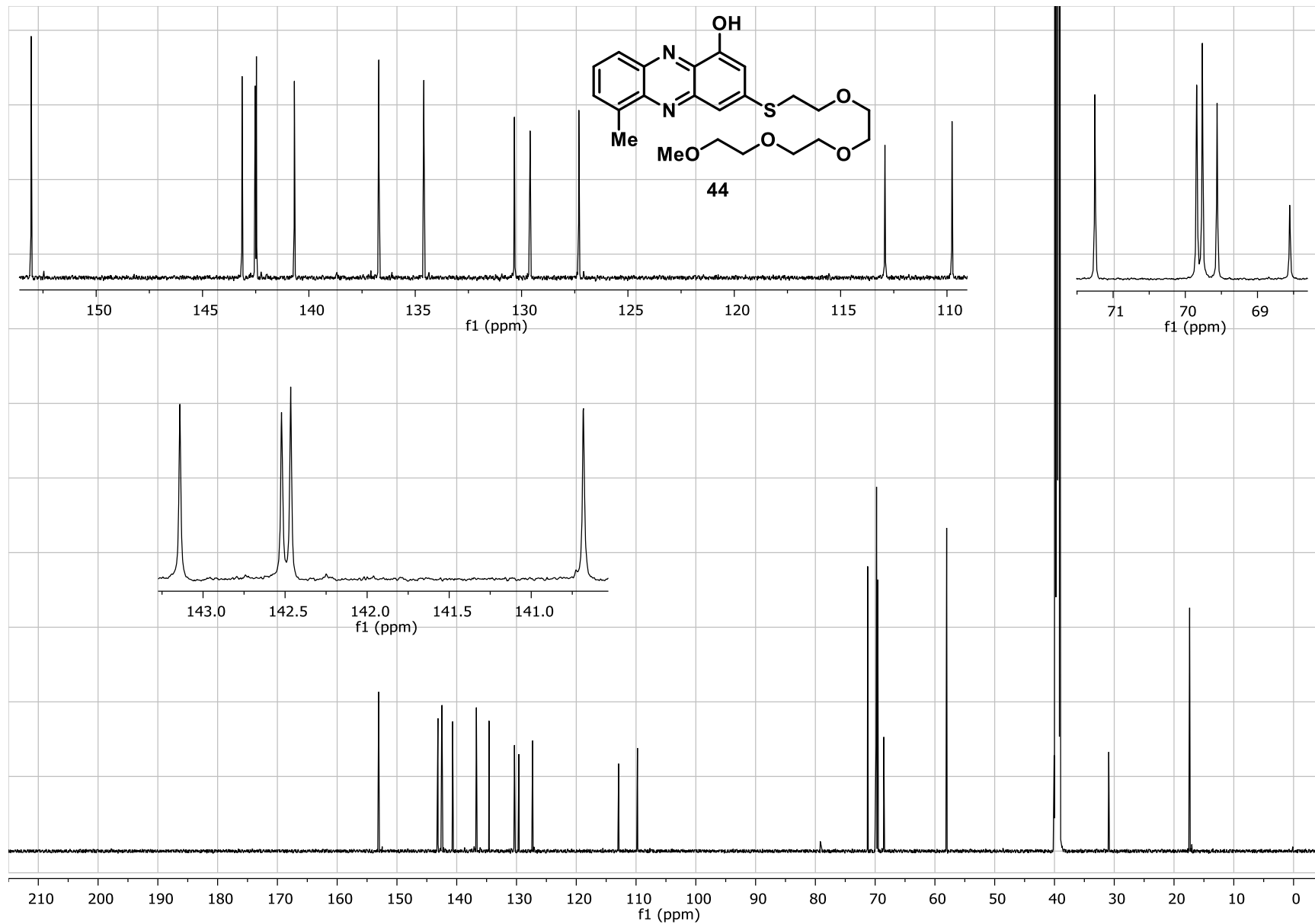
S105

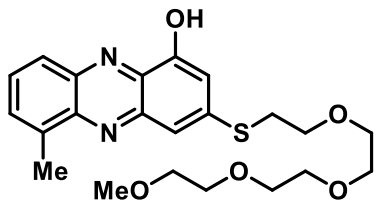




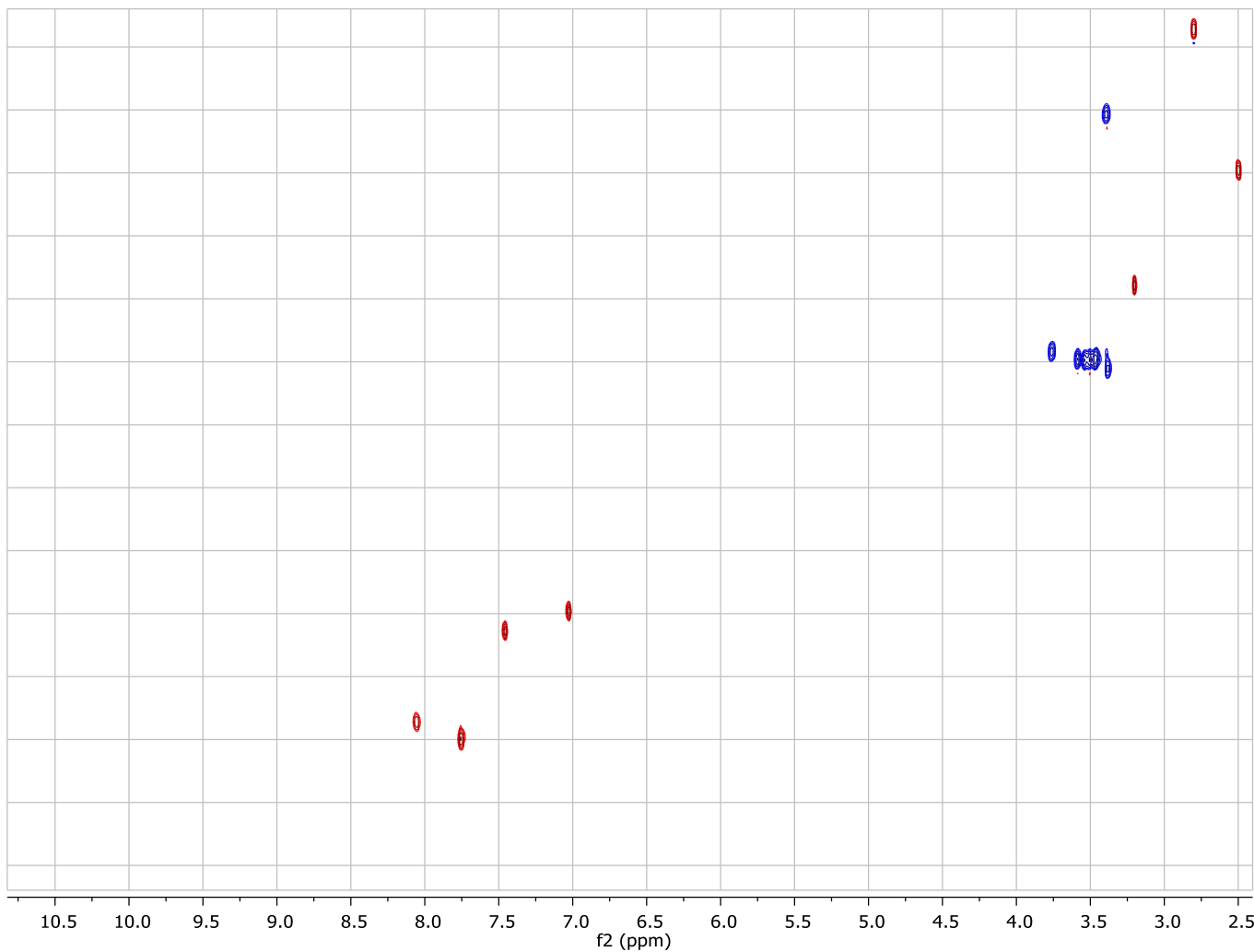
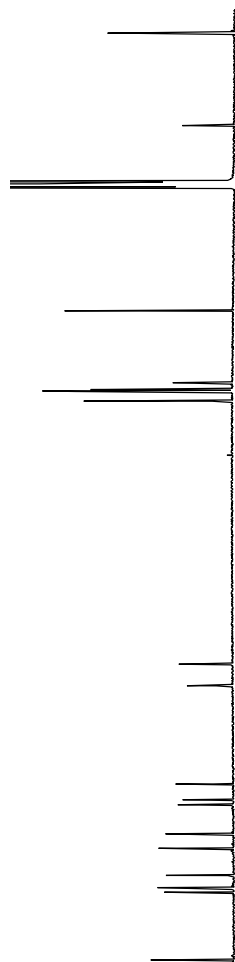
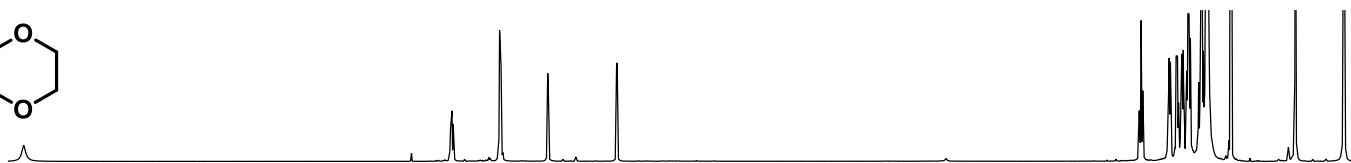
44







44

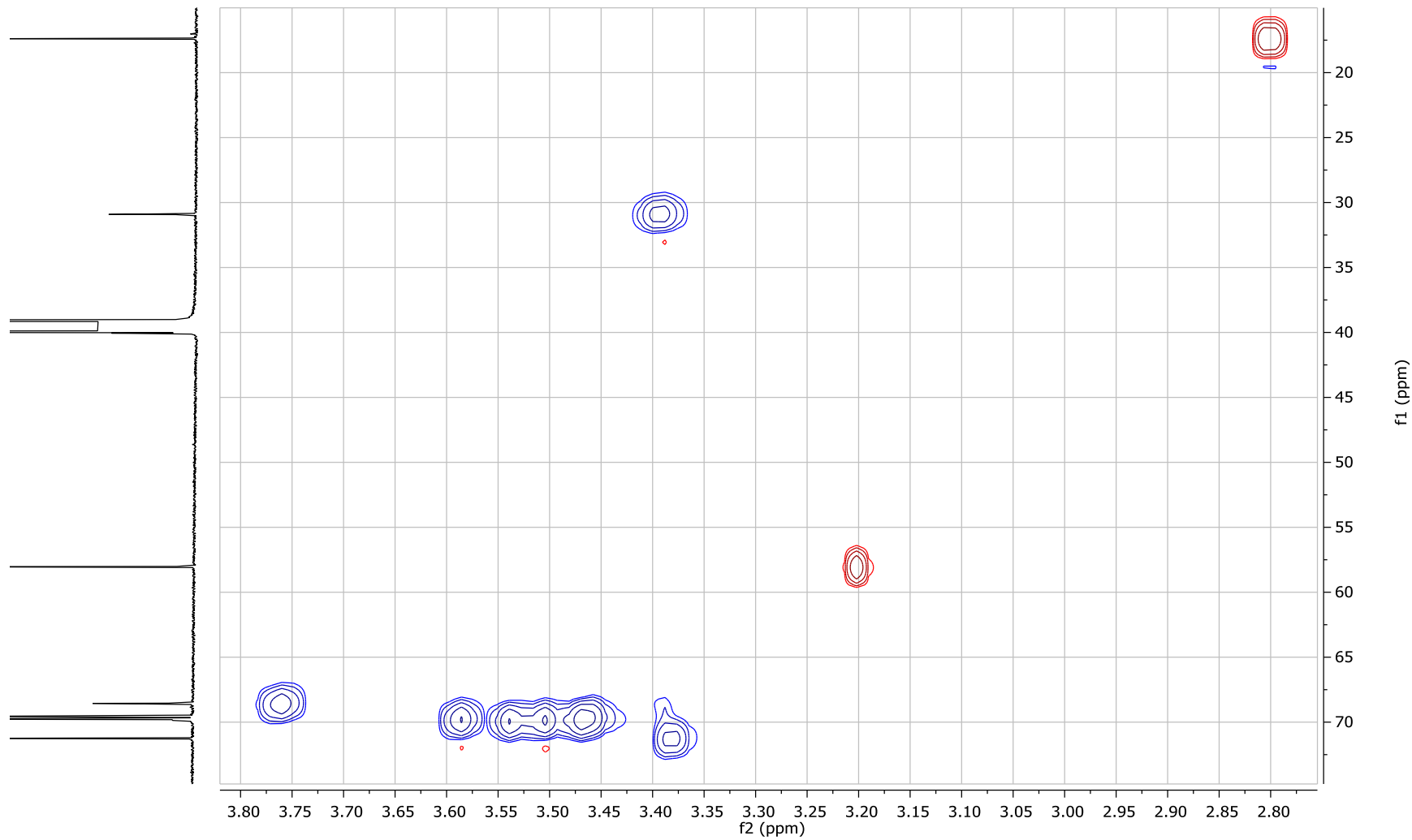
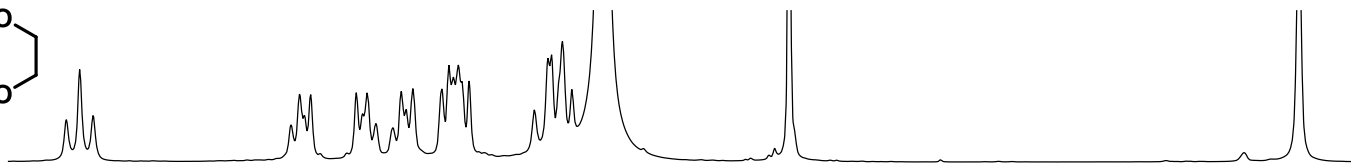
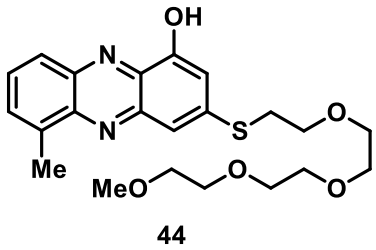


f1 (ppm)

f2 (ppm)

