

Appendix

Transneuronal Dpr12/DIP- δ interactions facilitate compartmentalized dopaminergic innervation of *Drosophila* mushroom body axons

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List of genotypes used in this study:

hsFLP is *y,w,hsFLP122*; tdT is *tdTomato*; mtdT-HA is *mtdTomato-3xHA*; 40A and G13 are FRTs on 2L and 2R respectively; R71G10 is *GMR71G10-Gal4*; R18H09 is *GMR18H09-Gal4*; R58E02 is *GMR58E02-Gal4*; R10G03 is *GMR10G03-Gal4*; R48H011 is *GMR48H011-Gal4*; G80 is *TubP-Gal80*. Males and females were used interchangeably but only the female genotype is mentioned.

Figure 1

(C) *y, w; R71G10, UAS-mCD8-GFP/+; R71G10, UAS-Dcr-2/+*
(D) *y, w; R71G10, UAS-mCD8-GFP/+; R71G10, UAS-Dcr-2/ TRiP.JF03210(dpr12-RNAi)*
(E) *y, w; R71G10, UAS-mCD8-GFP/+; UAS-Cas9.C*
(F) *y, w; R71G10, UAS-mCD8-GFP/+; UAS-Cas9.C/dpr12-gRNA*
(G-I) *y, w; UAS-mCD8-GFP/+; R71G10/+*
(J) *y, w; UAS-mCD8-GFP/+; R71G10/UAS-Dpr12*
(K-M) *y, w; 40A, dpr12 Δ^{50-81} , G13/40A, dpr12 Δ^{50-81} , G13; R71G10, UAS-mCD8-GFP/+*
(N) *y, w; 40A, dpr12 Δ^{50-81} , G13/40A, dpr12 Δ^{50-81} , G13; R71G10, UAS-mCD8-GFP/UAS-Dpr12*

Figure 2

(A-C, G-I) *hsFLP, UAS-mCD8-GFP; Gal80, FRT40A/FRT40A; R71G10, UAS-Dcr-2/+*
(D-F, J-L) *hsFLP, UAS-mCD8-GFP; Gal80, FRT40A/FRT40A; R71G10, UAS-Dcr-2/ TRiP.JF03210 (dpr12-RNAi)*
(O) *y, w; UAS-mCD8-GFP/+; R18H09/+*

Figure 3

(A-C) *y, w; Dpr12^{GFSTF}/+*
(D) *y, w; UAS-CD4-tdT/Dpr12^{GFSTF}; R18H09/+*
(E-G) *y, w;; DIP- δ ^{GFSTF}/+*
(H) *y, w; UAS-CD4-tdT/+; DIP- δ ^{GFSTF}/R18H09*

Figure 4

(A-C) *y, w; R71G10-QF2, QUAS-mtdT-HA/+; DIP- δ ^{T2A-Gal4}/+*
(D) *y, w; R71G10-QF2, QUAS-mtdT-HA/UAS-DIP- δ ; DIP- δ ^{T2A-Gal4}/+*
(E-G) *y, w; R71G10-QF2, QUAS-mtdT-HA/+; DIP- δ ^{T2A-Gal4}/DIP- δ ^{T2A-Gal4}*
(H) *y, w; R71G10-QF2, QUAS-mtdT-HA/UAS-DIP- δ ; DIP- δ ^{T2A-Gal4}/DIP- δ ^{T2A-Gal4}*
(J) *y, w; UAS-DIP- δ -RNAi/R71G10-QF2, QUAS-mtdT-HA; Repo-Gal4, UAS-mCD8-GFP/+*
(K) *y, w; UAS-DIP- δ -RNAi/R71G10-QF2, QUAS-mtdT-HA; OK107-Gal4*
(L) *C155-Gal4; UAS-DIP- δ -RNAi/R71G10-QF2, QUAS-mtdT-HA*
(M) *y, w; UAS-DIP- δ -RNAi/R71G10-QF2, QUAS-mtdT-HA; DIP- δ ^{T2A-Gal4}/+*

