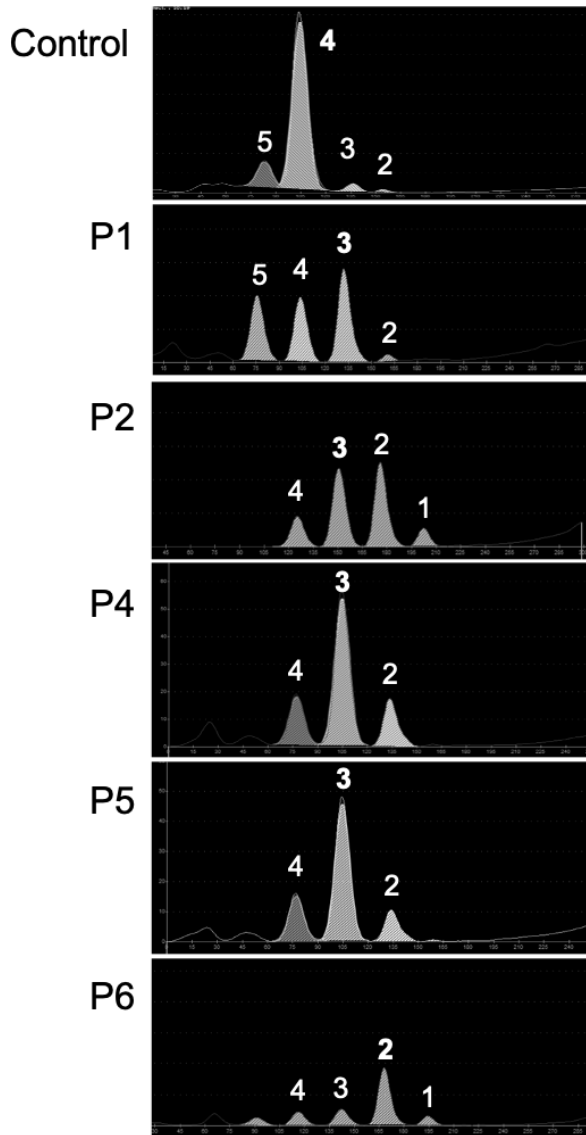


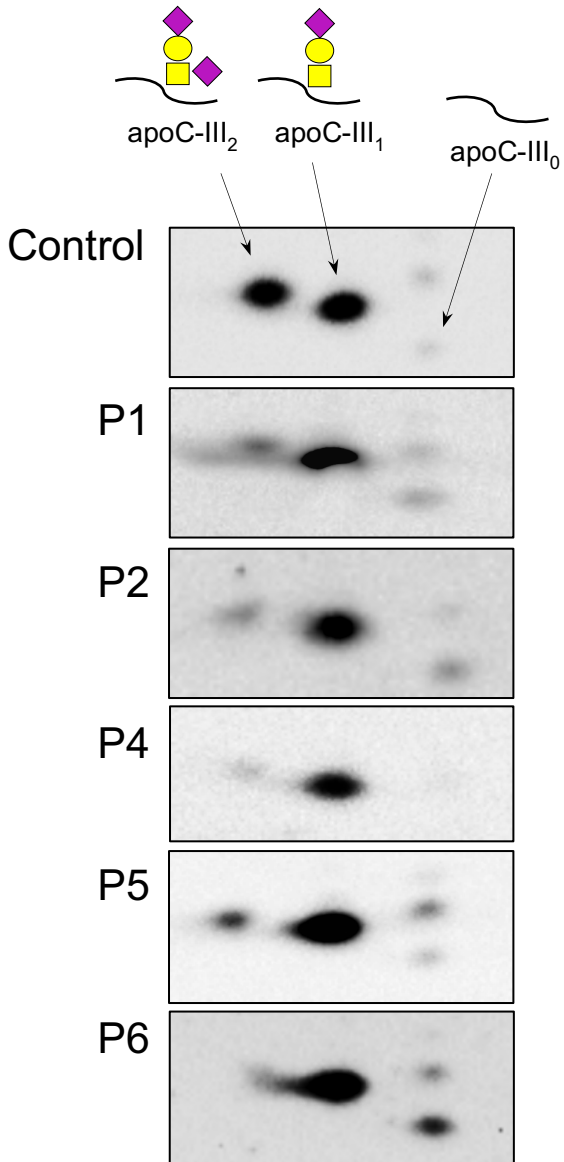
**Supplemental information**

**A mutation in *SLC37A4* causes a dominantly  
inherited congenital disorder of glycosylation  
characterized by liver dysfunction**