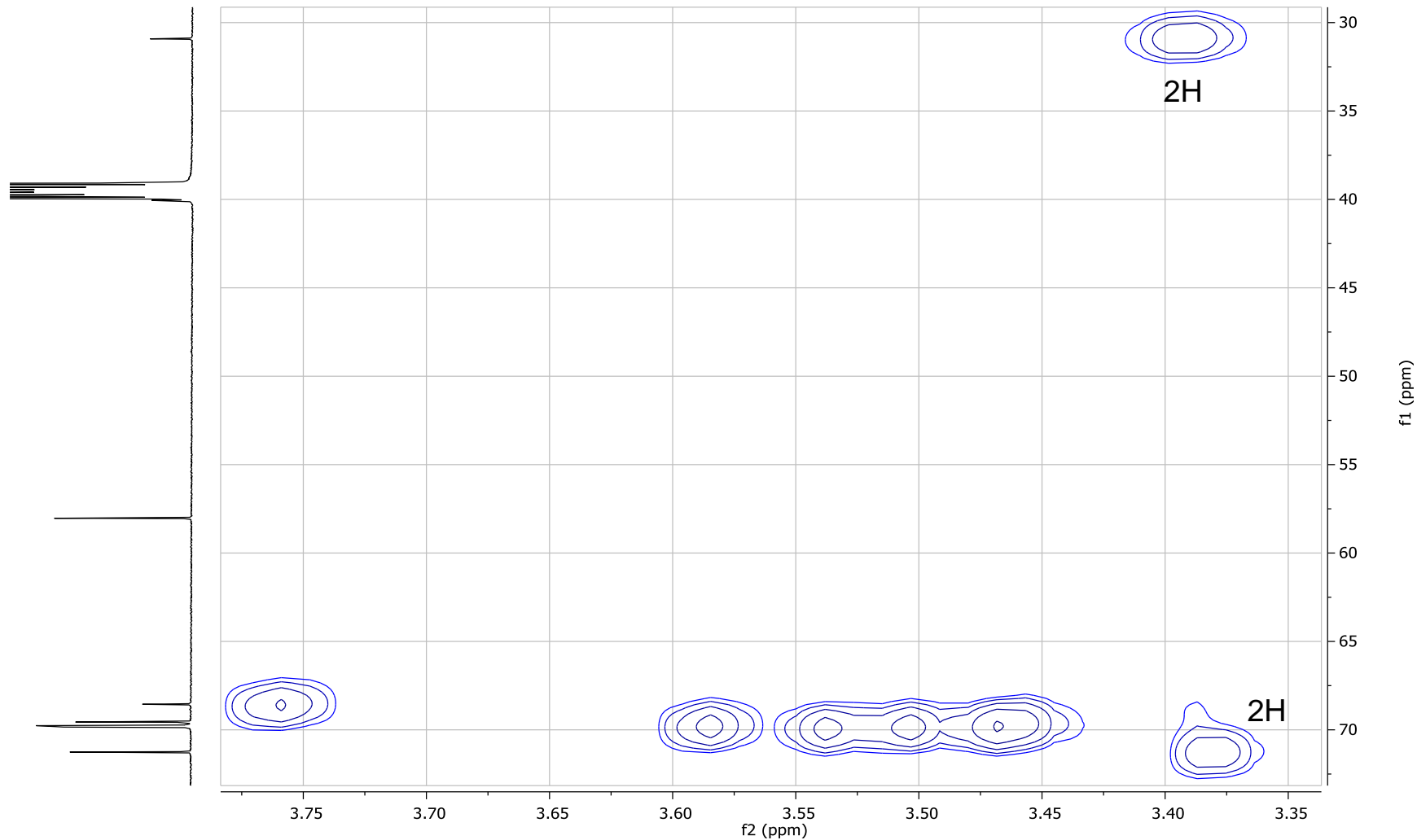
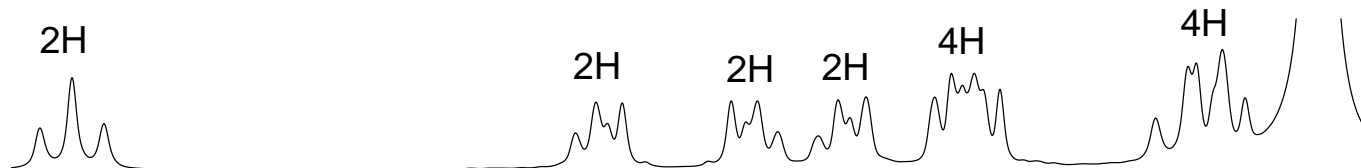
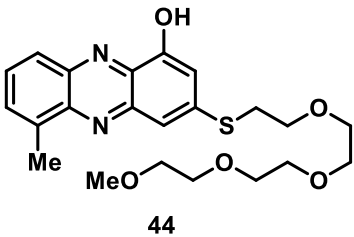
HSQC (full)

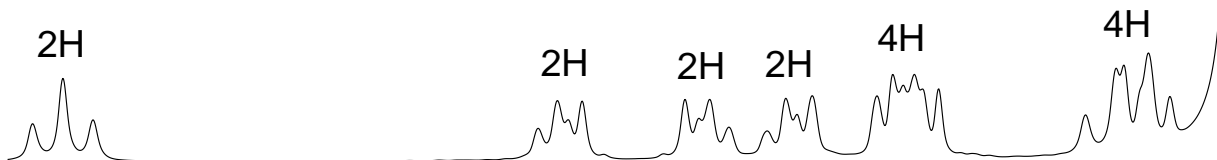
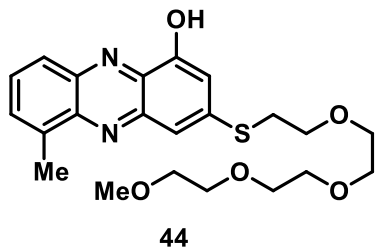
S109



HSQC (zoomed in)

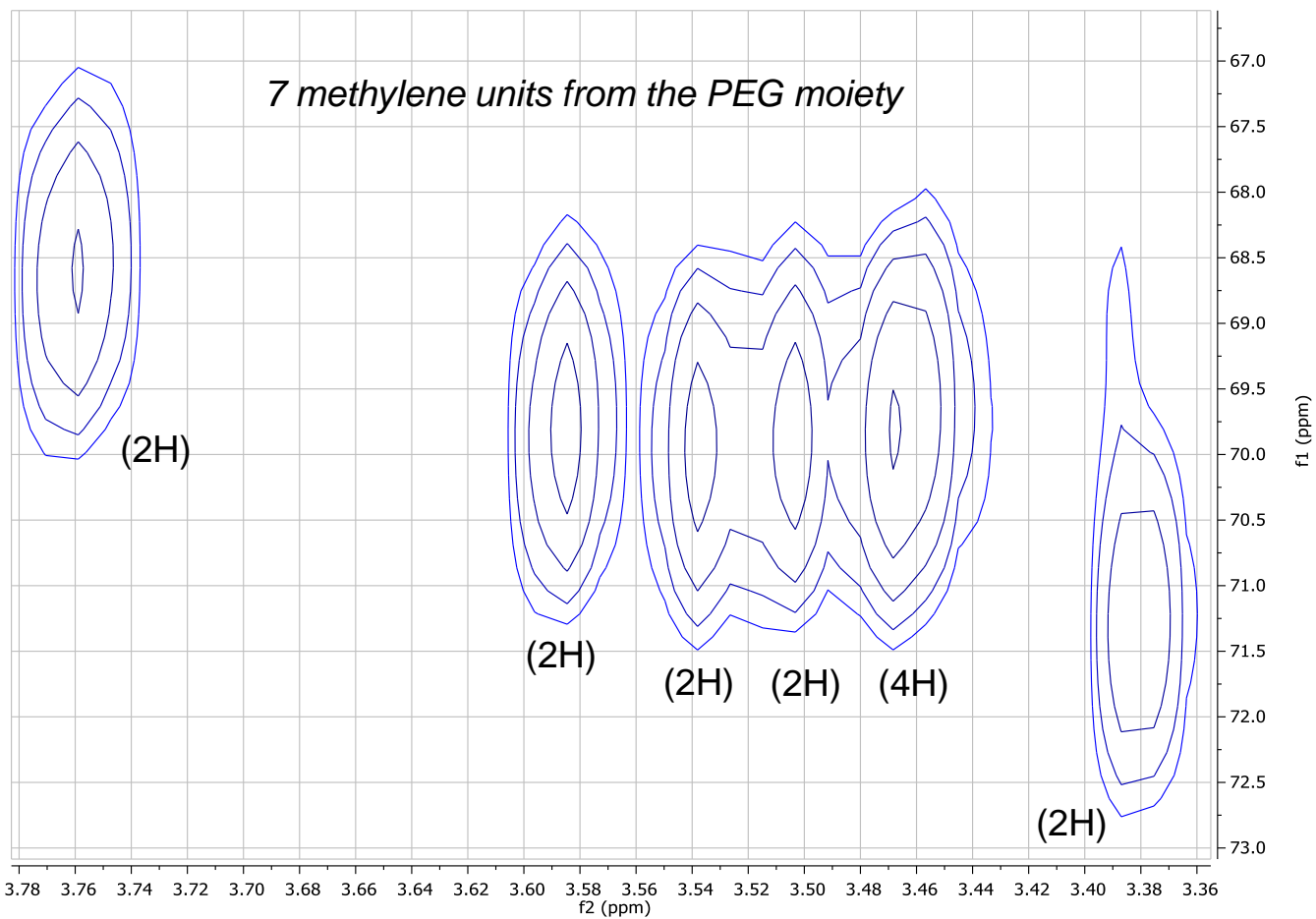
S110



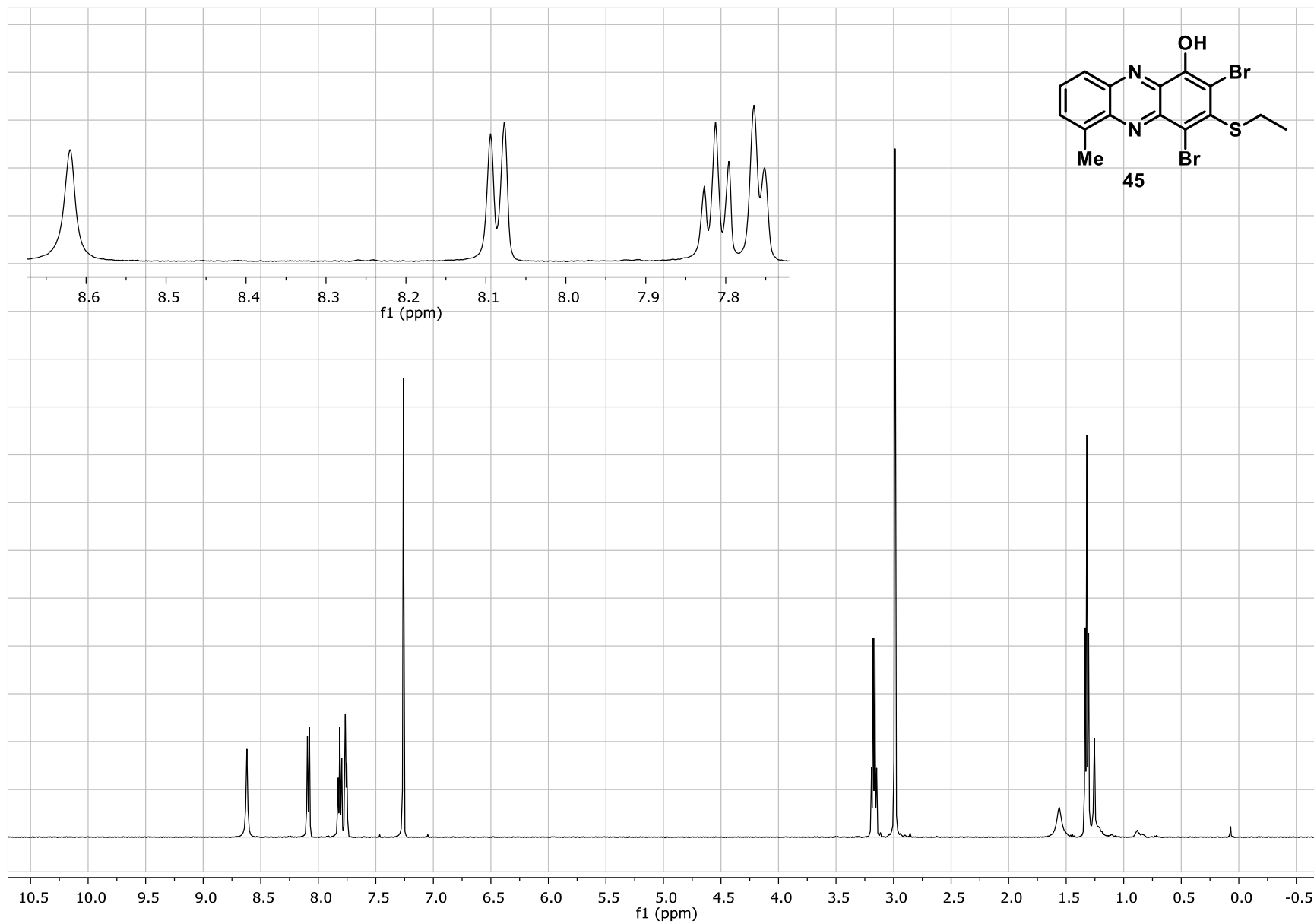


7 carbon atoms  
14 protons

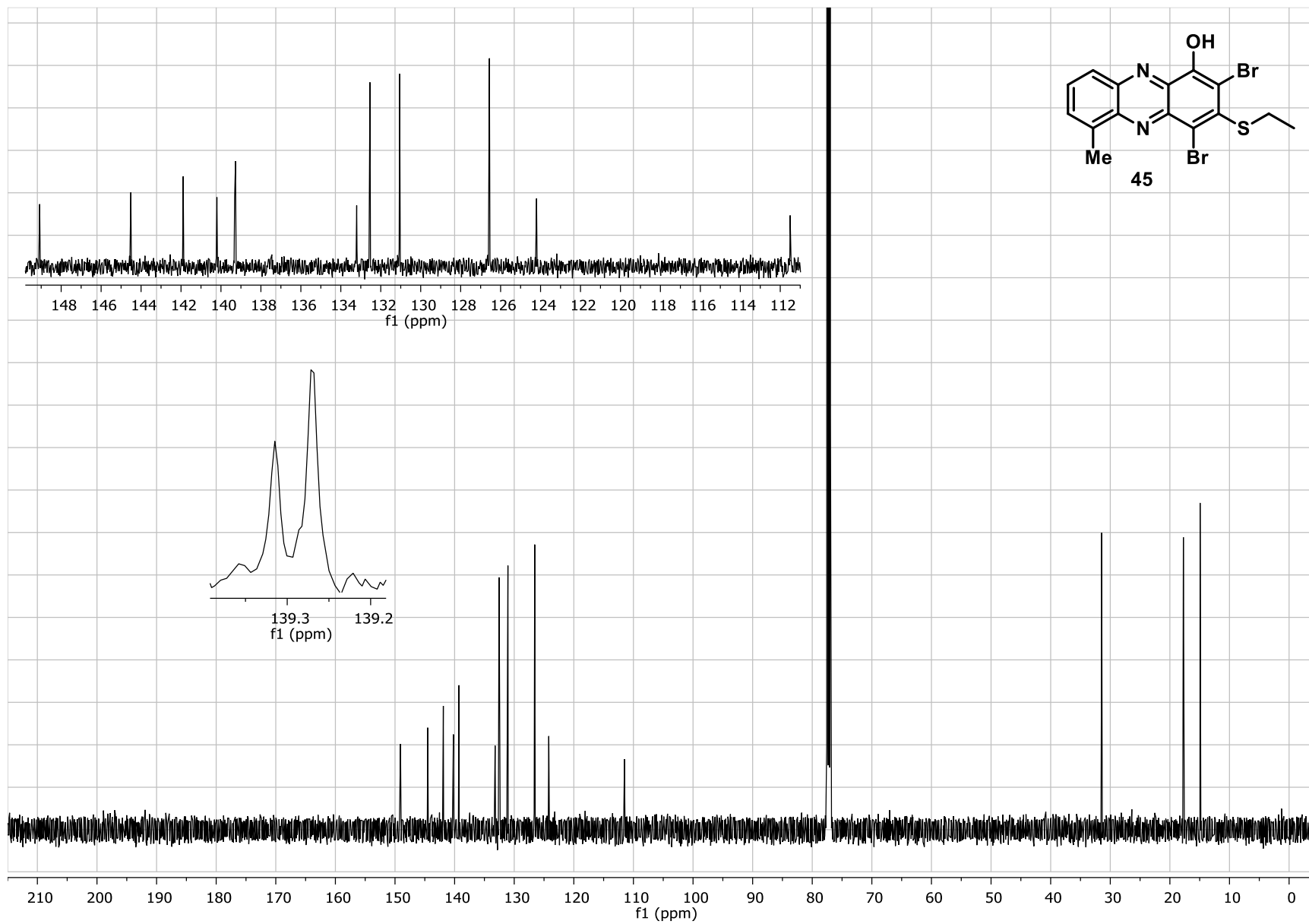
5 carbon signals  
(2 missing due to overlap ~70 ppm)



