Supporting Information for:

Peters plus syndrome mutations affect the function and stability of human β 1,3-glucosyltransferase

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Clone	Primer name	Sequence $(5^{\prime} \rightarrow 3^{\prime})$
D349N	AI73 f	AGTCATTGTGAATGATGATACATTAATAAGTATCTCCAGGCTCC
	AI74 r	GTATCATCATTCACAATGACTAACCATGCTGTTTTGTCCTG
G393E	KZ21 f	CACGGGAG <u>A</u> AGGAGGAATGGTCTTCAGCAGAG
	KZ22 r	ATTCCTCCTTCCCGTGATGTAGCTGTAGCC
G394E	KZ23 f	GGGAGGAGAAGGAATGGTCTTCAGCAGAGAAG
	KZ24 r	ACCATTCCT <u>T</u> CTCCTCCCGTGATGTAGCTGT
Q457R	KZ07 f	TTCTCATC <u>GG</u> GTTCCCATATCGTTCCACAAAC
	KZ08 r	ATGGGAAC <u>CC</u> GATGAGAAAGGTAGTCCTTAGG
T179S	KZ41 f	CGAGAATCCT <u>T</u> CAGTTTTTAAGTATCCAGACTTTG
	KZ42 r	CTTAAAAACTG <u>A</u> AGGATTCTCGGAAAAGGCATAATG
V245M	KZ43 f	GTACCAATGACATGGACTTCTACTGTGCTACCACATTC
	KZ44 r	GTAGAAGTCCA <u>T</u> GTCATTGGTACAAAACTCAGGCACTG
R337H	KZ45 f	TTTCTGAATC <u>A</u> TAGCCAGGACAAAACAGCATGG
	KZ46 r	GTCCTGGCTA <u>T</u> GATTCAGAAATCTTTCCAAAATGG
Y366*	KZ17 f	AGCTGTTA <u>A</u> GACTCCGGCGAGCCTGTGTTT
	KZ18 r	GCCGGAGTC <u>T</u> TAACAGCTAAGCAAGTGCTG
R412*	KZ19 f	AGTAAATGTTGATGCTACAGCAATGATGCTC
	KZ20 r	CTGTAGCATC <u>A</u> ACATTTACTGGCGAGAA
E132A	KZ49 f	CTTCTGTG <u>CC</u> GAAGAGACAAGAATACAGATTCCAAAACTCTTGG
	KZ50 r	CTTGTCTCTTC <u>GG</u> CACAGAAGAAAATCCAAGATGAATTTCTGCTA
D421A	KZ55 f	TCCCGATG <u>CC</u> ATGGTCCTGGGAATGTGCTTTAG
	KZ56 r	AGGACCAT <u>GG</u> CATCGGGAGCATCATTGCTGTAGC
CT1	AI210 f	GAGTTTTGTACCAATGACGTGGACTTCTAC
	AI214 r	AGCCAAACCAAAAGCCAGGGAG
	(pcDNA3.1)	GGCCGCGTCACCAGTGG
	ÄI213 r	
	(pSecTag2C)	
CT2	AI211 f	ACATTCCATTCTTTCTACCGCTTTGTAG
	AI214 r	AGCCAAACCAAAAGCCAGGGAG
	(pcDNA3.1)	GGCCGCGTCACCAGTGG
	AI213 r	
	(pSecTag2C)	

Table S1. Site-directed mutagenesis primers for human B3GLCT construct.

CT3	AI212 f	AAGCCAGTGAAGAAGAAGGATATTTTTGTTG
	AI214 r	AGCCAAACCAAAAGCCAGGGAG
	(pcDNA3.1)	GGCCGCGTCACCAGTGG
	AI213 r	
	(pSecTag2C)	
NT1	AI337 f	CTGAGTTTTG <u>A</u> ACCAATGACGTGGACTTCTACTGTG
	AI338 r	CGTCATTGGT <u>T</u> CAAAACTCAGGCACTGGGGTC
NT2	AI339 f	TGTGCTACC <u>TG</u> ATTCCATTCTTTCTACCGCTTTGTAGAAAGCC
	AI340 r	GAATGGAAT <u>CA</u> GGTAGCACAGTAGAAGTCCACGTCATTGG
NT3	AI341 f	GCTTTGTAGA <u>T</u> AGCCAGTGAAGAAGAAGGATATTTTTGTTGC
	AI342 r	TTCACTGGCTATCTACAAAGCGGTAGAAAAGAATGGAATG

Supporting Information Figure Legends:

Supporting information

Figure S1. Alignment of B3GLCT sequences from multiple species. Multiple sequence alignment of B3GLCT from human (UNIPROT Q6Y288), mouse (UNIRPOT Q8BHT6), zebrafish (UNIRPOTA0A068F9P7), fruitfly (UNIPROT X2JDC2), and starfish (NCBI REFSEQ XP_038056415.1) were aligned with the MAFFT software/algorithm. Red background highlights regions of 100% sequence identity within the shown sequences; red letters indicate regions of 100% sequence similarity; blue rectangles enclose sets of conserved residues. Annotated domains are highlighted under amino acid sequences.

