

Supplementary Data

A



Log Likelihood Ratio: 27 Information Content: 13.4 Relative Entropy: 13 Bayes Threshold: 3.45943

Name	Start	p-value	Sites
3. ASO2	12	3.88e-5	CCGGCCTCGG AAGCUCUCU
1. 196861965-C2	3	3.88e-5	GC A <u>GUCUCAU</u> GTCCACAGGC
2. 195513083-E10	10	7.24e-5	GAUGGTTCC A <u>GUCUCAU</u> UC

B



Log Likelihood Ratio: 33 Information Content: 10.1 Relative Entropy: 9.5 Bayes Threshold: 5.25873

Name	Start	p-value	Sites
3. P2-F2	4	2.97e-4	AGC A <u>GUCUC</u> CATGTCCCAG
1. P2-C2	3	2.97e-4	GC A <u>GUCUC</u> CATGTCCCAG
7. P2-E8	2	6.29e-4	G <u>GUUCUC</u> CTCCACAGCC
2. P2-E10	10	1.03e-3	GAUGGTTCC A <u>GUCUC</u> UCUUC
10. P2-F9	1	1.48e-3	<u>GUUCUC</u> ATCCCTTCTG

C



Log Likelihood Ratio: 31 Information Content: 10.4 Relative Entropy: 11.1 Bayes Threshold: 6.08215

Name	Strand	Start	p-value	Sites
11. ASO2	+	4	1.47e-4	TCC <u>GGCCTC</u> GGAAGCTCTC
8. P2-C4	+	11	1.47e-4	CCATGTCCCA <u>GGCCTC</u> CAGT
4. P2-C11	+	3	2.51e-4	TT <u>GGCCTG</u> TGGATGCTTT
7. P2-A11	+	11	4.58e-4	AAATGCTCTG <u>GGCCTC</u> ACTG

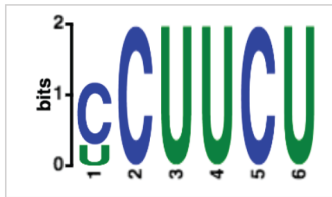
D



Log Likelihood Ratio: 55 Information Content: 16.7 Relative Entropy: 16 Bayes Threshold: 3.44518

Name	Strand	Start	p-value	Sites
6. P2-A6	+	4	3.25e-7	GAT TAAACAGATTAA TACA
1. P2-C6	+	5	3.25e-7	GGAT TAAACAGATTAA TAC
3. P2-E6	+	4	7.92e-6	CTT GTAAACAGATTAT CTTC
5. P2-C5	+	3	1.10e-5	CC TAGAAACAGCAA AGATT
10. P2-D6	+	3	1.53e-5	AA TTTAAACAGCAA TATTA

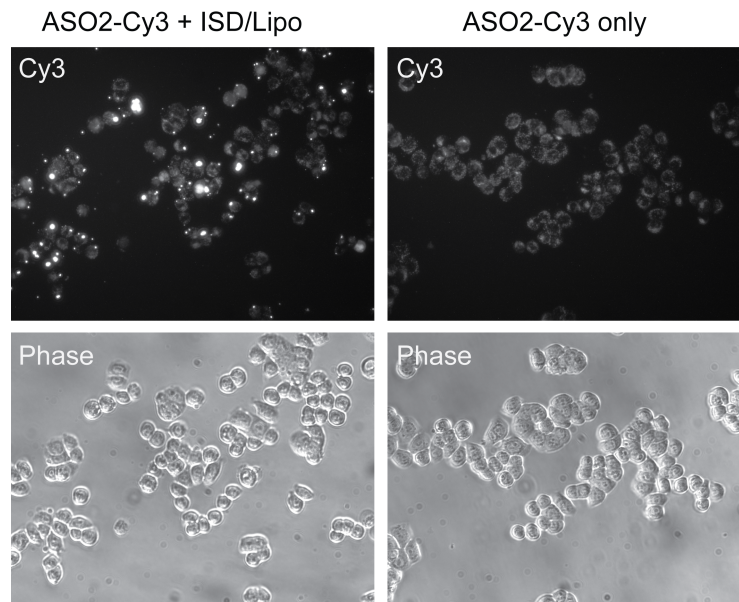
E



Log Likelihood Ratio: 50 Information Content: 11.2 Relative Entropy: 9 Bayes Threshold: 5.7039