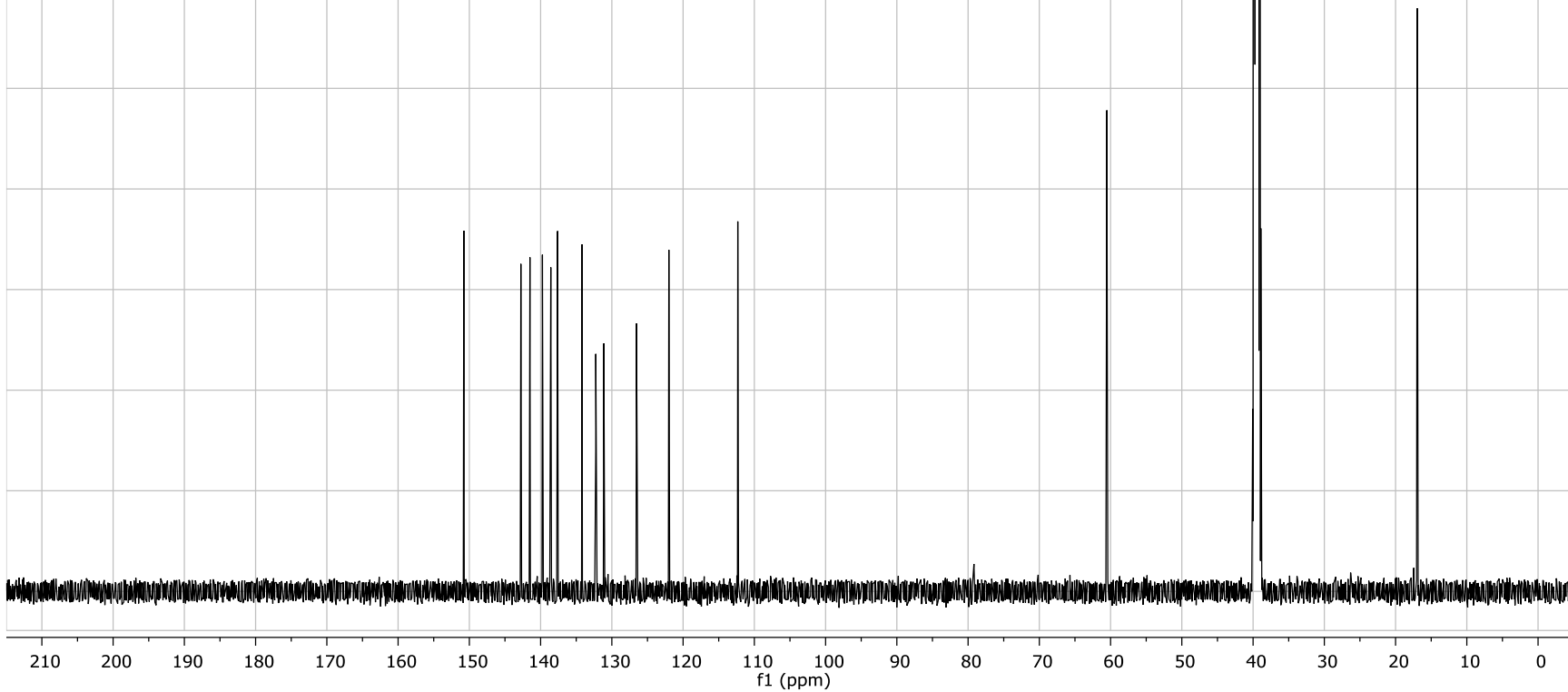
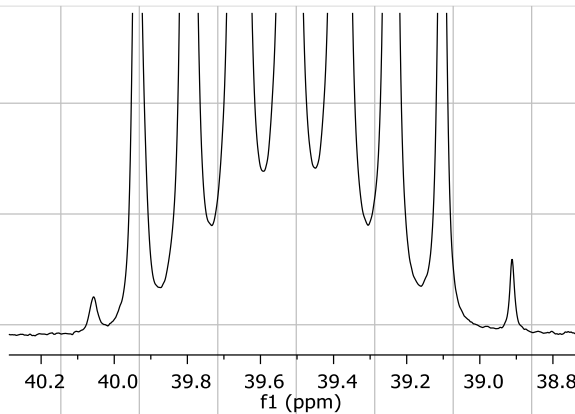
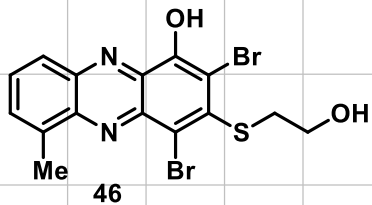


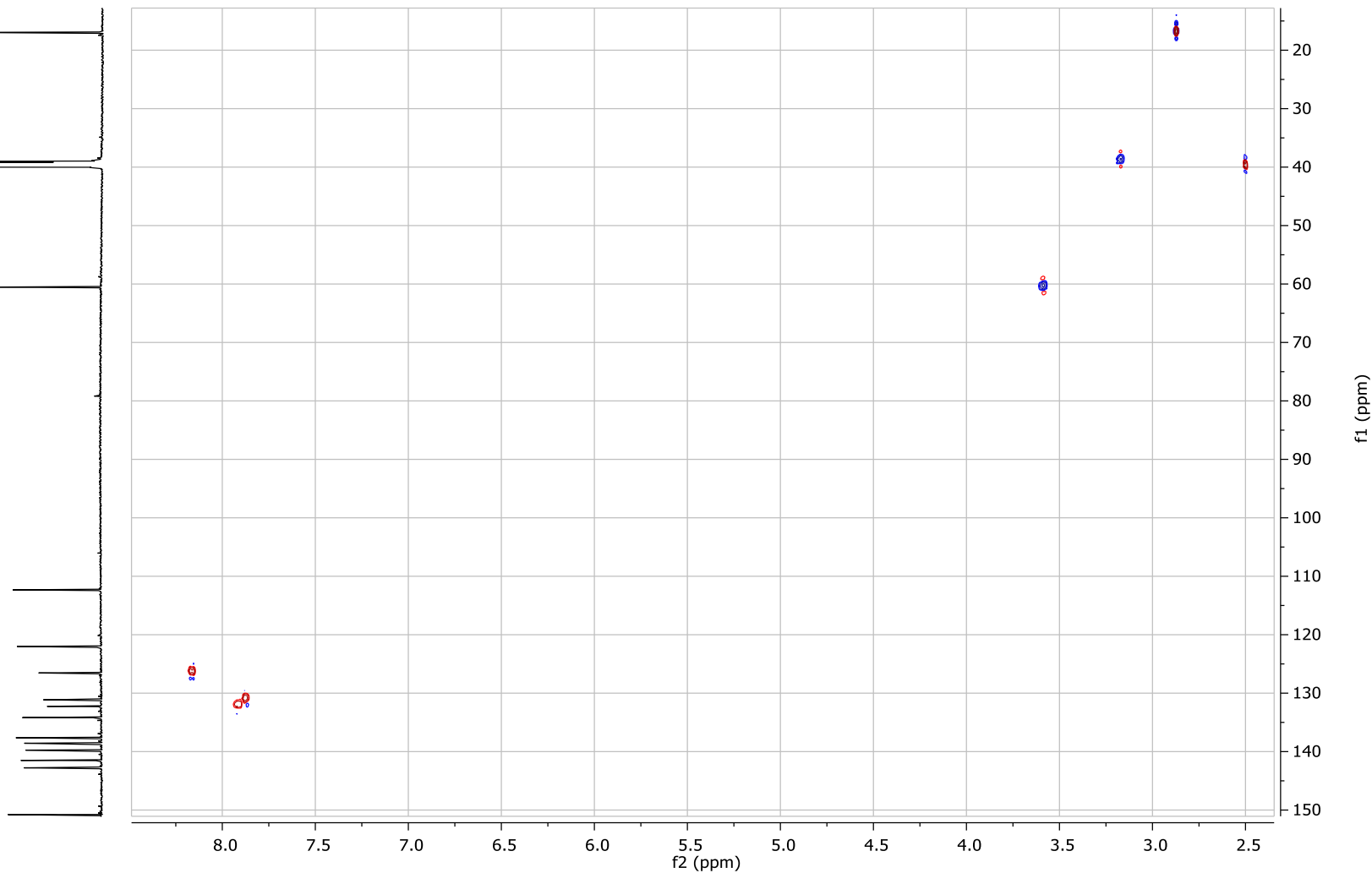
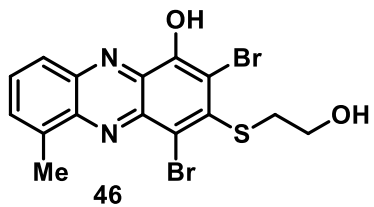


S113



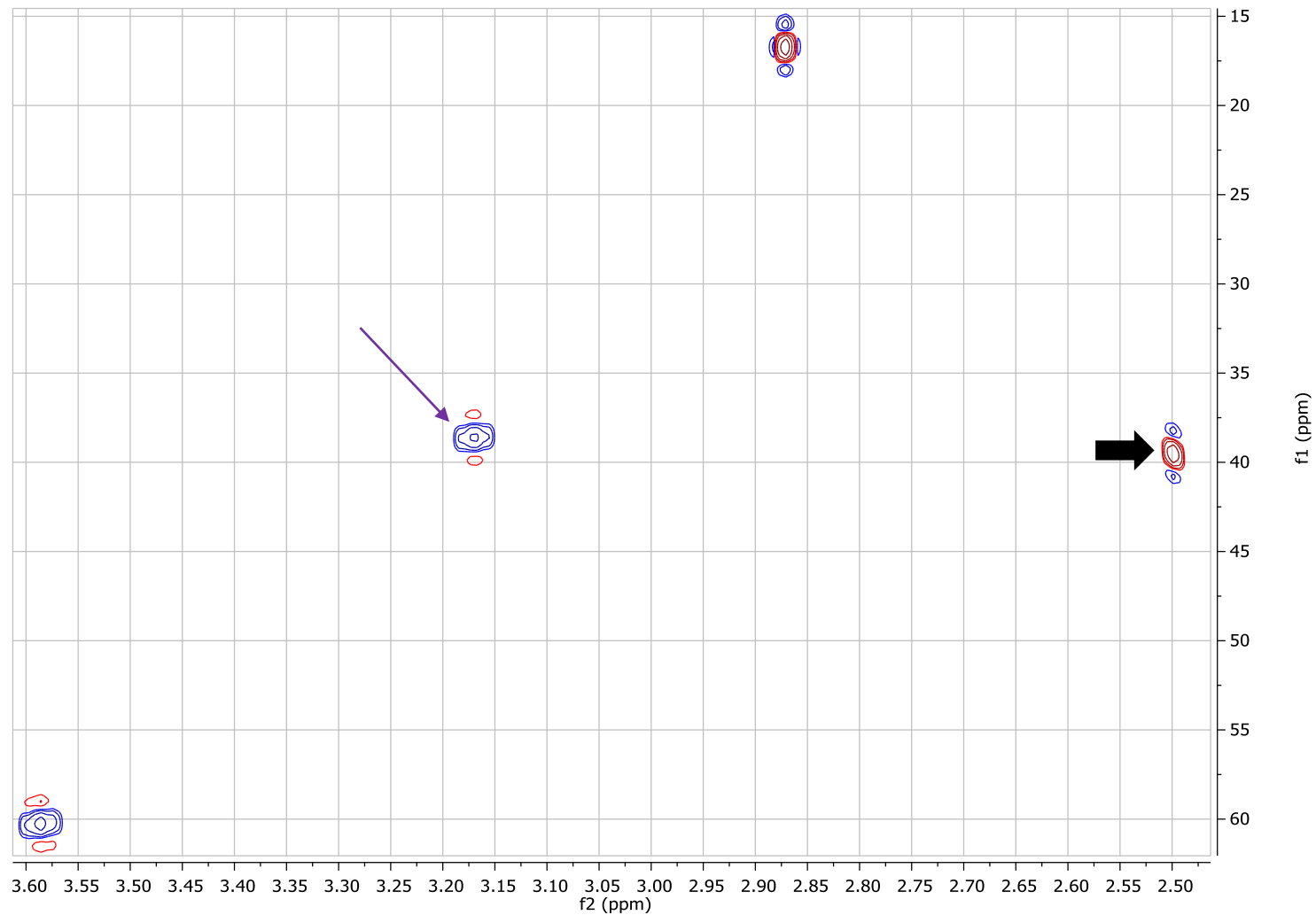
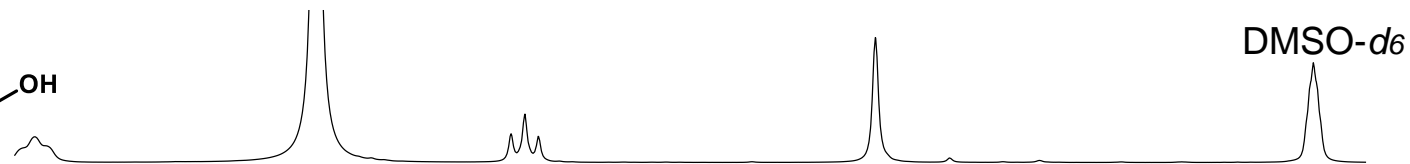
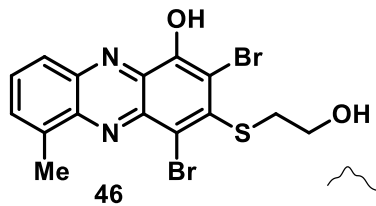