Figure 5

(A) *UAS-mCD8-RFP; R53C03-p65.AD/+; DIP- δ ^{GFSTF}/R24E12-GAL4.DBD*
(B) *UAS-mCD8-RFP; R53C03-p65.AD/UAS-DTi; DIP- δ ^{GFSTF}/R24E12-GAL4.DBD*

(C) y, w; UAS-CD4-tdT/+; DIP- δ ^{GFSTF}/R58E02
(D) y, w; UAS-CD4-tdT/UAS-DTi; DIP- δ ^{GFSTF}/R58E02
(E-G) hsFLP, UAS-mCD8-GFP; Gal80, FRT40A/FRT40A; *DIP- δ ^{T2A-Gal4}/+*

Figure 6

(A-D) y, w; Dpr12^{GFSTF}/+; *DIP- δ ^{T2A-Gal4}/DIP- δ ^{T2A-Gal4}*
(E-H) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13; DIP- δ ^{GFSTF}/+
(I) y, w; Dpr12^{GFSTF}/+; UAS-DIP- δ -T2A-tdT/+
(J) y, w; Dpr12^{GFSTF} /R30G08-p65.AD; R48B03-GAL4.DBD/ UAS-DIP- δ -T2A-tdT

Figure 7

(A) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/+; R10G03/UAS-mCD8-GFP
(B) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13; R10G03/UAS-mCD8-GFP
(C) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/+; R48H011/UAS-mCD8-GFP
(D) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13; R48H011/UAS-mCD8-GFP
(E) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/+; R18H09/UAS-mCD8-GFP
(F) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13; R18H09/UAS-mCD8-GFP
(G) w¹¹¹⁸
(H) w¹¹¹⁸; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13

Figure 8

(A) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- δ ^{T2A-Gal4}/DIP- δ ¹⁻¹¹⁹*
(B) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- δ ^{T2A-Gal4}/DIP- δ ¹⁻¹¹⁹*, UAS-DIP- α
(C) y,w; Dpr12^{GFSTF}/+; *DIP- δ ¹⁻¹¹⁹*, UAS-DIP- α /+
(D) y,w; Dpr12^{GFSTF}/+; *DIP- δ ¹⁻¹¹⁹*, UAS-DIP- α /DIP- δ ^{T2A-Gal4}
(E) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13; *DIP- δ ^{T2A-Gal4}*/ UAS-mtdT
(F) y, w; 40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *dpr12 Δ ⁵⁰⁻⁸¹*, G13; *DIP- δ ^{T2A-Gal4}*/ UAS-DIP- α -T2A-tdT

Figure EV1

(A)
y, w; R71G10, UAS-mCD8-GFP/+; R71G10, UAS-Dcr-2/ TRiP.JF03306(Dpr5-RNAi)
y, w; UAS-mCD8-GFP/+; R71G10/ TRiP.JF03172 (Dpr8-RNAi)
y, w; UAS-mCD8-GFP/+; R71G10/ TRiP.HMS00288 (Dpr9-RNAi)
y, w; R71G10, UAS-mCD8-GFP/+; R71G10, UAS-Dcr-2/ TRiP.JF02920 (Dpr10-RNAi)
y, w; R71G10, UAS-mCD8-GFP/+; R71G10, UAS-Dcr-2/ TRiP.JF03210(Dpr12-RNAi)
y, w; UAS-mCD8-GFP/+; R71G10/ TRiP.GL01238 (Dpr17-RNAi)
y, w; R71G10, UAS-mCD8-GFP/+; R71G10, UAS-Dcr-2/ TRiP.JF03283}attP2 (Dpr18-RNAi)
y, w; UAS-mCD8-GFP/+; R71G10/ TRiP.JF02923 (Dpr20-RNAi)
(D)
y, w; 40A, *Dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *Dpr12 Δ ⁵⁰⁻⁸¹*, G13; R71G10, UAS-mCD8-GFP/+
y, w; 40A, *Dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *Dpr12 Δ ⁵⁰⁻⁸¹*, G13; R71G10, UAS-mCD8-GFP/+
y, w; 40A, *Dpr12 Δ ⁵⁰⁻⁸¹*, G13/40A, *Dpr12 Δ ⁵⁰⁻⁸¹*, G13; R71G10, UAS-mCD8-GFP/+
y, w; UAS-mCD8-GFP/+; R71G10/+

