Supplementary Figures for

A porcine brain-wide RNA editing landscape

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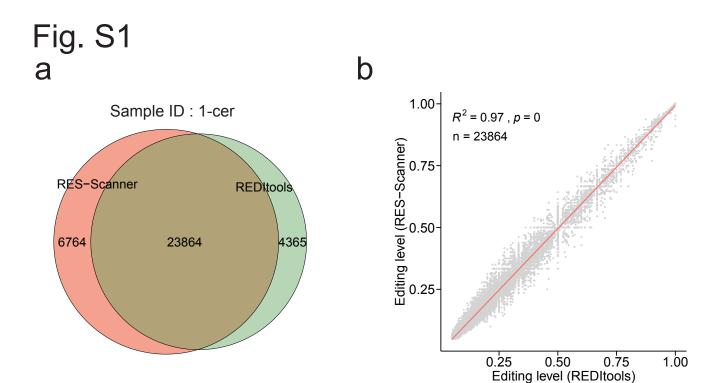


Fig. S1 | **Comparison of results from REDItools and RES-Scanner**. One sample (ID: 1-cer) from cerebellum is shown as an example. a, Venn diagram of RNA editing sites identified by two tools showing most of sites are overlapped. b, Correlations between editing levels of common sites identified by the two tools.

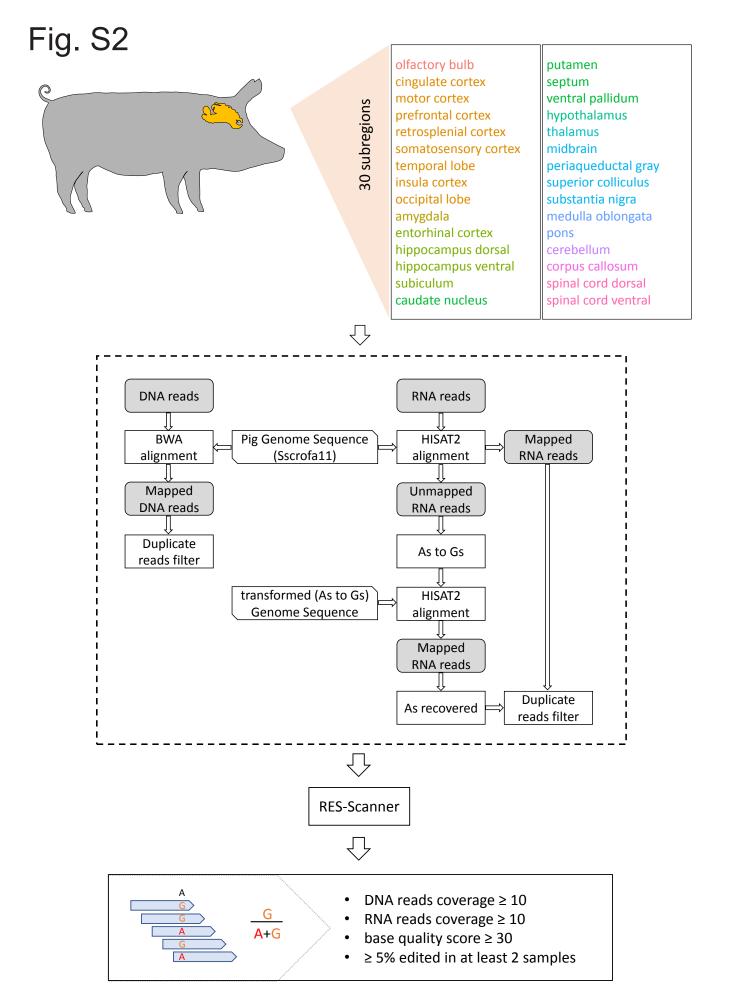


Fig. S2 | **The pipeline of A-to-I RNA editing analysis**. BWA (Burrows-Wheeler Aligner) is a program for mapping DNA reads against reference genome. HISAT2 (Hierarchical Indexing for Spliced Alignment of Transcripts) is a fast program for mapping RNA-seq reads against reference genome.

