



Supplementary Information for

Developmental influence on evolutionary rates and the origin of placental mammal tooth complexity

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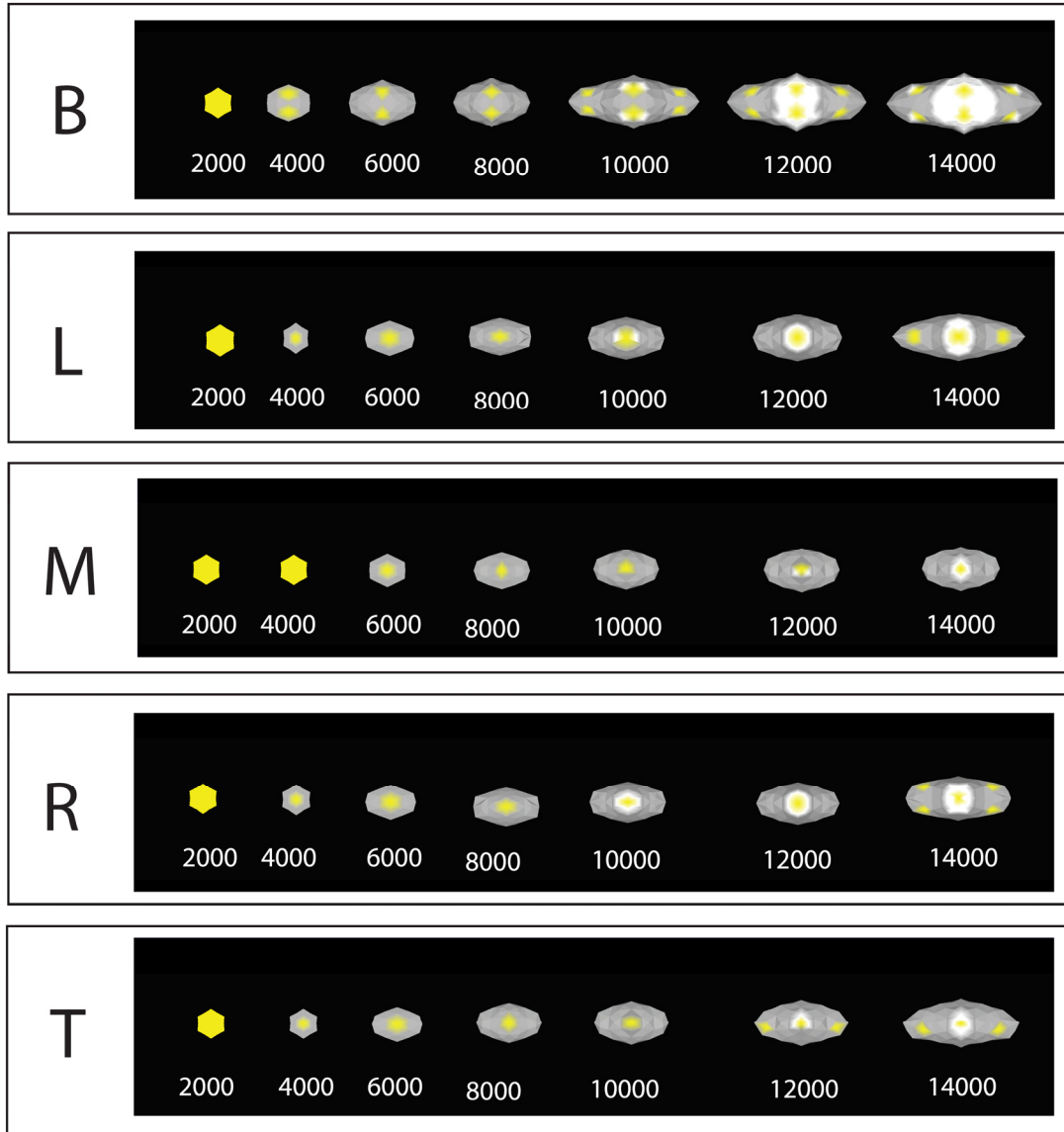


Figure S1. Occlusal view of enamel knot patterning histories. Enamel knots are indicated by yellow zones of epithelial differentiation. Numbers are model iterations (range 0–14000), a proxy for in silico developmental time. Knot pattern abbreviations: 'B', buccolingual (lateral); 'L', longitudinal; 'M', monocuspid; 'R', radial; 'T', triangular.

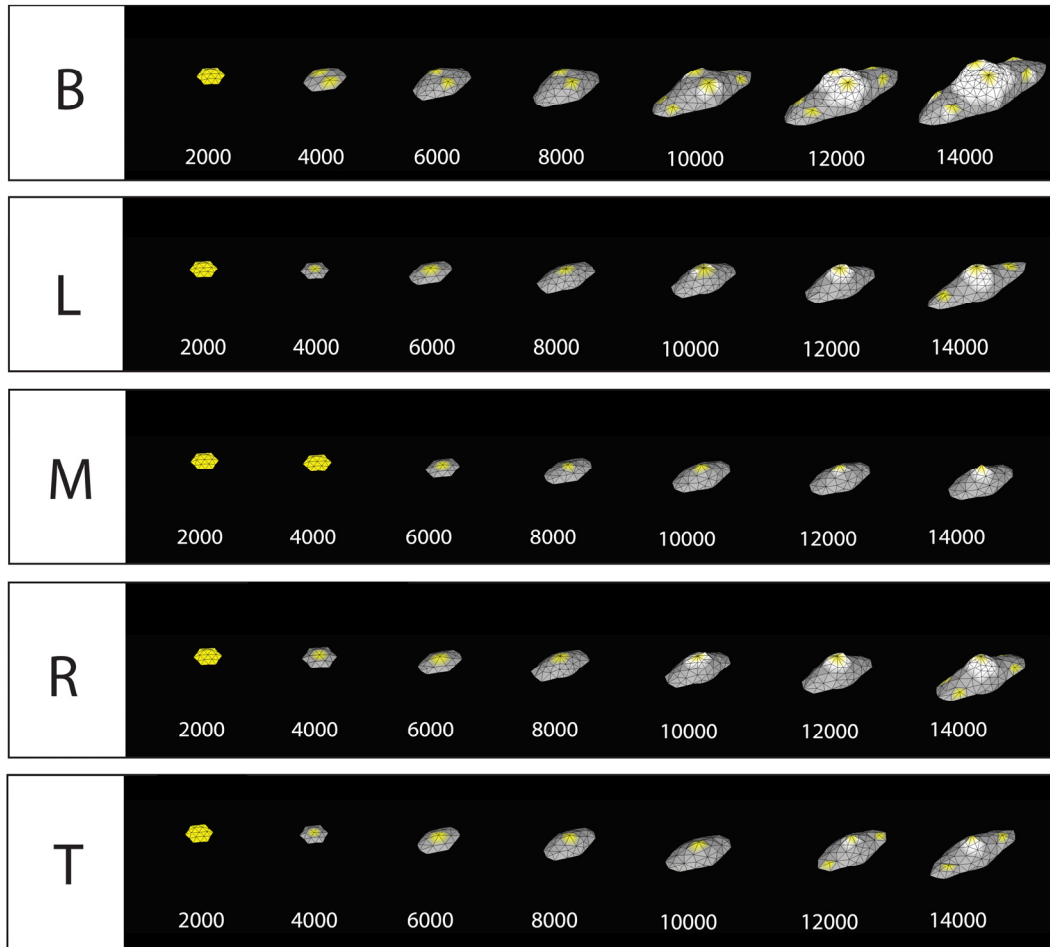


Figure S2. Oblique view of enamel knot patterning histories. Enamel knots are indicated by yellow zones of epithelial differentiation. Numbers are model iterations (range 0–14000), a proxy for developmental time. Knot pattern abbreviations: ‘B’, buccolingual (lateral); ‘L’, longitudinal; ‘M’, monocuspid; ‘R’, radial; ‘T’, triangular.