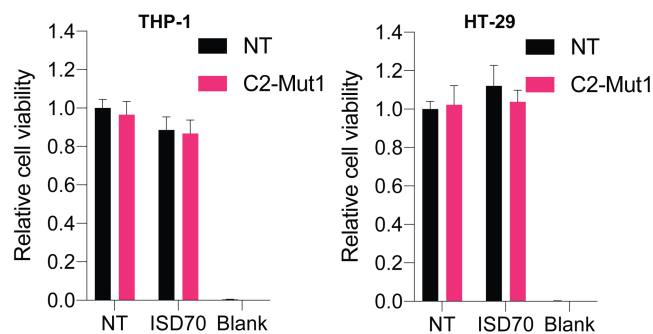
Name	Start	p-value	Sites
12. P2-D10	4	1.17e-3	CAC CCUUCU CTCTGGUCCC
8. P2-A8	1	1.17e-3	CCUUCU CTCTGGTCCC
6. P2-A9	15	1.17e-3	UGGTCCCATC CCUUCU
4. P2-A3	15	1.17e-3	CTGCTGCCCA CCUUCU
2. P2-H9	9	1.17e-3	GUCCCATC CCUUCU GCUGCC
1. P2-H8	14	1.17e-3	UCCTCCACAC CCUUCU C
11. P2-H2	14	2.11e-3	GCCTCCAGTG UCUUCU C
9. P2-G7	8	2.11e-3	UCCCAAC UCUUCU AACUCGU

Supplementary Figure S1. Multiple Em for Motif Elicitation (MEME) motif discovery in ASOs. (A) The two most potent cGAS inhibitors in our 80 ASOs screen, including ASO2 (i.e. ASO2, C2 and E10) were analysed with MEME. (B) The 10 most potent cGAS inhibitors in our 80 ASOs screen were analysed with MEME (see Supplementary Table S2). Five sequences were identified with the motif shown (noting that C2 and F2 are closely related sequences, but the other ones were not). (C, D) The 10 most potent TLR9 inhibitors in our 80 ASOs screen were analysed with MEME (see Supplementary Table S2), along ASO2. Two motifs are shown here, including one common with ASO2 (C), and an A-rich central motif (D). (E) MEME analysis of 17 ASOs with less than 10% inhibition of cGAS sensing in our 80 ASOs screen (see Supplementary Table S2). Eight ASOs shared the highlighted motif (noting that D10/A8 and A9/H9 are related sequences, respectively). (A-E) The figures are direct screenshots from the MEME website. The ASO names are provided as per their reference position in the plate (see Supplementary Table S2).



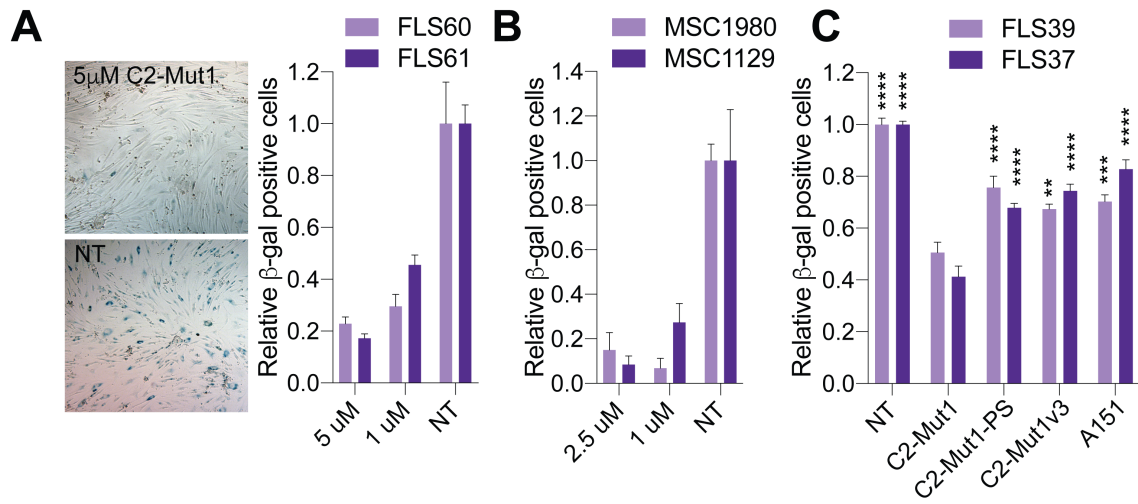
Supplementary Figure S2.

HT-29 cells were incubated overnight with 500 nM ASO2-Cy3, prior to ISD70+lipofectamine transfection or not, for 4 h. The cells were subsequently washed with PBS prior to imaging by inverted fluorescent microscopy. The images shown are representative of two independent experiments. While cytosolic fluorescence in ASO2-Cy3 only treated cells clearly confirmed spontaneous uptake of the ASOs by the cells, the occurrence of bright fluorescent puncta suggests an increased uptake of the labelled ASOs during the transfection of the lipofectamine-ISD complexes.