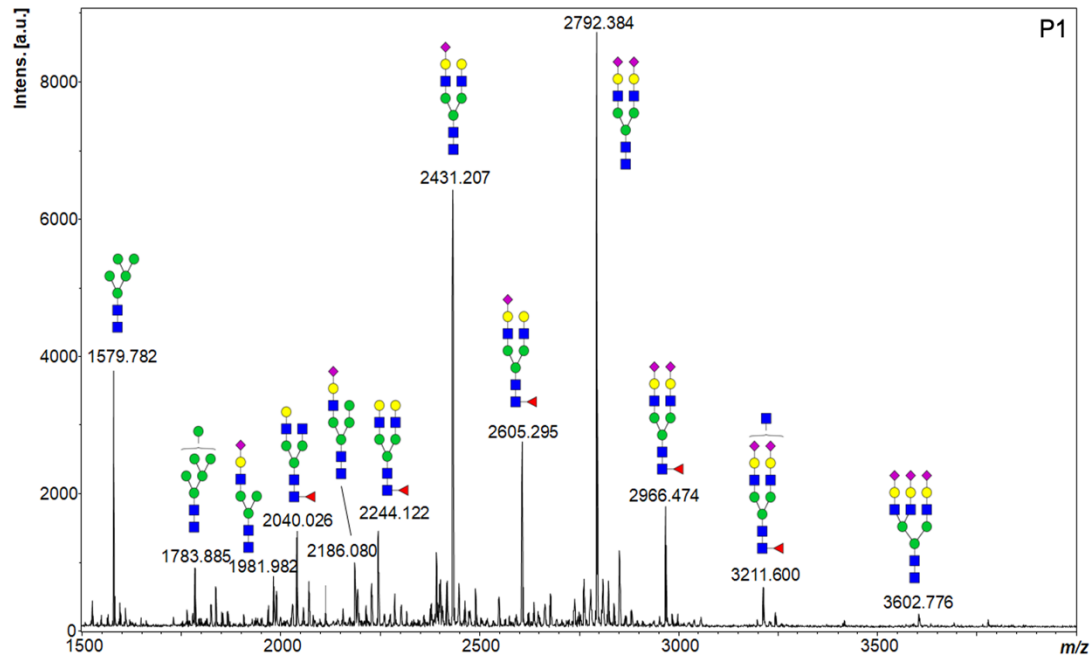
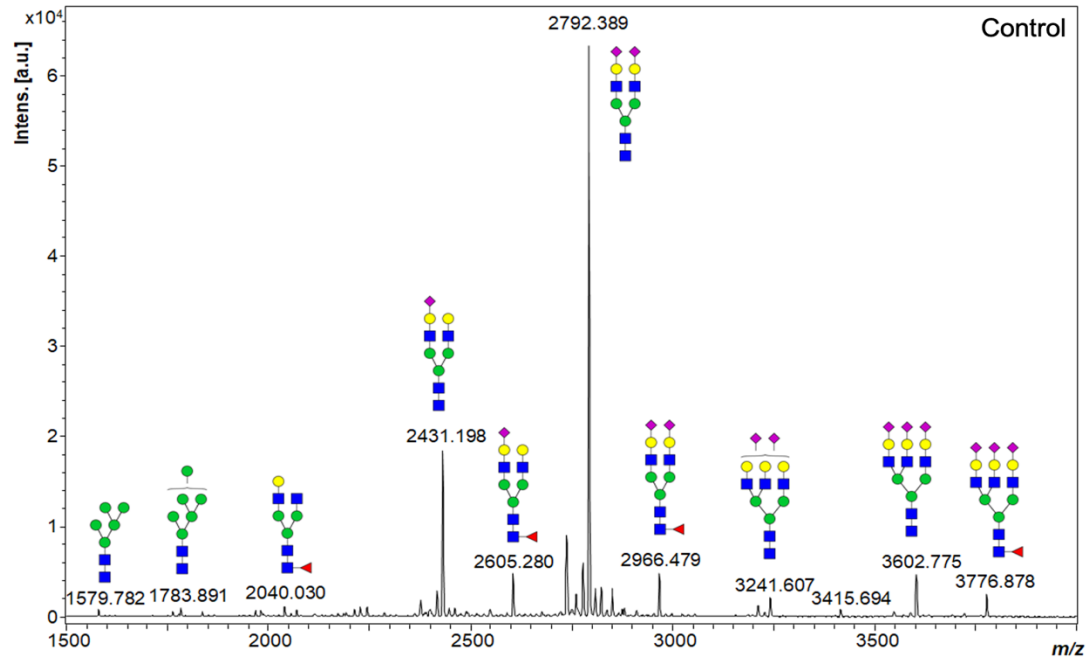
**Bobby G. Ng, Paulina Sosicka, François Fenaille, Annie Harroche, Sandrine Vuillaumier-Barrot, Mindy Porterfield, Zhi-Jie Xia, Shannon Wagner, Michael J. Bamshad, Marie-Christine Vergnes-Boiteux, Sophie Cholet, Stephen Dalton, Anne Dell, Thierry Dupré, Mathieu Fiore, Stuart M. Haslam, Yohann Huguenin, Tadahiro Kumagai, Michael Kulik, Katherine McGoogan, Caroline Michot, Deborah A. Nickerson, Tiffany Pascreau, Delphine Borgel, Kimiyo Raymond, Deepti Warad, University of Washington Center for Mendelian Genomics (UW-CMG), Heather Flanagan-Steet, Richard Steet, Michael Tiemeyer, Nathalie Seta, Arnaud Bruneel, and Hudson H. Freeze**

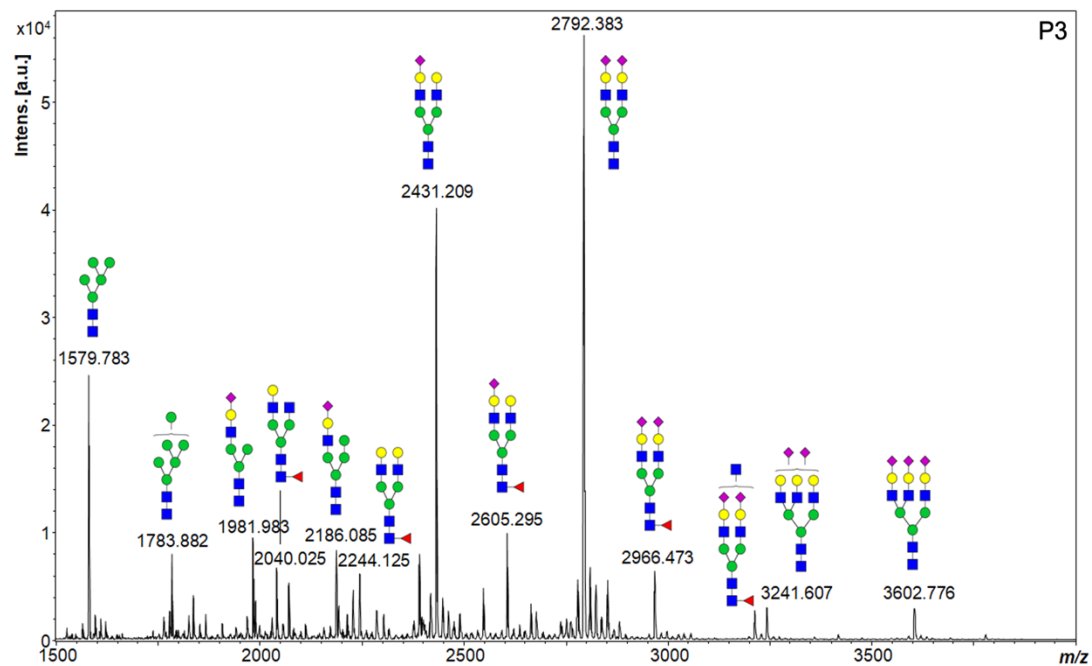
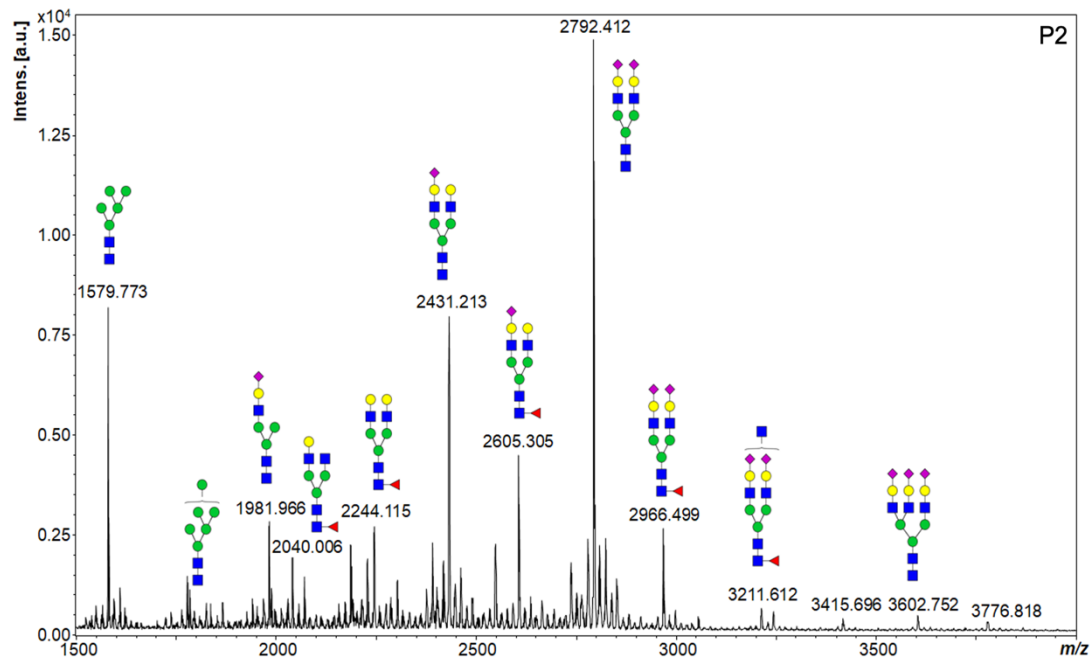