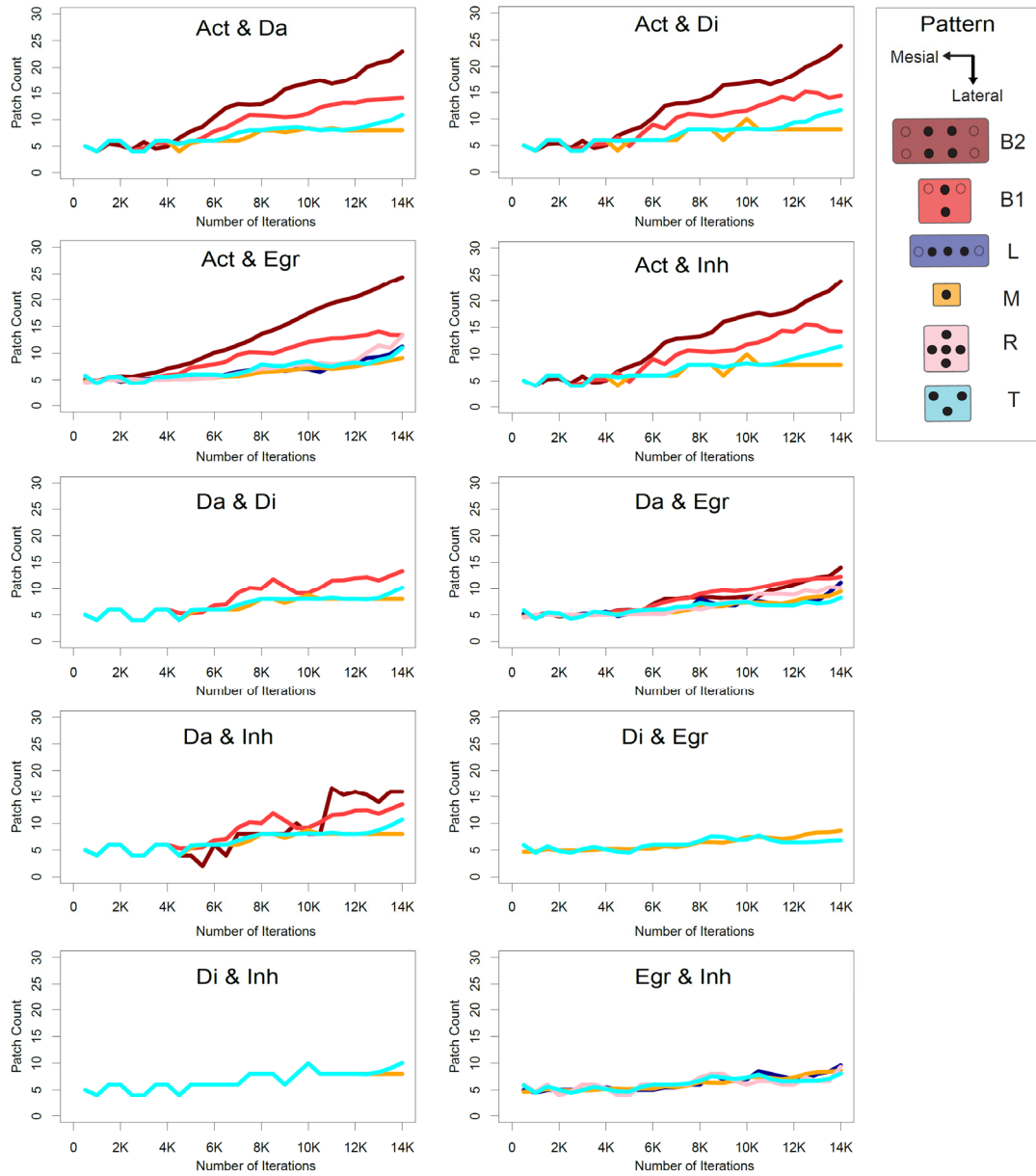


Figure S3. Developmental trajectories of tooth complexity for different enamel knot patterns. Tooth complexity measured with orientation patch count. Parameter abbreviations: Activator self-regulation (Act), activator diffusion rate (Da), inhibitor diffusion rate (Di), epithelial growth rate (Egr), and strength of inhibition (Inh).

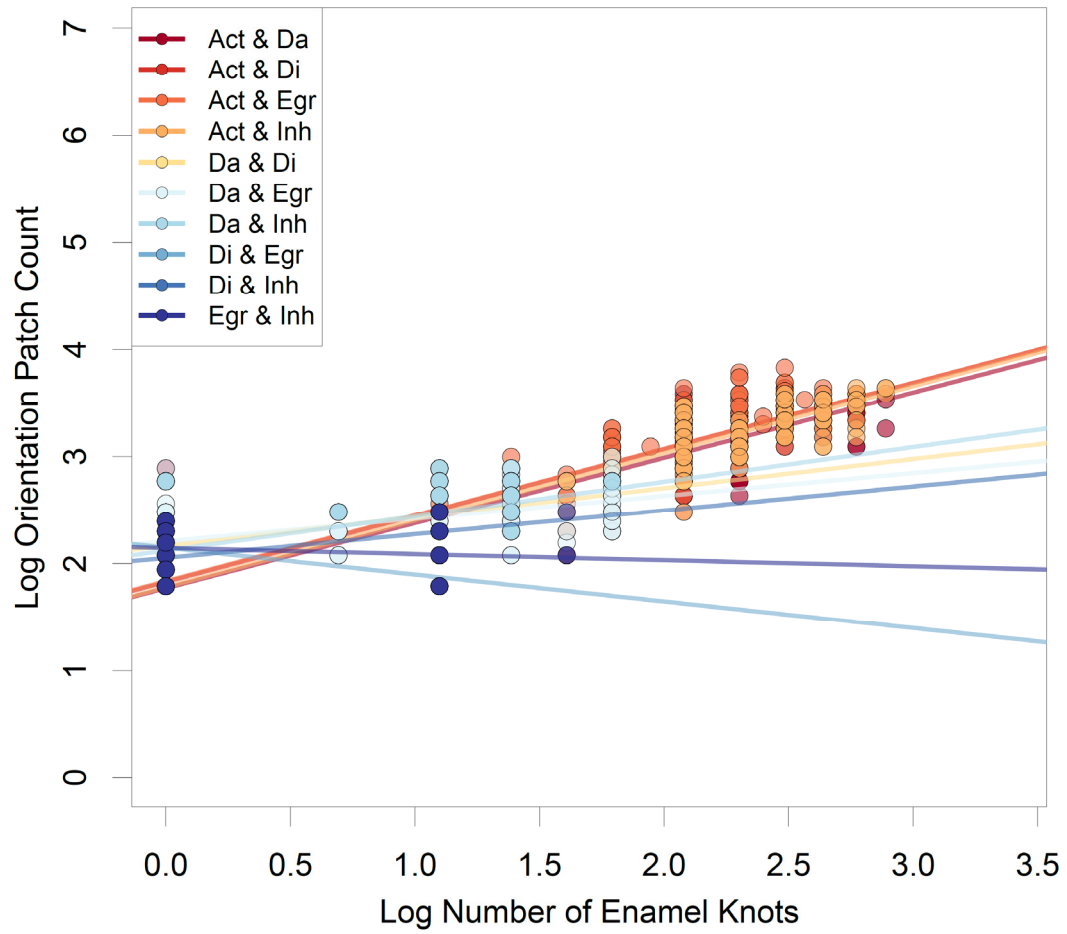


Figure S4. Log–Log major axis regressions of enamel knot number and orientation patch count for simulated tooth shapes (r score=0.69). Colors denote different developmental landscapes. Parameter abbreviations: Activator self-regulation (Act), activator diffusion rate (Da), inhibitor diffusion rate (Di), epithelial growth rate (Egr), and strength of inhibition (Inh).