HSQC (full)

S117

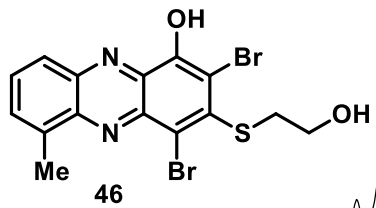


Carbon Signal  
at 38.9 ppm

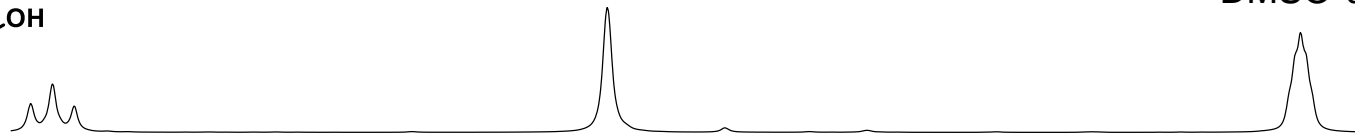
DMSO-*d*<sub>6</sub>

f1 (ppm)

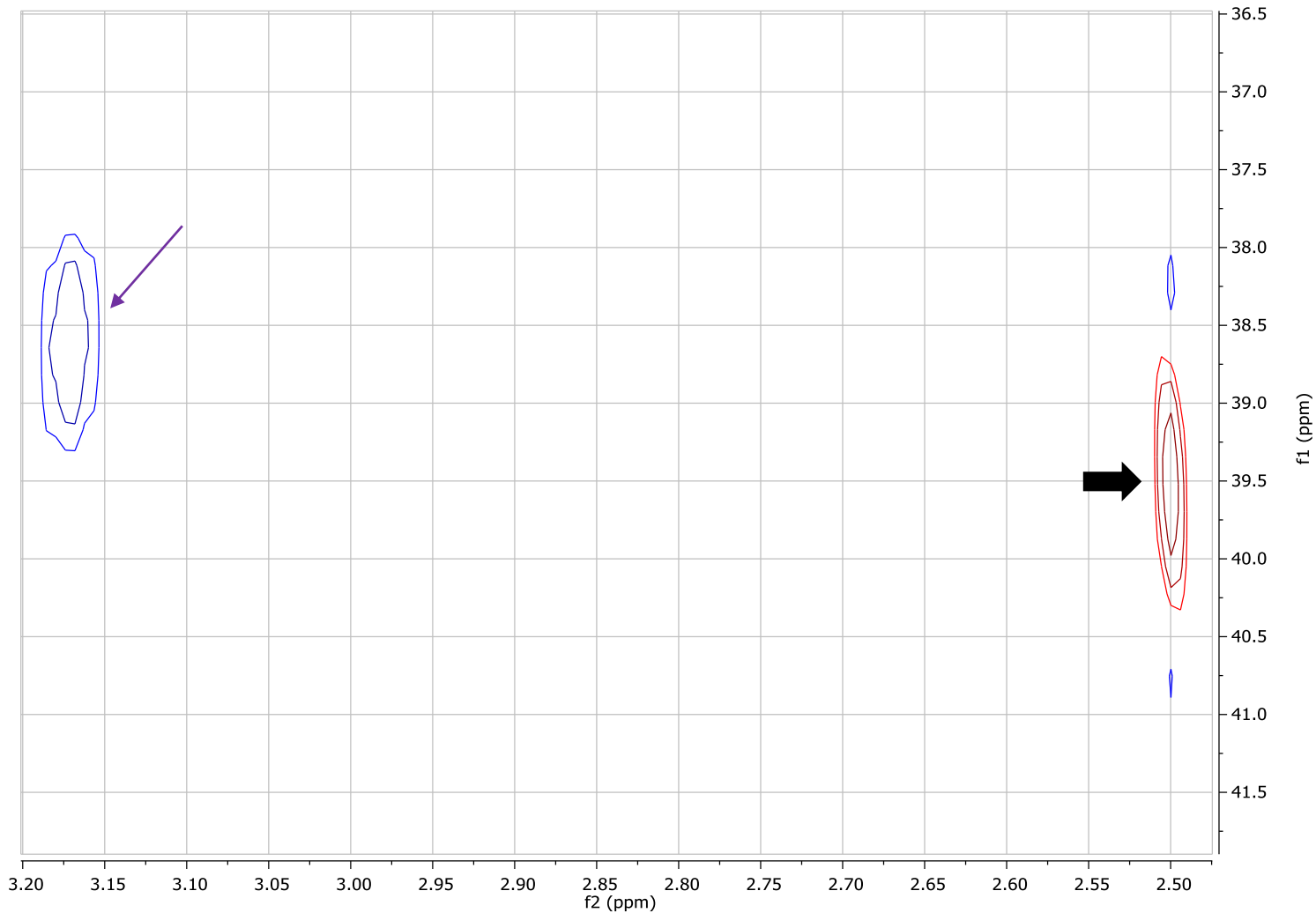
f2 (ppm)



DMSO-*d*<sub>6</sub>

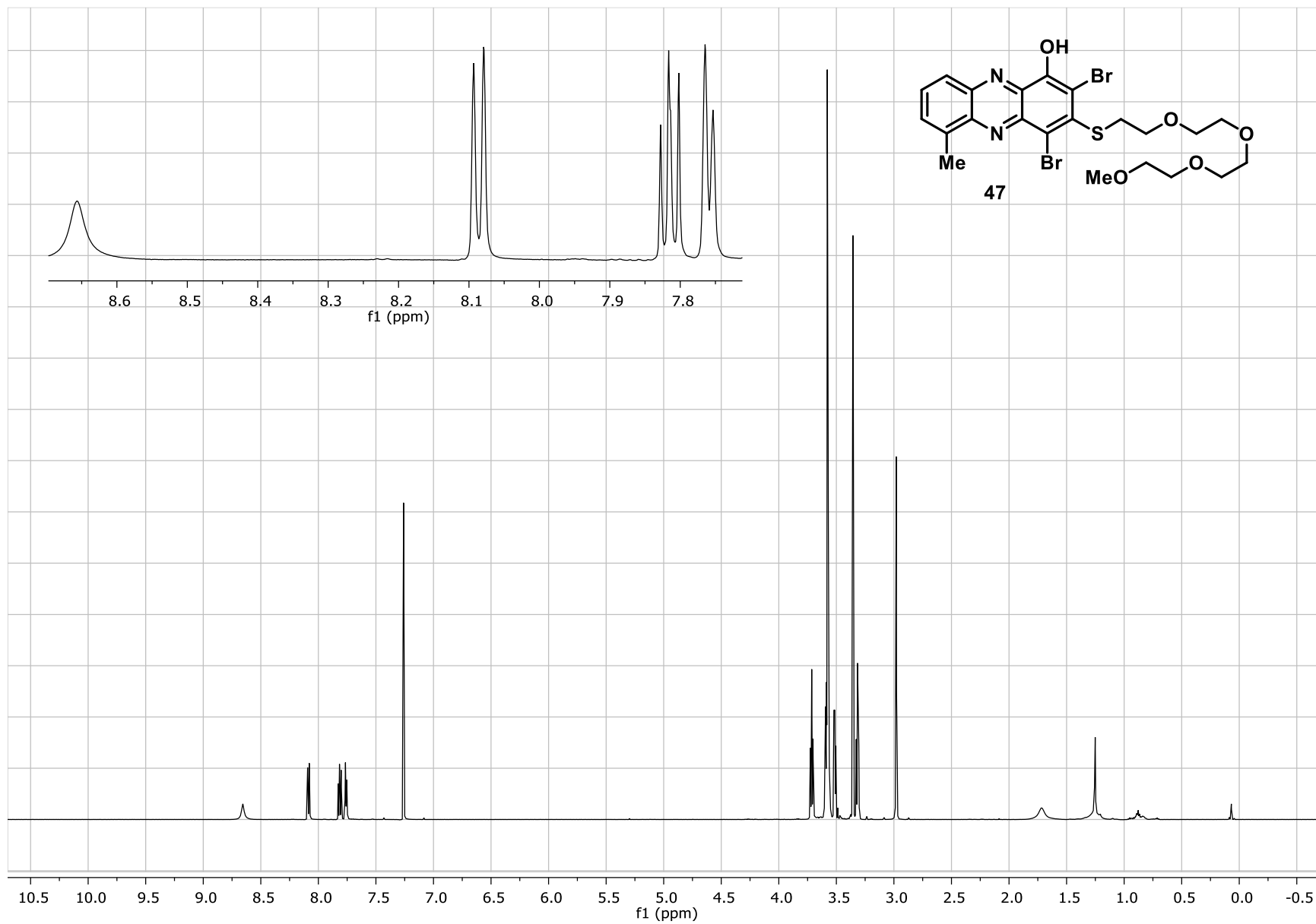


Carbon Signal  
at 38.9 ppm



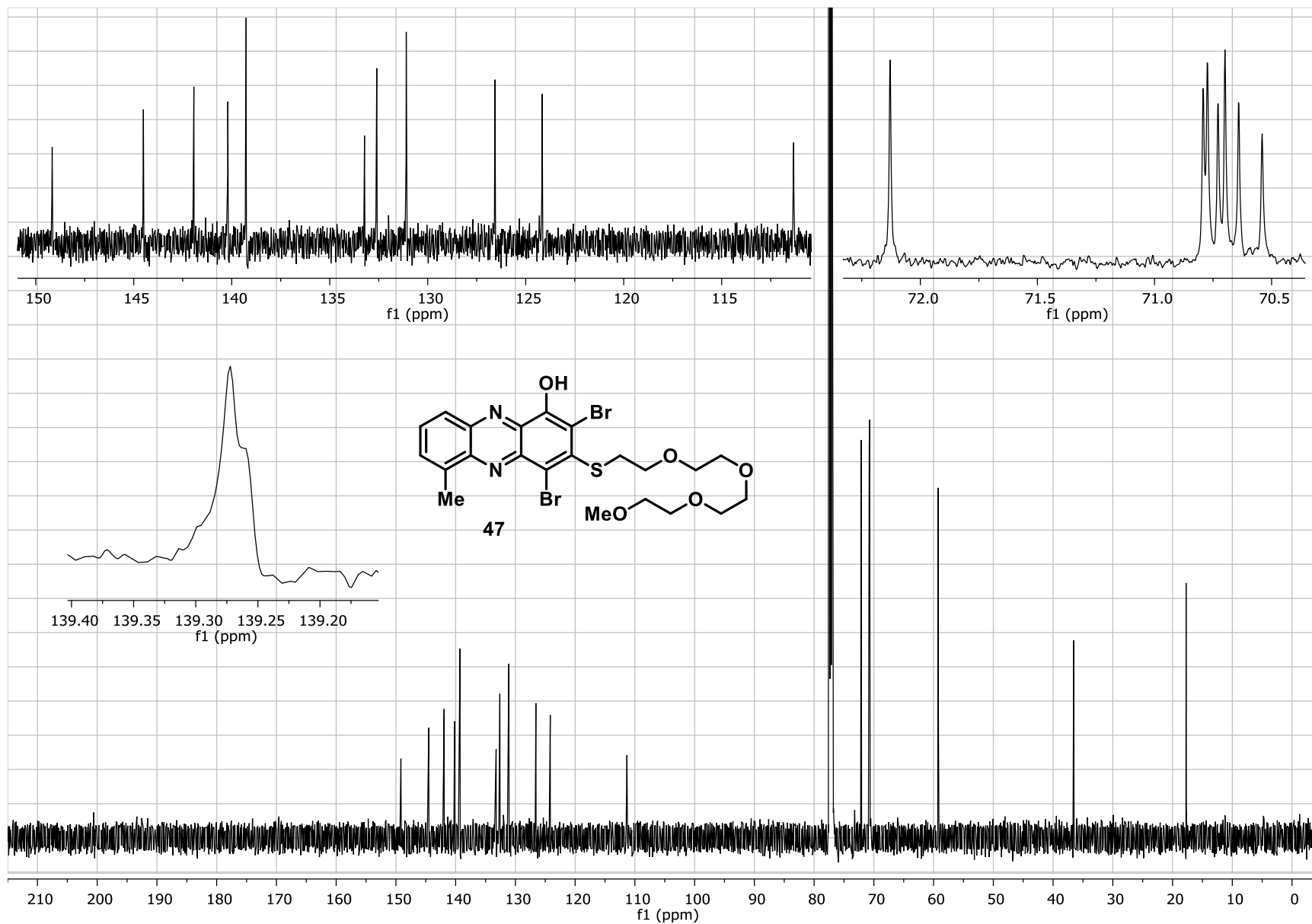
HSQC (zoomed in to assign carbon signal)

