

Supplementary Figure 1. N-glycan synthesis and processing. A Glc₃Man₉GlcNAc₂ precursor is synthesized on the cytoplasmic face of the endoplasmic reticulum on a dolichol-pyrophosphate precursor before being flipped to the ER lumen (as Man₅GlcNAc₂) and transferred to asparagine in the Asn-X-Ser/Thr sequon on nascent polypeptides through the oligosaccharyltransferase (OST) multiprotein complex. Glucosidases remove terminal glucose residues, calnexin and calreticulin assist in

folding, and the glycoprotein is interrogated by the protein folding and quality control machinery. Improperly folded proteins are retrotranslocated by the lectin EDEM, whereas properly folded proteins are marked with removal of the final glucose by glucosidase II. The glycoprotein is then transported to the cis-Golgi network and processed further by mannosidase II before addition of a terminal GlcNAc by GnT1 (which is defective in LEC1-mutant cells), initiating the first N-glycan branch. After additional processing, GnT2 adds GlcNAc to form the second branch of a biantennary GlcNAc₂Man₃GlcNAc₂ glycan, which is further modified by addition of a 'core' α 1,6 fucose by *FUT8*. This structure is transferred to the trans-Golgi network (TGN) for addition of a terminal galactose (which is absent in UDP-Gal transporter-mutant Lec8 cells) as well as a terminal sialic acid (which is absent in CMP-sialic acid transporter-mutant Lec2 cells). The fully matured glycoprotein is secreted extracellularly or transported to the cell surface. A parallel pathway targeting N-glycoproteins to the lysosome with mannose-6-phosphate is not shown.



Supplementary Figure 2. O-glycan synthesis. Synthesis of the major core (1–4) Oglycans is shown, with structures implicated in inflammatory bowel disease highlighted in red. Mucin-type O-glycosylation is initiated by a family of 20 polypeptide GalNAc transferases (ppGalNacTs) to form the Tn antigen, which is typically modified either by addition of galactose to form core 1 (also known as T antigen) and subsequently core 2 or extended core 1, or by addition of GlcNAc to form core 3 and subsequently core 4 structures. All cores can be further modified by Gal, GlcNAc, Fuc, Sia and sulfate to form terminal epitopes. Core 1-based and core 2-based structures are found in all tissues, whereas core 3-based and core 4-based structures are restricted to the intestinal epithelia.



Supplementary Figure 3. Select terminal glycan structures. (**A**) ABO(H) blood group antigens. These can be further classified as type-1 or -2, based on β3 or β4 linkage, respectively, of the terminal (H Ag) or subterminal (A/B Ag) galactose to the proximal GlcNAc (i.e. LacNAc) or type-3 or -4 based on β3 linkage to an O-GalNAc-linked glycan or a glycolipid, respectively. (**B**) Lewis antigens. These can be further modified by addition of sulfate to the 6-position of GlcNAc (6-Sulfo-), Gal (6'-Sulfo-), or 3-position of Gal (3'-Sulfo-).

Supplementary Table 1. Inflammatory bowel disease risk genes. Genes are divided by

those found in UC, CD, or both (IBD). Those genes implicated in regulating glycosylation are highlighted in red.

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| CD | UC | | | IBD | | |
|-----------|----------|-----------|------------|-----------|----------|---------|
| ADAM30 | ADA | ADCY3 | CXCR5 | IL21 | PLA2R1 | TRIB1 |
| AKAP1 | AHR | ALDH2 | DAP | IL23R | PRKAB1 | TRIM 35 |
| ATG16L1 | CALM3 | ARHGEF6 | DNMT3B | IL27 | PRKCB | TRIM8 |
| CD27 | CARD11 | ATG4B | DOK3 | IL2RA | PTGER4 | TSPAN14 |
| CHADL | DLD | ATXN 2 | DUSP1 | ΙΡΜΚ | PTGS2 | TST |
| CPEB4 | FAM55A | BACH2 | DUSP22 | IRF4 | PTK2B | TUBD1 |
| CREB5 | FAM55D | BRE | EDG1 | IRF8 | PTPRC | TYK2 |
| FASLG | GNA12 | BTBD8 | EPHX2 | IRGM | REL | VDR |
| FUT2 | HNF4A | C10orf58 | EPO | JAK2 | RELA | ZFP36L1 |
| GPX4 | IRF5 | C1GALT1C1 | ERAP1 | KIF21B | RORC | ZNF831 |
| HMHA1 | ITGAL | C1orf53 | ERAP2 | LIF | RPS6KA2 | ZPBP |
| IFNAR1 | ITIH4 | C5orf4 | FCGR2A/B | LIFR | RPS6KB1 | |
| IFNGR2 | JRKL | CARD9 | FCGR3A | LITAF | SELE | |
| IL31RA | MAML2 | CCDC88B | FLJ78302 | LOH12CR1 | SELL | |
| IL6ST | MANBA | CCL13 | FOS | LPXN | SELP | |
| JAZF1 | MAN2A1 | CCL2 | FOSL2 | LSP1 | SH2B3 | |
| L3MBTL2 | NFKB1 | CCL20 | FYB | LTF | SLC30A | |
| LACC1 | NFKBIZ | CCR1 | GALC | LY75 | SMAD3 | |
| LGALS9 | PLCL1 | CCR2 | GPR18 | MAP3K7IP2 | SMAD7 | |
| LTBR | PRKCD | CCR3 | GPR183 | MAP3K8 | SMURF1 | |
| NHP2L1 | RFTN2 | CCR5 | GPR35 | MARCH7 | SOCS1 | |
| NOD2 | SLC9A3 | CCR6 | GPR65 | MLH3 | SPRED2 | |
| NOS2 | TNFRSF14 | CD226 | НСК | MST1 | SPRY4 | |
| PMM1 | ZFP90 | CD28 | HGFAC | MUC19 | STAT1 | |
| PTPN22 | | CD40 | IBD5 locus | NDFIP1 | STAT3 | |
| RASGRP1 | | CD48 | ICOS | NFATC1 | STAT4 | |
| RIPK2 | | CD6 | ICOSLG | NFIL3 | TMEM180 | |
| SP140 | | CEBPB | IFIH1 | NFKB2 | TMEM258 | |
| SPRED1 | | CEBPG | IFNG | NKX2-3 | TNFAIP3 | |
| TAGAP | | CISD1 | IKZF1 | ORMDL3 | TNFRSF18 | |
| TEF | | CNTF | IL10 | OSM | TNFRSF4 | |
| TFSF11 | | CNTNAP2 | IL12B | OSMR | TNFRSF6B | |
| TNFRSF 1A | | CREM | IL15RA | PDCD1 | TNFRSF9 | |
| TNFSF18 | | CRTC3 | IL18RAP | PFKB4 | TNFSF15 | |
| UCN | | CTLA4 | IL1R1 | PHACTR2 | TNNI2 | |
| USP1 | | CTSZ | IL2 | PLA2G4A | TRAF3IP2 | |