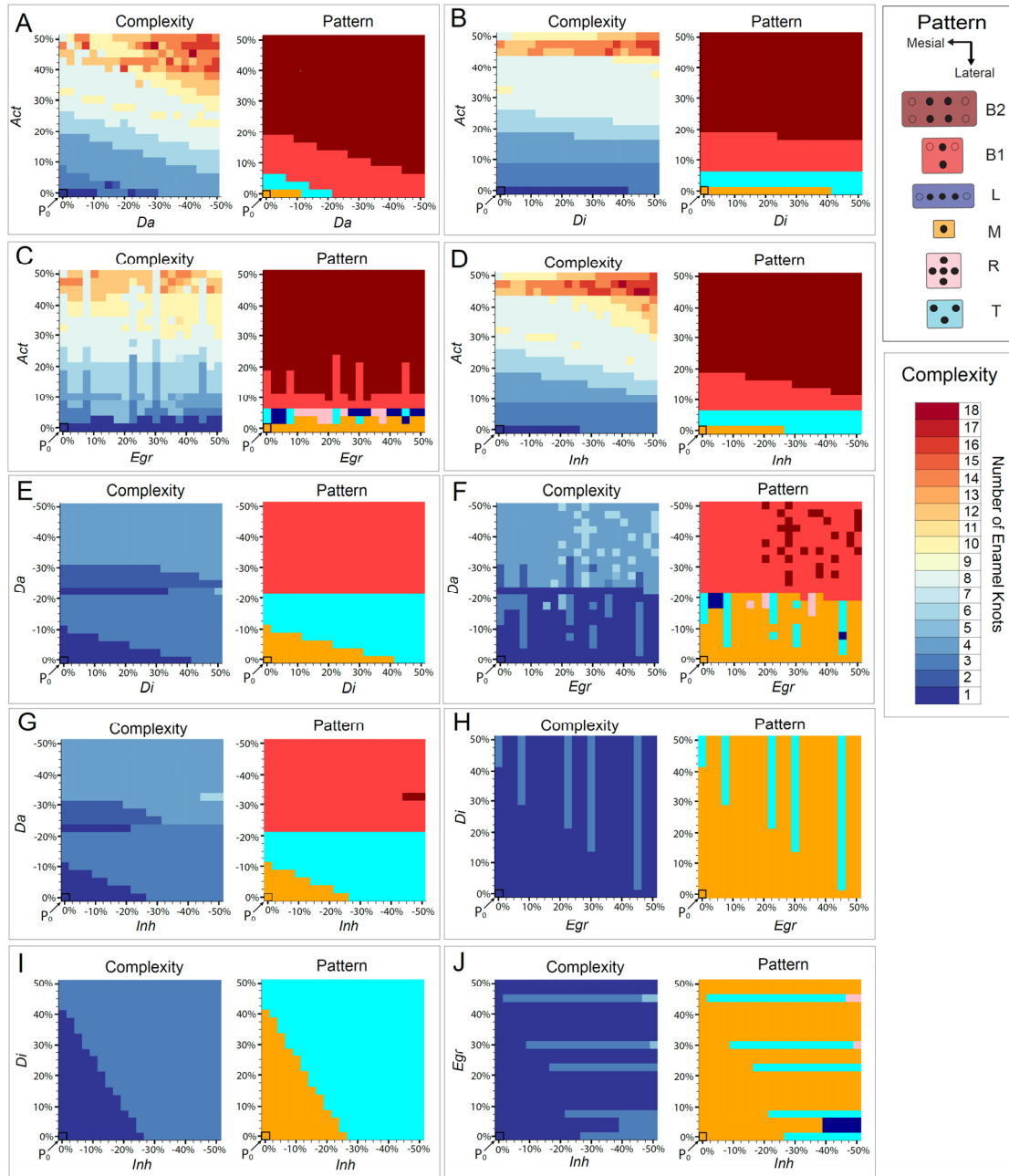


Figure S5. Heatmap representations of enamel knot number and pattern variation across different developmental landscapes. The starting parameter state (P_0) is indicated in the bottom left corner. Axis values denote percentage changes (\pm) in the parameter values relative to the ancestral state. Axis tick-marks denote the center of grid cells. Parameter abbreviations as follows: Activator self-regulation (Act), Activator diffusion rate (*Da*), inhibitor diffusion rate (*Di*), epithelial growth rate (*Egr*), and strength of inhibition (*Inh*).