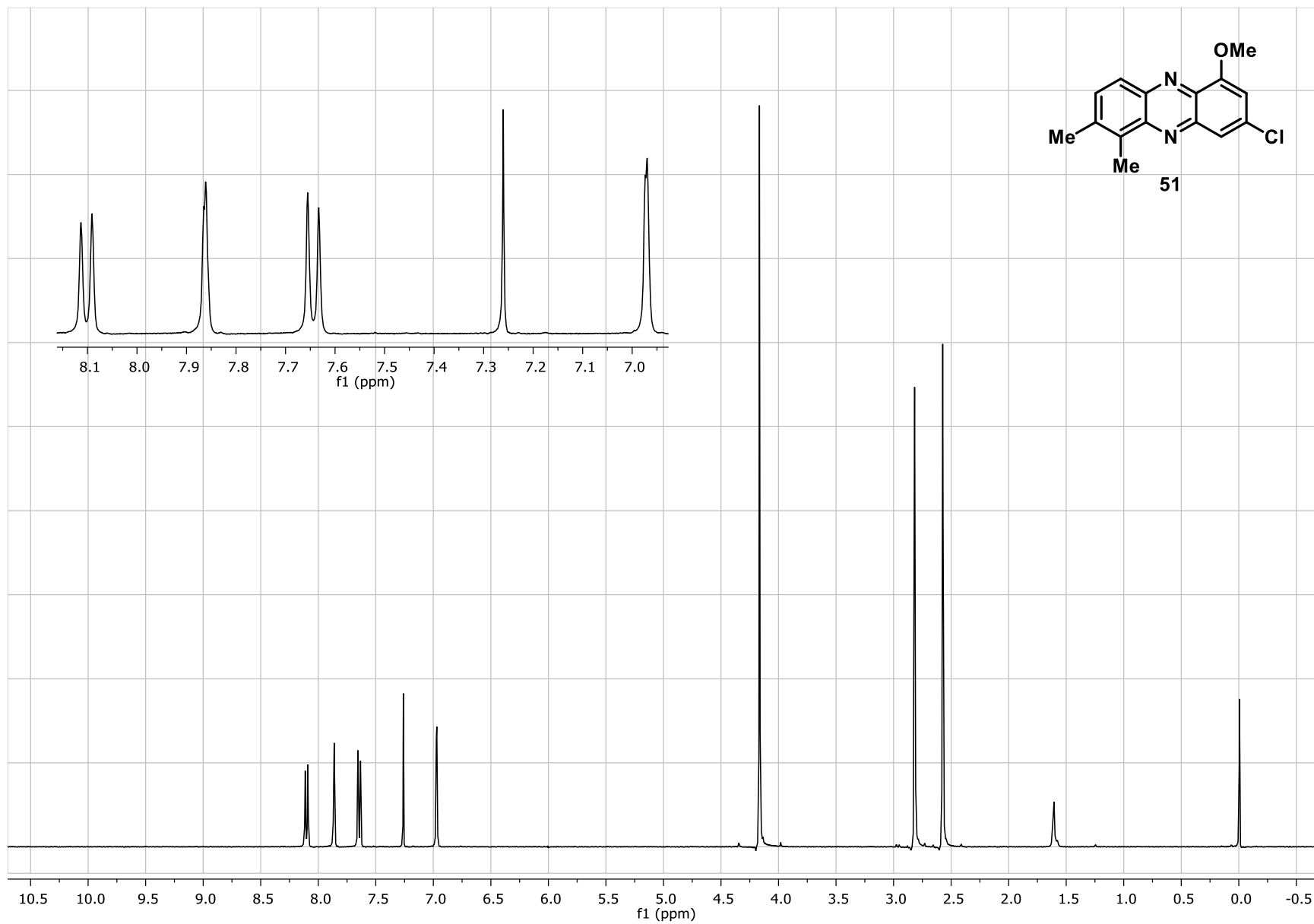
S119

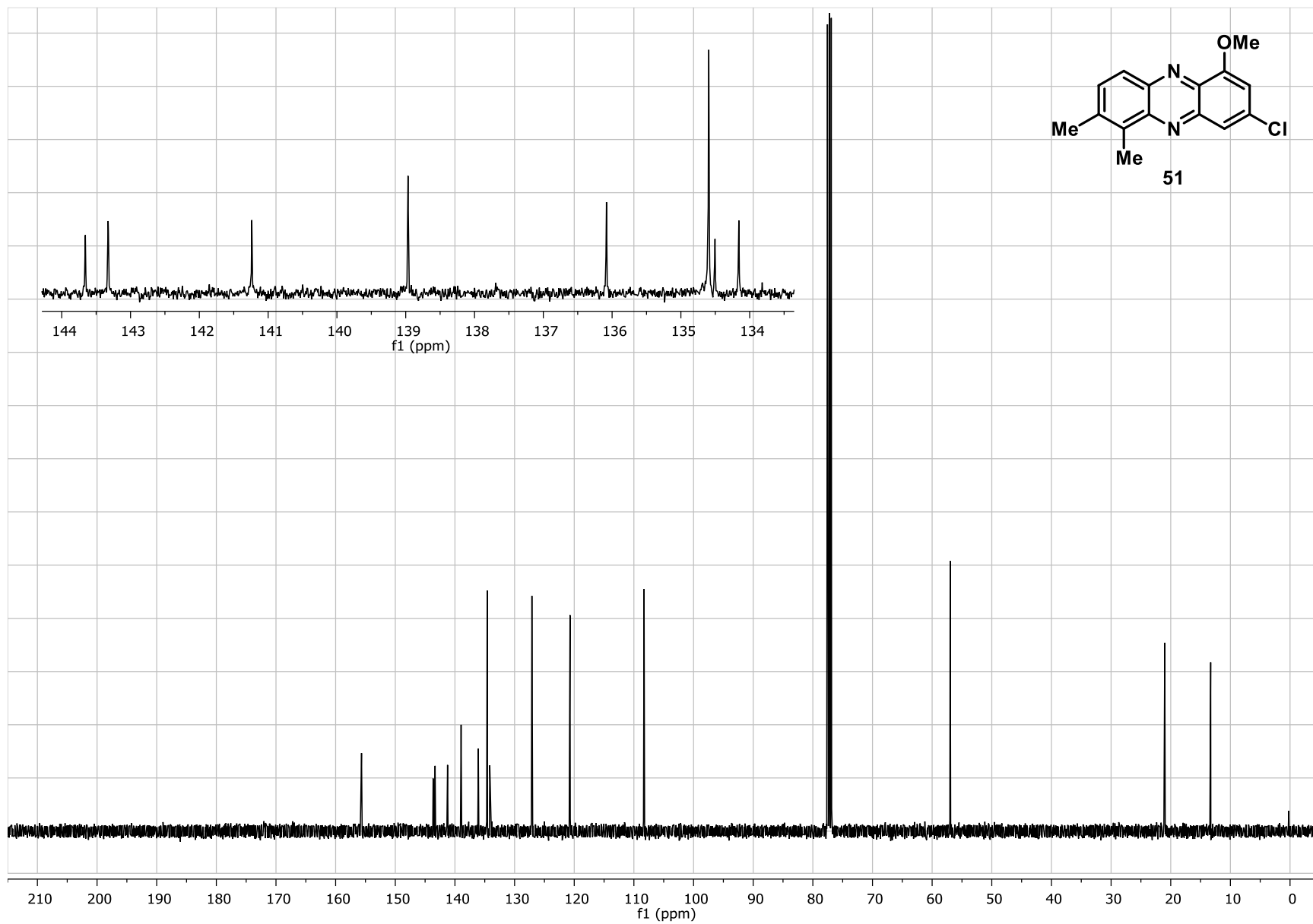


S120

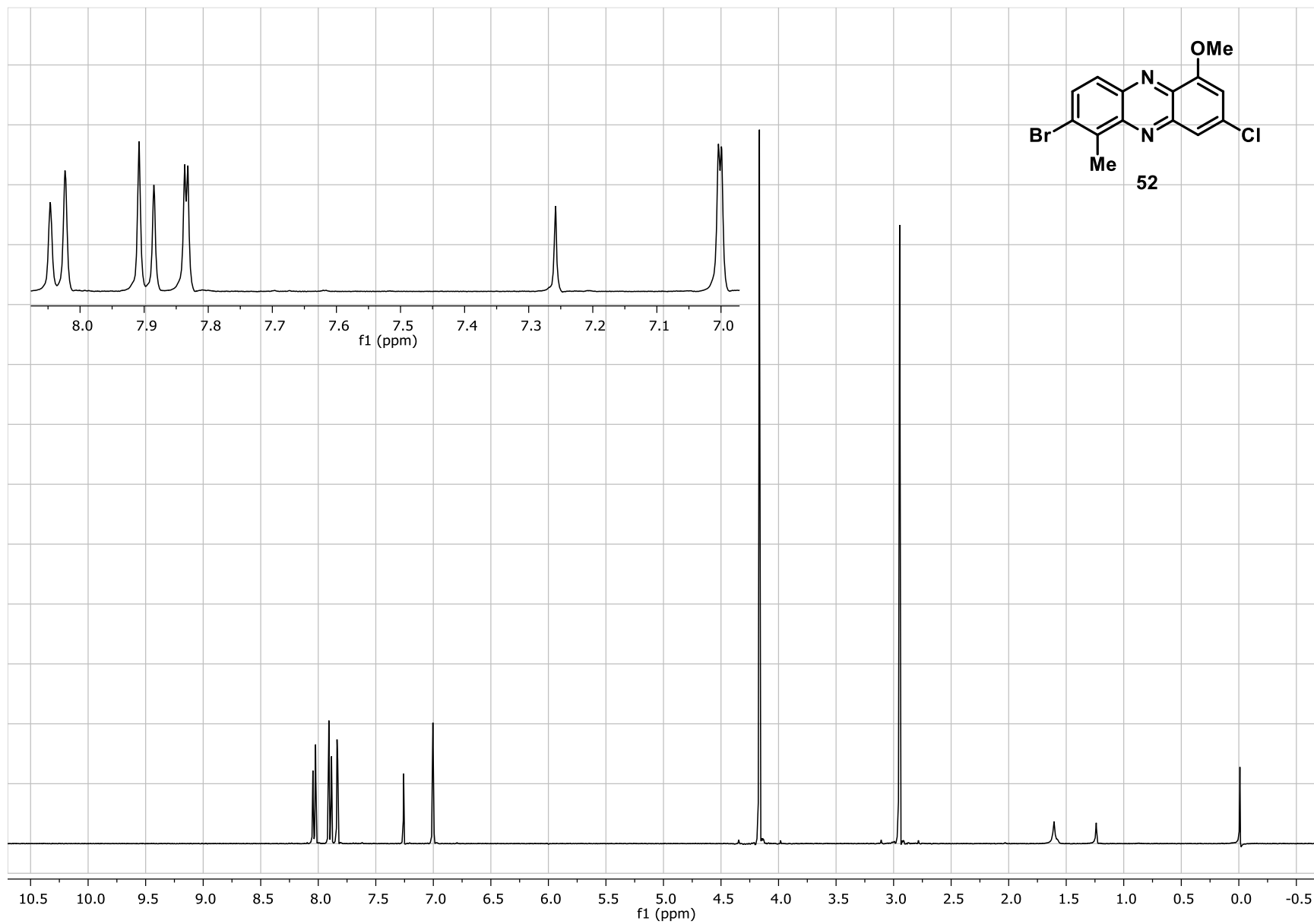


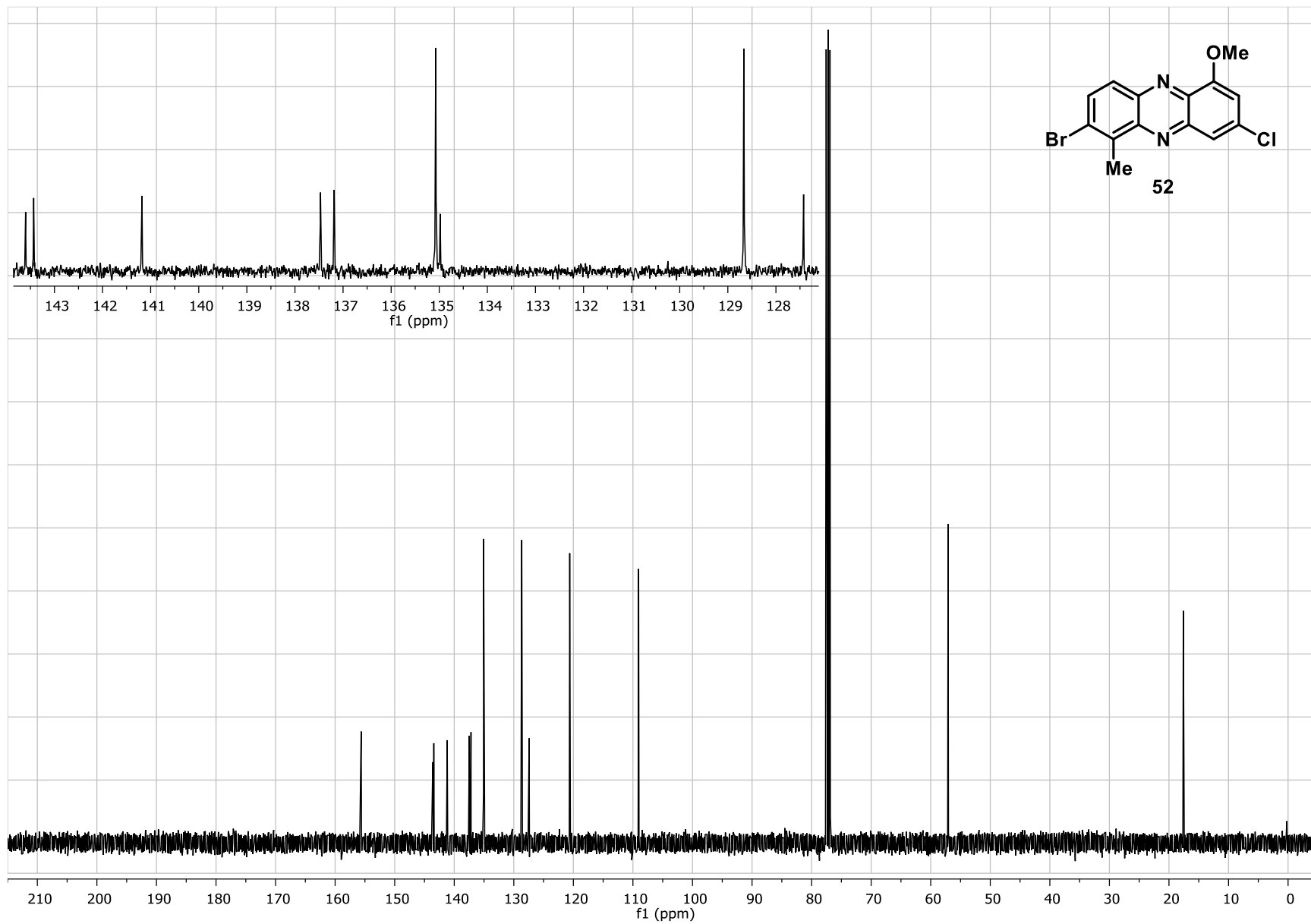




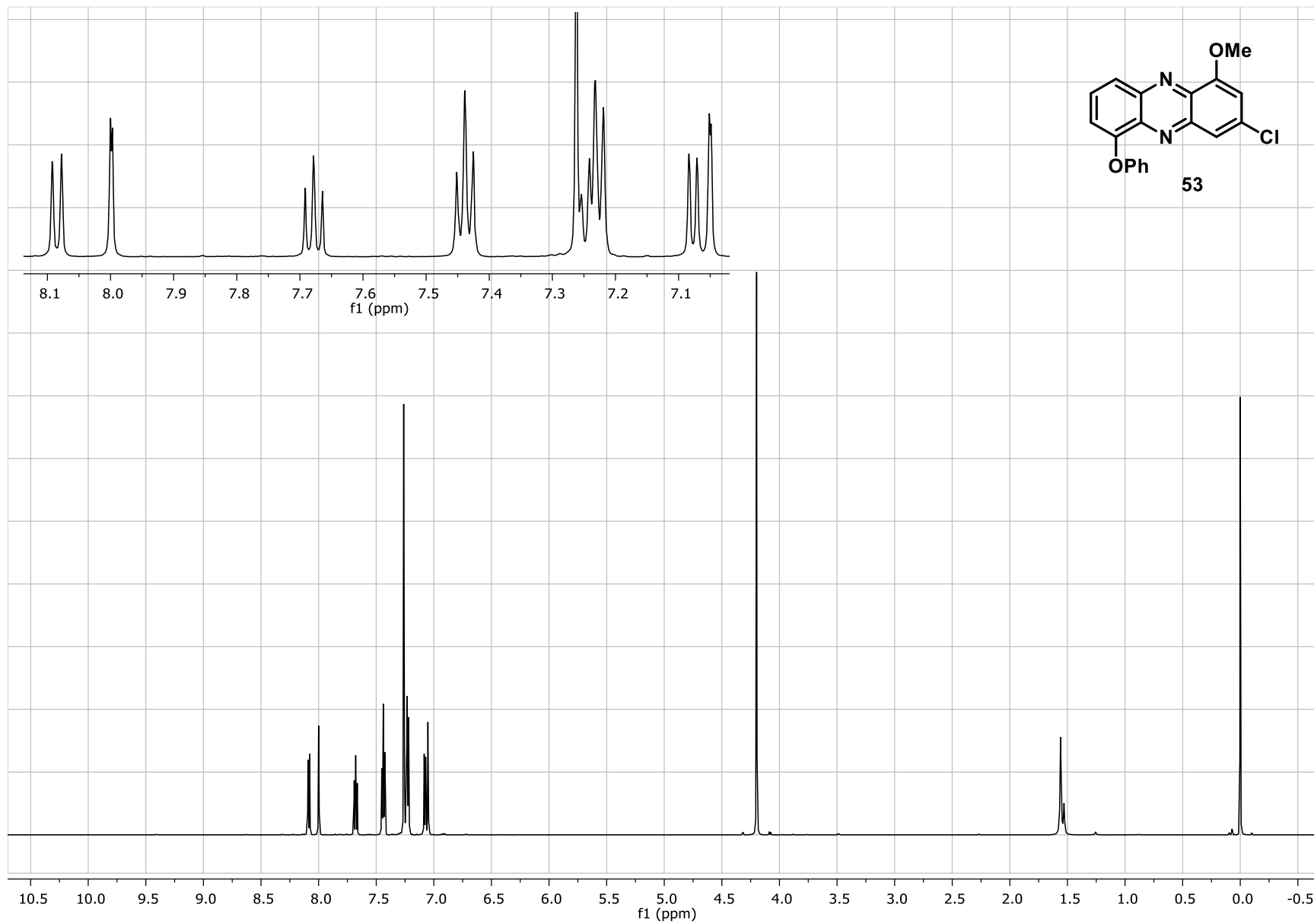


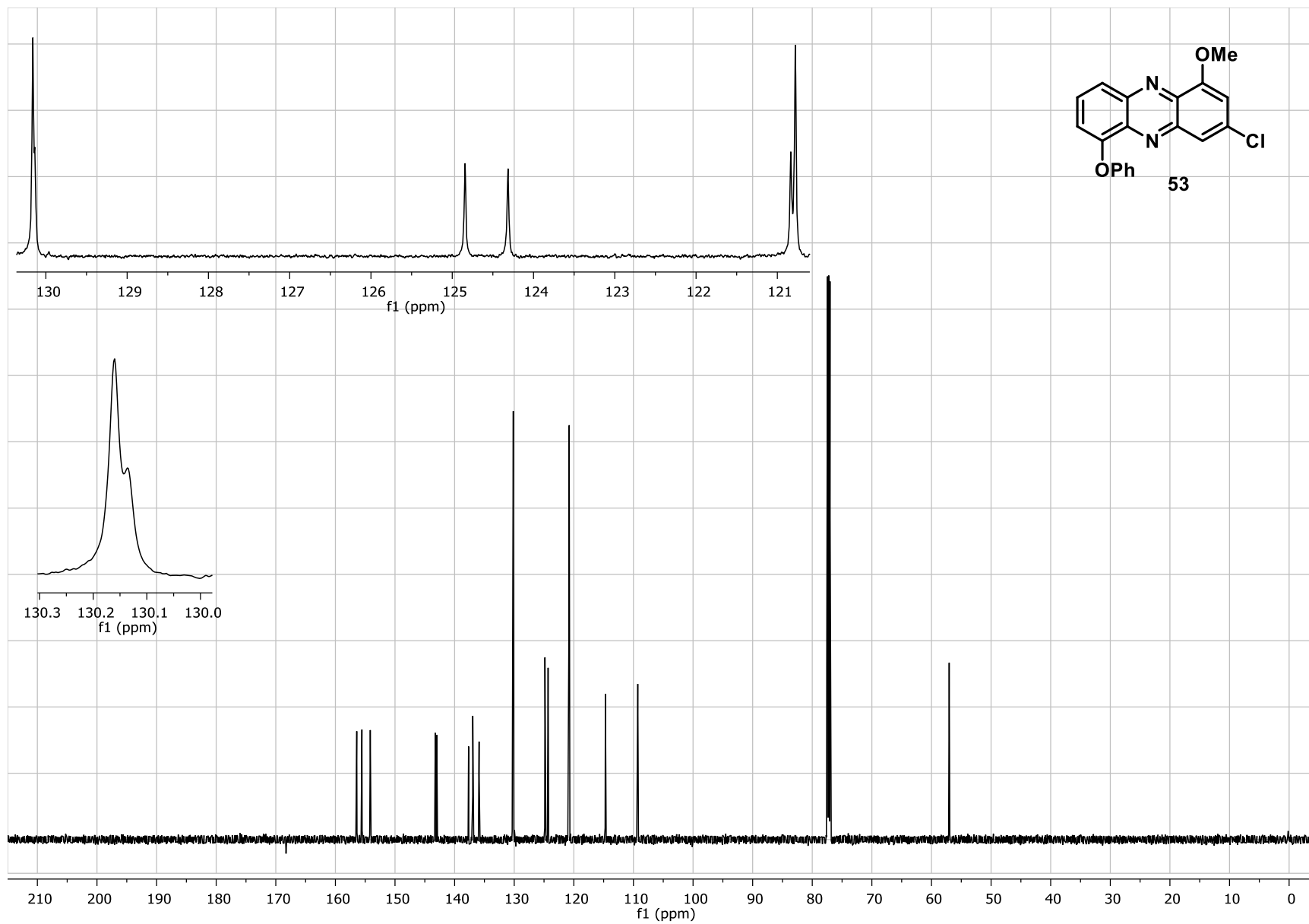