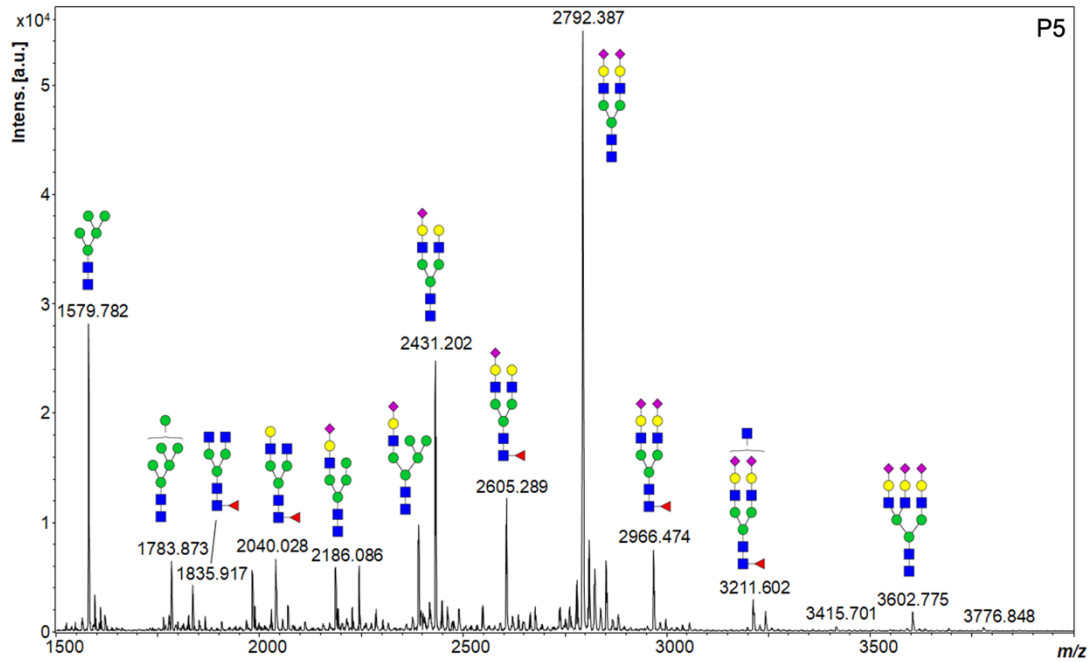
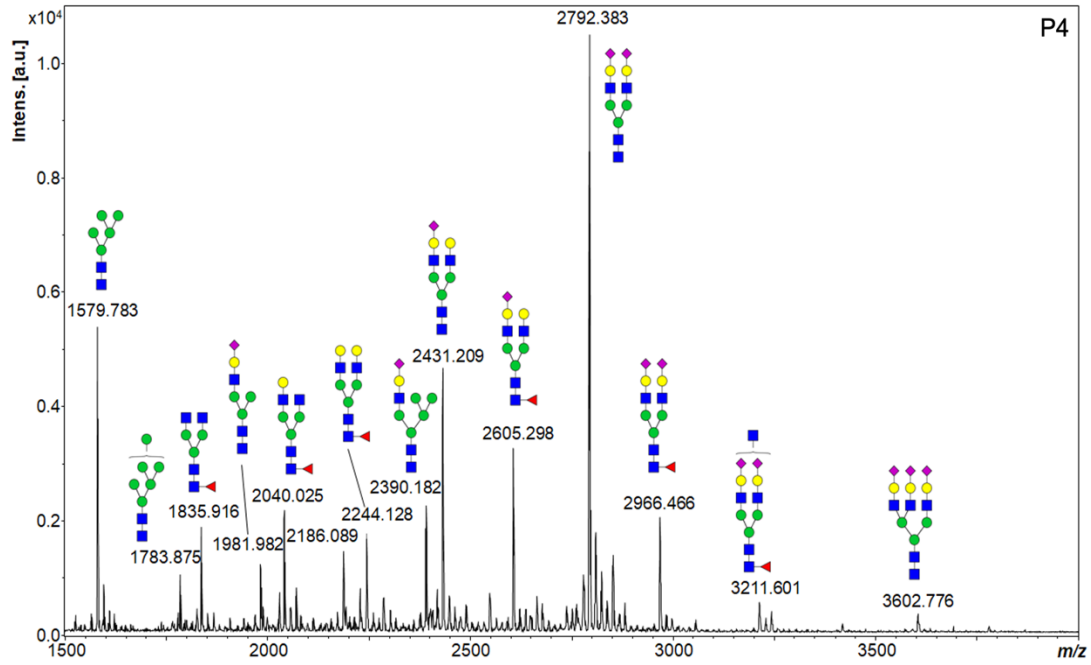
**Figure S1: Capillary zone electrophoresis of transferrin for Control, P1, P2, P4, P5, P6.** Glycosylated transferrin isoforms were separated by capillary zone electrophoresis with each peak labeled with the corresponding number of sialic acids. 0-Asialo, 1-Monosialo, 2-Disialo, 3-Trisialo, 4-Tetrasialo, 5-Pentasialo.