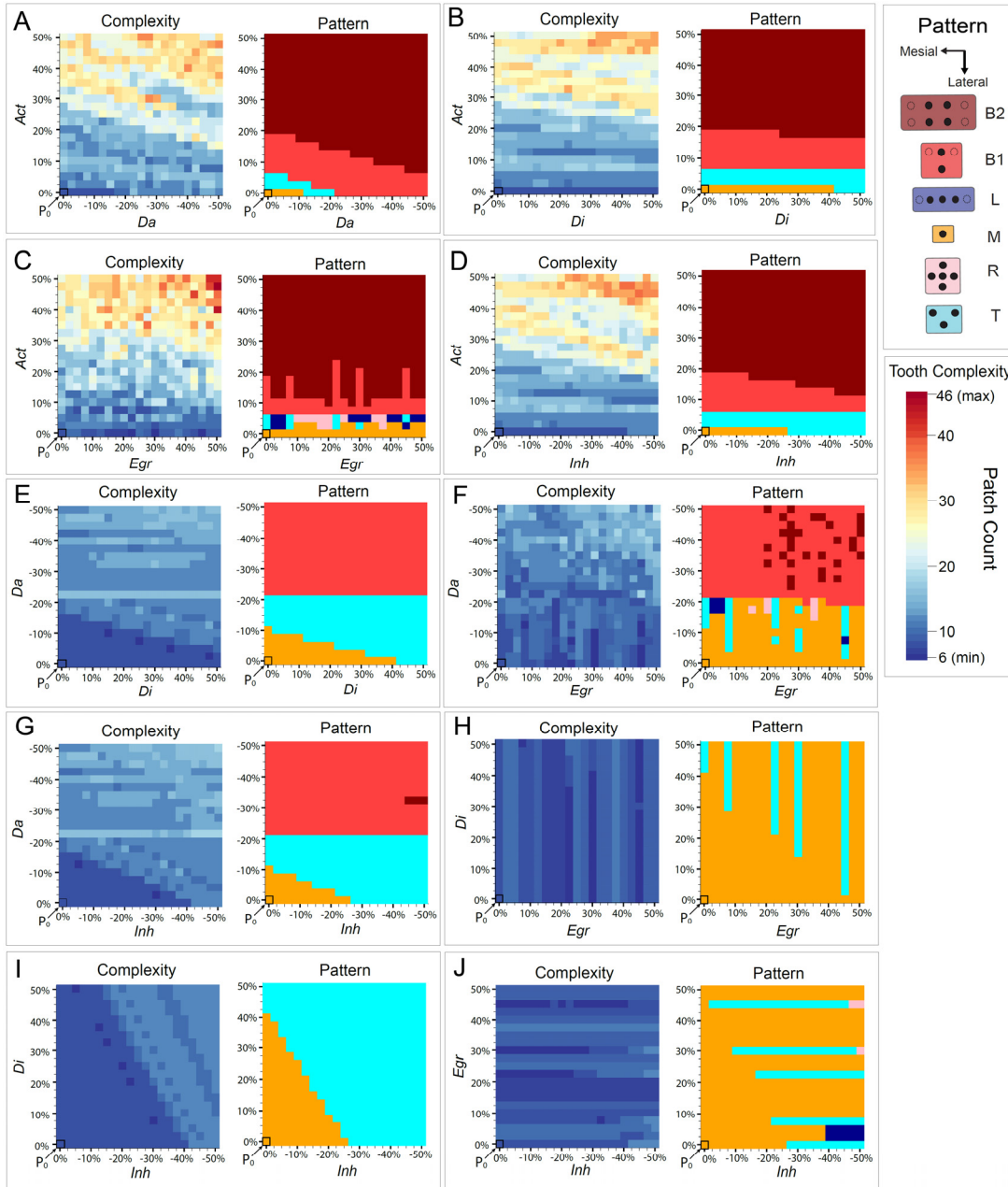


Figure S6. Heatmap representation of tooth surface complexity (orientation patch count) and knot pattern across different developmental landscapes. The starting parameter state (P_0) is indicated in bottom left corner. Axis values denote percentage changes (\pm) in parameter values relative to the ancestral state. Axis tick-marks denote the center of grid cells. Parameter abbreviations as follows: Activator self-regulation (Act), Activator diffusion rate (Da), inhibitor diffusion rate (Di), epithelial growth rate (Egr), and the strength of inhibition (Inh).

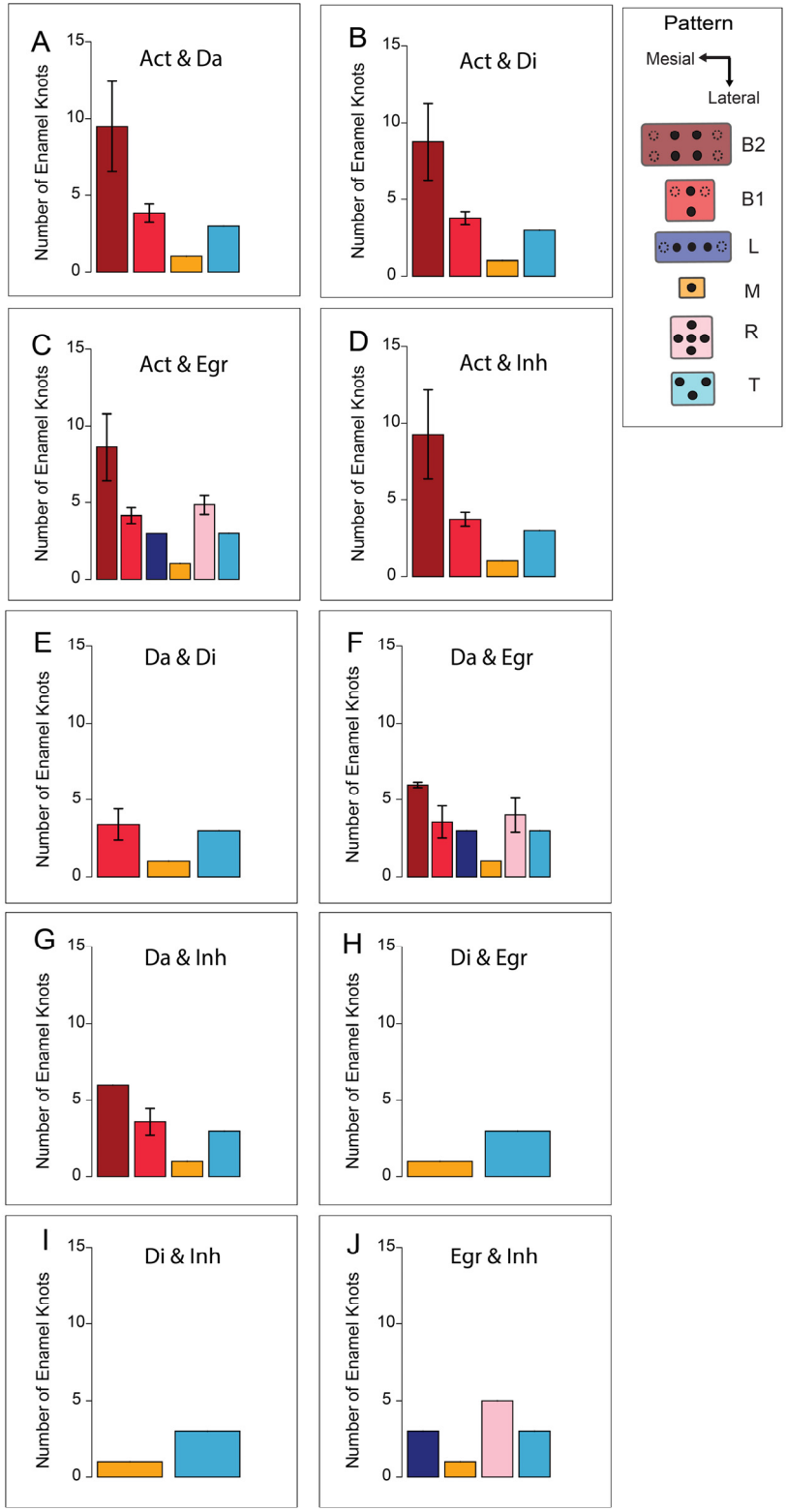


Figure S7. Bar plots of enamel knot number and pattern for different developmental landscapes. Error bars represent one standard deviation. Parameter abbreviations as follows: Activator self-regulation (Act), activator diffusion rate (Da), inhibitor diffusion rate (Di), epithelial growth rate (Egr), and the strength of inhibition (Inh).