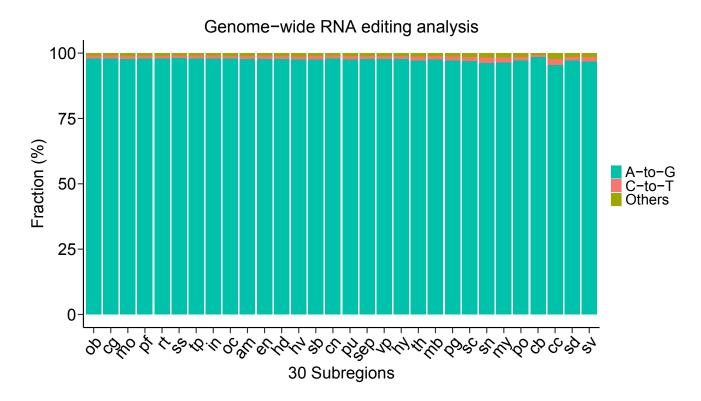


Fig. S3 | RNA editing type identified in 30 subregions of pig brain. olfactory bulb, ob; cingulate cortex, cg; motor cortex, mo; prefrontal cortex, pf; retrosplenial cortex, rt; somatosensory cortex, ss; temporal lobe, tp; insula cortex, in; occipital lobe, oc; amygdala, am; entorhinal cortex, en; hippocampus dorsal, hd; hippocampus ventral, hv; subiculum, sb; caudate nucleus, cn; putamen, pu; septum, sep; ventral pallidum, vp; hypothalamus, hy; thalamus, th; midbrain, mb; periaqueductal gray, pg; superior colliculus, sc; substantia nigra, sn; medulla oblongata, my; pons, po; cerebellum, cb; corpus callosum, cc; spinal cord dorsal, sd; spinal cord ventral, sv.

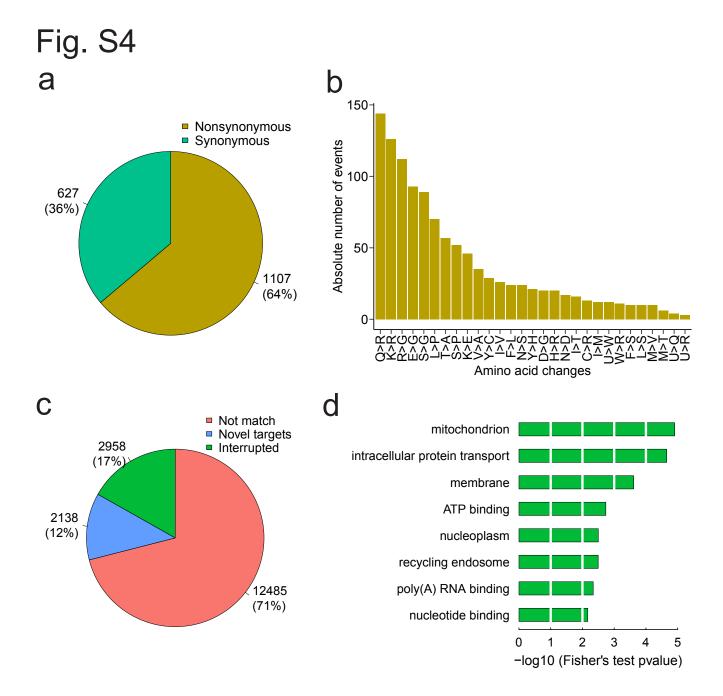


Fig. S4 | The A-to-I editing sites located in CDS and 3-UTRs. a, The number of nonsynonymous (recoding) and synonymous editing sites. b, Distribution of amino acid changes type. c, Distribution of editing sites relative to miRNA target sites in 3-UTRs. Not match, editing sites without miRNA target; Novel targets, editing sites which potentially create novel miRNA target; Interrupted, editing sites which potentially interrupt miRNA target. d, The top Gene ontology (GO) terms associated with genes harboring editing sites which potentially interrupt miRNA target.