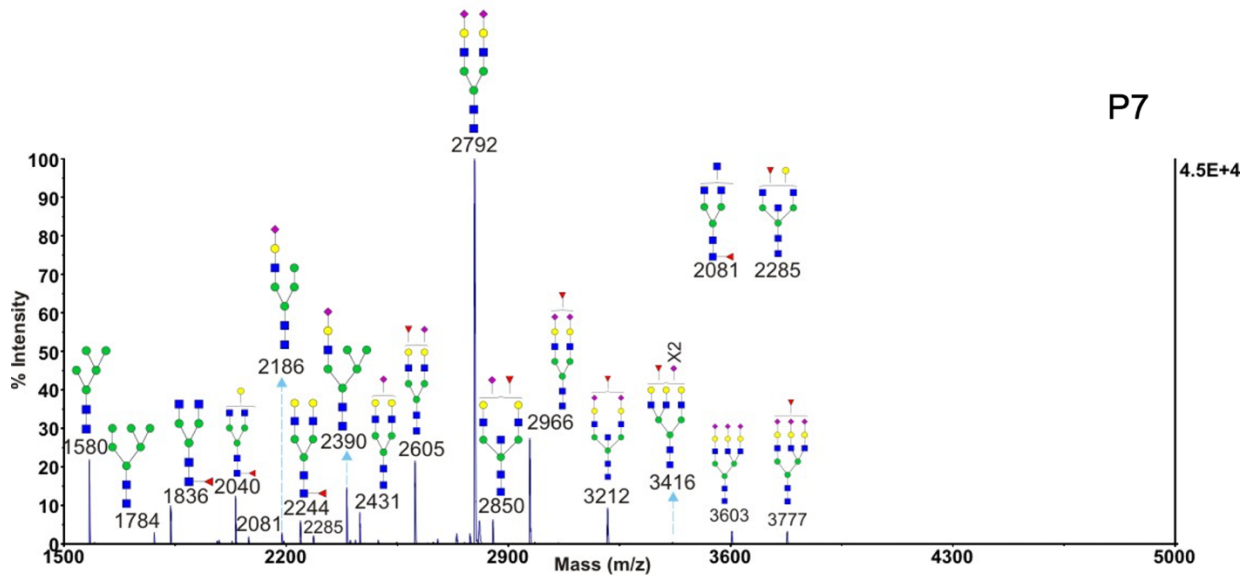
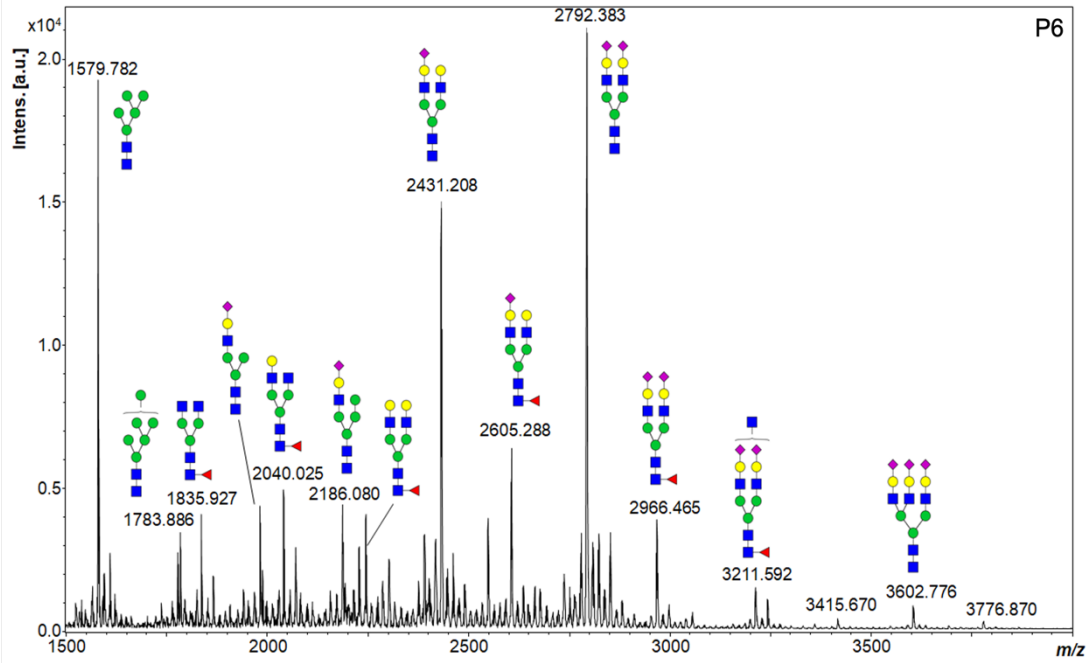


**Figure S2: 2-Dimensional gel electrophoresis of ApoCIII for Control, P1, P2, P4, P5, P6.** 2-DG electrophoresis of the serum O-linked glycoprotein, ApoCIII showing changes in the ratio of di-sialo (ApoCIII<sub>2</sub>), mono-sialo (ApoCIII<sub>1</sub>) and Asialo (ApoCIII<sub>0</sub>).