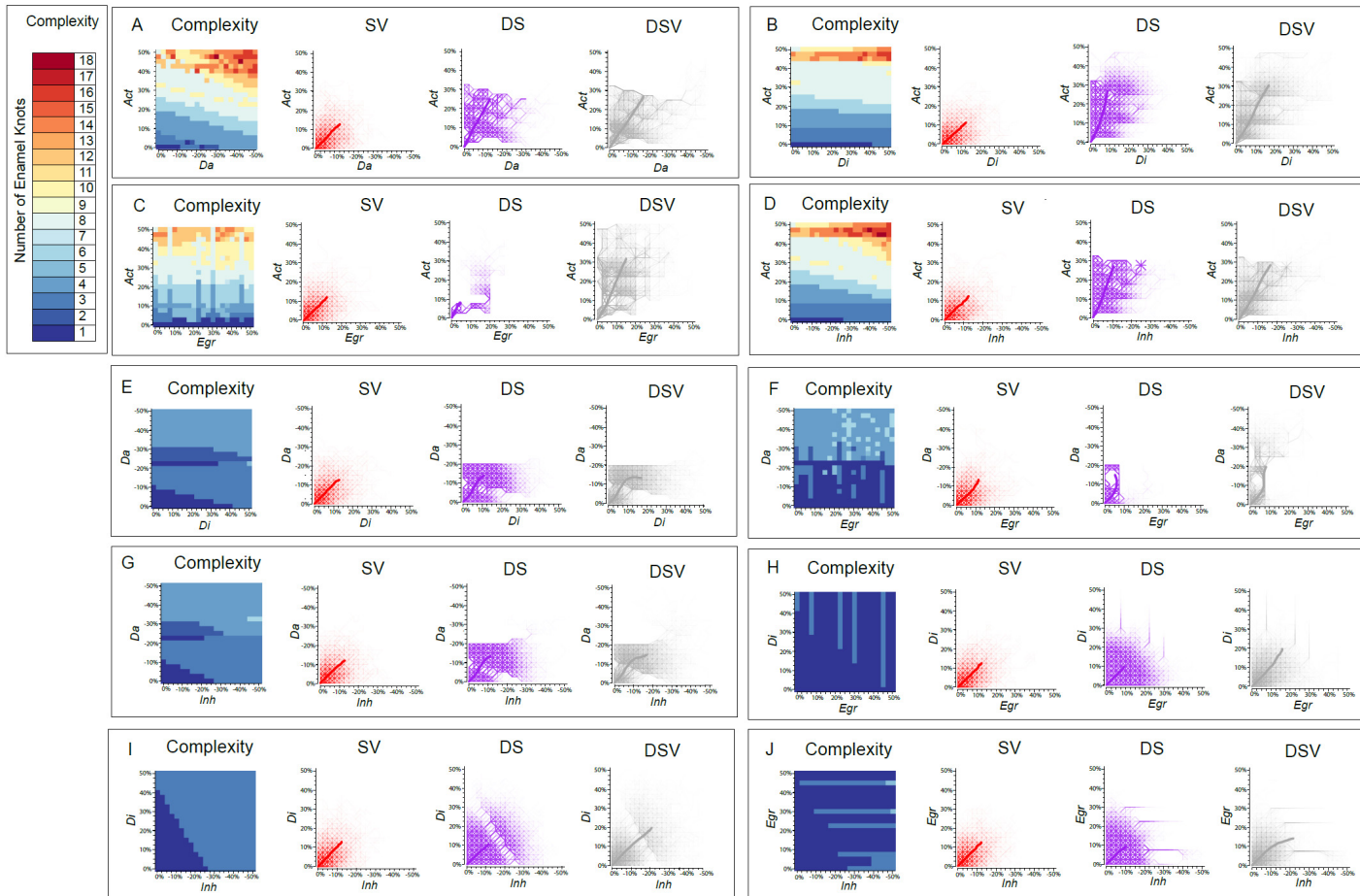


Figure S8. Evolutionary trajectories across different developmental landscapes under three evolutionary models. Bold lines are the average trajectory through the landscape (10,000 simulations, each run for a maximum of 30 generations). Landscapes are ordinated with the starting state (P_0) in the lower left corner. Faint lines are a random sample of 5,000 trajectories. Parameter abbreviations as follows: Activator self-regulation (Act), activator diffusion rate (Da), inhibitor diffusion rate (Di), epithelial growth rate (Egr), and strength of inhibition (Inh).

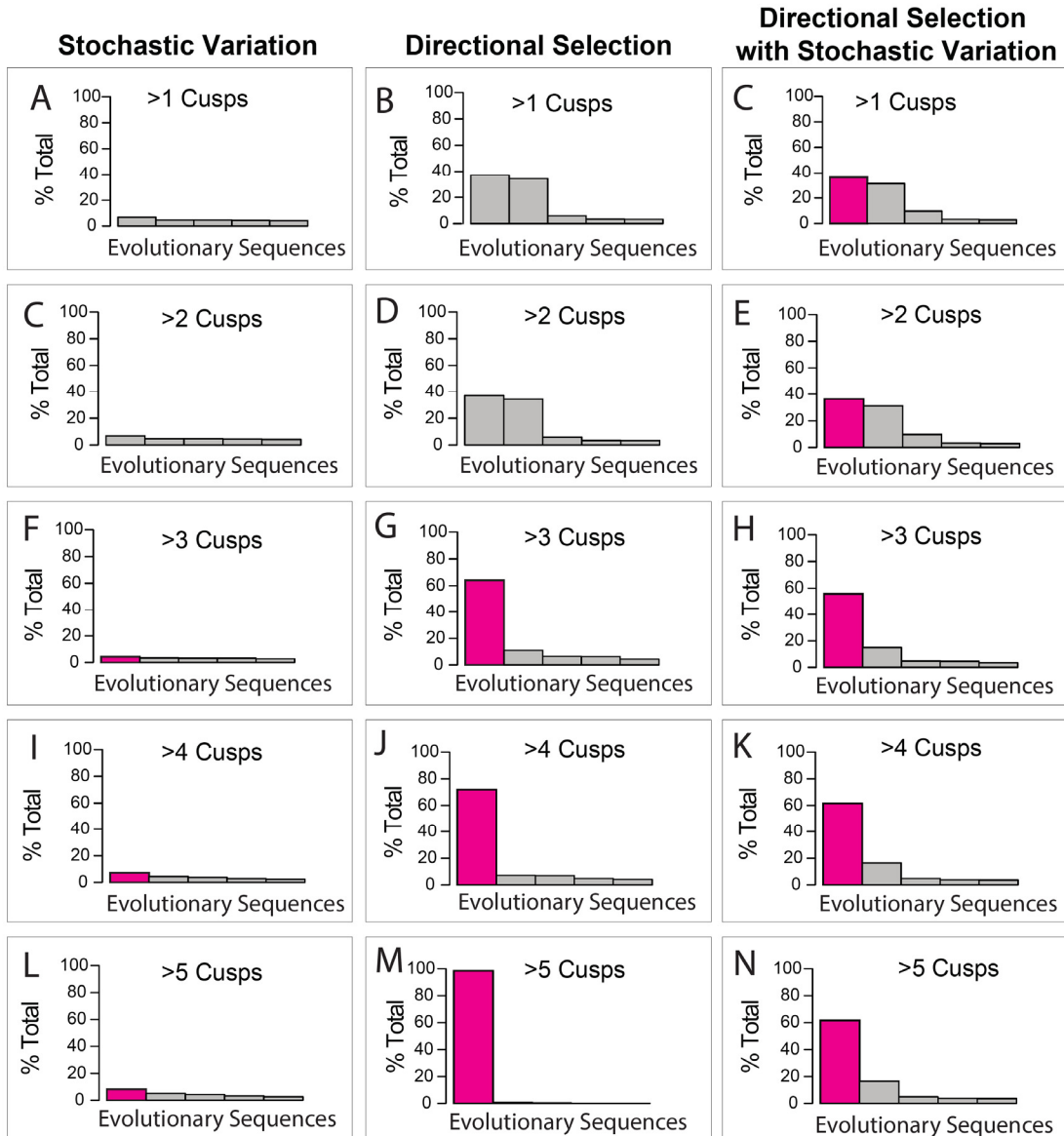
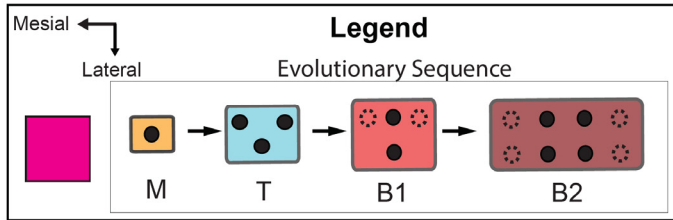


Figure S9. Frequency distributions of the most common evolutionary sequences for each evolutionary model at different tooth complexity levels: (A, C, F, I, L) stochastic variation (SV); (B, D, G, J, M) directional selection (DS), (C, E, H, K, N) directional selection combined with stochastic variation (DSV). Magenta highlights 'M>T>B1>B2' transformation sequence.

