

Supporting Information

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Steroids Enable Mesenchymal Stromal Cells to Promote CD8⁺ T Cell Proliferation via VEGF-C

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Supplementary Figures:



Figure S1. CD4⁺ T cells are not involved in immune promotion by Dex-treated MSCs. A) Flow cytometric analysis of the expressions of CD11b, CD11c, CD34, Sca-1, CD44 and CD140a in MSCs. B) Oil red O staining and Alizarin red S staining of MSCs cultured in adipogenic and osteogenic differentiation medium for 7 and 21 days, respectively. C) Splenocytes were activated by anti-CD3 and co-cultured with or without MSCs in the presence or absence of Dex at indicated concentrations, and the total cell numbers were counted. D-E) Flow cytometric analysis of the percentages of CD4⁺ T cells and CD8⁺ T cells among splenocytes. F) The proliferation of CD4⁺ T cells. CD4⁺ T cells isolated from the mouse spleen were stimulated with anti-CD3 and anti-CD28 with or without MSCs, in the presence or absence of Dex at indicated concentrations. Cell proliferation was assessed by ³H-

thy midine incorporation. Data are presented as mean \pm SEM. n.s., no significance, *p < 0.05 and **p < 0.01.



Figure S2. Expressions of VEGF family members in MSCs upon Dex treatment. A) Top 9 up-regulated genes in MSCs by Dex in the presence of activated T cells. Four groups were presented, including MSCs (G1), MSCs with Dex treatment (G2), MSCs co-cultured with CD8⁺ T cells (G3) and MSCs co-cultured with CD8⁺ T cells and with Dex treatment (G4). B-E) mRNA expressions of *Vegfc*, *Vegfd*, *Vegfa* and *Vegfb* in MSCs stimulated with activated T cell supernatant, with or without Dex addition. F) Efficiency of short hairpin RNA (shRNA)-mediated *Vegfc* knockdown in MSCs. NC, scramble shRNA control. Data are presented as mean \pm SEM. *p < 0.05 and **p < 0.01.



Figure S3. The influence of VEGF-C on the numbers of immune cells in the liver, blood and spleen of aGvHD mice. A-B) The numbers of lymphocytes (A) and CD8⁺ T cells (B) in the liver of aGvHD mice treated with PBS or VEGF-C. Lethally irradiated C3H × C57 BL/6 F1 mice were adoptively transferred with bone marrow cells (5×10^6) and splenocytes (5×10^7) from C57BL/6 mice to induce aGvHD. On day 7 post aGvHD induction, mice were *i.p.* injected with VEGF-C at 10 µg per mouse daily. For the BMT group, recipient mice were adoptively transferred with bone marrow cells (5×10^6), with or without VEGF-C administration. C-D) The numbers of lymphocytes (C) and CD8⁺ T cells (D) in peripheral blood of aGvHD mice treated with PBS or VEGF-C. E-F) Numbers of splenocytes (E) and CD8⁺ T cells (F) in the spleen of aGvHD mice treated with PBS or VEGF-C. Data are presented as mean ± SEM. *p < 0.05 and ***p < 0.001.



Figure S4. The effects of VEGF-C on CD8⁺T cell apoptosis and IFN- γ expression. A) Flow cytometric analysis of apoptotic cells among CD8⁺T cells activated with anti-CD3 and anti-CD28 for 48 hours, in the presence or absence of VEGF-C (800 ng/ml). B) Flow cytometric analysis of IFN- γ expression in CD8⁺T cells stimulated with anti-CD3 and anti-CD28 in the presence of VEGF-C. C) Flow cytometric analysis of the expressions of VEGFR2 and VEGFR3 on naïve CD8⁺T cells.



Figure S5. Activation and apoptosis in CD8⁺ T cells isolated from VEGFR3^{fl/fl} and VEGFR3^{fl/fl}CD8^{Cre} mice. A) Immunoblotting analysis of total p85, phosphorylated p85/p55, CDK2, CDK4, and CDK6 in CD8⁺ T cells stimulated with PMA and ionomycin. β-actin was used as a loading control. B) Flow cytometric analysis of the expression of VEGFR3 on CD4⁺ T cells co-cultured with MSCs, with or without Dex (10 ng/ml) additon. C) Immunoblotting analysis of Cyclin D1 expression in CD4⁺ T cells and CD8⁺ T cells co-cultured with MSCs, with or without Dex (10 ng/ml) additon. C) Immunoblotting analysis of Cyclin D1 expression in CD4⁺ T cells and CD8⁺ T cells co-cultured with MSCs, with or without Dex (10 ng/ml) addition. D) Immunoblotting analysis of the expressions of p-p85/p-p55, p-Akt and Cyclin D1 in CD4⁺ T cells stimulated with anti-CD3 and anti-CD28 and in the presence or absence of VEGF-C (800 ng/ml). E) Flow cytometric analysis of the expressions of the expressions of CD25 and CD69 on CD8⁺ T cells activated by anti-CD3 and anti-CD28. F-G) Flow cytometric analysis of apoptotic cells in anti-CD3 and anti-CD28 activated CD8⁺ T cells isolated from VEGFR3^{fl/fl} and VEGFR3^{fl/fl}CD8^{Cre} mice.



Figure S6. TNF α and Dex synergistically enhance VEGF-C expression in MSCs. A) The protein levels of VEGF-C in the supernatant of MSCs treated with PBS, LPS (100 ng/ml) or IFN- γ (10 ng/ml). B) The protein level of TNF α in the supernatant of CD8⁺ T cells stimulated with anti-CD3 and anti-CD28. C) *Vegfc* mRNA expressions in MSCs stimulated with LPS (20 ng/ml), IFN- γ (5 ng/ml) or TNF α (5 ng/ml) and treated with Dex at indicated concentrations. D) mRNA expressions of *Vegfc* in MSCs treated with TNF α (5 ng/ml) and Dex at indicated concentrations. E) mRNA expressions of *Vegfd* in MSCs treated with TNF α (10 ng/ml), Dex (10 ng/ml) or their combination. F) mRNA expressions of *Tnfr1* and *Tnfr2* in MSCs. G) Efficiency of siRNA-mediated *Tnfr1* knockdown in MSCs. NC: scramble siRNA control. Data are presented as mean \pm SEM. **p < 0.01.