Fig. S5

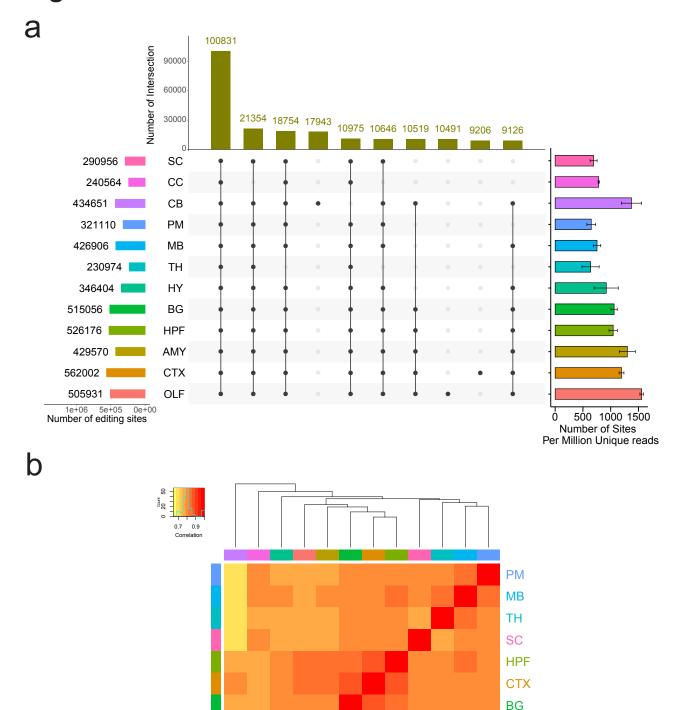


Fig. S5 | Landscape of RNA editing sites across pig brain regions. a, UpSetR plot showing the number of editing sites identified across 12 brain regions and the top ten intersection among regions. The number of A-to-I editing sites were normalized by uniquely mapped reads each sample on the right. b, Heatmap and dendrogram of correlations on the editing levels of different regions.

AMY
OLF
HY
CC
CB



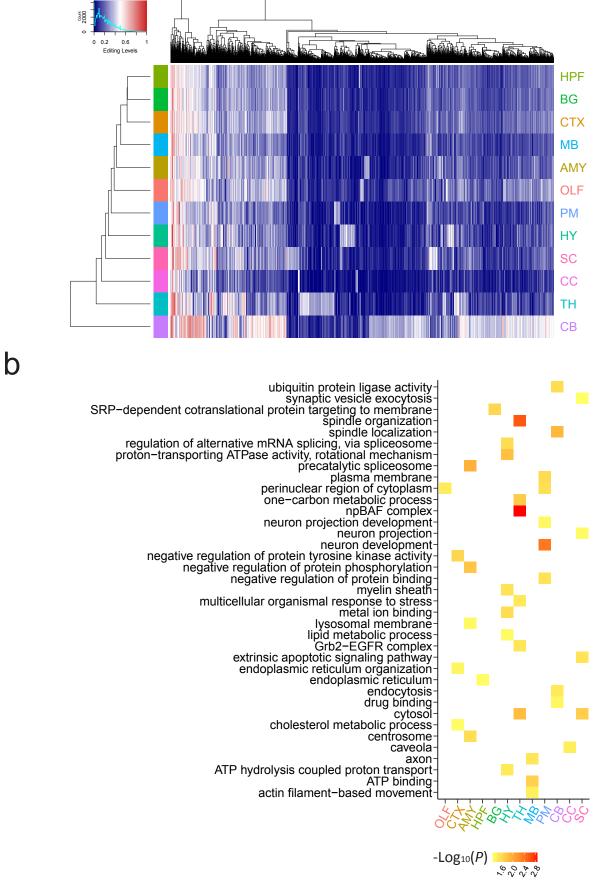


Fig. S6 | **Region-specific editing sites within the pig brain**. a, Heatmap showing region-specific editing sites with coverage ≥ 10 RNA reads in all regions. b, Go enrichment analysis of genes harboring at least one region-enriched editing site.

Fig. S7

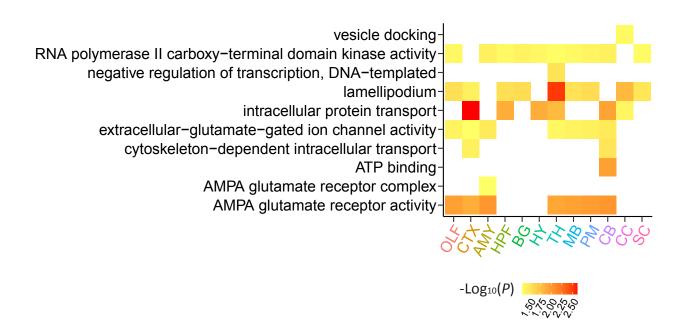
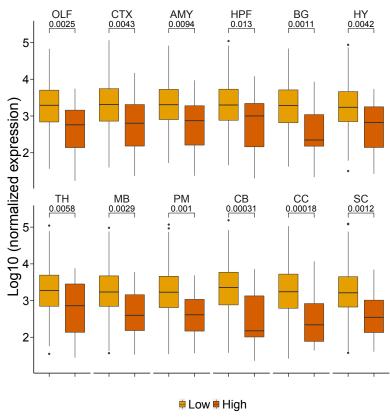