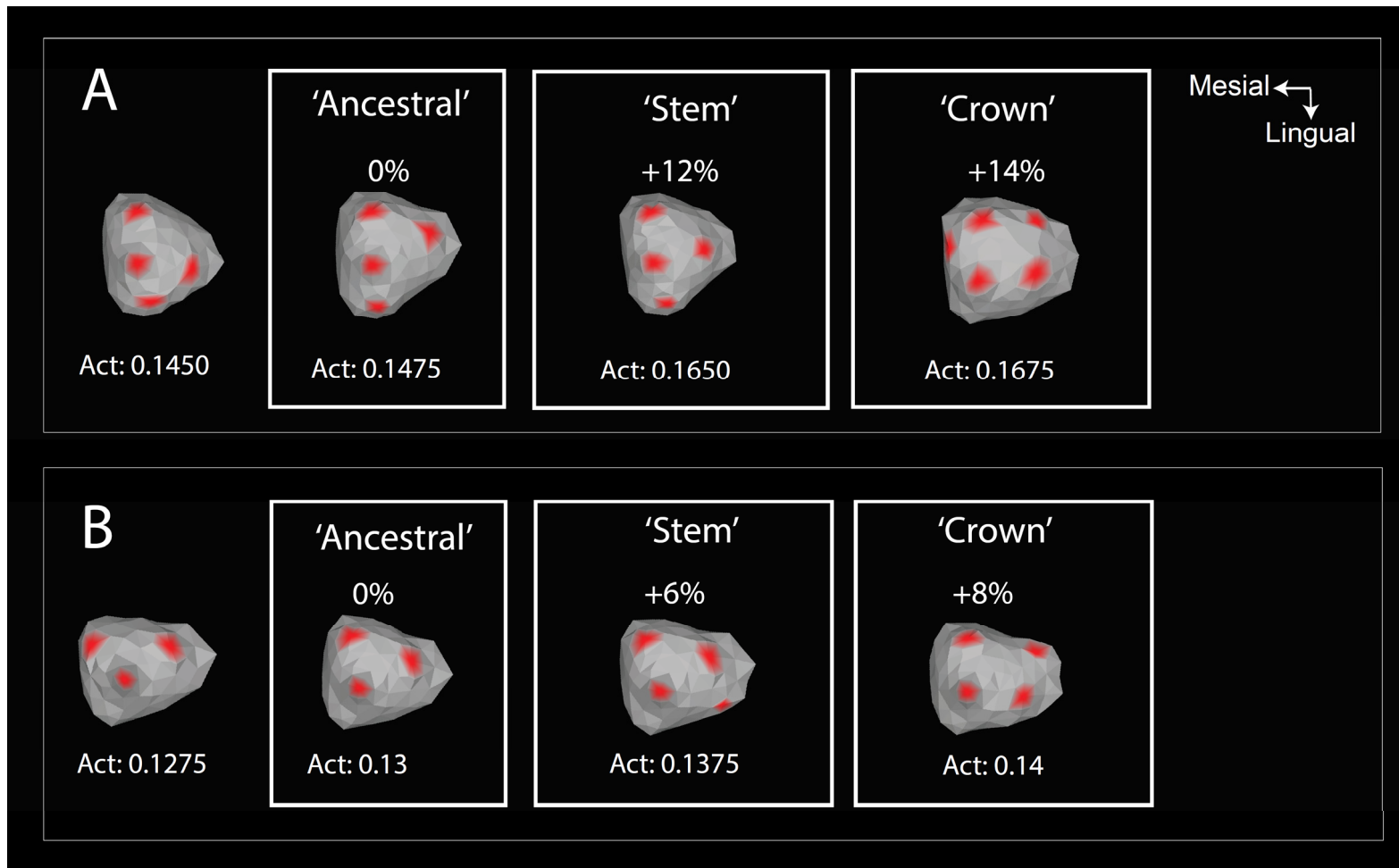


Figure S10. Sensitivity of different tooth bud growth biases to increases in activation (Act). (A) Enamel knot patterns originating from an 'ancestral' tooth bud with low levels of anterior growth, low levels of posterior growth, and a strong lingual bias in activator expression. (B) Enamel knot patterns originating from ancestral parameters as with 'A', but with more posterior growth. Red denotes growth factor concentration at view threshold=0.2.

