## **Supplemental Materials**

Supplement Table 1. Sequence of ORF8-scFv-1 and ORF8-scFv-2.

#### ORF8-scFv-1:

#### ORF8-scFv-2:

# **Supplementary information**

## Antibodies

| Reagent or Resource              | Source                    | Identifier      |
|----------------------------------|---------------------------|-----------------|
| HLA-A2-APC (BB7.2)               | BD Biosciences            | Cat# 561341     |
| HLA-A,B,C-APC (W6/32)            | Biolegend                 | Cat# 311410     |
| HLA-A2-APC (BB7.2)               | Biolegend                 | Cat# 343308     |
| β2-microglobulin-PE (2M2)        | Biolegend                 | Cat# 316306     |
| Fixable Viability Dye eFluor 780 | Thermo Scientific         | Cat# 65-0865-14 |
| CD8-BV421 (RPA-T8)               | BD Biosciences            | Cat# 562428     |
| MHC class I Rabbit mAb           | Abclonal                  | Cat# A8754      |
| HLA Class I ABC Antibody         | Proteintech               | Cat# 15240-1-AP |
| HLA Class I Antibody             | Immunoway                 | Cat# YT5837     |
| Calreticulin (D3E6) Rabbit mAb   | Cell Signaling Technology | Cat# 12238T     |
| LAMP1 (D2D11) Rabbit mAb         | Cell Signaling Technology | Cat# 9091T      |
| LC3A/B (D3U4C) Rabbit mAb        | Cell Signaling Technology | Cat# 12741S     |
| GM130 (D6B1) Rabbit mAb          | Cell Signaling Technology | Cat# 12480S     |
| Rab5 (C8B1) Rabbit mAb           | Cell Signaling Technology | Cat# 3547T      |
| HLA A2 antibody                  | Abcam                     | Cat# ab168405   |

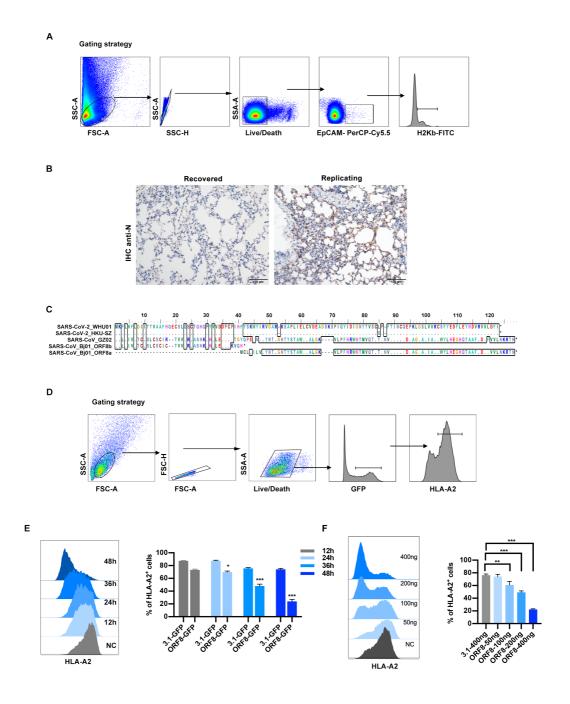
# **Chemicals, Peptides, and Recombinant Proteins**

| Reagent or Resource                       | Source            | Identifier       |
|---|-------------------|------------------|
| HLA-A cDNA ORF Clone, Human, C- Flag tag  | sinobiological    | Cat# HG13263-CF  |
| Calreticulin cDNA ORF Clone, Human, C-HA  | sinobiological    | Cat# HG13539-CY  |
| tag                                       |                   |                  |
| RAB5 cDNA ORF Clone                       | sinobiological    | Cat# HG14013-UT  |
| RAB7A cDNA ORF Clone, C- Flag tag         | sinobiological    | Cat# HG16168-NF  |
| Beta-2 microglobulin Protein, Human,      | sinobiological    | Cat# 11976-H08H  |
| Recombinant (His Tag)                     |                   |                  |
| Beclin 1 cDNA ORF Clone, Human, C-GFP tag | sinobiological    | Cat# HG11162-ACG |
|   |                   |                  |
| DSP                                       | Thermo Scientific | Cat# 22586       |
| Pepstatin                                 | TargetMol         | Cat# T3695       |
| DBeQ                                      | TargetMol         | Cat# T1969       |

| MG-132                   | TargetMol | Cat# T2154   |
|--------------------------|-----------|--------------|
| Aloxistatin (E64d)       | TargetMol | Cat# T6040   |
| Bafilomycin A1           | Selleck   | Cat# S1413   |
| Recombinant Human GM-CSF | Peprotech | Cat# 300-03  |
| Recombinant Human IL-4   | Peprotech | Cat# 200-04  |
| Recombinant Human TNF-α  | Peprotech | Cat# 300-01A |
| Recombinant Human IL-2   | Peprotech | Cat# 200-02  |
| Recombinant Human IL-10  | Peprotech | Cat# 200-10  |

