

## **Supplementary Information for**

Translational shutdown and evasion of the innate immune response by SARS-CoV-2 NSP14 protein

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**Fig. S1** NSP1 but not NSP14 inhibits poly(A) RNA nuclear export and total poly(A) RNA level. (A) Representative confocal images of RNA-FISH for poly(A) RNA detection. 293T cells were transfected with plasmids encoding indicated HA-tagged viral proteins for 24 h. Cells were fixed, stained by oligo(dT)-Cy5 probes to detect poly(A) RNA, anti-HA antibody for viral proteins, and Hoechst dye for the nucleus, and analyzed by confocal microscopy. White arrowheads show examples for transfected cells. Images represent > 40 HA-positive cells for each condition. Data represent two independent experiments.

(B) Fluorescence intensity of total poly(A) RNA in individual cells is shown for the indicated experimental condition. Data represent two independent experiments. \*\*\*p < 0.001 by unpaired Student's t test. ns, not significant.





For (A) to (C), Data are shown as mean  $\pm$  SD of three biological repeats. \*\*\*p < 0.001 by unpaired Student's t test. ns, not significant.



Fig. S3 Coronavirus NSP14 inhibits protein translation.

293T cells were transfected with increasing amounts of plasmid DNA encoding HA-tagged NSP14 proteins from different coronaviruses. After 24 h, cells were puromycin-labeled for 15 min. Puromycin incorporation was determined by immunoblotting using anti-puromycin antibody (Puro). HA-tagged NSP14 proteins were detected by anti-HA antibody (HA). EV, empty vector.

SARS-CoV-1 NSP14 SARS-CoV-2 NSP14	1 AENVTGLFKDCSKIITGLHPTQAPTHLSVDIKFKTEGLCVDIPGIPKDMTYRRLISMMGFKMNYQVNGYP 70 1 AENVTGLFKDCSKVITGLHPTQAPTHLSVDTKFKTEGLCVDIPGIPKDMTYRRLISMMGFKMNYQVNGYP 70 * ** * * * * * * * * * * * * * * * * *
SARS-CoV-1 NSP14 SARS-CoV-2 NSP14	71 NMFITREEAIRHVRAWIGFDVEGCHATRDAVGTNLPLQLGFSTGVNLVAVPTGYVDTENNTEFTRVNAKP 140 71 NMFITREEAIRHVRAWIGFDVEGCHATREAVGTNLPLQLGFSTGVNLVAVPTGYVDTPNNTDFSRVSAKP 140 * *
SARS-CoV-1 NSP14	141 PPGDQFKHLIPLMYKGLPWNVVRIKIVQMLSDTLKGLSDRVVFVLWAHGFELTSMKYFVKIGPERTCCLC 210
SARS-CoV-2 NSP14	141 PPGDQFKHLIPLMYKGLPWNVVRIKIVQMLSDTLKNLSDRVVFVLWAHGFELTSMKYFVKIGPERTCCLC 210
SARS-CoV-1 NSP14	211 DKRATCFSTSSDTYACWNHSVGFDYVYNPFMIDVQQWGFTGNLQSNHDQHCQVHGNAHVASCDAIMTRCL 280
SARS-CoV-2 NSP14	211 DRRATCFSTASDTYACWHHSIGFDYVYNPFMIDVQQWGFTGNLQSNHDLYCQVHGNAHVASCDAIMTRCL 280
SARS-CoV-1 NSP14	281 AVHECFVKRVDWSVEYPIIGDELRVNSACRKVQHMVVKSALLADKFPVLHDIGNPKAIKCVPQAEVEWKF 350
SARS-CoV-2 NSP14	281 AVHECFVKRVDWTIEYPIIGDELKINAACRKVQHMVVKAALLADKFPVLHDIGNPKAIKCVPQADVEWKF 350
SARS-CoV-1 NSP14	351 YDAQPCSDKAYKIEELFYSYATH <mark>H</mark> DKFTDGVCLFWNCNVDRYPANAIVCRFDTRVLSNLNLPGCDGGSLY 420
SARS-CoV-2 NSP14	351 YDAQPCSDKAYKIEELFYSYATH <mark>S</mark> DKFTDGVCLFWNCNVDRYPANSIVCRFDTRVLSNLNLPGCDGGSLY 420
SARS-CoV-1 NSP14	421 VNKHAFHTPAFDKSAFTNLKQLPFFYYSDSPCESHGKQVVSDIDYVPLKSATCITRCNLGGAVCRHHANE 490
SARS-CoV-2 NSP14	421 VNKHAFHTPAFDKSAFVNLKQLPFFYYSDSPCESHGKQVVSDIDYVPLKSATCITRCNLGGAVCRHHANE 490
SARS-CoV-1 NSP14	491 YRQYLDAYNMMISAGFSLWIYKQFDTYNLWNTFTRLQ 527
SARS-CoV-2 NSP14	491 YRLYLDAYNMMISAGFSLWVYKQFDTYNLWNTFTRLQ 527