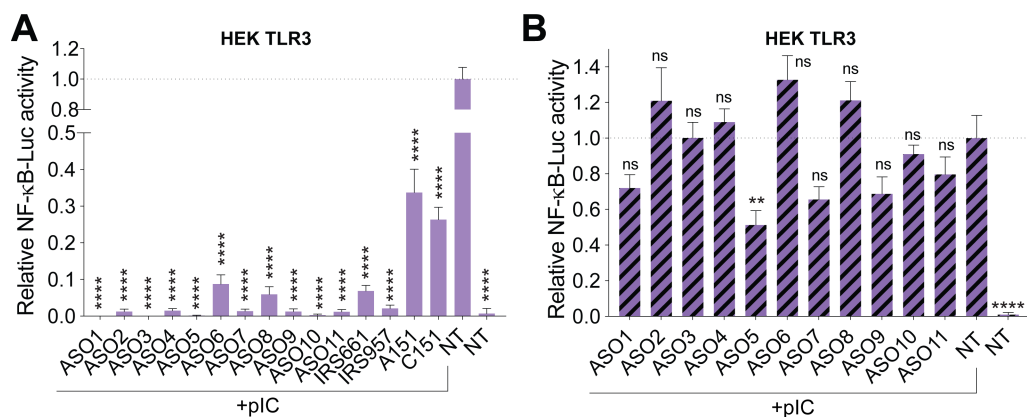


Supplementary Figure S3.

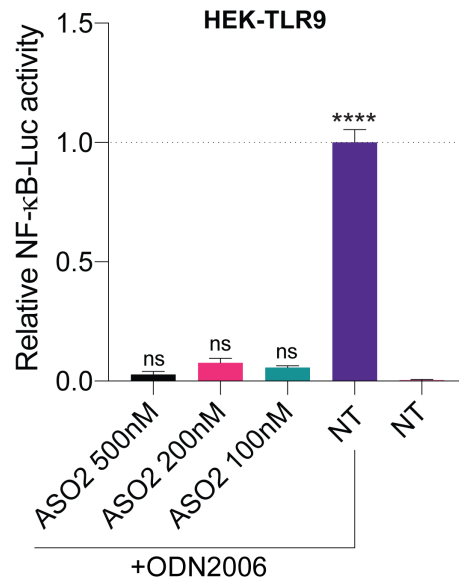
THP-1 and HT-29 cells were incubated for 6 h with 100 nM or 187.5 nM C2-Mut1, respectively, prior to overnight transfection or not with ISD70. The next day, 1X resazurin solution was added to each well, and cell viability was measured after 4-5 h at 37°C. Data were normalised to the NT condition (no ASO, no ISD70), after background correction with blank condition. Data shown are averaged from two independent experiments in biological triplicate (\pm s.e.m).



Supplementary Figure S4. (A) Primary FLS cells from two RA patients were cultivated for 15 days in the presence of indicated doses of naked ASOs, prior to being fixed and analysed for β -galactosidase staining. The numbers of β -galactosidase positive cells were normalised to the NT condition. Left panel: representative images of NT and 5 μ M C2-Mut1 conditions; 40-50% of the cells in NT conditions were positive for β -galactosidase staining. Data shown are averaged from three replicates for each condition (\pm s.e.m), for each independent donor. (B) Primary bone marrow-derived MSCs from two different patients were cultivated for 2 weeks in the presence of indicated doses of naked C2-Mut1, prior to being fixed and analysed for β -galactosidase staining. The numbers of β -galactosidase positive cells were normalised to the NT condition. Data shown are averaged from six replicates for each condition (\pm s.e.m), for each independent donor. (C) Primary FLS cells from two control patients were cultivated for 7 days in the presence of 2.5 μ M naked ASOs, prior to being fixed and analysed for β -galactosidase staining. The numbers of β -galactosidase positive cells were normalised to the NT condition. Data shown are averaged from at least five replicates for each condition (\pm s.e.m), for each independent donor. Ordinary two-way ANOVA with Sidak's multiple comparison tests relative to the "C2-Mut1 only" condition of each donor are shown.



Supplementary Figure S5. HEK-TLR3 cells expressing a NF- κ B-luciferase reporter were treated with 500 nM (A) or 100 nM (B) indicated ASOs/ODNs for 20-50 min prior to stimulation or not (non-treated [NT]) with poly(I:C) (at 1 μ g/ml [A] or 0.5 μ g/ml [B]). NF- κ B-luciferase levels were measured after overnight incubation and were normalised to the "pIC" condition, after background correction with NT condition. (A, B) Data are averaged from two independent experiments in biological triplicate (\pm s.e.m and one-way ANOVA with Dunnett's multiple comparison tests to the condition "pIC" are shown). ** $P \leq 0.01$, **** $P \leq 0.0001$. ns: non-significant.



Supplementary Figure S6. Inhibition of TLR9 sensing by ASO2 is preserved with decreasing amounts of ASO2. HEK-TLR9 cells expressing a NF-κB-luciferase reporter were treated with indicated amount of ASO2 (100-500 nM) for 50 min prior to stimulation or not (non-treated [NT]) with 200 nM ODN2006. NF-κB-luciferase levels were measured after overnight incubation. NF-κB-luciferase levels were normalised to “ODN2006 only” condition, after background correction with the NT condition. Data shown are averaged from two independent experiments in biological triplicate (\pm s.e.m and one-way ANOVA with Dunnett’s multiple comparison tests to the condition “NT” are shown). **** $P < 0.0001$. ns: non-significant.

