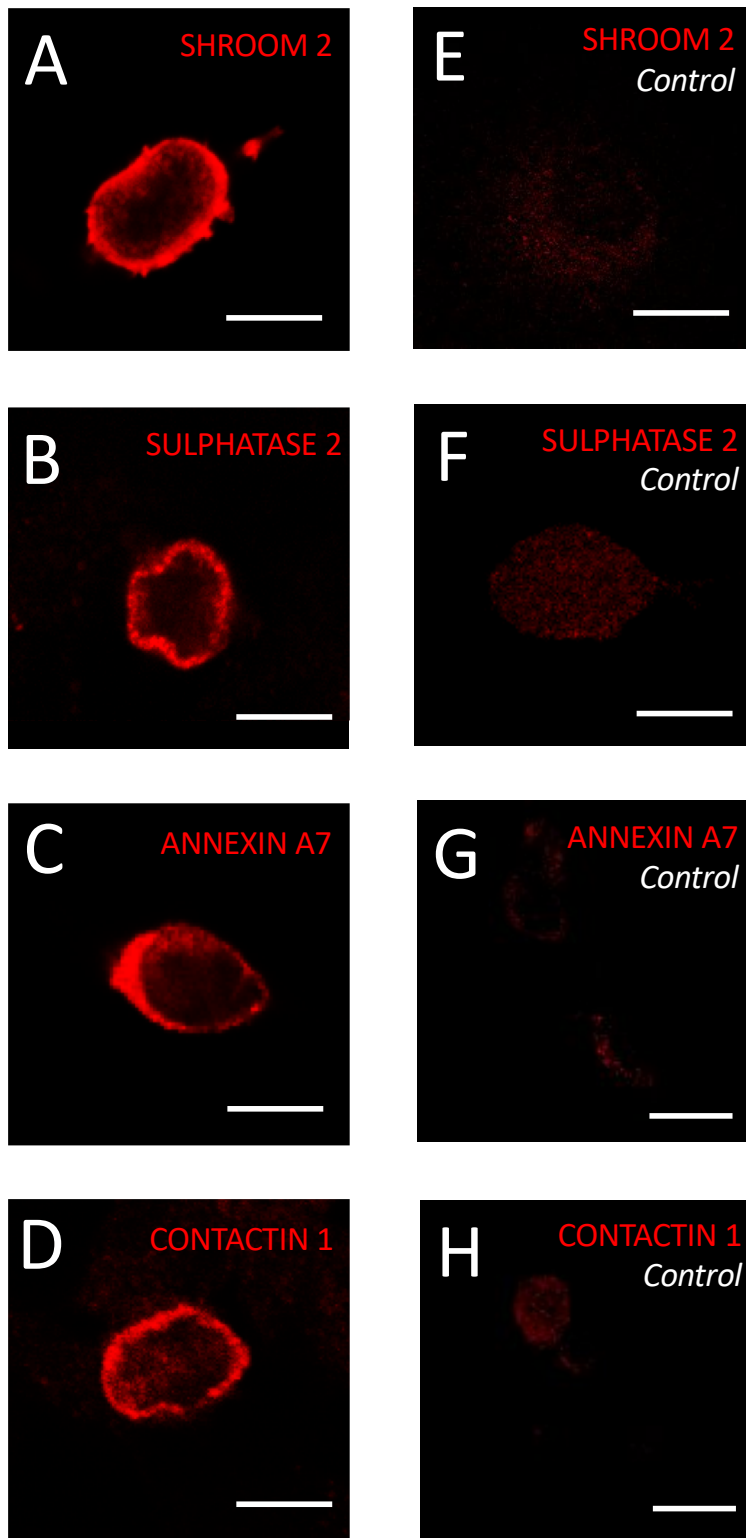


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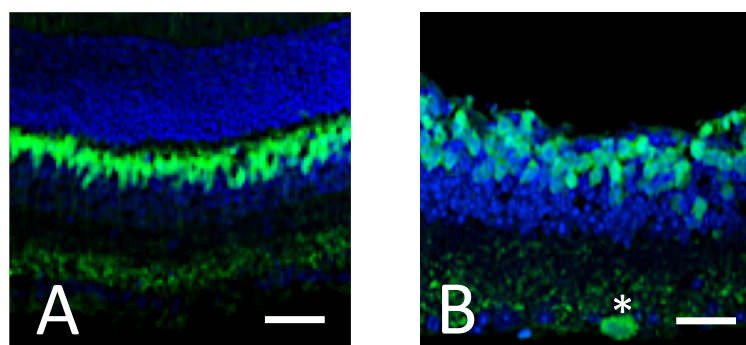
“Retinal bipolar cell gene expression during retinal degeneration: implications for optogenetic visual restoration”

Supplementary Figures & Tables

February 2021



(Fig S1.)



(Fig S2.)

Figure S1. Validation of antibodies. ICC images of HEK293T cells transfected to express the protein corresponding to the labelled antibody. As can be seen, all four stain at a level far above that of a non-transfected control. Note staining pattern of all four antibodies is consistent with protein in or around the cell membrane.

A – D Appropriately transfected HEK293T cells stained with the respective antibody

E – H Untransfected HEK293T cells stained with the respective antibody

Scale bars - 10µm

Figure S2. L7.Cre.EYFP mice express EYFP in rod and cone type 2 & 6 bipolar cells as well as a small subset of bistratified retinal ganglion cells (circa 1,500 in retina, denoted by asterix).

Blue = DAPI, Green = YFP Scale bars = 15µm

(A) On a *Pde6b*<sup>wt/wt</sup> background

(B) On a *Pde6b*<sup>rd1/rd1</sup> background

Probe/gene name	log <sub>2</sub> fold change	p value
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Rod Specific		
<i>Gnat1</i>	-21.14	0.0002
<i>Pdc</i>	-1.83	0.0421
<i>Pde6b</i>	-3.47	0.0004
<i>Nrl</i>	-2.79	0.0380
<i>Sag</i>	-6.01	0.0009
<i>Slc24a1</i>	-5.27	0.0231
<i>Pde6g</i>	-3.02	0.0473
<i>Reep6</i>	-2.56	0.0380