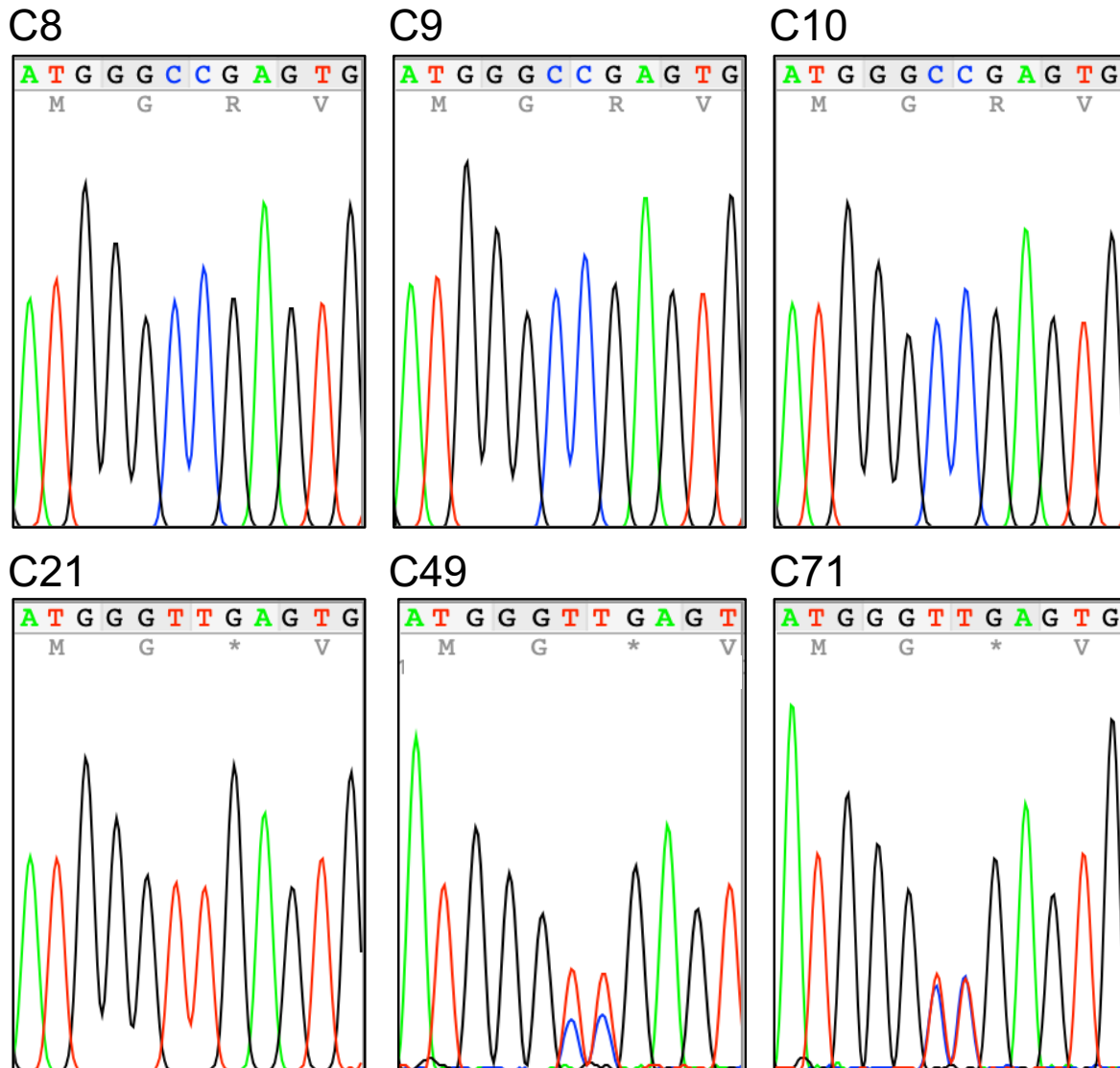




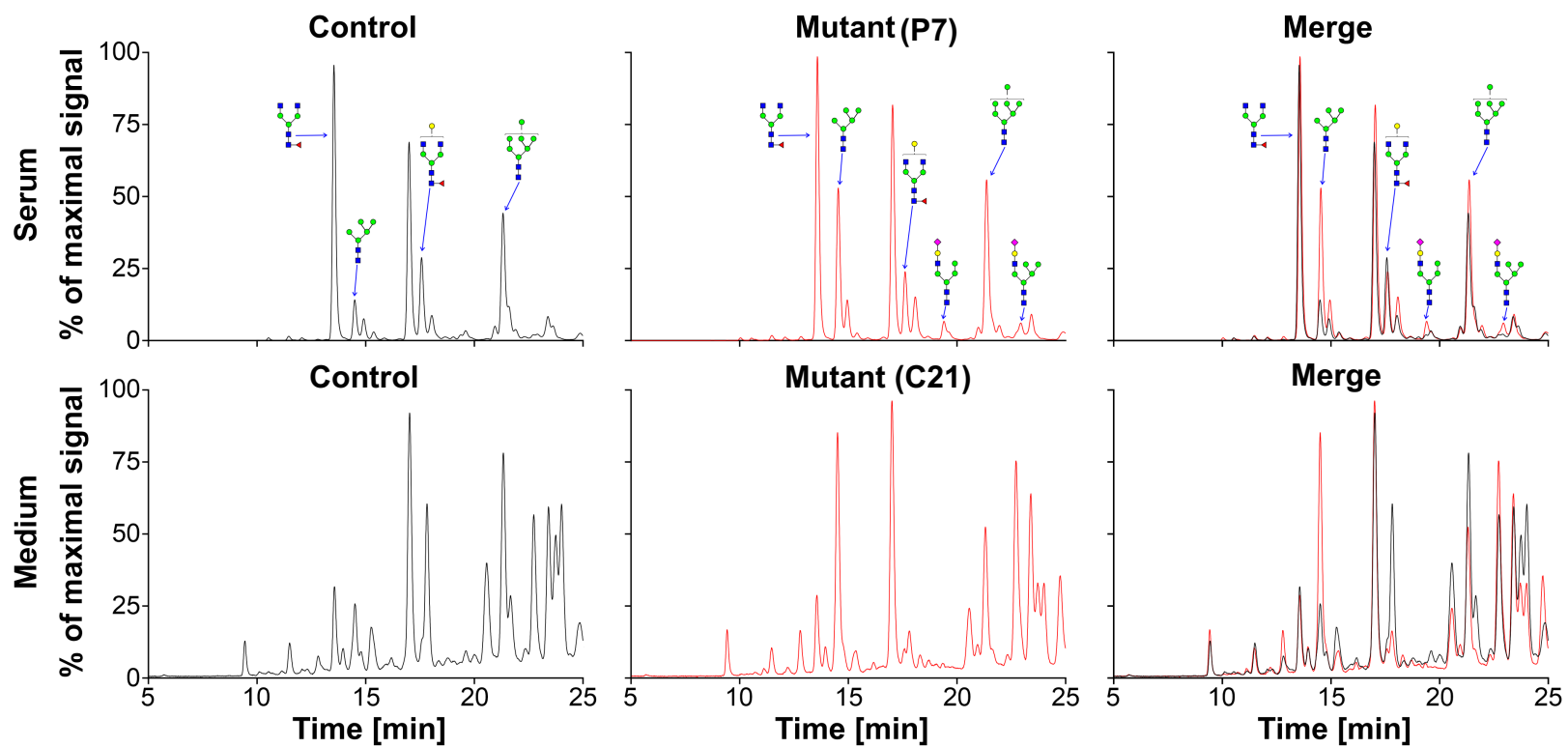


**Figure S3: Characterization of N-glycan abnormalities in serum from affected individuals.** MS spectra of N-glycans from serum glycoproteins for control and all seven affected unrelated individuals showing N-glycan abnormalities in serum. Specifically, the accumulation of multiple species of both high mannose (peaks 1579.8, 1783.9 m/z) and hybrid type N-glycans was observed (peaks 1981.9, 2186.1, 2390.2 m/z).