# **Critical Commercial Assays**

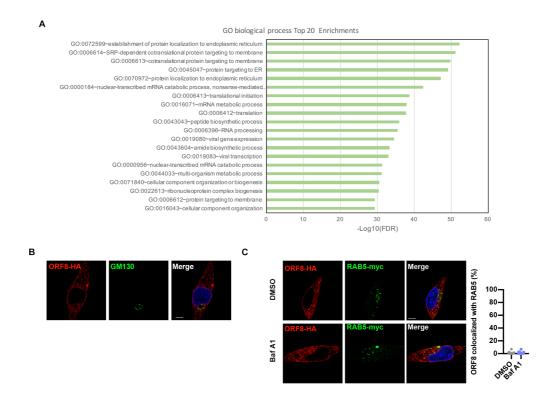
| Reagent or Resource                              | Source            | Identifier           |
|--|-------------------|----------------------|
| Sybr Green PCR Master Mix                        | Thermo Scientific | Cat# 4309155         |
| QuickSwitch Quant HLA-A*02:01<br>Tetramer Kit-PE | MBL               | Cat# TB-7300-K1      |
| Human CD8 T Lymphocyte Enrichment<br>Set-DM      | BD Biosciences    | Cat# 557941          |
| Lipofectamin RNAiMAX Transfection<br>Reagent     | Thermo Scientific | Cat# 13778150        |
| Lipofectamin 2000 Transfection Reagent           | Thermo Scientific | Cat# 11668019        |
| Lysosome Isolation Kit                           | Millipore Sigma   | Cat# LYSISO1         |
| ProteoSilver <sup>TM</sup> Silver Stain Kit      | Millipore Sigma   | Cat# PROTSIL1        |
| Human IFN-γ ELISpot assay kits                   | Dakewe            | Cat# DKW22-1000-096s |
| FITC Annexin V Apoptosis Detection Kit with PI   | Biolegend         | Cat# 640914          |



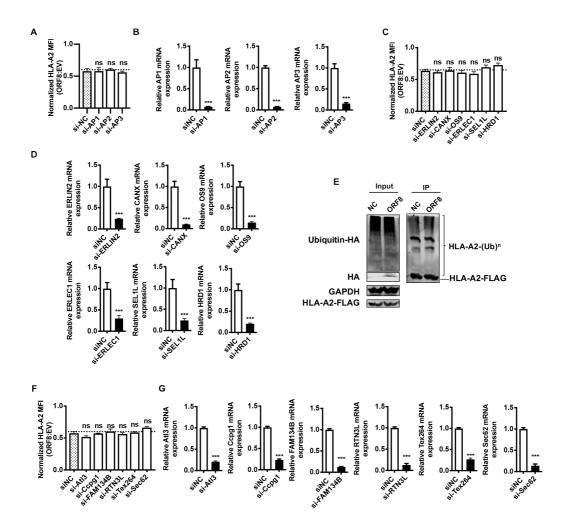
# Supplement Fig. 1 Surface expression of MHC-I is downregulated in ORF8-expressing cells.

(A) Gating strategy for FACS analysis of H2Kb expression on lung epithelial cells of mice. (B) hACE2 mice were intranasal infected with 4×10³ PFU (recovered), 4×10⁴ PFU (replicating) SARS-CoV-2 virus, or uninfected as control. At day 6 post infection, lung tissues were collected for IHC staining. (C) The multiple alignment was created based on the amino acid sequences of the encoded protein of orf8 gene, including 1 from SARS-CoV-2\_WHU01, 1 from SARS-CoV-2\_HKU-SZ with L84S

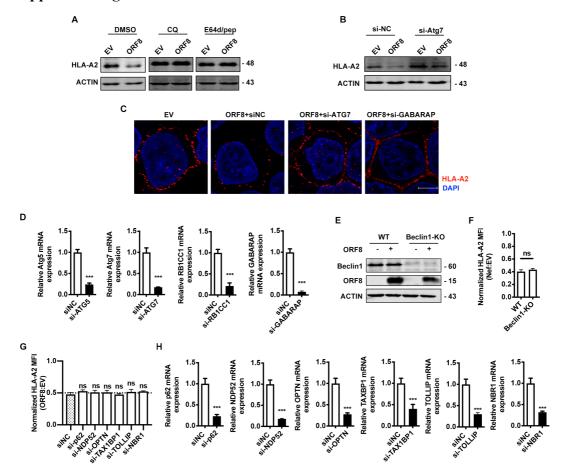
mutation, 1 from SARS-CoV\_GZ02, and 1 from SARS-CoV\_BJ01 orf8a, 1 from SARS-CoV\_BJ01 orf8b. The similarity shading with the color was referred to the chemistry of each amino acid at that position. **(D)** Gating strategy for FACS analysis of MHC-I expression on HEK293T cells used in this study. **(E)** GFP, or ORF8-GFP expressing plasmids were transfected into HEK293T cells, respectively. Cells were harvested for flow cytometry analysis at the indicated time points. Frequency of HLA-A2 $^+$  cells (gated on GFP $^+$  cells) were shown (n=5). **(F)** Different doses of GFP, or ORF8-GFP expressing plasmids were transfected into HEK293T cells, respectively. After 48 hours cells were harvested for flow cytometry. Frequency of HLA-A2 $^+$  cells (gated on GFP $^+$  cells) were shown (n=5). Data were shown as mean  $\pm$  SD (error bars). t test and one way ANOVA was used. P < 0.05 indicates statistically significance difference. \* indicates P < 0.05; \*\* indicates P < 0.01; \*\*\* indicates P < 0.001.