S123

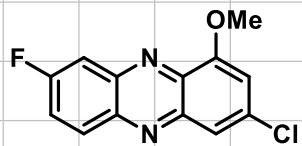




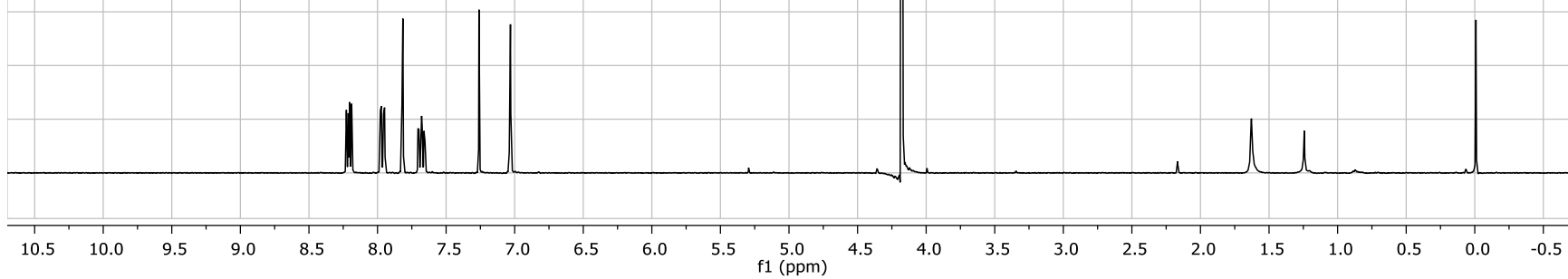
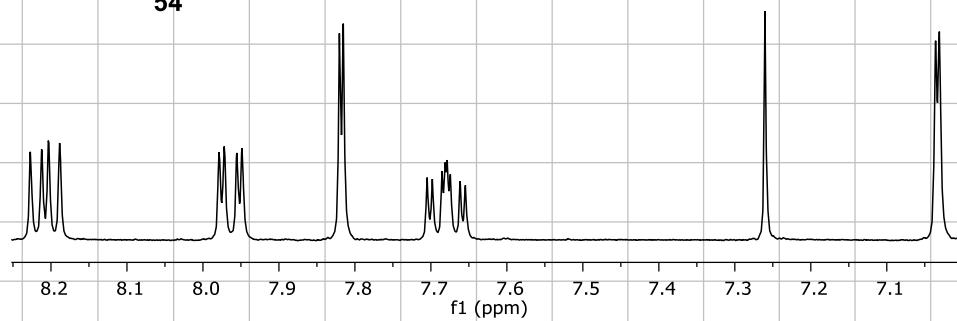
S125





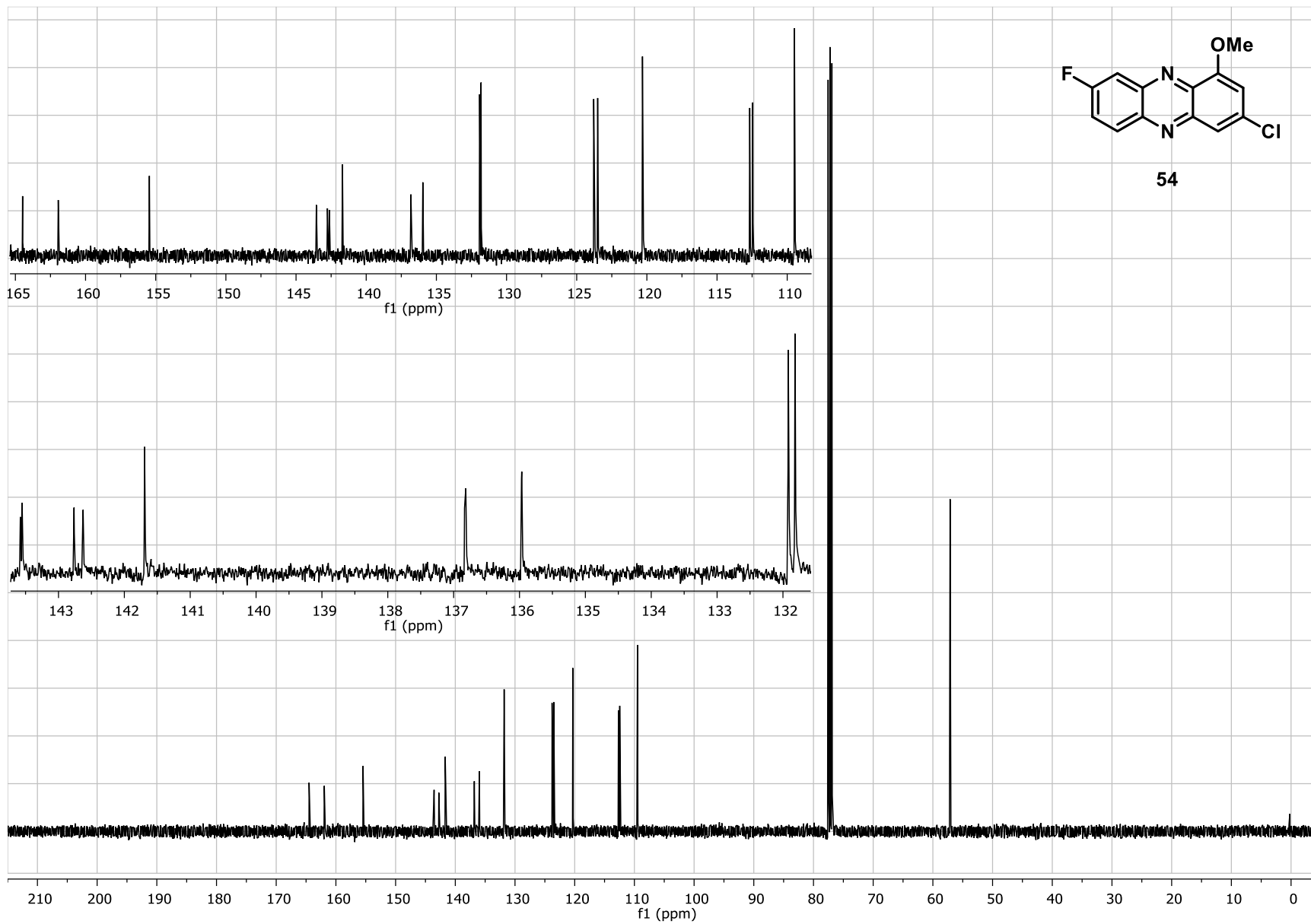


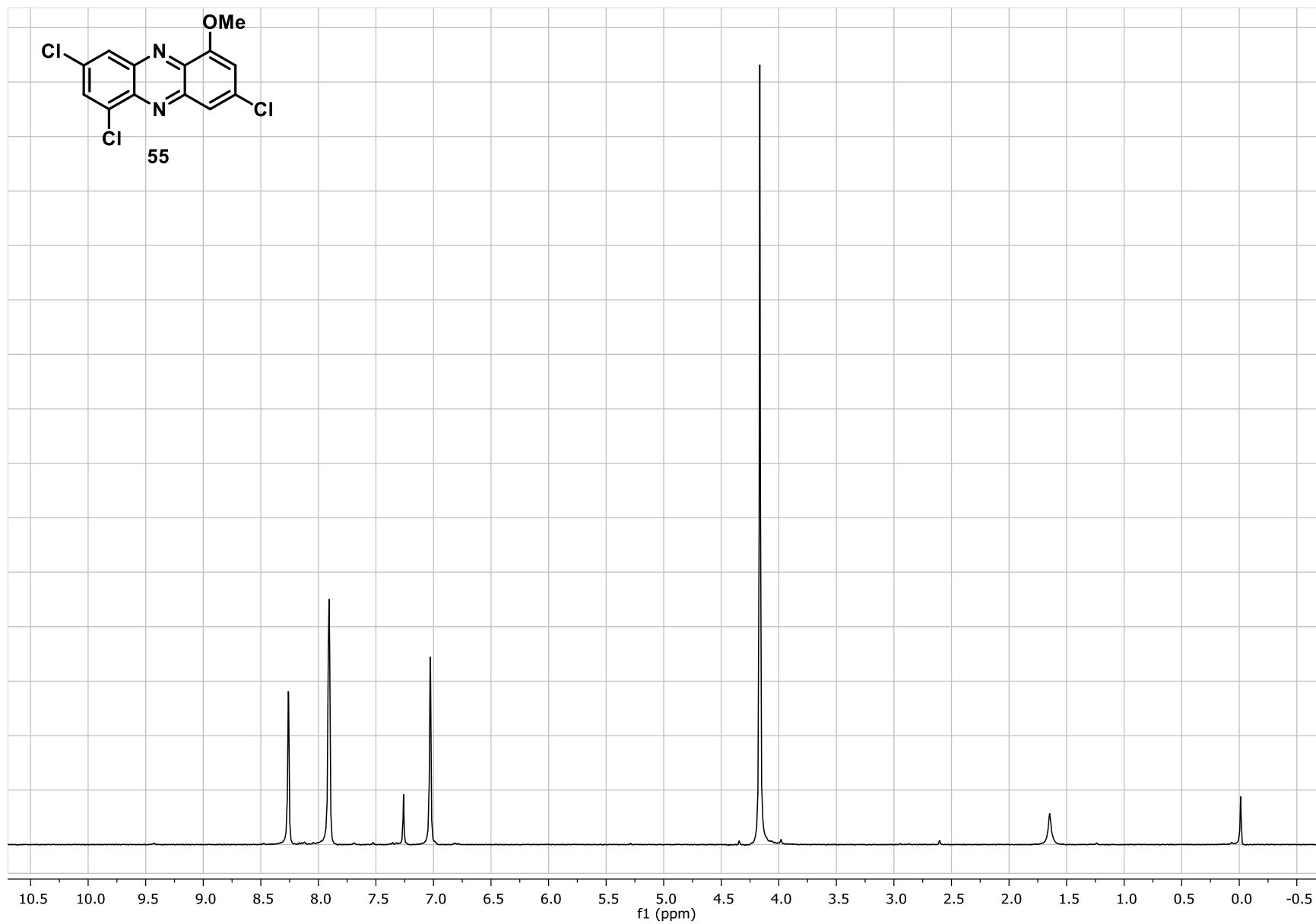
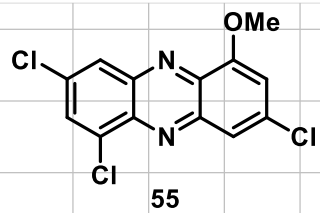
54