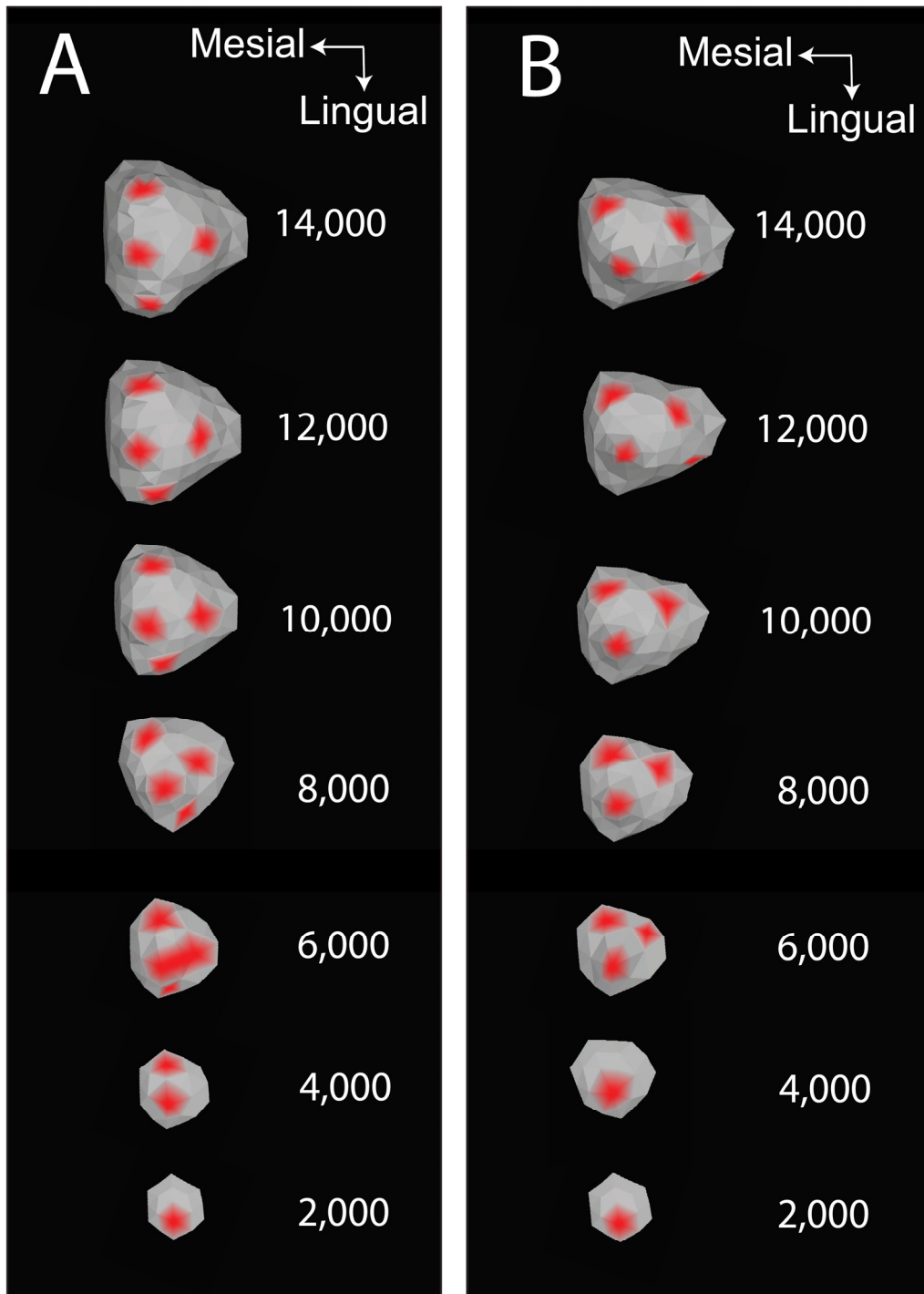


Figure S11. Patterning changes leading to distinct origins of posterolingual knots in ToothMaker simulations. (A) Formation of a lingual cingulum-like structure on an ancestral tooth subject to low-levels of anterior and posterior growth, and a strong lingual bias in activator expression. (B) As for (A), but with increased posterior growth. Red denotes growth factor concentration at view threshold=0.2. Numbers indicate the model iteration number, a proxy for developmental time.

Table S1. ToothMaker parameter settings used to generate the initial tooth state (P_0) for constructing the developmental landscapes and for the ancestral state in trait evolution simulations. Blue rows denote parameters that were varied in the pairwise simulations.

Parameter	Abbreviation	Class	Starting Value
Protein degradation rate	Deg	Degradation	0.076
Growth factor threshold	Set	Growth factor	0.95
Growth factor secretion rate	Sec	Growth factor	0.14
Growth factor diffusion rate	Ds	Growth factor	1.0
Activator self-regulation	Act	Activator	0.1
Activator diffusion rate	Da	Activator	0.05
Buccal bias	Bbi	Activator	0.1
Lingual bias	Lbi	Activator	0.1
Initial activator concentration	Ina	Activator	0.0
Initial inhibitor threshold	Int	Inhibitor	0.19
Inhibition of activator	Inh	Inhibitor	10.0
Inhibitor diffusion rate	Di	Inhibitor	1.0
Epithelial proliferation rate	Egr	Epithelium	0.01
Downward vector of growth	Dgr	Epithelium	10500.0
Differentiation rate	Dff	Epithelium	0.0002
Traction between neighboring cells	Adh	Epithelium	0.005
Young's modulus (stiffness)	Rep	Epithelium	1.5
Mechanical traction from the borders to the nucleus	Ntr	Epithelium	0.000013
Anterior bias	Abi	Epithelium	5.0
Posterior bias	Pbi	Epithelium	5.0
Mesenchymal proliferation rate	Mgr	Mesenchyme	1000.0
Border growth, amount of mesenchyme in the anterior–posterior region.	Bgr	Mesenchyme	0.9
Mesenchyme mechanical resistance	Boy	Mesenchyme	0.1
Radius of initial conditions	Rad	Initial size	3.0
Distance from 0 where the borders are defined	Swi	Initial size	1.4
Width of border	Bwi	Initial size	2.4

Table S2. Combinations of ToothMaker parameters used to construct developmental landscapes (blue). Parameter abbreviations: Activator self-regulation (Act), activator diffusion rate (Da), inhibitor diffusion rate (Di), epithelial growth rate (Egr), and inhibition of activation (Inh). P_{\min} : minimum parameter value; P_{\max} : maximum parameter value; Step: step size increment for each grid cell. Underlined values represent initial conditions (P_0).

Parameter	Act P_{\min} : <u>0.1</u> P_{\max} : <u>0.15</u> Step: 2.5×10^{-3}	Da P_{\min} : 2.5×10^{-2} P_{\max} : <u>5×10^{-2}</u> Step: 1.25×10^{-3}	Di P_{\min} : <u>1</u> P_{\max} : 1.5 Step: 2.5×10^{-2}	Egr P_{\min} : <u>1.0×10^{-2}</u> P_{\max} : 1.5×10^{-2} Step: 2.5×10^{-4}	Inh P_{\min} : 5 P_{\max} : <u>10</u> Step: 0.25
Act P_{\min} : <u>0.1</u> P_{\max} : 0.15 Step: 2.5×10^{-3}					
Da P_{\min} : 2.5×10^{-2} P_{\max} : <u>5×10^{-2}</u> Step: 1.25×10^{-3}					
Di P_{\min} : <u>1</u> P_{\max} : 1.5 Step: 2.5×10^{-2}					
Egr P_{\min} : <u>1.0×10^{-2}</u> P_{\max} : 1.5×10^{-2} Step: 2.5×10^{-4}					
Inh P_{\min} : 5 P_{\max} : <u>10</u> Step: 0.25					

Table S3. Variation in cusp pattern and cusp number on the upper second molar for 93 species of fossil and extant mammalian taxa. PPM=Protocone–Paracone–Metacone.

Taxon	Order/Clade	Status	Cusp Pattern	Cusp Number	PPM Angle
<i>Abdounodus hamdii</i>	Paenungulatomorpha	Fossil	T	6	64
<i>Adapis parisiensis</i>	Primates	Fossil	T	4	66
<i>Ailurus fulgens</i>	Carnivora	Extant	B2	13	85
<i>Ailuropoda melanoleuca</i>	Carnivora	Extant	B2	>13	88
<i>Amblysomus hottentotus</i>	Afrosoricida	Extant	T	4	65
<i>Apheliscus insidiosus</i>	Macroscelidea	Extant	B1	3	93
<i>Archaeonycteris brailloni</i>	Chiroptera	Fossil	B1	5	88
<i>Archaeotherium mortoni</i>	Artiodactylomorpha	Fossil	T	5	75
<i>Artiocetus clavis</i>	Artiodactyla	Fossil	T	3	NA
<i>Basilosaurus cetoides</i>	Artiodactyla	Fossil	L	6	NA
<i>Bos taurus</i>	Artiodactyla	Extant	B2	8	89
<i>Cantius abditus</i>	Primates	Fossil	T	8	64
<i>Caperea marginata</i>	Artiodactyla	Extant	E	0	NA
<i>Castor canadensis</i>	Rodentia	Extant	B2	7	90
<i>Cavia porcellus</i>	Rodentia	Extant	B2	5	90
<i>Cocomys lingchaensis</i>	Glirres	Fossil	B1	8	81
<i>Cymbalophus cuniculus</i>	Perissodactyla	Fossil	T	4	76
<i>Cynocephalus volans</i>	Dermoptera	Extant	T	8	67
<i>Dasypus novemcinctus</i>	Cingulata	Extant	M	1	NA
<i>Dawsonolagus antiquus</i>	Glirres	Fossil	T	4	70
<i>Diacodexis antunesi</i>	Artiodactyla	Fossil	B1	5	80
<i>Didelphis virginiana</i>	Didelphimorphia	Extant	T	7	67
<i>Didolodus multicuspis</i>	Meridiungulata	Fossil	T	6	70
<i>Dormaalocyon latouri</i>	Carnivoraformes	Fossil	B1	5	99
<i>Dromiciops gliroides</i>	Microbiotheria	Extant	B1	6	82
<i>Echinops telfairi</i>	Afrosoricida	Extant	B1	4	95
<i>Ectocion parvus</i>	Euungulata	Fossil	T	8	60
<i>Equus caballus</i>	Perissodactyla	Extant	B2	9	97
<i>Erinaceus europaeus</i>	Eulipotyphla	Extant	T	5	55
<i>Eritherium azzouzororum</i>	Tethytheria	Fossil	B2	5	97
<i>Galeopterus variegatus</i>	Dermoptera	Extant	T	8	65
<i>Gomphos elkema</i>	Glirres	Fossil	T	4	62
<i>Gracilocyon solei</i>	Carnivoraformes	Fossil	B1	6	86
<i>Haldanodon exspectatus</i>	Docodonta	Fossil	T	2	NA
<i>Hapalops elongatus</i>	Pilosa	Fossil	M	1	NA
<i>Henkelotherium guimarotae</i>	Dryolestida	Fossil	T	4	43
<i>Hippopotamus amphibius</i>	Artiodactyla	Extant	B2	7	81
<i>Homo sapiens</i>	Primates	Extant	T	4	70