<i>Esrrb</i>	-2.00	0.0153

Probe/gene name	log <sub>2</sub> fold change	p value
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Rod contentious		
<i>Gpnmb</i>	-4.14	0.0035
<i>Ldha</i>	-4.61	0.0014
<i>Car2</i>	-2.94	0.0004
<i>Adcy1</i>	-2.97	0.0004

Table S1. (A) Genes with rod specific ontological annotation; (B) Genes with literature suggesting contentiousness due to low bipolar compared to photoreceptor protein expression. A positive log fold change represents upregulation in degenerate samples. "*p* value" represents the adjusted *p*-value following False Detection Rate (FDR) testing.

Symbol	Probe/gene name	log <sub>2</sub> fold Δ	p value
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A - Probes upregulated in degeneration			
<i>Igfn1</i>	Immunoglobulin-like and fibronectin type III domain containing 1	4.10	0.0181
<i>Cgn</i>	Cingulin	3.52	0.0138
<i>Msrb2</i>	Methionine sulfoxide reductase B2	3.03	0.0344
<i>Cabyr</i>	Calcium binding tyrosine-(Y)-phosphorylation regulated	2.25	0.0380
<i>Efnb1</i>	Ephrin-B1	2.15	0.0312
<i>Ccdc171</i>	Coiled-coil domain containing 171	1.99	0.0458
<i>Phc1</i>	Polyhomeotic homolog 1	1.93	0.0276
<i>Sfxn1</i>	Sideroflexin 1	1.92	0.0195
<i>Fam19a3</i>	Family with sequence similarity 19 (chemokine (C-C motif)-like) A3	1.87	0.0458
<i>Shroom2</i>	Shroom family member 2	1.81	0.0416
<i>Sfrs1</i>	Serine/arginine-rich splicing factor 1	1.72	0.0417
<i>Sept4</i>	Septin 4	1.54	0.0324
<i>Sulf2</i>	Sulfatase 2	1.43	0.0344