Supplementary Table S1. Various oligonucleotides used in this study (all in 5'-3'). UPPERCASE alone for DNA, 'm' indicates 2'OMe base, and * denotes the phosphorothioate backbone.

Name	Sequence
[cGAS]ASO1-146	mA*mU*mG*mG*mC*C*T*T*T*C*G*T*G*C*mC*mA*mA*mG*mG
[cGAS]ASO2-168	mU*mC*mC*mG*mG*C*C*T*G*G*A*A*G*C*mU*mC*mU*mC*mU
[cGAS]ASO3-200	mG*mC*mA*mU*mU*C*G*T*G*G*G*A*A*mG*mC*mC*mU*mU
[cGAS]ASO4-277	mG*mG*mC*mC*mG*A*A*C*T*T*T*C*G*mC*mC*mU*mU*mA
[cGAS]ASO5-521	mG*mG*mU*mC*mU*T*G*G*C*T*T*C*G*T*G*mG*mA*mG*mC*mA
[cGAS]ASO6-616	mG*mG*mA*mG*mC*T*T*G*A*G*G*C*C*mC*mA*mG*mG*mC
[cGAS]ASO7-697	mG*mG*mU*mG*mG*T*C*A*A*A*C*G*mC*mU*mU*mU*mC
[cGAS]ASO8-793	mC*mA*mU*mU*mA*G*G*T*G*A*G*A*A*mU*mC*mU*mU*mC
[cGAS]ASO9-829	mU*mU*mC*mU*mG*G*G*A*A*C*T*T*C*A*mG*mU*mU*mU*mA
[cGAS]ASO10-1098	mU*mG*mA*mU*mU*C*A*A*A*G*C*A*G*mG*mG*mU*mU*mA
[cGAS]ASO11-1185	mC*mU*mU*mU*mA*G*T*G*T*A*G*T*T*G*mC*mU*mU*mC*mC
ASO11Mut1	mC*mU*mU*mU*mA*G*T*G*T*A*G*T*T*G*mU*mC*mU*mC*mU
ASO11Mut2	mU*mC*mC*mG*mG*G*T*G*T*A*G*T*T*G*mC*mU*mU*mC*mC
ASO2up	mU*mC*mC*mG*mG*C*C*T*G*G*A*G*T*C*mU*mC*mC*mA*mU
ASO2down	mU*mC*mC*mG*mG*C*C*T*G*G*A*G*A*mU*mA*mU*mC*mG
ASO2-Mut1	mU*mC*mC*mG*mG*C*C*T*G*G*G*A*G*A*mU*mC*mU*mC*mU
C2	mG*mC*mA*mG*mU*C*T*C*A*T*G*T*C*mC*mA*mG*mG*mC
C2-Mut1	mG*mC*mG*mG*mU*A*T*C*A*T*G*T*C*mC*mA*mG*mG*mC
C2-Mut2	mG*mC*mG*mG*mU*A*T*A*C*A*G*G*T*C*mC*mA*mG*mG*mC
C2-Mut1v1	mG*mC*mU*mG*mU*T*T*C*A*T*G*T*C*mC*mA*mG*mG*mC
C2-Mut1v2	mG*mC*mU*mG*mU*G*T*C*A*T*G*T*C*mC*mA*mG*mG*mC
C2-Mut1v3	mG*mC*mC*mG*mU*T*T*C*A*T*G*T*C*mC*mA*mG*mG*mC
C2-Mut1v4	mG*mC*mC*mG*mU*G*T*C*A*T*G*T*C*mC*mA*mG*mG*mC
C2-ASO2-A	mG*mC*mG*mG*mU*A*T*C*A*T*A*G*T*C*mU*mC*mC*mA*mU
C2-ASO2-B	mG*mC*mG*mG*mU*A*T*C*A*T*C*A*G*A*mU*mA*mU*mC*mG
C2-Mut1-PS	G*C*G*G*T*A*T*C*A*T*G*T*C*C*A*G*G*C
[C2]Mut1-dC	mG*mG*mU*A*T*C*C*C*C*C*C*C*C*C*C*C*C
[C2]Mut1-v3-dC	mC*mG*mU*T*T*C*C*C*C*C*C*C*C*C*C*C
dC20	C*C*C*C*C*C*C*C*C*C*C*C*C*C*C
ASO2-Cy3	mU*mC*mC*mG*mG*C*C*T*G*G*A*A*G*C*mU*mC*mU*mC*mU/3Cy3Sp/
A151	T*T*A* G*G*G* T*T*A* G*G*G* T*T*A* G*G*G* T*T*A* G*G*G
C151	T*T*C* A*A*A* T*T*C* A*A*A* T*T*C* A*A*A* T*T*C* A*A*A
IRS661	T*G*C* T*T*G* C*A*A* G*C*T* T*G*C* A*A*G* C*A
IRS957	T*G*C* T*T*G* A*C*A* T*C*C* T*G*G* A*G*G* G*G*T* T*G*T
[HPRT]551	mC*mU*mU*mC*mC*A*A*A*T*A*A*G*A*mC*mA*mU*mU*mC
[HPRT]660	mC*mU*mU*mC*mG*T*G*G*G*T*C*T*T*mU*mU*mC*mA*mC
[HPRT]661	mA*mC*mU*mU*mC*G*T*G*G*G*T*C*T*mU*mU*mU*mC*mA
[HPRT]662	mC*mA*mC*mU*mU*C*G*T*G*G*G*T*C*mU*mU*mU*mU*mC
[HPRT]664	mA*mA*mC*mA*mC*T*T*C*G*T*G*G*G*T*mC*mC*mU*mU*mU
[HPRT]665	mC*mA*mA*mC*mA*C*T*T*C*G*T*G*G*G*mU*mC*mC*mU*mU
[HPRT]666	mC*mC*mA*mA*mC*A*C*T*T*C*G*T*G*G*G*mG*mU*mC*mC*mU
ODN 2006	T*C*G* T*C*G* T*T*T* T*G*T* C*G*T* T*T*T* G*T*C* G*T*T
ODN 1826	T*C*C* A*T*G* A*C*G* T*T*C* C*T*G* A*C*G* T*T
ISD45-FWD	TAC AGA TCT ACT AGT GAT CTA TGA CTG ATC TGT ACA TGA TCT ACA
ISD45-REV	TGT AGA TCA TGT ACA GAT CAG TCA TAG ATC ACT AGT AGA TCT GTA
ISD70-FWD	CCA TCA GAA AGA GGT TTA ATA TTT TTG TGA GAC CAT CGA AGA GAG AAA GAG ATA AAA CTT TTT TAC GAC T
ISD70-REV	AGT CGT AAA AAA GTT TTA TCT CTT TCT CTC TTC GAT GGT CTC ACA AAA ATA TTA AAC CTC TTT CTG ATG G