<i>Hyopsodus paulus</i>	Perissodactyla	Fossil	T	6	56
<i>Icaronycteris menui</i>	Chiroptera	Fossil	B1	5	88
<i>Ictidomys tridecemlineatus</i>	Rodentia	Extant	B2	3	87
<i>Lama glama</i>	Artiodactyla	Extant	B2	7	91
<i>Lemur catta</i>	Primates	Extant	T	5	78
<i>Leptictis dakotensis</i>	Leptictida	Fossil	B1	4	80
<i>Loxodonta africana</i>	Proboscidea	Extant	B2	17	95
<i>Lycophocyon hutchisoni</i>	Carnivora	Fossil	T	3	72
<i>Maelestes gobiensis</i>	Cimolesta	Fossil	B1	4	91
<i>Manis pentadactyla</i>	Pholidota	Extant	E	0	NA
<i>Meshippus bairdi</i>	Perissodactyla	Fossil	T	5	73
<i>Mesonyx obtusidens</i>	Mesonychia	Fossil	T	4	67
<i>Metacheiromys marshi</i>	Palaeanodonta	Fossil	E	0	NA
<i>Moeritherium lyonsi</i>	Proboscidea	Fossil	B2	5	81
<i>Morganucodon watsoni</i>	Morganucodonta	Fossil	L	3	NA
<i>Myotis lucifugus</i>	Chiroptera	Extant	T	6	73
<i>Notharctus tenebrosus</i>	Primates	Fossil	T	7	65
<i>Nycteris thebaica</i>	Chiroptera	Extant	T	7	76
<i>Ocepeia grandis</i>	Paenungulatomorpha	Fossil	T	5	63
<i>Orycteropus afer</i>	Tubulidentata	Extant	B2	4	90
<i>Oryctolagus cuniculus</i>	Lagomorpha	Extant	B2	4	89
<i>Paramys delicatus</i>	Rodentia	Fossil	T	4	47
<i>Phenacodus intermedius</i>	Euungulata	Fossil	T	6	62
<i>Phosphatherium escullei</i>	Proboscidea	Fossil	B2	5	90
<i>Plesiadapis ploegi</i>	Plesiadapiformes	Fossil	T	6	61
<i>Procavia capensis</i>	Hyracoidea	Extant	T	5	62
<i>Protictis microlestes</i>	Carnivoramorpha	Fossil	T	6	66
<i>Protolipterna ellipsodontoides</i>	Litopterna	Fossil	T	6	63
<i>Protungulatum donnae</i>	Eutheria	Fossil	T	6	67
<i>Pteronotus parnellii</i>	Chiroptera	Extant	B2	7	84
<i>Pteropus giganteus</i>	Chiroptera	Extant	B1	2	NA
<i>Ptilocercus lowii</i>	Scandentia	Extant	T	3	60
<i>Pucadelphys andinus</i>	Marsupialiformes	Fossil	B1	9	80
<i>Purgatorius titusi</i>	Primates	Fossil	T	4	79
<i>Rattus norvegicus</i>	Rodentia	Extant	B2	6	NA
<i>Rhinopoma hardwickii</i>	Chiroptera	Extant	B1	7	91
<i>Rhombomylus turpanensis</i>	Glires	Fossil	T	5	50
<i>Rhynchocyon cirnei</i>	Macroscelidea	Extant	T	4	73
<i>Saccopteryx bilineata</i>	Chiroptera	Extant	B2	7	101
<i>Saimiri sciureus</i>	Primates	Extant	T	4	70
<i>Solenodon paradoxus</i>	Euliptophyla	Extant	B1	6	88

<i>Sorex araneus</i>	Euliptophyla	Extant	T	7	74
<i>Sus scrofa</i>	Artiodactyla	Extant	B2	6	80
<i>Tachyglossus aculeatus</i>	Monotremata	Extant	E	0	NA
<i>Talpa europaea</i>	Euliptophyla	Extant	B2	6	80
<i>Tamandua tetradactyla</i>	Pilosa	Extant	E	0	NA
<i>Tarsius syrichta</i>	Primates	Extant	T	3	78
<i>Todralestes variabilis</i>	Eutheria	Fossil	T	4	78
<i>Tribosphenomys minutus</i>	Glires	Fossil	T	4	74
<i>Trichechus manatus</i>	Sirenia	Extant	B2	6	87
<i>Tupaia glis</i>	Scandentia	Extant	T	6	70
<i>Tursiops truncatus</i>	Artiodactyla	Extant	M	1	NA
<i>Uintacyon rudis</i>	Carnivoraformes	Fossil	B1	4	83
<i>Vulpavus ovatus</i>	Carnivoraformes	Fossil	T	3	69
<i>Zalambdalestes lechei</i>	Eutheria	Fossil	T	4	78

Table S4. ToothMaker parameters states used to generate 'ancestral' placental tooth shapes. 'Ancestral state 1' = Fig. 5A, 'Ancestral state 2' = Fig. 5B.

Parameter	Abbreviation	Ancestral State 1	Ancestral State 2
Protein degradation rate	Deg	0.076	0.076
Growth factor threshold	Set	0.95	0.95
Growth factor secretion rate	Sec	0.14	0.14
Growth factor diffusion rate	Ds	1.0	1.0
Activator self-regulation	Act	0.1475	0.13
Activator diffusion rate	Da	0.06	0.06
Buccal bias	Bbi	0.00001	0.00001
Lingual bias	Lbi	0.5	0.5
Initial activator concentration	Ina	0.0	0.0
Initial inhibitor threshold	Int	0.19	0.19
Inhibition of activator	Inh	11.0	11.0
Inhibitor diffusion rate	Di	1.50	1.50
Epithelial proliferation rate	Egr	0.012	0.012
Downward vector of growth	Dgr	10500	10500
Differentiation rate	Dff	0.0002	0.0002
Traction between neighboring cells	Adh	0.005	0.005
Young's modulus (stiffness)	Rep	1.5	1.5
Mechanical traction from the borders to the nucleus	Ntr	0.00013	0.00013
Anterior bias	Abi	0.2	0.2
Posterior bias	Pbi	2.5	3.0
Mesenchymal proliferation rate	Mgr	1000	1000
Border growth, amount of mesenchyme in the anterior–posterior region.	Bgr	0.9	0.9
Mesenchyme mechanical resistance	Boy	0.1	0.1
Radius of initial conditions	Rad	3.0	3.0
Distance from 0 where the borders are defined	Swi	1.5	1.5
Width of border	Bwi	3.0	3.0