

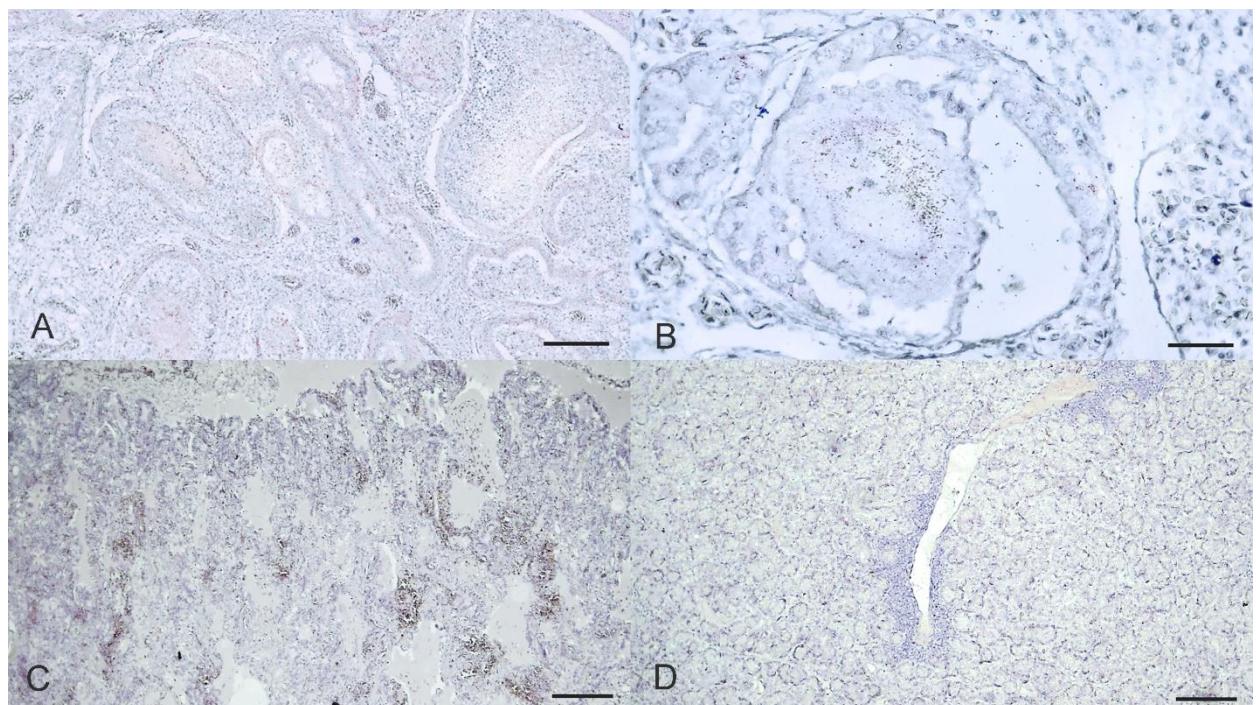
**Epizootic reptilian ferlavirus infection in individuals and multiple snake colonies with additional evidence of the virus in male genital tract**

**Chutchai Piewbang<sup>1,2</sup>, Sabrina Wahyu Wardhani<sup>2,3</sup>, Panida Poonsin<sup>1,2</sup>, Poowadon Chai-in<sup>4</sup>, Sitthichok Lacharoje<sup>1</sup>, Thanyarat Saengdet<sup>5</sup>, Taksa Vasaruchapong<sup>6</sup>, Suwimon Boonrungsiman<sup>4</sup>, Piyaporn Kongmakee<sup>7</sup>, Wijit Banlunara<sup>1</sup>, Anudep Rungsipipat<sup>1</sup>, Tanit Kasantikul<sup>8</sup>, Somporn Techangamsuwan<sup>1,2,\*</sup>**