B - Probes downregulated in degeneration			
<i>Nrm</i>	Nurim (nuclear envelope membrane protein)	-1.40	0.0416
<i>Gm362</i>	Predicted gene 362	-1.46	0.0138
<i>Pde9a</i>	Phosphodiesterase 9A	-1.55	0.0416
<i>Ndufb2</i>	NADH dehydrogenase (ubiquinone) 1 beta subcomplex, 2	-1.55	0.0414
<i>Gabrg2</i>	Gamma-aminobutyric acid (GABA) A receptor, gamma 2	-1.56	0.0494
<i>Anxa7</i>	Annexin A7	-1.56	0.0244
<i>Fam195a</i>	Family with sequence similarity 195, member A	-1.61	0.0053
<i>Atox1</i>	Antioxidant 1 copper chaperone	-1.62	0.0414
<i>Socs5</i>	Suppressor of cytokine signaling 5	-1.64	0.0149
<i>Csrnp2</i>	Cysteine-serine-rich nuclear protein 2	-1.65	0.0276
<i>Rnf11</i>	Ring finger protein 11	-1.75	0.0244
<i>Cntn1</i>	Contactin 1	-1.79	0.0494
<i>COXIII</i>	Mitochondrial cytochrome oxidase II subunit	-1.83	0.0244
<i>Ap3m2</i>	Adapter-related protein complex 3, mu 2 subunit	-1.85	0.0231
<i>Rnf11</i>	Ring finger protein 11	-1.88	0.0060
<i>Mif</i>	Macrophage migration inhibitory factor	-1.91	0.0337
<i>Pde1c</i>	Phosphodiesterase 1C, calmodulin-dependent 70kDa	-1.96	0.0053
<i>Pcdha7</i>	Protocadherin alpha 7	-1.98	0.0264
<i>Extl3</i>	Exostosin-like glycosyltransferase 3	-2.05	0.0291
<i>Unc13a</i>	Unc-13 homolog A	-2.11	0.0060
<i>Gm5478</i>	Predicted pseudogene 5478	-2.13	0.0103
<i>Tmsb10</i>	Thymosin beta 10	-2.15	0.0494
<i>Pgk1</i>	Phosphoglycerate kinase 1	-2.30	0.0414
<i>Stmnd1</i>	Stathmin domain containing 1	-2.42	0.0034
<i>Mt2</i>	Metallothionein 2	-2.50	0.0126
<i>Yaf2</i>	YY1 associated factor 2	-2.54	0.0473
<i>Clip1</i>	CAP-GLY domain containing linker protein 1	-2.75	0.0060
<i>Slc7a3</i>	Solute carrier family 7 (cationic amino acid transporter, Y+ system), 3	-2.87	0.0103
<i>Abca1</i>	ATP-binding cassette, sub-family A (ABC1), member 1	-3.10	0.0171
<i>Them6</i>	Thioesterase superfamily member 6	-3.24	0.0011
<i>Hdac9</i>	Histone deacetylase 9	-3.69	0.0029
<i>Ptpr</i>	Protein tyrosine phosphatase, receptor type, R	-3.72	0.0135
<i>Naa11</i>	N(alpha)-acetyltransferase 11, NatA catalytic subunit	-3.83	0.0004
<i>Lynx1</i>	Ly6/neurotoxin 1	-8.26	0.0135
<i>Mt1</i>	Metallothionein 1	-10.73	0.0015

Table S2. Differentially expressed genes in a microarray study comparing FACS-isolated EYFP-positive cells from degenerate and non-degenerate L7-Cre EYFP mouse retinas. Statistically significant, differentially expressed, probes are listed according to log fold change. A positive log fold change represents upregulation in the degenerate samples. “*p* value” represents the adjusted p-value following False Detection Rate (FDR) testing.