Figure EV2

- (A) hsFLP, UAS-mCD8-GFP; Gal80, FRT40A/FRT40A; R71G10, UAS-Dcr-2/+
 (B) hsFLP, UAS-mCD8-GFP; Gal80, FRT40A/FRT40A; R71G10, UAS-Dcr-2/
 TRiP.JF03210 (Dpr12-RNAi)
 (D) y, w; UAS-mCD8-GFP/R30G08-p65.AD; R48B03-GAL4.DBD/ +
 (E) y, w; UAS-mCD8-GFP/+; R18H09/ +
 (F) y, w; UAS-mCD8-GFP/+; R48H011/+

Figure EV3

- (C-D) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- $\delta^{T2A-Gal4}$* /*DIP- δ^{1-119}*
 (E) y, w; R71G10-QF2, QUAS-mtdT-HA/UAS-*DIP- δ* ; *DIP- $\delta^{T2A-Gal4}$* /*DIP- $\delta^{T2A-Gal4}$*
 (F) C155-Gal4; UAS-Cas9.P2/+
 (G) C155-Gal4; UAS-Cas9.P2/TKO.GS02451(*gRNA DIP- δ*)
 (H) y, w; R71G10, UAS-mCD8-GFP/ +; UAS-Cas9.C/+
 (I) y, w; R71G10, UAS-mCD8-GFP/ TKO.GS02451(*gRNA DIP- δ*); UAS-Cas9.C/+

Figure EV4

- (A-C) y, w; QUAS-GFP, UAS-mtdT-HA/UAS-tdT; R58E02/R58E02-QF2
 (D-F) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- $\delta^{T2A-Gal4}$* /+

Figure EV5

- (A) y, w; 40A, *dpr12 Δ^{50-81}* , G13/+; R10G03/UAS-mCD8-GFP
 (B) y, w; 40A, *dpr12 Δ^{50-81}* , G13/40A, *dpr12 Δ^{50-81}* , G13; R10G03/UAS-mCD8-GFP
 (D) y, w; 40A, *dpr12 Δ^{50-81}* , G13/+; R48H011/UAS-mCD8-GFP
 (E) y, w; 40A, *dpr12 Δ^{50-81}* , G13/40A, *dpr12 Δ^{50-81}* , G13; R48H011/UAS-mCD8-GFP
 (G) y, w; 40A, *dpr12 Δ^{50-81}* , G13/+; R18H09/UAS-mCD8-GFP
 (H) y, w; 40A, *dpr12 Δ^{50-81}* , G13/40A, *dpr12 Δ^{50-81}* , G13; R18H09/UAS-mCD8-GFP