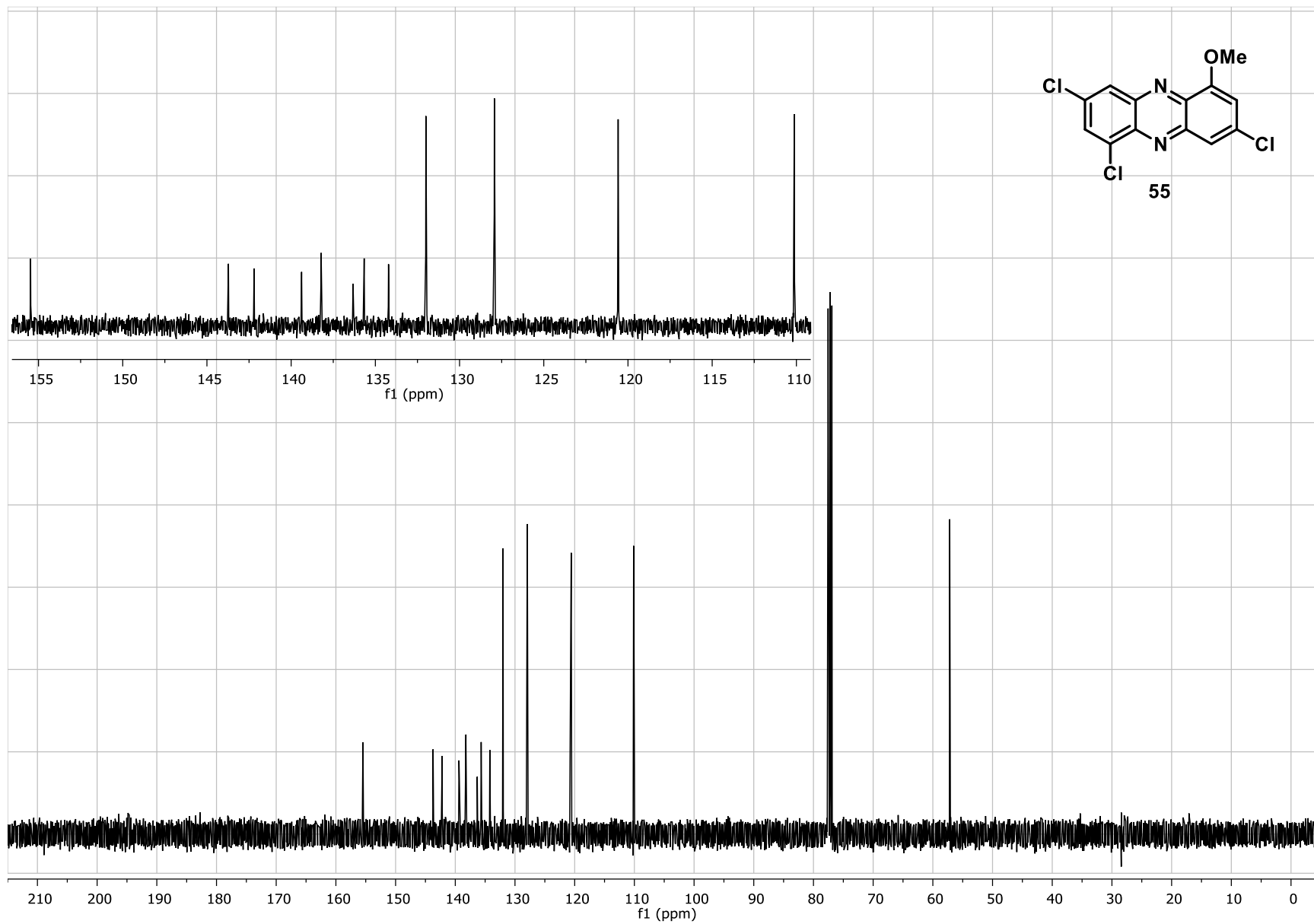
S128

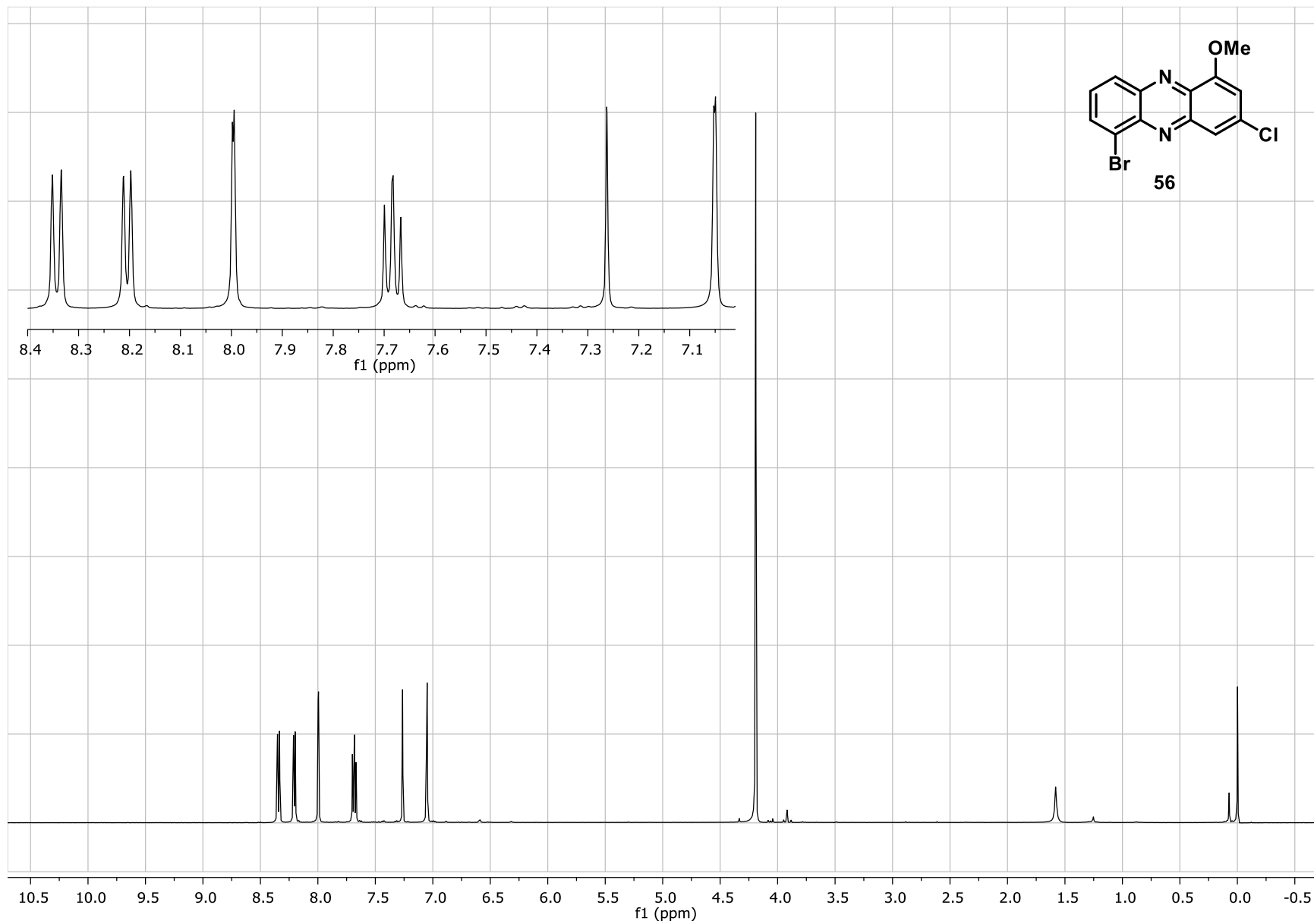


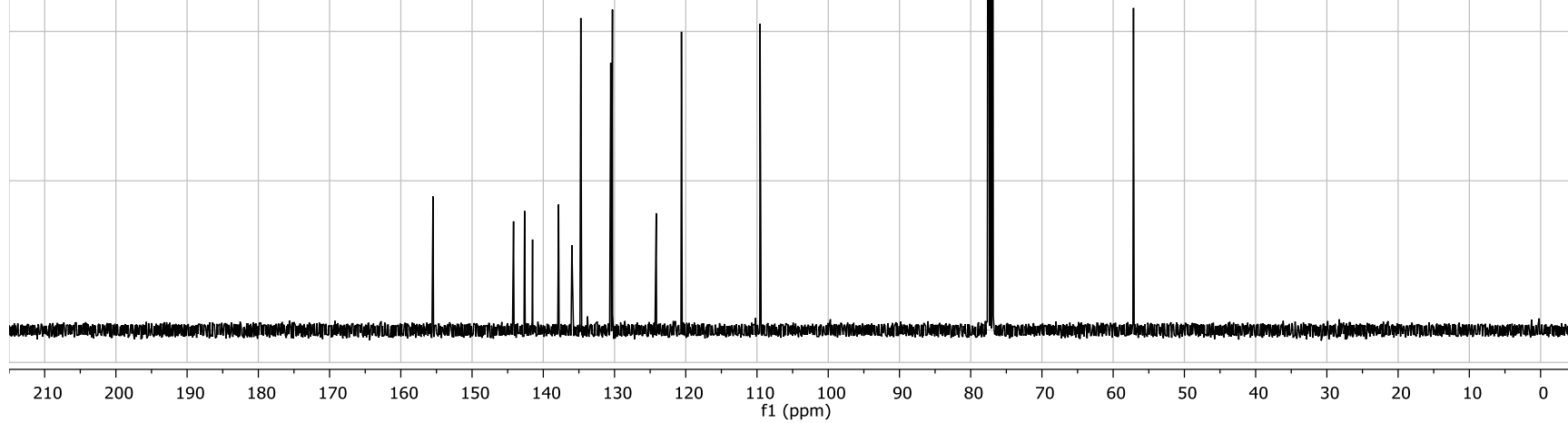
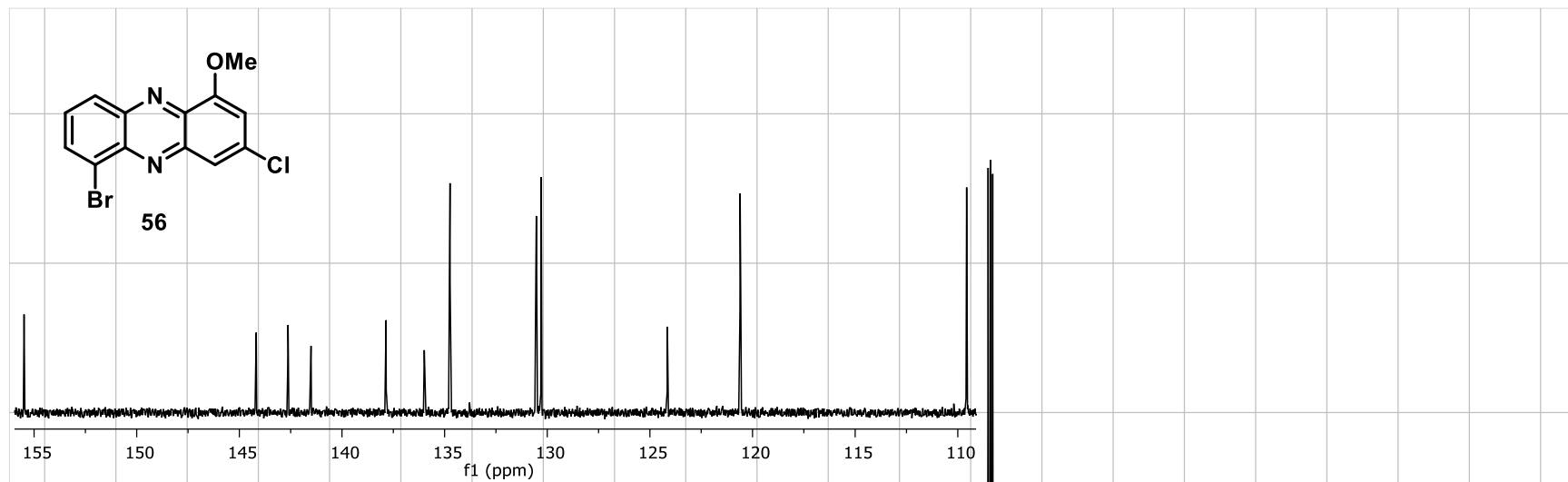
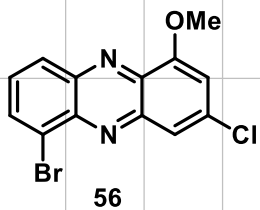