**Fig. S4** Primary sequence alignment of SARS-CoV and SARS-CoV-2 NSP14 protein sequences using SnapGene. The protein sequences share 95% sequence identity and 99% sequence similarity. Similar and dissimilar residues are highlighted in blue and red, respectively. Residues at the NSP10-NSP14 interface are highlighted by red asterisks. Sequences of NSP14 in SARS-CoV (accession number ADC35510) and SARS-CoV-2 (QNQ17295) were used for the analysis.

CoV NSP10 CoV-2 NSP10	1 AGNATEVPANSTVLSFCAFAVDPAKAYKDYLASGGQPITNCVKMLCTHTGTGQAITVTPEANMDQESFGG 70 1 AGNATEVPANSTVLSFCAFAVDAAKAYKDYLASGGQPITNCVKMLCTHTGTGQAITVTPEANMDQESFGG 70 *** * * * * * * * * * * * * *
CoV NSP10 CoV-2 NSP10	71 ASCCLYCRCHIDHPNPKGFCDLKGKYVQIPTTCANDPVGFTLRNTVCTVCGMWKGYGCSCDQLREPLMQ 139 71 ASCCLYCRCHIDHPNPKGFCDLKGKYVQIPTTCANDPVGFTLKNTVCTVCGMWKGYGCSCDQLREPMLQ 139 * * *

**Fig. S5** Primary sequence alignment of SARS-CoV and SARS-CoV-2 NSP10 protein sequences using SnapGene. The protein sequences share 97% sequence identity and 99% sequence similarity. Similar and dissimilar residues are highlighted in blue and red, respectively. Residues at the NSP10-NSP14 interface are highlighted by red asterisks. Sequences of NSP10 in SARS-CoV (accession number ADC35510) and SARS-CoV-2 (QNQ17295) were used for the analysis.



Fig. S6 NSP10 mutants that disrupt the NSP10-NSP14 interaction fail to enhance the translation inhibition activity of NSP14.

(A) 293T cells were co-transfected with FLAG-tagged NSP10 and HA-tagged NSP14 or its mutants for 24 h and puromycin-labeled for 15 min. Puromycin incorporation was determined by immunoblotting. EV, empty vector.

(B) SARS-CoV NSP10-NSP14 interaction interface. NSP10 (red) interacts with the N-terminal ExoN domain of NSP14 (green). K43 and H80 are highlighted in red boxes. Zinc ion is represented as gray spheres. Figure adapted from (1).

(C) Effects of SARS-CoV NSP10 mutations on the NSP10-NSP14 interaction and on the ExoN activity of NSP14 (2). Effects of mutations on the replication of SARS-CoV were determined by plaque assay at 24 hours post infection (2).

(D) 293T cells were co-transfected with FLAG-tagged NSP10 and HA-tagged NSP14NSP14 or their mutants for 24 h and puromycin-labeled for 15 min. Puromycin incorporation was determined by immunoblotting.

## SI References

- 1. Y. Ma *et al.*, Structural basis and functional analysis of the SARS coronavirus nsp14nsp10 complex. *Proc Natl Acad Sci U S A* **112**, 9436-9441 (2015).
- M. Bouvet *et al.*, Coronavirus Nsp10, a critical co-factor for activation of multiple replicative enzymes. *J Biol Chem* 289, 25783-25796 (2014).