Figure EV6

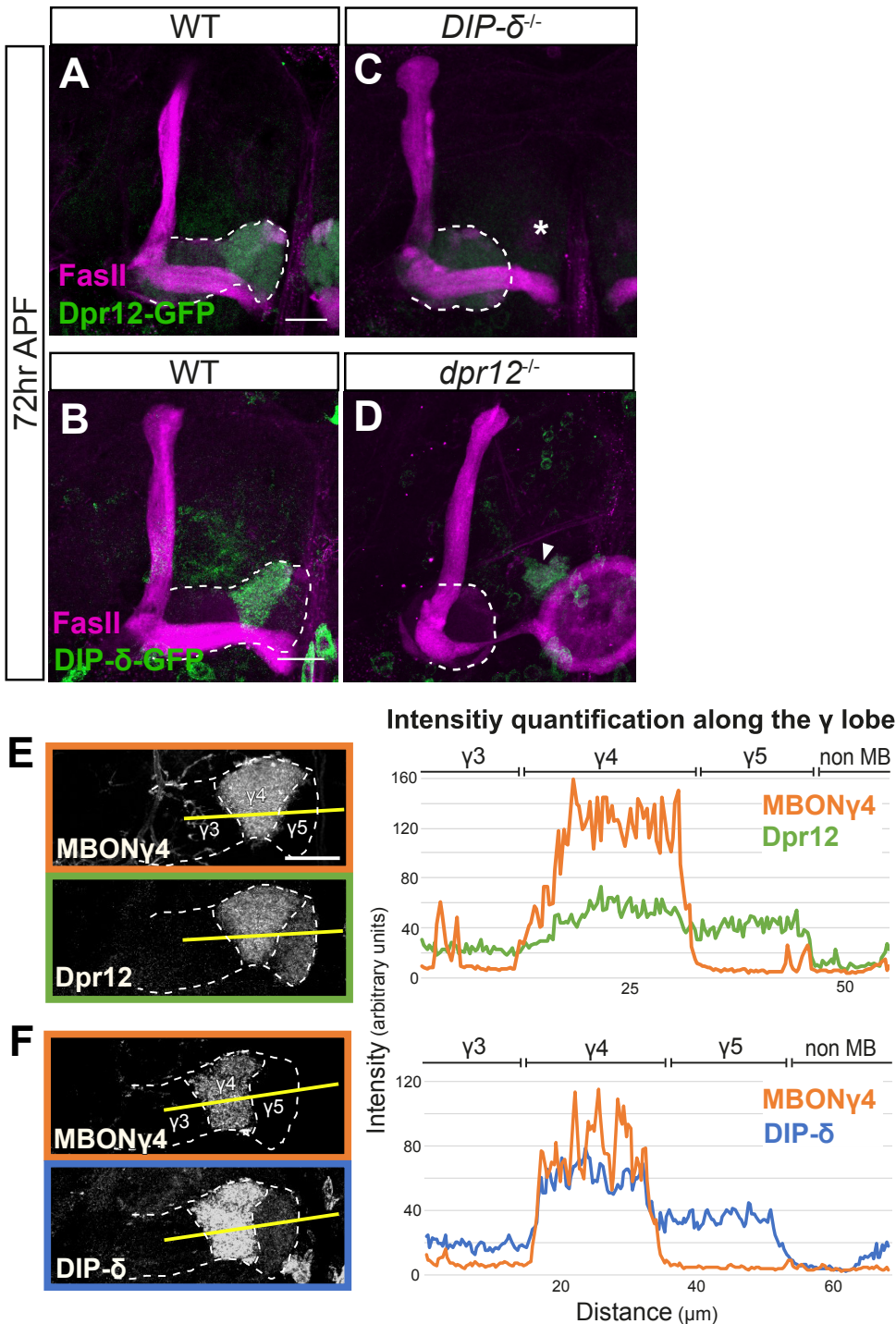
- (A) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- δ^{1-119}* /*DIP- δ^{1-119}*
 (B) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- δ^{1-119}* /*DIP- δ^{1-119}* , UAS-DIP- α
 (C) y, w; R71G10-QF2, QUAS-mtdT-HA/+; *DIP- $\delta^{T2A-Gal4}$* /*DIP- $\delta^{T2A-Gal4}$*
 (D) y, w; R71G10-QF2, QUAS-mtdT-HA/UAS-FasII; *DIP- $\delta^{T2A-Gal4}$* /*DIP- $\delta^{T2A-Gal4}$*