Supplementary Table S2. Eighty ASO screen data. The targeted gene names are provided in brackets (e.g. [CDKN2B-AS1]), followed by the reference position in the target RNA. ASOs were synthesised with the following modifications: UPPERCASE alone for DNA, ‘m’ indicates 2’OMe base modifications, and * denotes the phosphorothioate backbone. The “Position” column denotes the reference position of the ASOs in the 96 well plate used in the screen. The concentration of the ASOs used in each screen is indicated. Averaged NF-κB-luciferase or IP-10 levels from each screen are given relative to each of the ISD70 (cGAS), ODN2006 (TLR9), R848 (TLR7/8 – Alharbi et al., *Nucleic Acids Res.* 2020) conditions, as percentages (TLR7/9/cGAS) or fold increases (TLR8). **Bold** denotes the 10 strongest cGAS inhibitors used for motif discovery in Supplementary Figure S1B. *Italic* denotes the 17 ASOs with less than 10% inhibition of the “ISD70 only” condition used for motif discovery in Supplementary Figure S1E. Underlined denotes the 10 most potent TLR9 inhibitors used for motif discovery in Supplementary Figure S1C and S1D. **Blue** denotes the 16 most potent TLR9 inhibitors and **red** the 16 weakest inhibitors of TLR9 used in Figure 4G. **Yellow highlight** denotes the 4 ASOs inhibiting TLR7/9 and cGAS by less than 30%.