Probe	Log <sub>2</sub> fold Δ	p value
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Receptor		
<i>Grm6</i>	1.054	0.8573

Gα subunits		
<i>Gna11</i>	1.04	0.7896
<i>Gna11</i>	-1.07	0.8574
<i>Gna14</i>	-1.08	0.7764
<i>Gnai1</i>	-1.09	0.9360
<i>Gnai2</i>	-1.04	0.8810
<i>Gnai3</i>	1.01	0.9863
<i>Gnao1</i>	-1.23	0.7900
<i>Gnao1</i>	1.02	0.9835
<i>Gnaq</i>	-1.40	0.0539
<i>Gnaq</i>	-1.03	0.8832

Gβγ subunits		
<i>Gnb3</i>	1.91	0.1685
<i>Gnb4</i>	-1.05	0.9335
<i>Gnb5</i>	-1.01	0.9670
<i>Gng2</i>	-1.10	0.8044
<i>Gng3</i>	1.34	0.8414
<i>Gng5</i>	1.04	0.8175
<i>Gng7</i>	1.64	0.6041
<i>Gng7</i>	1.07	0.7679
<i>Gng10</i>	-1.08	0.7839
<i>Gng11</i>	-1.05	0.8035
<i>Gng13</i>	-1.13	0.7109

Bipolar cell-specific		
<i>Cabp5</i>	1.48	0.5394
<i>Pcp2</i>	-1.12	0.8044
<i>Prkca</i>	-1.09	0.7514
<i>Scgn</i>	1.40	0.8044
<i>Scgn</i>	-1.02	0.9746
<i>Vsx1</i>	1.04	0.9102
<i>Vsx2</i>	1.21	0.8056
<i>Vsx2</i>	1.10	0.9021

Kinases		
<i>Grk4</i>	1.13	0.7241
<i>Grk4</i>	-1.07	0.7896
<i>Grk5</i>	1.09	0.7918
<i>Grk5</i>	1.04	0.8642

Housekeeping genes		
<i>Actb</i>	1.45	0.6114
<i>Actb</i>	-1.31	0.7128
<i>Actb</i>	-1.03	0.9801
<i>Actb</i>	-1.03	0.9802

Probe	Log <sub>2</sub> fold Δ	p value
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Intracellular effectors		
<i>Adcy1</i>	-2.97	0.0004
<i>Adcy1</i>	1.05	0.8044
<i>Adcy2</i>	1.77	0.5721
<i>Adcy3</i>	1.01	0.9480
<i>Adcy8</i>	1.06	0.8086
<i>Adcy9</i>	1.05	0.8035
<i>Pde1b</i>	-1.37	0.5764
<i>Pde1b</i>	-1.41	0.6178
<i>Pde1c</i>	-1.96	0.0053
<i>Pde1c</i>	-1.22	0.1861
<i>Pde1c</i>	-1.09	0.7629
<i>Pde1c</i>	1.19	0.7932
<i>Pde1c</i>	-1.10	0.8337
<i>Pde1c</i>	1.00	0.9986
<i>Pde3a</i>	-1.03	0.8542
<i>Pde3a</i>	1.01	0.9441
<i>Pde3a</i>	1.00	1.0000
<i>Pde3b</i>	-1.02	0.9259
<i>Pde4a</i>	-1.12	0.5721
<i>Pde4b</i>	-1.26	0.4073
<i>Pde4b</i>	-1.20	0.7671
<i>Pde4b</i>	1.27	0.8311
<i>Pde4b</i>	-1.02	0.9409
<i>Pde4d</i>	-2.05	0.5154
<i>Pde4d</i>	-1.00	0.9918
<i>Pde7a</i>	-1.06	0.7887
<i>Pde7b</i>	-1.16	0.6154
<i>Pde8b</i>	-1.02	0.8908
<i>Pde9a</i>	-1.55	0.0416
<i>Plcb3</i>	1.07	0.7536
<i>Plcb4</i>	1.24	0.8044

TRP channels		
<i>Trpc6</i>	-1.05	0.7749
<i>Trpc7</i>	-1.29	0.7887
<i>Trpm1</i>	1.19	0.7847
<i>Trpm1</i>	-1.13	0.8035
<i>Trpm1</i>	1.09	0.8220
<i>Trpm1</i>	1.19	0.8401