Fig. S7 \mid Go enrichment analysis of genes harboring highly edited (>75%) CDS or 3-UTR residing sites





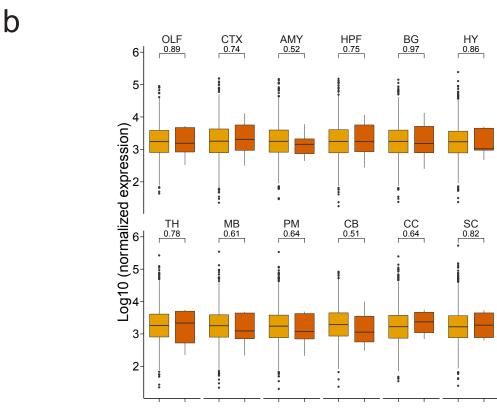


Fig. S8 | Box plots showing expression level of genes with highly and lowly edited sites. a, CDS-residing sites. b, 3-UTR residing sites. The significance of differences in expression was assessed by Wilcoxon test.

■ Low **■** High

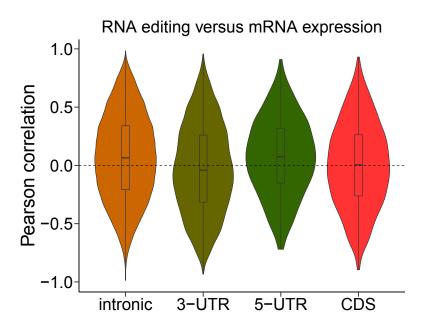


Fig. S9 | Pearson correlation between RNA editing and mRNA expression.

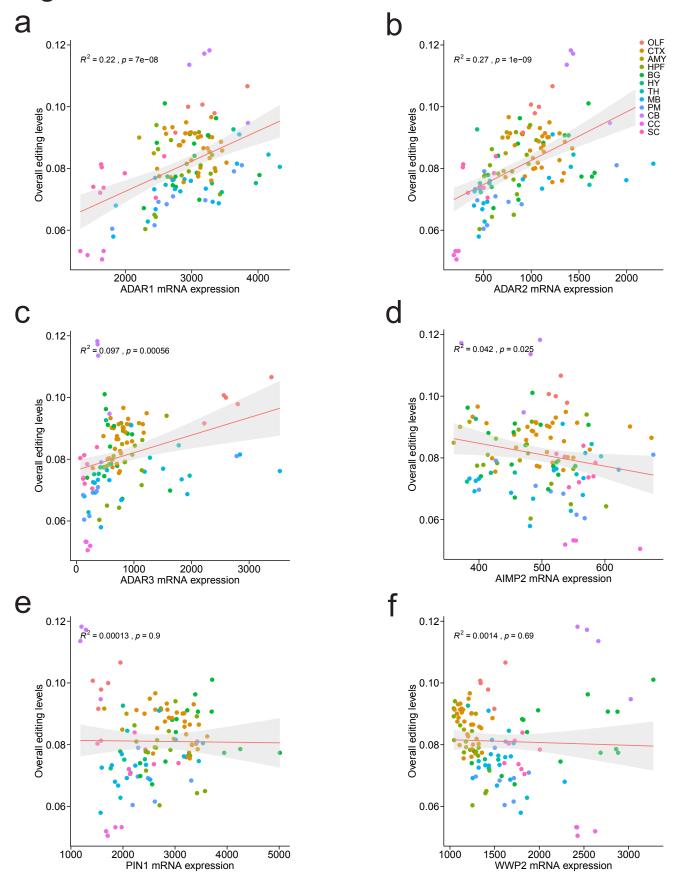
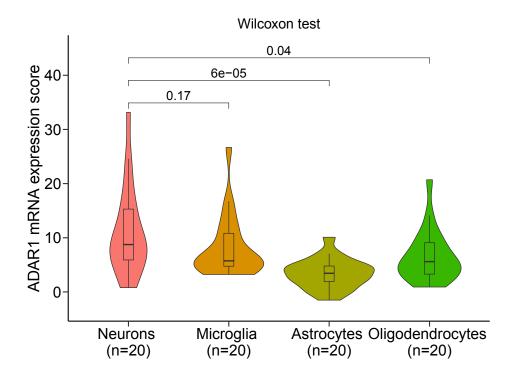