[Transcript Name] and Seq Name	Sequence	Position	cGAS-100nM	TLR9-500nM	TLR7-100nM	TLR8-500nM
[CDKN2B-AS1]132	mU*mU*mA*mA*mA*mA*T*A*A*T*C*T*A*G*T*T*mU*mG*mA*mA*mG	B5	52.20	98.18	25.66	6.43
[CDKN2B-AS1]1415	mG*mU*mG*mU*mC*C*T*T*C*A*T*G*C*T*T*mU*mG*mG*mA*mU	F3	58.18	98.99	18.05	1.60
[CDKN2B-AS1]1519	mA*mG*mA*mA*mA*mA*G*A*A*G*C*A*A*G*A*mU*mU*mC*mA*mA	F5	49.09	68.12	42.99	2.78
[CDKN2B-AS1]1522	mC*mC*mU*mA*mG*A*A*G*A*G*A*G*A*A*mA*mG*mA*mU*mU	C5	60.90	42.44	57.20	4.94
[CDKN2B-AS1]1528	mG*mU*mC*mA*mA*A*C*C*T*A*G*A*A*G*mA*mA*mG*mC*mA	E4	68.94	68.13	23.64	1.85
[CDKN2B-AS1]1773	mG*mA*mU*mU*mA*A*A*A*C*A*G*A*T*T*A*mA*mU*mA*mC*mA	A6	89.36	44.97	21.92	1.48
[CDKN2B-AS1]1774	mG*mG*mA*mU*mU*A*A*A*A*C*A*G*A*T*T*mA*mA*mU*mA*mC	C6	50.11	27.79	27.16	1.67
[CDKN2B-AS1]1775	mA*mG*mG*mA*mU*T*A*A*A*A*C*A*G*A*T*mU*mA*mA*mU*mA	G4	88.54	63.48	36.28	2.63
[CDKN2B-AS1]2130	mA*mG*mA*mU*mU*A*T*C*T*T*C*T*T*T*mA*mA*mU*mU*mU	F6	55.80	78.72	37.10	6.26
[CDKN2B-AS1]2131	mA*mA*mG*mA*mU*T*A*T*C*T*T*C*T*T*mU*mA*mA*mU*mU	G5	73.50	59.77	42.09	5.62
[CDKN2B-AS1]2132	mA*mA*mA*mG*mA*T*T*A*T*C*T*T*C*T*T*mU*mU*mA*mA*mU	E5	74.59	63.14	49.28	5.04
[CDKN2B-AS1]2133	mA*mA*mA*mA*mG*A*T*T*A*T*C*T*T*C*T*mU*mU*mU*mA*mA	B6	57.68	80.95	47.41	4.68
[CDKN2B-AS1]2134	mG*mA*mA*mA*mA*G*A*T*T*A*T*C*T*T*C*mU*mU*mU*mU*mA	H5	51.49	61.11	34.44	1.87
[CDKN2B-AS1]2137	mU*mG*mU*mG*mA*A*A*G*A*T*T*A*T*C*mU*mU*mC*mU*mU	G6	40.72	62.16	34.51	4.97
[CDKN2B-AS1]2138	mU*mU*mG*mU*mG*A*A*A*G*A*T*T*A*T*mC*mU*mU*mC*mU	H6	69.36	61.15	35.02	1.94
[CDKN2B-AS1]2139	mC*mU*mU*mG*mU*G*A*A*A*G*A*T*T*A*mU*mC*mU*mU*mC	E6	33.75	34.91	85.33	6.02
[CDKN2B-AS1]2196	mG*mG*mU*mG*mG*C*C*A*G*A*G*A*G*A*A*mC*mG*mU*mC*mA	D4	48.58	97.04	39.07	0.98
[CDKN2B-AS1]2198	mA*mA*mG*mG*mU*G*G*C*C*A*C*A*G*G*C*mA*mA*mC*mG*mU	H3	60.48	84.28	57.85	0.78
[CDKN2B-AS1]2218	mA*mG*mG*mC*mC*T*C*A*G*T*G*T*C*T*mU*mC*mU*mC*mC	B3	62.85	81.24	44.73	4.66
[CDKN2B-AS1]2219	mC*mA*mG*mG*mC*C*T*C*A*G*T*G*T*C*mU*mU*mC*mU*mC	H2	99.38	66.99	54.11	3.89
[CDKN2B-AS1]2223	mG*mU*mC*mC*mC*A*G*G*C*C*T*C*A*G*mU*mG*mU*mC*mU	G3	74.22	67.61	19.01	2.17