Supplement Fig.2 ORF8 interacting with endoplasmic reticulum (ER). (A) The top 20 significant enrichments of the gene ontology (GO) biological process terms. Transformed false discovery rate (FDR) was indicated at the X-axis. (B) Localization of SARS-CoV-2 ORF8 (red) relative to GM130 (green, the top panel). ORF8-HA expressing plasmid was transfected into HEK293T cells. At 24 hours after transfection, co-localization was visualized by confocal microscopy. Scale bars, 5μm. (C) Localization of SARS-CoV-2 ORF8 (red) relative to RAB5 (green). ORF8-HA expressing plasmid together with RAB5-myc expressing plasmid were transfected into HEK293T cells. At 24 hours after transfection, co-localization was visualized by confocal microscopy. Scale bars, 5μm.



Supplement Fig.3 ORF8 mediates MHC-I trafficking from ER to lysosome for degradation. (A) GFP (EV) or ORF8-GFP expressing plasmids, and the indicated siRNAs were transfected into HEK293T cells. MFI of HLA-A2 (gated on GFP<sup>+</sup> cells) was normalized to GFP group (n=5). (B) The knockdown efficiency of indicated siRNAs (n=3). (C) GFP (EV) or ORF8-GFP expressing plasmids, and the indicated siRNAs were transfected into HEK293T cells. MFI of HLA-A2 (gated on GFP<sup>+</sup> cells) was normalized to GFP group (n=5). (D) The knockdown efficiency of indicated siRNAs (n=3). (E) HLA-A2-FLAG and ubiquitin-HA expressing plasmids, in combination with ORF8-HA expressing plasmid or empty vector were transfected into HEK293T cells. Cells were treated with MG132 (10µM) for 12 h before harvest. HLA-A2 ubiquitination was analyzed by Co-IP with anti-Flag-tag beads and followed by western botting. (F) GFP (EV) or ORF8-GFP expressing plasmid, and the indicated siRNAs were transfected into HEK293T cells. MFI of HLA-A2 (gated on GFP<sup>+</sup> cells) was normalized to 3.1-GFP (n=5). (G) The knockdown efficiency of indicated siRNAs (n=3). Data were shown as mean  $\pm$  SEM (error bars). t test was used. P < 0.05 indicates statistically significance difference. \*\*\* indicates P < 0.001.



# Supplement Fig.4 ORF8 mediates MHC-I degradation though autophagy pathway.

(A) GFP (EV) or ORF8-GFP expressing plasmid were transfected into HEK293T cells. Before harvest, cells were treated with chloroquine (CQ) (50 µM), E64d (10ug/mL), or pepstatin A (pep) (10ug/mL) for 6 hours. The total HLA-A2 protein expression was analyzed by western blotting (n=3). (B) The GFP (EV) or ORF8-GFP expressing plasmids, and the indicated siRNAs were transfected into HEK293T cells. The total HLA-A2 protein expression was analyzed by western blotting (n=3). (C) The ORF8-HA expressing plasmid and the indicated siRNAs were transfected into HEK293T cells. At 48 hours after transfection, HLA-A2 localization was visualized by confocal microscopy. **(D)** The knockdown efficiency of indicated siRNAs (n=3). (E) GFP (EV) or ORF8-GFP expressing plasmids were transfected into HEK293T cells (WT), or Beclin1 knockout HEK293T cells. At 48 hours after transfection, cells were collected for western blotting (n=3). (F) GFP (EV) or Nef-GFP expressing plasmids were transfected into HEK293T cells (WT), or Beclin1 knockout HEK293T cells. MFI of HLA-A2 (gated on GFP<sup>+</sup> cells) was normalized to GFP group (n=5). (G) The GFP (EV) or ORF8-GFP expressing plasmids, in combination with the indicated siRNAs were transfected into HEK293T cells. MFI of HLA-A2 (gated on GFP<sup>+</sup> cells) was normalized to GFP group (n=5). (H) The knockdown efficiency of indicated siRNAs (n=3). t test was used. \*\*\* indicates P < 0.001.