Fig. S10 | **Known enzymes associated with A-to-I RNA editing**. Correlations between overall editing levels at all editing sites and mRNA expression levels of ADAR1 (a), ADAR2 (b), ADAR3 (c), AIMP2 (d), PIN1 (e) and WWP2 (f). R^2 values were calculated by robust linear regressions on overall editing levels and normalized expression. Gray shaded areas, 95% confidence interval.

a



b

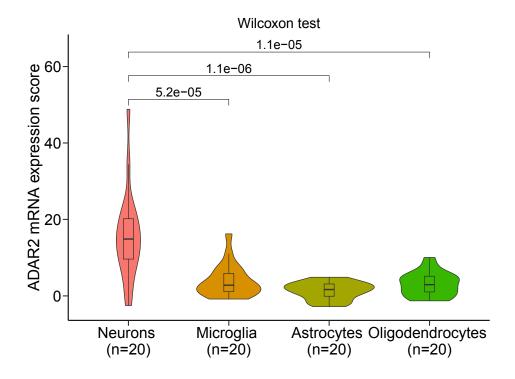


Fig. S11 | The ADARs mRNA expression scores of neurons, microglia, astrocytes and oligodendrocytes. a, ADAR1. b, ADAR2. The significance of differences between expression scores of ADAR1 or ADAR2 in neurons and non-neuronal cells was assessed by Wilcoxon test.