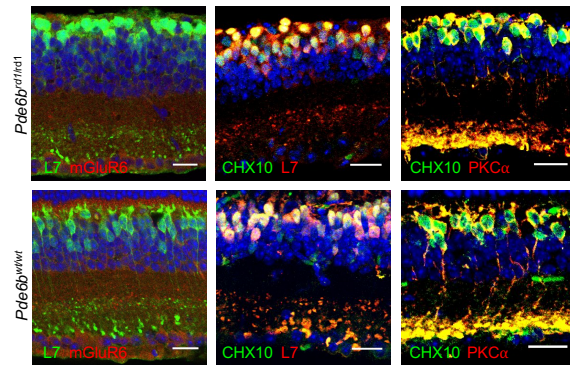
Arrestin		
<i>Arb2</i>	-1.11	0.8673

<i>Gapdh</i>	-1.51	0.1950
<i>Rplp0</i>	-1.15	0.8035
<i>Rplp0</i>	1.02	0.9900

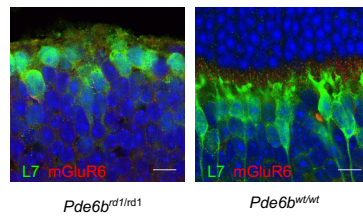


Table S3. In a microarray study comparing FACS-isolated EYFP-positive cells from degenerate and non-degenerate L7-Cre EYFP mouse retinas, genes related to intracellular communication, the native on-bipolar signal transduction cascade. A selection of “housekeeping” genes are included for comparison. A positive log fold change represents upregulation in degenerate samples. “p value” represents the adjusted p-value following False Detection Rate (FDR) testing. Note that multiple probes for some genes are reported in the above table, for clarity, all probes are labelled only with the associated gene name.

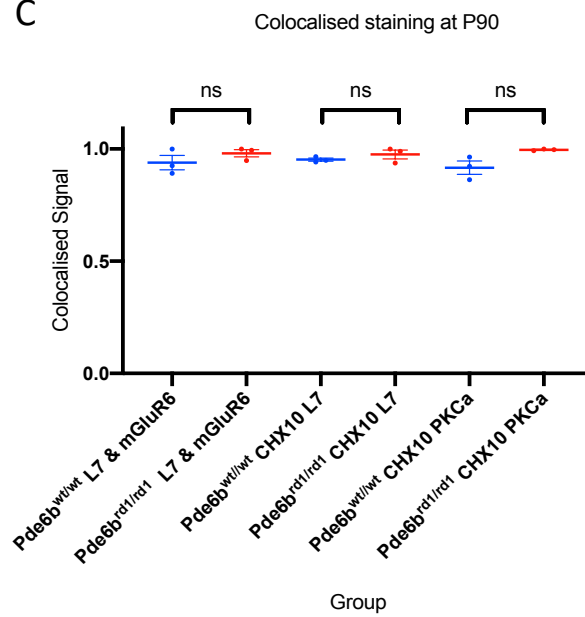
A



B



C



(Fig S3.)

Figure S3.

- A. Immunohistochemical staining of retina from *Pde6b<sup>rd1/rd1</sup>* and *Pde6b<sup>wt/wt</sup>* mice culled at P90 for proteins associated with the bipolar signalling cascade (mGluR6, PKC $\alpha$  & L7 – none of which demonstrated significant changes in gene expression between *Pde6b<sup>rd1/rd1</sup>* and *Pde6b<sup>wt/wt</sup>* - see table S3). In all panels, DAPI staining is in blue, other colours as indicated in panels scale bar = 20 $\mu$ m main panels.
- B. Higher magnification views of mGluR6 staining in the outer plexiform layer . Note particularly the punctate pattern of staining of the bipolar cells dendrites in the wild type. Note L7 staining covers cell bodies and dendrites. Scale bars =10 $\mu$ m.
- C. Colocalised pixels above threshold - a semiquantitative index of protein expression based on immunohistochemical staining. Normalised to highest value over all retinae stained for the same proteins. t-tests did not reveal a difference in mean values between *Pde6b<sup>rd1/rd1</sup>* and *Pde6b<sup>wt/wt</sup>* retinae stained for the same proteins ( $p>0.05$ ). NS = not-significant.