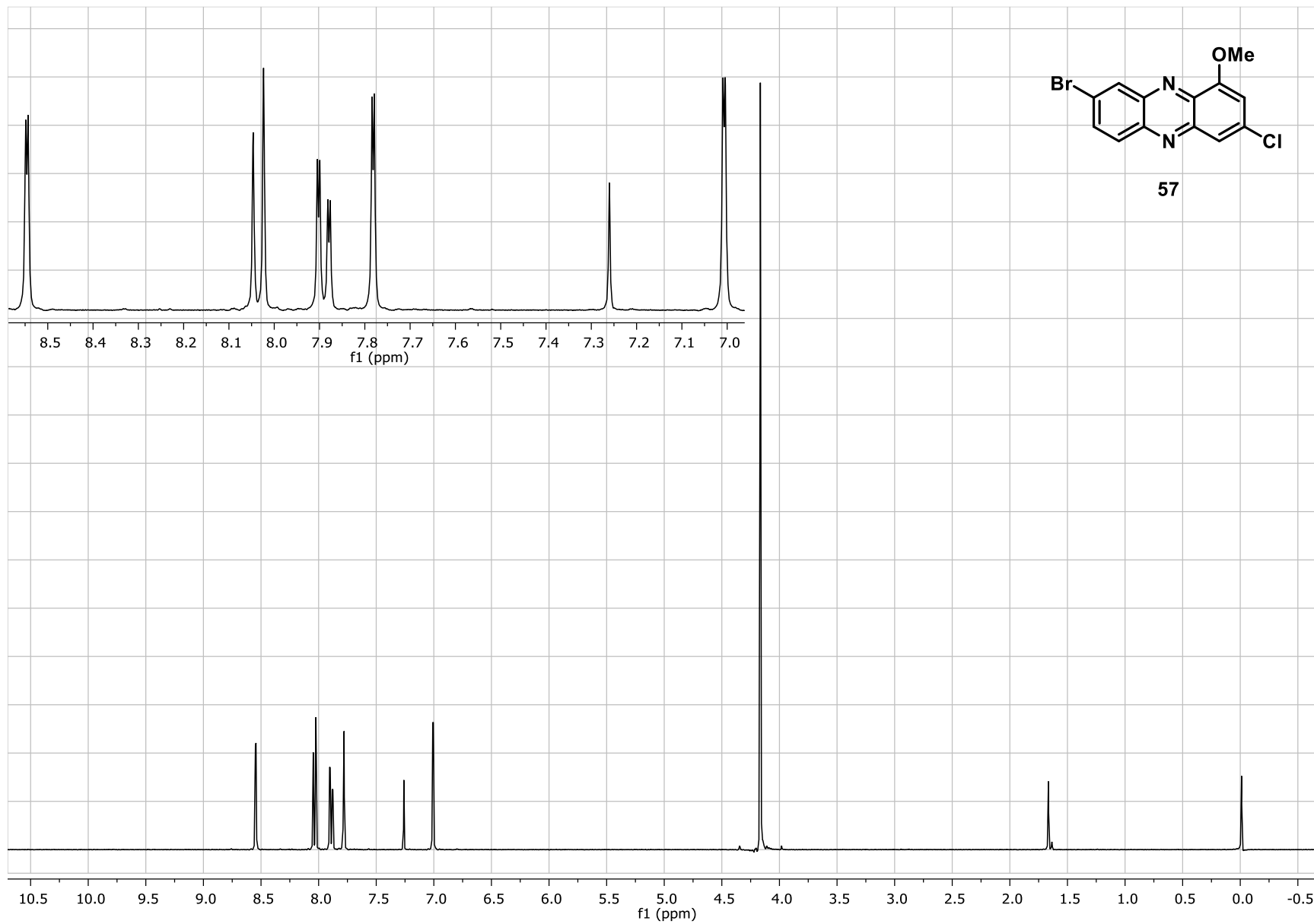


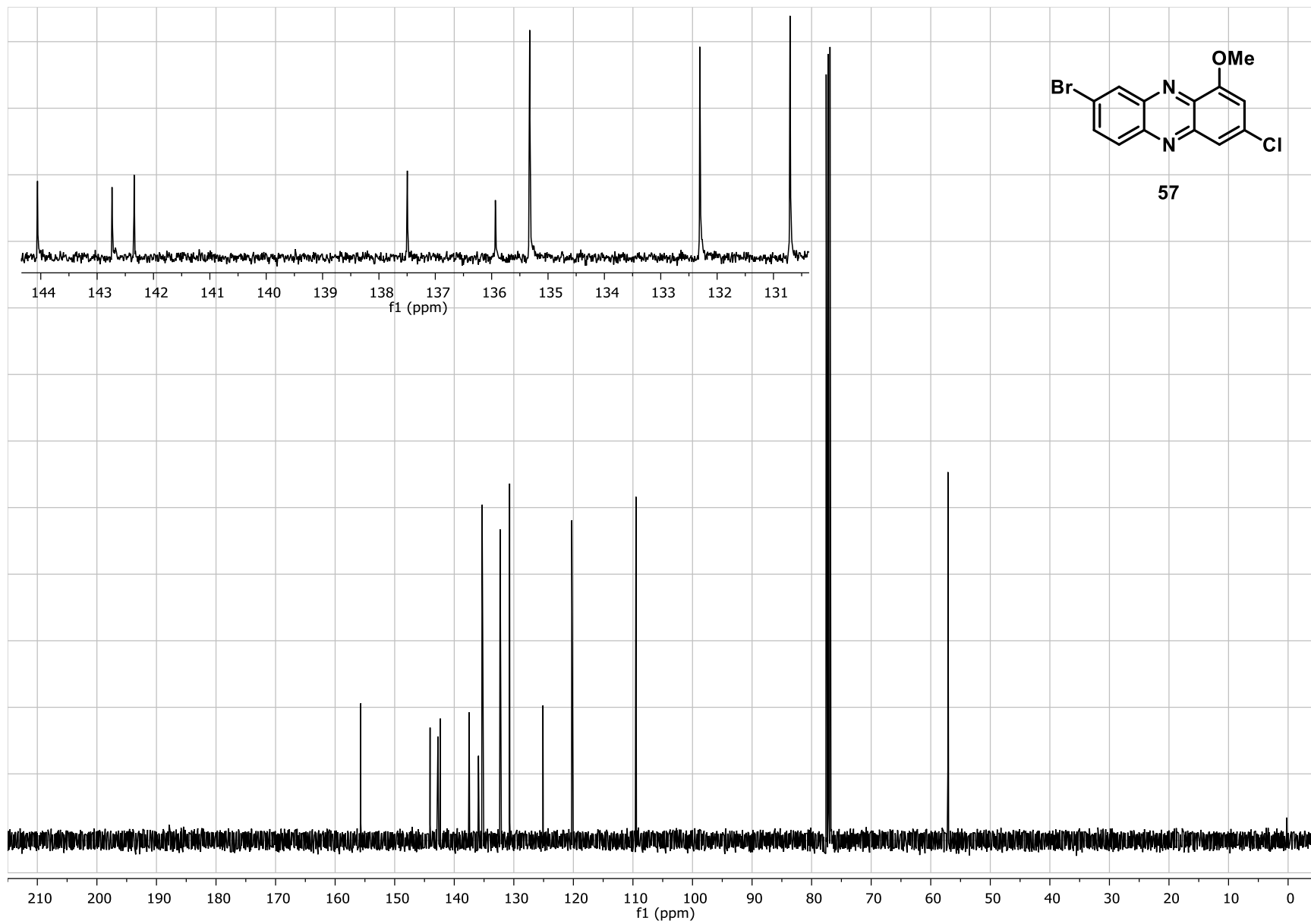
S130

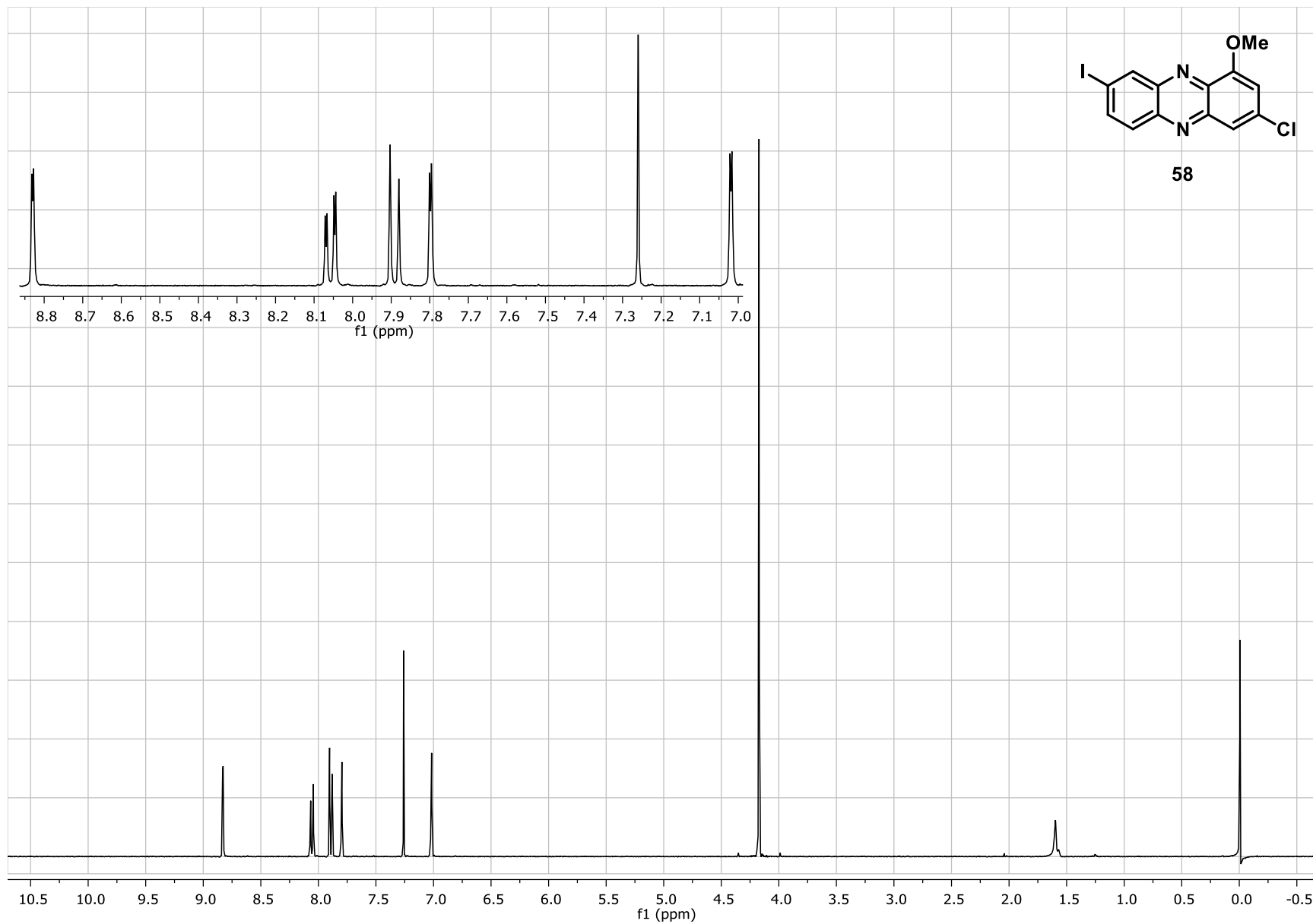




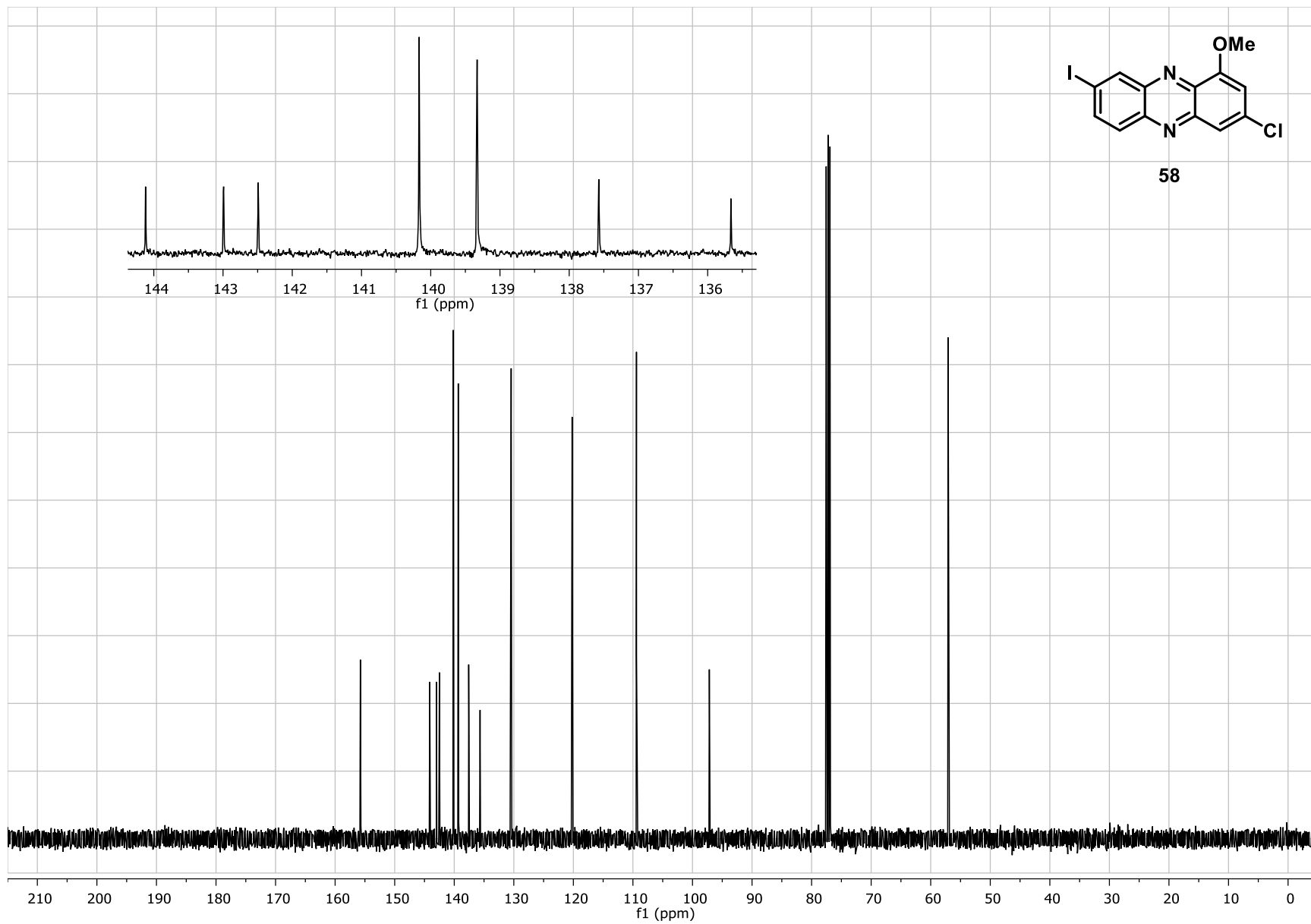












S137