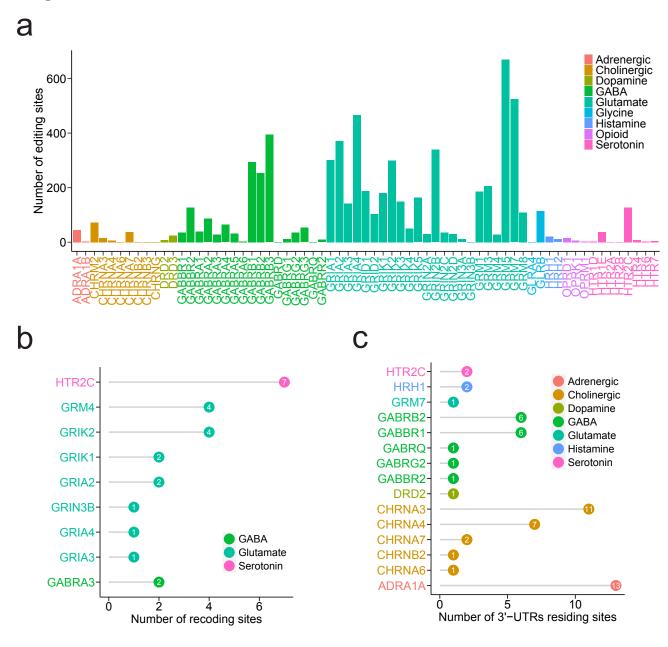
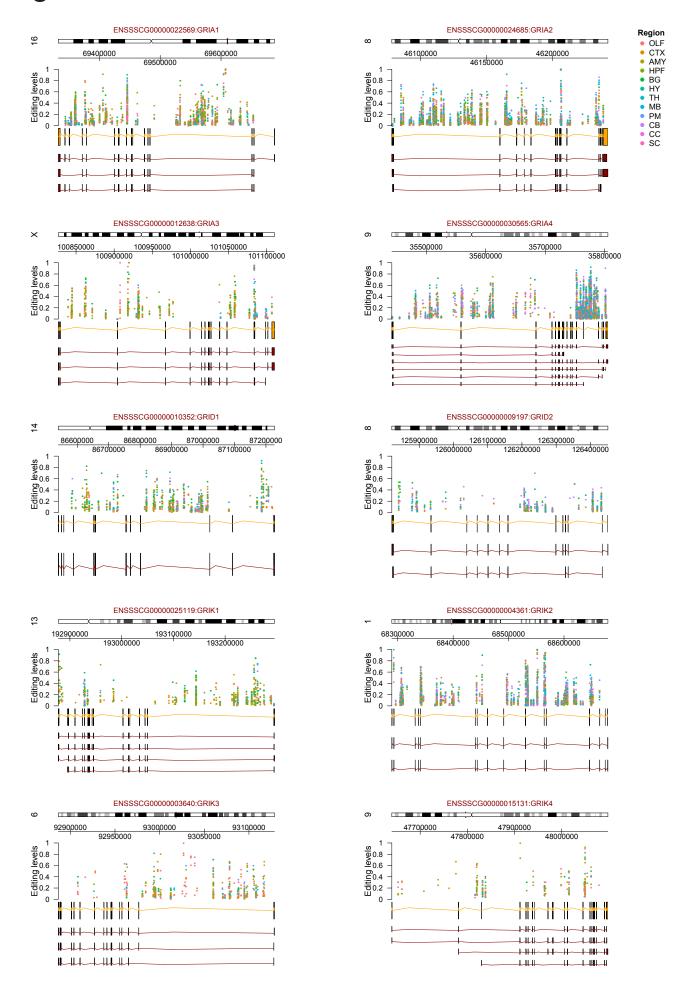
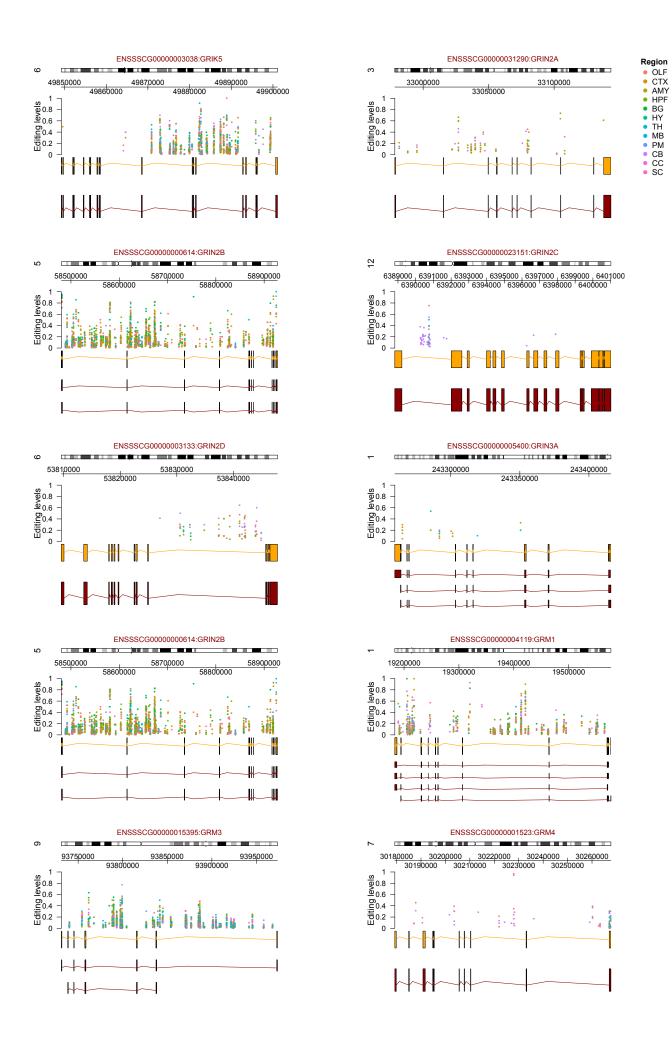


Fig. S12 | Distribution of the A-to-I RNA editing sites located in major neurotransmitter receptors. a, RNA editing sites located in neurotransmitter receptors, including adrenergic, cholinergic, dopamine, GABA, glutamate, glycine, histamine, opioid and serotonin receptors. b, The number of recoding editing sites. c, The number of editing sites located in 3-UTRs.





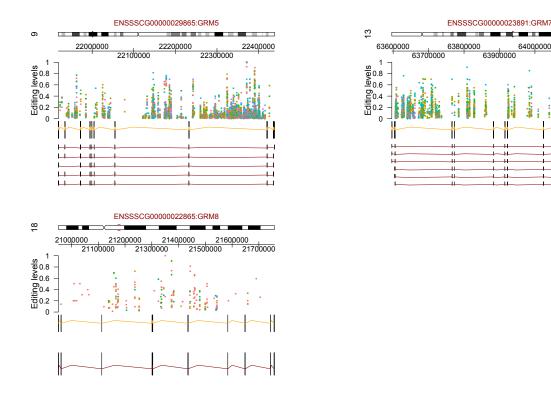


Fig. S13 | **Detailed information of editing sites located in glutamate receptors**. Olfactory bulb, OLF; Cerebral cortex, CTX; Amygdala, AMY; Hippocampal formation, HPF; Basal ganglia, BG; Hypothalamus, HY; Thalamus, TH; Midbrain, MB; Pons and medulla, PM; Cerebellum, CB; Corpus callosum, CC; Spinal cord, SC.