[CDKN2B-AS1]2227	mC*mC*mA*mU*mG*T*C*C*A*G*G*C*C*T*mC*mC*mA*mG*mU	C4	36.34	48.79	41.39	1.46
[CDKN2B-AS1]2230	mU*mC*mU*mC*mC*A*T*G*T*C*C*A*G*G*mC*mC*mU*mC*mC	C3	94.78	118.93	43.45	9.17
[CDKN2B-AS1]2231	mG*mU*mC*mU*mC*C*A*T*G*T*C*C*A*G*mG*mC*mC*mU*mC	D2	84.16	77.80	23.75	1.25
[CDKN2B-AS1]2232	mA*mG*mU*mC*mU*C*C*A*T*G*T*C*C*A*mG*mG*mC*mC*mU	B2	88.13	122.20	35.77	2.28
[CDKN2B-AS1]2233	mC*mA*mG*mU*mC*T*C*A*T*G*T*C*C*mA*mG*mG*mC*mC	A2	75.92	55.13	47.75	3.81
[CDKN2B-AS1]2234	mG*mC*mA*mG*mU*C*T*C*A*T*G*T*C*C*mC*mA*mG*mG*mC	C2	10.02	90.04	20.30	2.25
[CDKN2B-AS1]2235	mA*mG*mC*mA*mG*T*C*T*C*A*T*G*T*C*mC*mC*mA*mG*mG	F2	28.75	73.18	25.50	4.93
[CDKN2B-AS1]2236	mA*mA*mG*mC*mA*G*T*T*C*A*T*G*T*mC*mC*mC*mA*mG	G2	107.34	69.71	35.69	2.62
[CDKN2B-AS1]2237	mA*mA*mA*mG*mC*A*G*T*T*C*A*T*G*mU*mC*mC*mC*mA	E2	70.95	87.17	27.16	3.38
[CDKN2B-AS1]495	mA*mG*mU*mG*mG*C*A*C*A*T*A*C*A*C*mA*mC*mC*mC*mU	B4	59.69	86.07	41.95	4.48
[CDKN2B-AS1]611	mG*mU*mG*mU*mU*T*T*A*A*T*T*T*G*mU*mA*mG*mA*mG	H4	66.24	98.50	23.63	1.88
[CDKN2B-AS1]613	mC*mA*mG*mU*mG*T*T*T*T*A*A*T*T*mU*mG*mU*mA*mG	F4	60.94	92.59	62.68	6.73
[CDKN2B-AS1]626	mA*mU*mU*mU*mC*C*A*A*T*G*C*C*A*mG*mU*mG*mU*mU	A4	52.58	63.65	32.05	4.09
[CDKN2B-AS1]627	mU*mA*mU*mU*mU*C*C*A*A*T*G*C*C*mA*mG*mU*mG*mU	E3	80.99	71.20	30.54	3.87
[CDKN2B-AS1]645	mA*mA*mU*mU*mU*A*A*G*C*A*T*G*A*A*mU*mA*mU*mU*mA	D6	67.63	53.26	46.75	5.02
[CDKN2B-AS1]79	mA*mA*mA*mA*mU*A*A*G*G*G*G*A*A*T*A*mG*mG*mG*mA	D5	94.10	67.44	68.45	0.71
[CDKN2B-AS1]80	mU*mA*mA*mA*mA*T*A*A*G*G*G*A*A*T*mA*mG*mG*mG*mG	A5	110.58	64.58	40.95	1.30
[CDKN2B-AS1]831	mA*mU*mA*mU*mC*T*G*C*T*G*C*C*A*C*mC*mU*mU*mC*mU	A3	108.01	73.46	47.52	5.89
[CDKN2B-AS1]832	mA*mA*mU*mA*mU*C*T*G*C*T*G*C*C*A*mC*mC*mU*mU*mC	D3	83.17	70.72	43.66	4.45
[LINC-PINT]101	mU*mC*mC*mC*mA*T*C*C*T*T*C*T*G*C*mU*mG*mC*mC*mA	D9	86.62	59.80	43.08	10.88
[LINC-PINT]102	mG*mU*mC*mC*mC*A*T*C*C*T*T*C*T*G*mC*mU*mG*mC*mC	H9	113.87	101.22	22.47	5.07
[LINC-PINT]103	mG*mG*mU*mC*mC*C*A*T*C*C*T*T*C*T*mG*mC*mU*mG*mC	F9	45.23	88.75	20.75	1.87