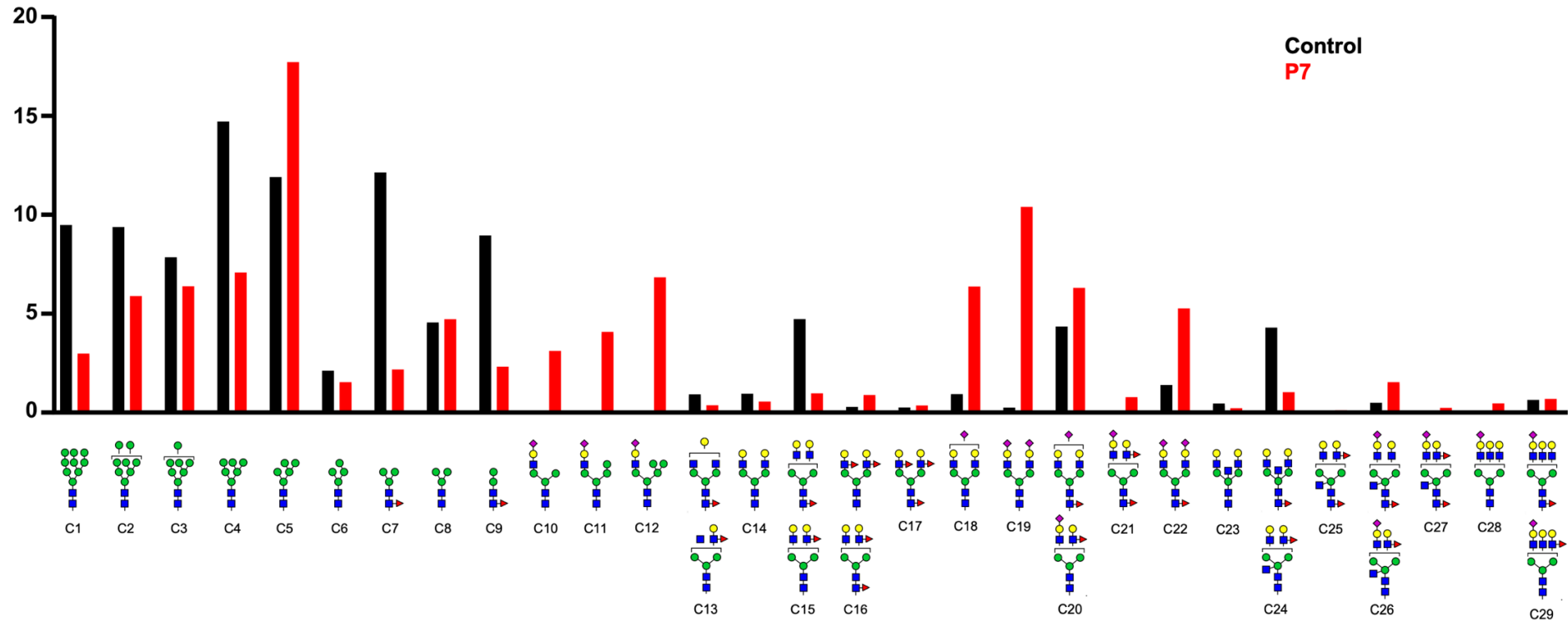


**Figure S4: Sanger sequencing of Huh7 base edited clones.** Sanger sequencing isogenic Huh7 base edited clones for the c.1267C>T [p.Arg423\*] in *SLC37A4*. Clones C8, C9, C10 underwent the same editing process as C21, C49, C71 but no edit was performed.

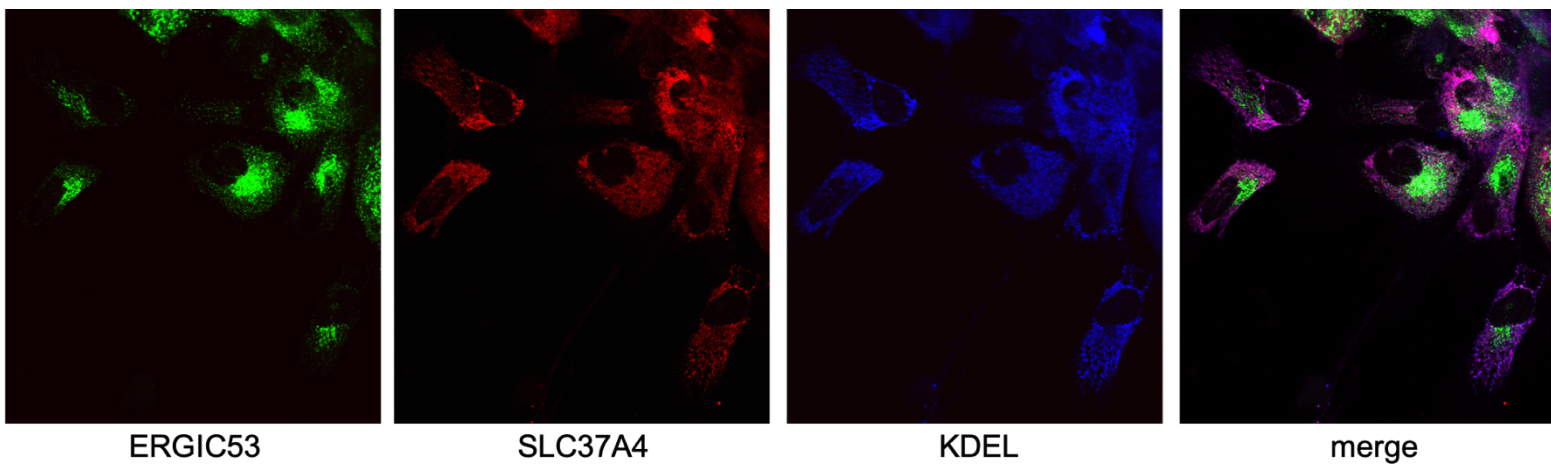


**Figure S5: HPLC analysis of N-glycans.** N-glycans were purified from Control and P7 serum (upper panel) as well as Huh7 Control and C21 medium (lower panel). Samples were run on HPLC and detected by fluorescence.