Region

OLF CTX AMY HPF

BG HY TH MB PM CB CC SC

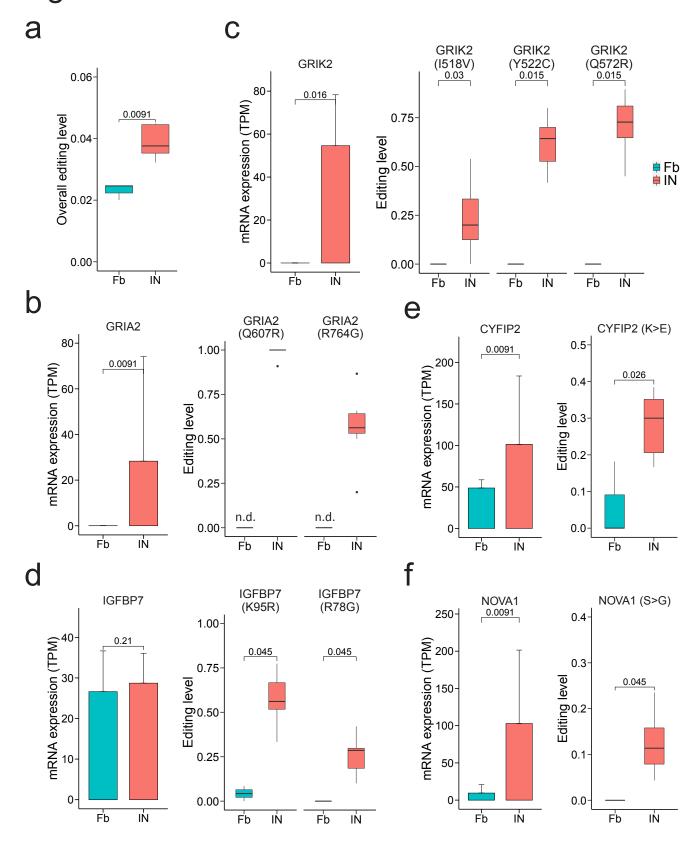


Fig. S14 | Increased editing from fibroblasts to induced neurons. a, Overall editing level of fibroblasts and neurons. The mRNA expression and recoding events of GRIA2 (b), GRIK2 (c), IGFBP7 (d), CYFIP2 (e) and NOVA1 (f). Not detected, n.d.; Fibroblasts, Fb; induced neurons, IN. The significance of differences was assessed by Wilcoxon test.

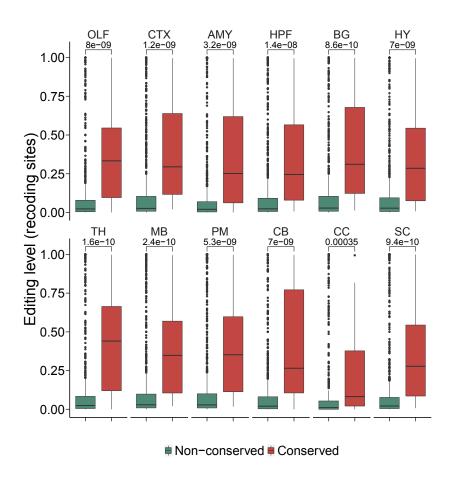


Fig. S15 | Box plot showing editing level of conserved and non-conserved recoding sites. The significance of differences was assessed by Wilcoxon test.

Fig. S16

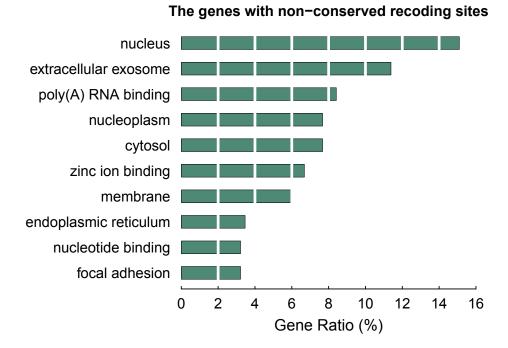


Fig. S16 | Bar plot showing categories of genes with non-conserved recoding sites. Only the top ten categories are shown (ranked by gene ratio).