Appendix Figure S1

- (A) y, w; Dpr12^{GFSTF}/+
 (B) y, w;; DIP- δ ^{GFSTF}/+
 (C) y, w; Dpr12^{GFSTF}/+; *DIP- $\delta^{T2A-Gal4}$* /*DIP- $\delta^{T2A-Gal4}$*
 (D) y, w; 40A, *dpr12 Δ^{50-81}* , G13/40A, *dpr12 Δ^{50-81}* , G13; DIP- δ ^{GFSTF}/+
 (E) y, w; UAS-CD4-tdT/Dpr12^{GFSTF}; R18H09/+
 (F) y, w; UAS-CD4-tdT/+; DIP- δ ^{GFSTF}/R18H09

Appendix Figure S2

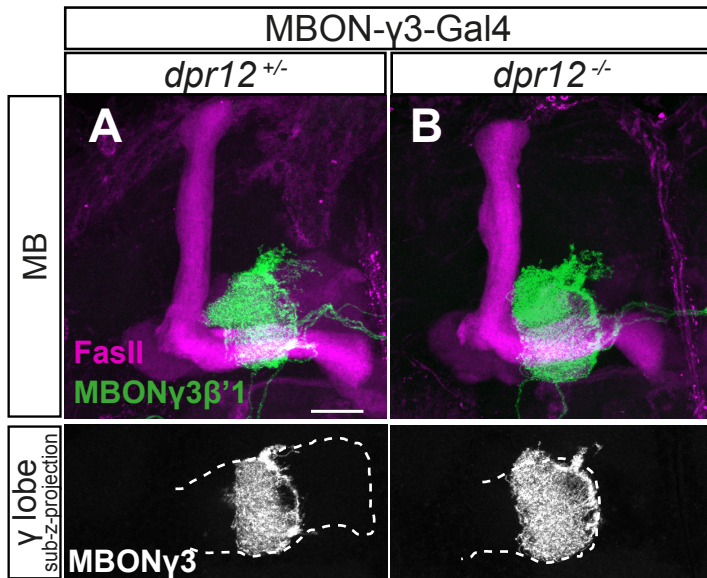
- (A) y,w; 40A, *dpr12 Δ^{50-81}* , G13/+; R94B10-GAL4.DBD, R52G04-p65.AD/ UAS-mCD8-GFP
 (B) y,w; 40A, *dpr12 Δ^{50-81}* , G13/40A, *dpr12 Δ^{50-81}* , G13; R94B10-GAL4.DBD, R52G04-p65.AD/ UAS-mCD8-GFP



Appendix figure S1. Additional data regarding Dpr12 and DIP- δ protein localization, related to Figures 3 and 6.

(A-D) Confocal z-projections of brains expressing either MiMIC mediated Dpr12^{GFSTF} (Dpr12-GFP; A,C) or DIP- δ ^{GFSTF} (DIP- δ -GFP; B,D) fusion proteins, in either WT (A-B), *DIP- δ ^{T2A-Gal4}* homozygous mutant (C) or *dpr12 ^{Δ 50-81}* homozygous mutant (D) animals at 72hr APF. Green is GFP, magenta is FasII. Dashed line depicts the medial γ -lobe, as determined by FasII staining. Asterisk demarcates the distal part of the lobe; arrowhead demarcates DIP- δ expression at the distal part of the lobe. Scale bar is 20 μ m.

(E-F) Intensity quantification of the indicated channels in the confocal z-projections that appear in Figure 3D (for E) and Figure 3H (for F). Intensity (arbitrary units) was quantified along a defined line that crosses the γ lobe from γ 3 until it exits the MB (marked in yellow; distance is shown in μ m). The MBON- γ 4 channel (GMR18H09-Gal4>CD4-tdT) is shown in orange (E, F), Dpr12-GFP in green (E) and DIP- δ -GFP in blue (F). The rough division to γ -lobe zones is indicated above the graphs. Scale bar is 20 μ m.



Appendix figure S2. MBON- γ 3 innervation of the MB γ -lobe in Dpr12 mutant brains, related to Figure 7.

(A-B) Confocal z-projections of *dpr12* ^{Δ 50-81} heterozygous (A) or homozygous (B) mutant brains expressing membrane-bound GFP (CD8-GFP) driven by MB083C (MBON γ 3-Gal4) that is expressed in MBON γ 3 β '1 (MBON-09). The greyscale channels are sub-z-projections comprised of slices restricted to the γ -lobe (which is outlined in white).

