for Respiratory Syncytial Virus

Supplementary information

Supplementary Figure 1. NMR studies of FsIIm. A, Chemical shift assignments for FsIIm. B,
Statistical analysis of structure calculations. C, Hα chemical shift deviations from random coil values, showing mostly negative values typical of helical secondary structure. D, 600 MHz ¹H

NMR spectrum of FsIIm in H₂O/D₂O (9:1), pH 5.0.



10 in the Methods section is shown.



Supplementary Figure 3. **Analytical results for V-306**. *Top*, Analytical UPLC of **V-306** (see Methods section for experimental details). *Bottom*, High resolution electrospray ionization (ESI)-MS of **V-306**.



Supplementary Figure 4. EM and DLS data for V-306. A, Negative staining transmission electron micrograph of SVLPs formed by the V-306 lipopeptide dissolved in tris(hydroxymethyl)amino-methane (Tris) buffer containing 0.9% NaCl, pH 7.4. Method: V-306 in buffer was applied to glow discharged carbon-coated copper EM grid (400 mesh) and left for 50 s. Excess solution was removed with blotting paper, and then washed with 2 drops of water. The sample was stained with 2% uranyl acetate 3x for 15 min then air dried for 30 min. The sample was then examined by TEM. Scale bar 5x100 nm. B, V-306 lipopeptide was dissolved in PBS (0.5 mg/mL) at pH 7.4, and analyzed in a Wyatt DynaPro DLS instrument at 25°C. Size distributions are shown by regularization as intensity distributions. The average radius, % polydispersity and polydispersity index (PDI) are given below.





Supplementary Figure 5. Determination of Abs in human sera recognizing V-306. *Left*, Optimized ELISA calibration for determination of Abs in human sera that bind to peptide (V-306), carrier+peptide (V-306) and lipopeptide carrier (shown in Supplementary Figure 2). *Right*, Ab titre in each of 20 different sera. For experimental details see Methods.

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Supplementary Figure 6. Representative histopathology pictures. Each picture shows typical histopathology slides following immunizations with different doses of **V-306** (once with adjuvant), with FI-RSV and with PBS (negative control). Scale bar 500 μm.



15µg+Adju-Phos PBS

FI-RSV



Supplementary Figure 7. Nucleotide and deduced primary sequences of the VH and VL

regions of mAb 10D11. The predicted CDRs are underlined.

60 VH domain

Nt sequence

CAGGTGCAACTGAAGGAGTCAGGACCTGGCCTGGTGGCGCCCTCACAGAGCCTGTCCATTACCTGCA CTGTCTCTGGGTTCTCATTAACC<u>AGCTATGATATAAGC</u>TGGATTCGCCAGCCACCAGGAAAGGGTCT 65 GGAGTGGCTTGGA<u>ATAATATGGAATGGTGGAGGCACAAATTATAATTCAGCTTTCATGTCC</u>AGACTG AGCATCAACAAGGACAATTCCAAGAGCCAAGTATTCTTAAAAATGAACAGTCTGCAAACTGATGATA CAGCCATATATTACTGTGTAAGA<u>GGTGATTATGATTACGCCTGGTTTGATTAC</u>TGGGGCCAAGGGAC TCTGGTCACTGTCTCTGCA

70 Amino acid sequence

QVQLKESGPGLVAPSQSLSITCTVSGFSLT<u>SYDIS</u>WIRQPPGKGLEWLG<u>IIWNGGGTNYNSAFMS</u>RLSINK DNSKSQVFLKMNSLQTDDTAIYYCVR<u>GDYDYAWFDY</u>WGQGTLVTVSA

75

VL domain

Nt sequence

AACATTATGATGACACAGTCGCCATCATCTCTGACTGTGTCTGCAGGAGAAAAGGTCACTATGAGCT 80 GT<u>AAGTCCAGTCAAAGTGTTTTATACGGTTCAAATCAGAAGAACTACTTGGCC</u>TGGTACCAGCAGAA ACCAGGGCAGTCTCCTAAACTGCTGATCTAC<u>TGGGCATCCACTAGGGATTCT</u>GGTGTCCCTGATCGCT TCACAGGCAGTGGATCTGGGACAGATTTTACTCTTACCATCAGCAGTGTACAAGCTGAAGACCTGGC AGTTTATTATTGT<u>CATCAATACCTCTTCTCGTGGACG</u>TTCGGTGGAGGCACCAAGCTGGAAATCAAA

85 Amino acid sequence

NIMMTQSPSSLTVSAGEKVTMSC<u>KSSQSVLYGSNQKNYLA</u>WYQQKPGQSPKLLIY<u>WASTRDS</u>GVPDRF TGSGSGTDFTLTISSVQAEDLAVYYC<u>HQYLFSWT</u>FGGGTKLEIK

Supplementary Figure 8. Determination of dissociation constants (*K*_D) using Surface Plasmon

Resonance (SPR). A, For 10D11+Fsllm. Double referenced SPR sensorgram of interaction of

95 10D11 with Fslim at concentrations of 3, 6, 12, 24 and 48 nM, with single-cycle kinetic measurement (black) and 1+1 fit (red) using a 1:1 interaction model. B, Palivizumab+Fslim. Double referenced SPR sensorgram of interaction of Palivizumab with Fslim. Sensorgrams (black) and fit (red) to a 1:1 binding model, for 8 injections of the peptide in duplicate at concentrations of 7.8, 15.6, 31.25, 62.5, 125, 250, 500 and 1000 nM.