[LINC-PINT]106	mU*mC*mU*mG*mG*T*C*C*A*T*C*C*T*mU*mC*mU*mG*mC	B10	48.75	56.60	29.42	5.25
[LINC-PINT]108	mU*mC*mU*mC*mU*mG*T*C*C*A*T*C*C*mC*mU*mU*mC*mU	A9	104.65	100.51	78.25	11.42
[LINC-PINT]109	mC*mU*mC*mU*mC*T*G*T*C*C*A*T*C*mC*mC*mU*mU*mC	B9	80.22	89.85	84.97	10.10
[LINC-PINT]110	mU*mC*mU*mC*mU*C*T*G*T*C*C*A*T*mC*mC*mC*mU*mU	G9	92.39	74.25	43.82	11.27
[LINC-PINT]111	mU*mU*mC*mU*mC*T*G*T*C*C*A*mU*mC*mC*mC*mU	E9	77.41	98.01	26.22	9.51
[LINC-PINT]112	mC*mU*mU*mC*mU*C*T*G*T*C*C*A*mU*mC*mC*mC	B8	82.64	64.35	56.20	9.21
[LINC-PINT]113	mC*mC*mU*mU*mC*T*C*T*G*T*C*C*mC*mA*mU*mC*mC	A8	103.79	55.47	60.90	3.51
[LINC-PINT]114	mC*mC*mC*mU*mU*C*T*C*T*G*T*C*mC*mC*mA*mU*mC	C8	84.38	62.09	49.59	8.12
[LINC-PINT]115	mA*mC*mC*mU*mU*T*C*T*G*T*C*mC*mC*mA*mU	G8	89.81	32.00	43.37	7.57
[LINC-PINT]116	mC*mA*mC*mC*T*T*C*T*G*mU*mC*mC*mA	D10	97.19	102.85	73.30	5.97
[LINC-PINT]1222	mU*mU*mA*mG*mC*T*T*G*T*C*mG*mU*mU*mC*mC	C9	63.80	78.17	31.67	8.41
[LINC-PINT]126	mG*mU*mC*mU*mC*T*C*A*A*C*C*mU*mU*mC*mU*mC	H8	114.36	77.09	25.01	7.17
[LINC-PINT]127	mG*mG*mU*mC*mU*C*T*C*A*A*C*C*mU*mU*mC*mU	D8	76.95	77.91	23.66	3.72
[LINC-PINT]128	mG*mG*mG*mU*mC*T*C*C*A*A*C*mC*mC*mU*mU*mC	E8	37.83	94.51	21.47	1.23
[LINC-PINT]1284 KL	mU*mC*mC*mA*A*C*T*T*A*A*mC*mU*mC*mG*mU	G7	99.64	86.73	40.79	13.35
[LINC-PINT]1315 KL	mG*mC*mA*mA*mG*G*C*A*G*A*G*A*A*C*mU*mC*mC*mA*mG	H7	30.64	53.46	19.99	2.69
[LINC-PINT]148 .1	mA*mA*mA*mU*mG*T*C*T*G*G*C*C*T*mC*mA*mC*mU*mG	A11	66.46	46.72	46.45	1.56
[LINC-PINT]1497 .1	mG*mA*mU*mG*mG*T*T*C*A*G*T*C*C*mU*mC*mU*mU*mC	E10	26.05	80.47	22.32	1.01
[LINC-PINT]2673 .1	mU*mU*mG*mG*mC*T*G*T*G*G*A*T*G*C*mU*mU*mU*mG*mU	C11	68.35	40.71	21.43	0.94
[LINC-PINT]2690	mU*mU*mU*mG*mA*A*A*T*T*C*A*G*A*G*mA*mU*mU*mU*mG	H11	84.15	54.33	35.07	4.93
[LINC-PINT]2755	mC*mU*mU*mU*mA*T*A*T*A*A*A*G*mC*mU*mA*mC*mU	F11	92.54	84.43	54.83	6.37
[LINC-PINT]283 .1	mU*mG*mA*mU*mG*A*T*G*T*T*G*C*A*G*mG*mA*mG*mG*mC	G10	85.85	65.76	25.50	1.57
[LINC-PINT]2990	mC*mA*mC*mU*mG*T*A*T*T*T*A*T*A*mC*mA*mG*mA*mA	E11	69.47	117.42	58.13	4.86
[LINC-PINT]384	mU*mG*mA*mC*mA*A*A*A*A*T*A*A*T*mA*mA*mC*mA*mG	G11	88.79	53.61	36.68	2.57
[LINC-PINT]412 KL	mG*mU*mU*mC*mA*G*T*C*A*G*A*T*G*C*mU*mG*mG*mG*mA	A7	93.74	77.50	17.46	1.70
[LINC-PINT]501 KL	mU*mG*mU*mU*mU*C*C*G*G*A*G*A*G*mC*mA*mA*mU*mG	B7	57.81	71.47	30.90	2.04
[LINC-PINT]523 KL	mU*mG*mA*mC*mA*T*T*T*G*T*G*C*T*mC*mC*mU*mA*mC	C7	79.34	118.29	33.56	4.34
[LINC-PINT]524 .1	mA*mU*mG*mA*mC*A*T*T*G*T*G*G*C*mU*mC*mC*mU*mA	H10	99.38	107.12	32.53	1.20
[LINC-PINT]587 KL	mA*mG*mC*mC*mG*A*A*C*A*G*A*G*G*A*mG*mC*mG*mU*mC	D7	39.92	57.46	27.70	3.73
[LINC-PINT]727 .1	mG*mU*mC*mC*mG*T*A*C*T*C*A*C*C*mC*mA*mC*mC*mG	B11	86.09	58.15	22.79	3.70
[LINC-PINT]83 .1	mC*mA*mA*mG*mC*C*C*A*G*C*G*T*T*C*mC*mU*mC*mC*mG	F10	77.10	93.79	45.20	4.44
[LINC-PINT]877 KL	mC*mC*mC*mU*mA*A*T*G*T*T*T*C*T*mC*mU*mC*mC*mA	E7	89.09	86.47	58.39	8.39
[LINC-PINT]935 KL	mG*mC*mG*mU*mA*G*T*T*T*C*T*T*T*mC*mU*mC*mC*mC	F7	50.62	52.04	27.23	4.12
[LINC-PINT]937 .1	mU*mG*mG*mC*mG*T*A*G*T*T*T*C*T*mU*mC*mC*mU*mC	D11	70.97	79.89	32.81	4.25
[LINC-PINT]94	mC*mC*mU*mU*mC*T*G*T*G*C*A*G*mC*mC*mC*mA	A10	104.33	96.69	85.57	3.71
[LINC-PINT]98	mC*mA*mU*mC*mC*T*T*G*T*G*C*mC*mA*mA*mG*mC	F8	70.60	65.09	54.34	8.23
[LINC-PINT]99	mC*mC*mA*mU*mC*C*T*T*G*C*T*G*mC*mC*mA*mA*mG	C10	85.43	92.72	52.57	5.67