Green and grey are CD8-GFP, magenta is FasII. Scale bar is 20 μ m.

Color	Type	Name	Projection Pattern	<i>neuPrint</i> Body ID
●	γ-KC	KCym	Medial γ-lobe	415511327
●		KCym	Medial γ-lobe	415835070
●		KCym	Medial γ-lobe	415835178
●		KCym	Medial γ-lobe	415848228
●		KCym	Medial γ-lobe	415852518
●		KCym	Medial γ-lobe	415908516
●		KCym	Medial γ-lobe	416189410
●		KCym	Medial γ-lobe	416189500
●		KCym	Medial γ-lobe	416598862
●		KCym	Medial γ-lobe	415494049
●	γ5 PAM-DAN	PAM01	γ5 compartment	925799233
●		PAM01	γ5 compartment	1202397143
●		PAM01	γ5 compartment	5812982779
●		PAM15	γ5β'2a compartment	5813096699
●	γ4 PAM-DAN	PAM08	γ4 compartment	580317662
●		PAM08	γ4 compartment	988852391
●	γ4>γ1γ2 MBON	MBON05	γ4>γ1γ2 compartment	799586652
●	γ5β'2a MBON	MBON01	γ5β'2a compartment	612371421

Appendix Table S1. EM-skeletons used for model generation.

A list of the cell types, names, projection patterns, color coding and *neuPrint* IDs for all the EM-based skeletons used to construct the models in Figure 1P and Movie EV1 (see methods for additional details).

Driver		Expressing cells	Expression during development			Source
Name in manuscript	Formal name		Larva	Pupa	Adult	
γ-Gal4	R71G10-Gal4	γ-KCs consistently α/β-KCs stochastically	✓	✓	✓	Chr. III: Bl. #39604 Chr. II: Alyagor et al., 2018
γ-QF2	R71G10-QF2	γ-KCs consistently α/β-KCs stochastically	✓	✓	✓	This study
DIP-δ-Gal4	DIP-δ-T2A-Gal4	DIP-δ-expressing cells including PAM-DANs targeting γ4/5 zones	✓	✓	✓	This study
PAM-DAN-Gal4	R58E02-Gal4	All PAM-DANs	pPAMs*	After 48hr APF	✓	BDSC #41347
PAM-DAN-γ3-Gal4	MB441B	PAM-DANs targeting γ3	X	ND	✓	BDSC #68251
PAM-DAN-γ4-Gal4	R10G03	PAM-DANs targeting γ4	ND	After 72hr APF	✓	BDSC #48271
PAM-DAN-γ5-Gal4	R48H11	PAM-DANs targeting γ5	ND	After 72hr APF	✓	BDSC #50396
MBON-γ4-Gal4	R18H09	MBON-γ4>γ1γ2 (MBON-05)	X	Not at 24hr APF (ND for other time points)	✓	BDSC #48830
MBON-γ4-Gal4	MB298B	MBON-γ4>γ1γ2 (MBON-05)	ND	After 24hr APF	✓	BDSC #68309
MBON-γ3-Gal4	MB083C	MBON-γ3β'1 (MBON-09)	ND	ND	✓	BDSC #68287
Pan-Glia-Gal4	Repo-Gal4	All glia	✓	✓	✓	BDSC #7415
Pan-KC-Gal4	OK107-Gal4	All KCs (γ, α/β and α'/β')	✓	✓	✓	BDSC #854
Pan-Neuron-Gal4	C155-Gal4	All neurons	✓	✓	✓	BDSC #458

Appendix Table S2. List of drivers used in the current study.

The driver name (in the manuscript and formally), the cells in which it is expressed, and expression throughout development are depicted. ND; not determined. *pPAMs are primary-PAMs which are different from the adult PAM-DANs (data not shown).