Fig. S17

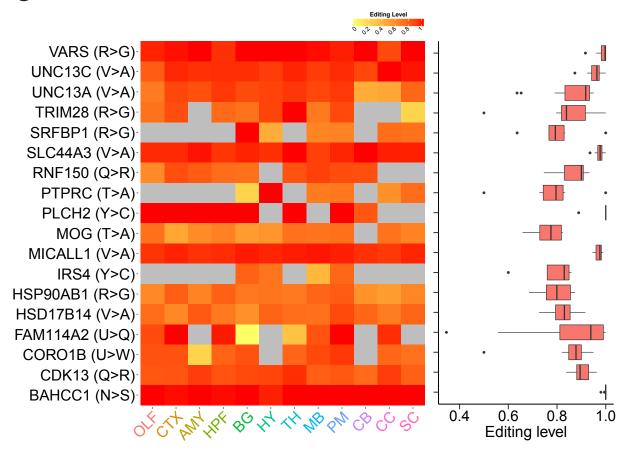


Fig. S17 | The highly edited non-conserved recoding sites in pig brain. The editing sites were > 75% edited in at least one brain region.

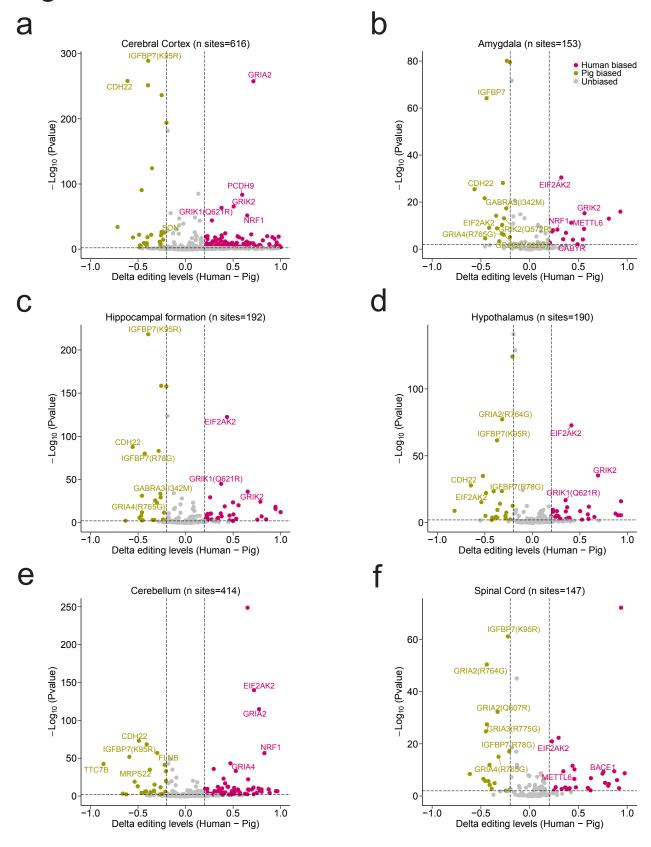


Fig. S18 | **Volcano plots showing species-biased editing sites in pig and human brain**. (a) cerebral cortex, (b) amygdala, (c) hippocampal formation, (d) hypothalamus, (e) cerebellum and (f) spinal cord. The X-axis represents the differential editing levels between pig and human. The Y-axis represents negative log₁₀ (Fisher's exact test p-value). For top differentially editing sites in genic region, their respective orthologous genes were indicated.

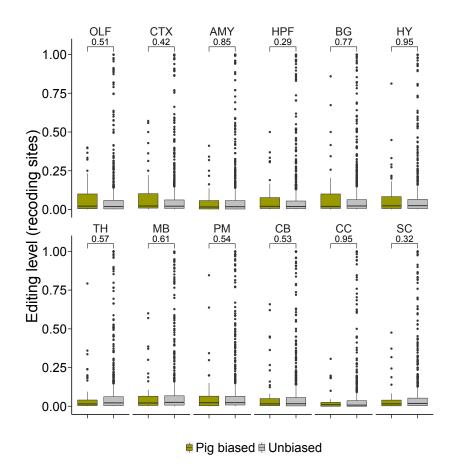


Fig. S19 | Box plot showing editing level of recoding sites located in genes showing pig-biased or unbiased expression. The significance of differences was assessed by Wilcoxon test.