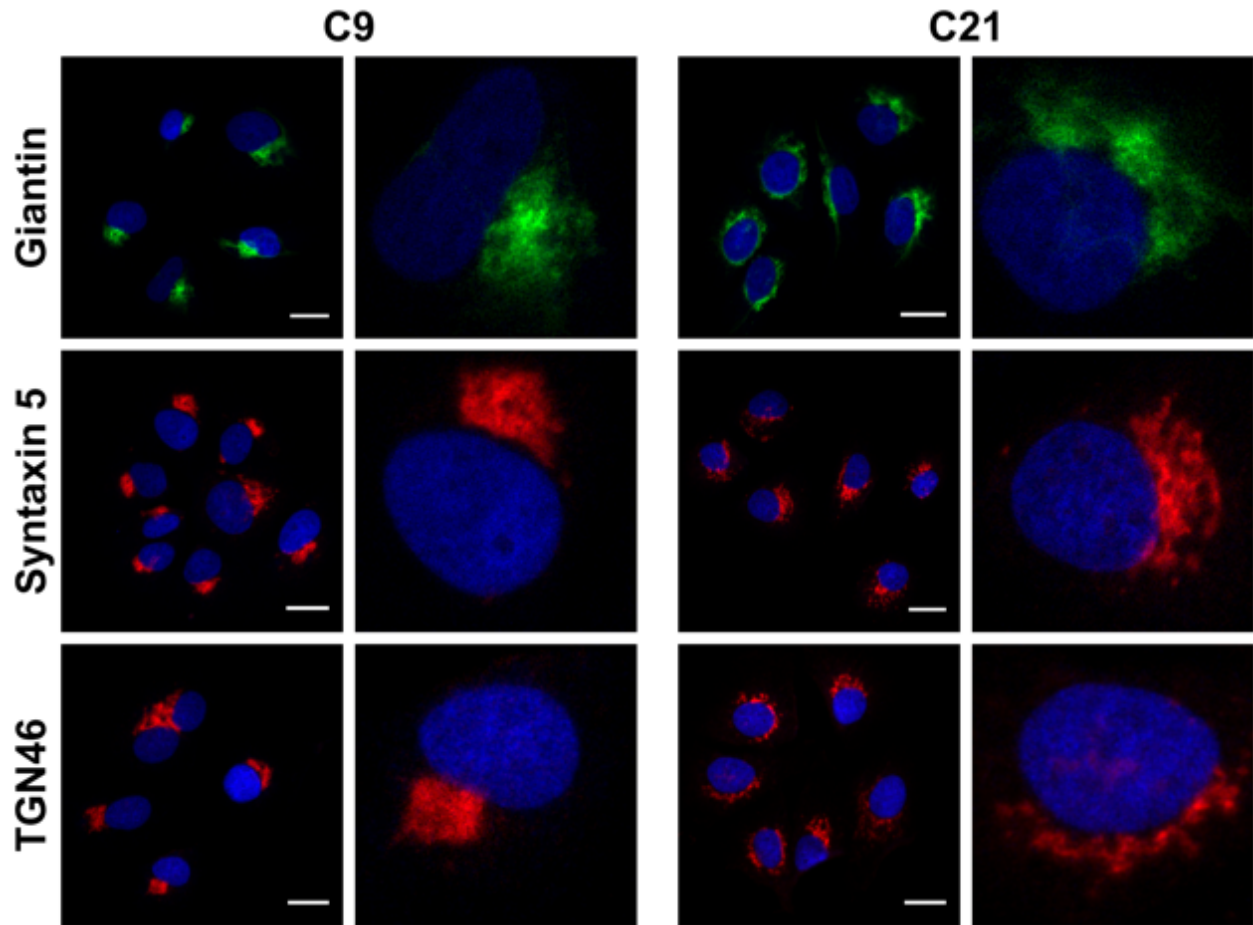
Glycan abundance [% of total profile]



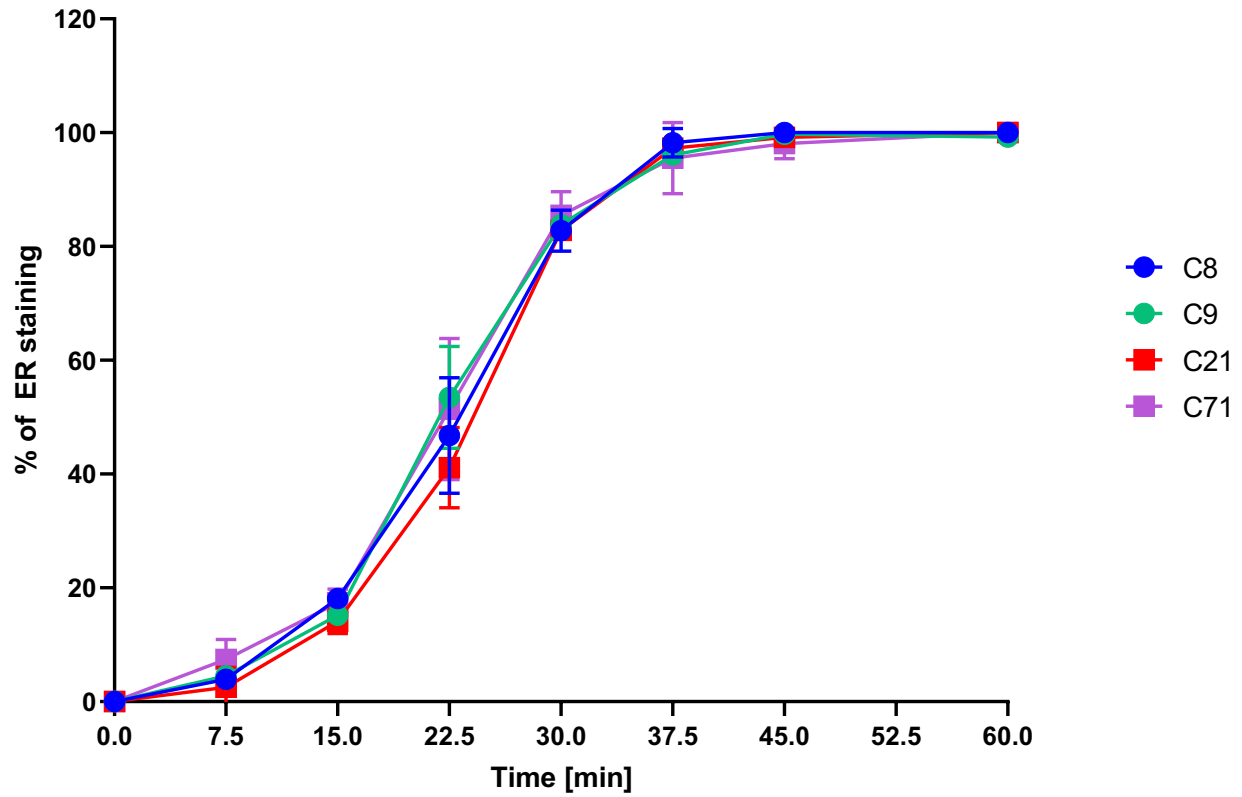
**Figure S6: Nanospray ionization mass spectrometry (NSI-MSn) analysis of N-glycans released from iPSC-derived hepatocytes.** Control and P7 iPSCs were differentiated to hepatocytes. N-glycans were released from total cell-associated proteins and analyzed after permethylation.