**Supplementary Table S1.** Primer sets used for whole genome characterization of reptilian ferlavirus

<b>Direction</b>	<b>Primer name</b>	<b>Primer sequences (5'-3')</b>
<b>Forward</b>	FerP1_44F	TAAGTTGAAATTAGGATCA
<b>Reverse</b>	FerP1_1381R	GACATGTTGGCTC
<b>Forward</b>	FerP2_912F	TATGGTATAAACACACACGCTC
<b>Reverse</b>	FerP2_1748R	CACCAGTCTCCACATTGAT
<b>Forward</b>	FerP3_1728F	CAATCAATGTGGAGACTG
<b>Reverse</b>	FerP3_2836R	GGTAATACGGGAACACA
<b>Forward</b>	FerP4_2789F	ATATTGAGGTYTGGGA
<b>Reverse</b>	FerP4_4137R	CCTGTGATGAYACTTTCA
<b>Forward</b>	FerP5_3960F	GAAACACCAGATGCAC
<b>Reverse</b>	FerP5_5128R	TCAAACATRACCCCTA
<b>Forward</b>	FerP6_4964F	TAG GAG CAA AGY TCTT
<b>Reverse</b>	FerP6_5962R	TGRGTTGCCATTG
<b>Forward</b>	FerP7_5854F	TGATCCAGATYGAGATG
<b>Reverse</b>	FerP7_7367R	AGTGAGAATGTDGGTAT

<b>Forward</b>	FerP8o_7532F	AAATCCTGCAGTACCGTGGCA
<b>Reverse</b>	FerP8o_7532R	AGATATCTGTGTAAACTCCTG
<b>Forward</b>	FerP8i_7647F	GCACTCTACCCCAGTGTGG
<b>Reverse</b>	FerP8i_8045R	TTCTATCCAGCTTATTCTTAT
<b>Forward</b>	FerP1_44F	GCAGAGATTTCTCTTCTT
<b>Reverse</b>	FerP1_1381R	AGCTCTCATTTGTATGTCAT
<b>Forward</b>	FerP9o_9618	TAGAGGCTGTTACTGCTGC
<b>Reverse</b>	FerP9o_10244	CATCTGGCAAATAATCTGCC
<b>Forward</b>	FerP9i_9661F	ATGAGAGCYGCACARGT
<b>Reverse</b>	FerP9i_10226R	ACTGCRGAGACTCTCAC
<b>Forward</b>	FerP12_10393F	GTGAGAGTCTCYGCAGT
<b>Reverse</b>	FerP12_11024R	CTRGCACCTGTCACRGA
<b>Forward</b>	FerP13_11008F	TAGACTGGGCATCRGATCC
<b>Reverse</b>	FerP13_11933R	CCRAGBACCCAATATCTRCT
<b>Forward</b>	FerP14_11693F	AARTGGACAATTCHGG
<b>Reverse</b>	FerP14_13691R	TCTGAGTGAACRARTGA
<b>Forward</b>	FerP15_13453F	CYTGGGTAGGRAATGA
<b>Reverse</b>	FerP15_14384R	AGGGRAAATATGAAYAAG A
<b>Forward</b>	FerP16_14306F	ATCCAATAAGAATAAGCTGGAT
<b>Reverse</b>	FerP16_15376R	GACCAAAGGATCGGAA
<b>Forward</b>	FerP17_8094F	ACTCAACC GG GTGTGTTAG
<b>Reverse</b>	FerP17_9801R	GTATGGTTCATCAGCCTCCAT
<b>Forward</b>	FerP18_7329F	GCAGAGATTTCTCTTCTT
<b>Reverse</b>	FerP18_7804R	AGCTCTCATTTGTATGTCAT



**Supplementary Fig. S1.** Reptilian ferlavirus infection. *In situ* hybridization. Epididymis. (A &B). Big-eyed viper (colony A). No hybridization signals were detected in the sections incubating with feline bocavirus probe. Lung (C.) and kidney(D). Boa constrictor (ferlavirus-PCR negative sections). No hybridization signals were found in the ferlavirus-PCR negative sections incubating with ferlavirus ISH probe. Bars indicate 250  $\mu$ m.