Probe	Log <sub>2</sub> fold Δ	p value
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Kainate type		
<i>Grik1</i>	1.57	0.6786
<i>Grik1</i>	1.07	0.7896
<i>Grik1</i>	1.13	0.7934
<i>Grik1</i>	1.09	0.8785
<i>Grik1</i>	-1.03	0.9705
<i>Grik4</i>	-1.01	0.9784

Vglut transporter		
<i>Slc17a6</i>	-1.05	0.8679
<i>Slc17a6</i>	-1.01	0.9865
<i>Slc17a7</i>	-1.05	0.8592
<i>Slc17a8</i>	-1.08	0.7415

Probe	Log <sub>2</sub> fold Δ	p value
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AMPA type		
<i>Gria1</i>	-1.39	0.7587
<i>Gria1</i>	1.05	0.8440
<i>Gria1</i>	1.04	0.8681
<i>Gria1</i>	1.04	0.8708
<i>Gria2</i>	1.27	0.6526
<i>Gria2</i>	1.21	0.7415
<i>Gria3</i>	-1.04	0.8035
<i>Gria3</i>	-1.06	0.9259
<i>Gria4</i>	1.07	0.7587

NMDA type		
<i>Grin2c</i>	-1.02	0.9094

*GABA and Glycine receptor subunits*

GABA		
<i>Gabra1</i>	-1.05	0.8824
<i>Gabra3</i>	1.07	0.8194
<i>Gabrb1</i>	1.01	0.9632
<i>Gabrb2</i>	1.11	0.6142
<i>Gabrb2</i>	1.01	0.9870
<i>Gabrb3</i>	1.12	0.7500
<i>Gabrb3</i>	-1.11	0.8013
<i>Gabrb3</i>	-1.03	0.9382
<i>Gabrg2</i>	-1.56	0.0494
<i>Gabrg2</i>	-1.48	0.6302
<i>Gabrg2</i>	-1.36	0.6308
<i>Gabrg2</i>	1.06	0.7420
<i>Gabrg2</i>	-1.06	0.8721
<i>Gabrg2</i>	-1.10	0.9477
<i>Gabrr1</i>	-1.56	0.5706
<i>Gabrr1</i>	-1.04	0.8718
<i>Gabrr2</i>	-1.16	0.7896

Glycine		
<i>Gla2</i>	-1.06	0.8015
<i>Gla4</i>	1.09	0.7753
<i>Glrb</i>	1.07	0.7969

Table S4. In a microarray study comparing FACS-isolated EYFP-positive cells from degenerate and non-degenerate L7-Cre EYFP mouse retinas, genes relating to glutamate, glycine and gaba receptors;. A positive log fold change represents upregulation in degenerate samples. "P value" represents the adjusted p-value following False Detection Rate (FDR) testing. Note that multiple probes for some genes are reported in the above table, for clarity, all probes are labelled only with the associated gene name.

<b>Protein of Interest</b>	<b>Antibody (source &amp; product number)</b>	<b>Plasmid used in validation ICC</b>
<b>ANNEXINa7</b>	Abcam ab197586	Source bioscience plasmid no. 3157926
<b>CONTACTIN1</b>	Abcam ab66265	Sino biologic plasmid no. MG50933
<b>SHROOM2</b>	Atlas hpa051646	GenScript plasmid no. OMu14281D
<b>SULPHATASE2</b>	Abcam ab113405	Addgene plasmid no. 13008
<b>CHX10</b>	Abcam ab16141	See Kim et al [REFERENCE Kim et al 2008]
<b>PKC<math>\alpha</math></b>	Abcam ab32376	See Ruether et al [REFERENCE Ruether et al 2010]

*(Table S5)*

Table S5. Antibodies and plasmids used in their validation

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