**Figure S7: ERGIC53 and SLC37A4 do not colocalize.** Immunofluorescence staining showing no colocalization of SLC37A4 with ERGIC53 in iPSC differentiated hepatocytes from P7.



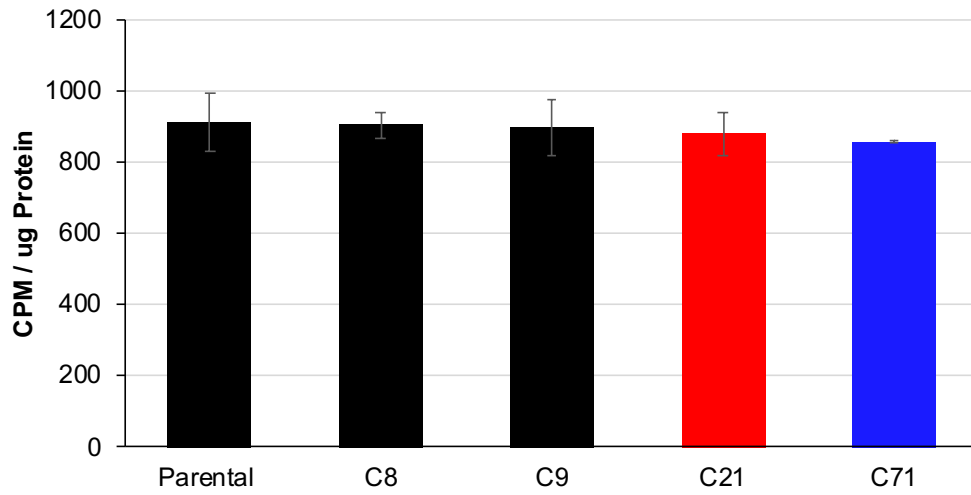
**Figure S8: Immunofluorescence for the Golgi morphology.** Multiple markers covering the three Golgi compartments were used to confirm abnormalities seen in the Golgi morphology. cis-Golgi/medial marker giantin, medial marker syntaxin 5 and trans marker Golgi network TGN46.



**Figure S9: Brefeldin A induced retrograde transport.**

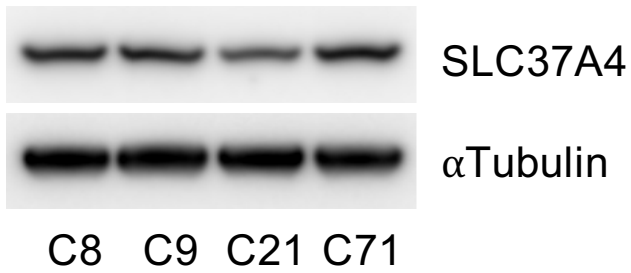
Brefeldin A induced retrograde transport was assayed in each cell line and showed no difference in retrograde transport rate.

### SLC37A4 Transport Activity in Huh7 Edited cells

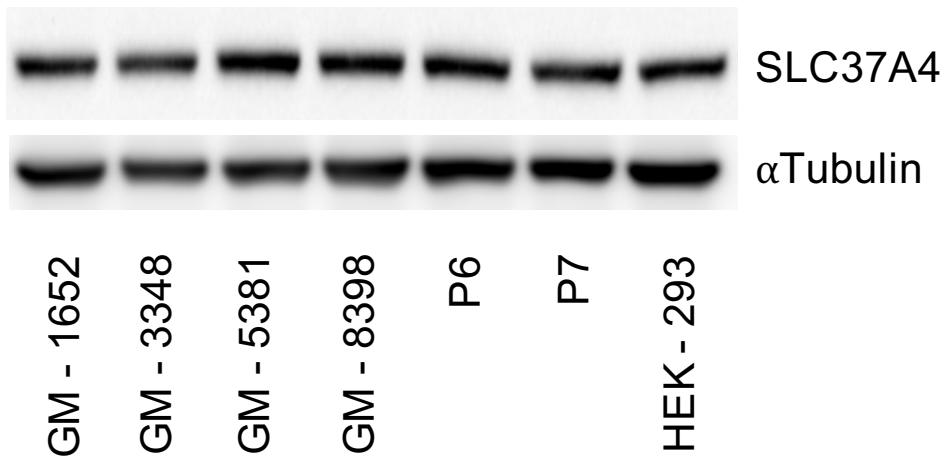


**Figure S10: SLC37A4 transport activity in CRISPR edited Huh7 cells.** SLC37A4 transport activity showing no difference in the ER specific uptake of  $^3\text{H}$ -Glc-6P into Huh7 control or edited cells.

A



B



**Figure S11: SLC37A4 protein expression.** Western blot analysis of SLC37A4 expression in (A) Huh7 control (C8, C9) and edited (C21, C71) cells. (B) Expression of SLC37A4 in control GM – 1652, GM – 3348, GM – 5381 and P6, P7. Alpha tubulin was used to normalize protein levels.



**Table S1: List of primary and secondary antibodies used in immunofluorescence.**

<b>Antibody</b>	<b>Cat. Number</b>	<b>Source</b>	<b>Dilution</b>	<b>Company</b>
anti-SLC37A4	PA5-58599	Rabbit	1:100	Life Technologies
anti-GM130	610822	Mouse	1:250	BD Biosciences
anti-giantin Alexa Fluor 488	308701	Rabbit	1:250	BioLegend
anti-syntaxin 5	SC-365124	Mouse	1:100	Santa Cruz Biotechnology Inc.
anti-TGN46	GTX-74290	Sheep	1:100	Genetex
anti-KDEL	ADI-SPA-827- D	Mouse	1:100	Enzo Life Sciences Inc.
anti- $\beta$ 4-GalT1	HPA-010807- 100uL	Rabbit	1:100	Sigma Aldrich
anti-Sec31A	612350	Mouse	1:100	BD Biosciences
anti-rabbit Alexa Fluor 405	A31556	Goat	1:100	Life Technologies
anti-rabbit Alexa Fluor 488	AB150077	Goat	1:500	Abcam
anti-rabbit Alexa Fluor 568	A11011	Goat	1:250	Life Technologies
anti-sheep Alexa Fluor 488	AB150178	Donkey	1:250	Abcam
anti-mouse Alexa Fluor 546	A11003	Goat	1:250	Life Technologies