Figure S2. Time-dependent *in vitro* enzyme assay to determine the linear phase of the transfer reaction catalyzed by B3GLCT. 0.5 μ g purified B3GLCT, 50 μ M UDP-Glc and three different concentrations of pNP-Fuc were analyzed. Reactions were stopped by freezing on dry ice. All reactions were incubated at 37 °C.

Figure S3. Neither the B3GLCT C-GT nor N-GT-like domains expressed alone. (A) Domain map of human B3GLCT with amino acids cutoff for N-GT-like domain and C-GT domain constructs. Each N-GT-like domain (NT1, NT2, NT3) or C-GT domain construct (CT1, CT2, CT3) contained amino acids ending or beginning as indicated. SP, signal peptide; N-GT-like, N-terminal GT-like domain; C-GT, C-terminal GT domain; REEL, C-terminus containing KDEL-like motif. (B) Plasmids encoding ADAMTS20 TSR2-8-Myc-His₆ and GFP were co-transfected into WT or *B3GLCT^{-/-}* HEK293T cells. Rescue experiments were performed by co-transfection

with plasmids encoding full-length WT B3GLCT (wtB3), or the CT1, CT2 or CT3 constructs, all with the REEL ER-retention motif. Media and cell lysates were analyzed by Western blot probed with anti-Myc (red, 50 kDa) to detect ADAMTS20 TSR2-8 (TSR2-8), and anti-B3GLCT for B3GLCT WT or mutants (green, 50 kDa, mutants expected <50kDa). Anti-GFP for transfection and loading control (red, 25 kDa). (C) Plasmids encoding WT, NT1, NT2, and NT3, all generated with the N-His6-B3GLCT construct, were co-transfected into WT HEK293 cells with GFP plasmid. Media and cell lysates were analyzed by Western blot probed with anti-His₆-tag (red, WT 50 kDa, mutants <50 kDa) and anti-GFP for transfection (green, 25 kDa).

Figure S4. Coomassie blue staining of purified WT and PTRPLS/PTRPLS-like mutant enzymes. Bovine serum albumin (BSA) standards and purified B3GLCT WT, PTRPLS (black) and PTRPLS-like (blue) mutants were separated by 10% SDS-PAGE and stained with Coomassie blue for imaging. For purified enzymes, 5 μ L of eluate was loaded in each lane.

Figure S5. B3GLCT with PTRPLS mutations did not rescue secretion of ADAMTS20 TSR2-8 while PTRPLS-like mutations did. Plasmids encoding ADAMTS20 TSR2-8-Myc-His₆ (TSR2-8) and GFP were co-transfected into wild type (WT) and *B3GLCT* knockout (*B3GLCT^{-/-}*) HEK293T cells. PTRPLS mutations are colored in black and PTRPLS-like mutations in blue. Rescue experiments were performed by co-transfection with a plasmid encoding full-length B3GLCT WT, G393E, and G394E (**A**), Q457R and T179S (**B**), and V245M and R337H (**C**) (B3GLCT) with serial dilutions. Serial dilutions of B3GLCT plasmids were performed starting with 0.24 μ g of plasmid diluted 5-, 10-, and 20-fold. Media and cell lysates were analyzed by Western blot probed with anti-Myc (red) to detect ADAMTS20 TSR2-8, antiB3GLCT to detect endogenous or transfected B3GLCT (green, 50 kDa), and anti-GFP for transfection and loading control (green, 25 kDa).

Figure S6. B3GLCT mutants G393E and Q457R destabilized the enzyme. Melting curve (**A**) and first derivative showing the melting peak (**B**) of WT and mutants were analyzed using the thermo shift protein stability assays described in Experimental Procedures. 10% glycerol in TBS was used as negative control to monitor the background. Arrows in (**B**) point to reduced melting temperature of G393E (blue) and Q457R (orange).

Figure S7. PTRPLS nonsense mutations at Y366 and R412 result in deletion of important catalytic residues. Ribbon presentation of structures for B3GLCT C-GT Y366* (A) and R412*
(B). Light blue ribbon represented the backbone of B3GLCT C-GT. Light blue, residues without mutations; salmon, residues at which had PTRPLS mutations; yellow, residues at which had PTRPLS-like mutations (blue letters).

Figure S8. Internal core structures of B3GLCT C-GT aligned with the cores of MFNG and B3GNT2. Protein backbone alignment between B3GLCT C-GT model (tan), MFNG (PDB 2J0B, blue) and B3GNT2 (PDB 7JHN, purple). Structure alignment was visualized with Chimera.

Figure S9. Sequence alignment revealed that B3GLCT C-GT has conserved motifs of GT31 family enzymes. Amino acid sequence of human B3GLCT (UNIPROT Q6Y288) was aligned to

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the sequences of human B3GNT2 (UNIPROT Q9NY97) and mouse MFNG (UNIPROT 009008). DxD motif, dark blue; Glycine rich loop, magenta; xED motif, purple; C-His, orange.

Figure S10. B3GLCT is active in the absence of divalent cations. *In vitro* enzyme assays of B3GLCT with UDP-Glc and Fuc-*O*-TSR3 were supplied with various divalent metal ions at three different concentrations. 10x stock solutions of MnCl₂, MgCl₂, CaCl₂ and EDTA were supplemented to each reaction to make final in-reaction concentrations as described. Statistical analysis was performed with One-way ANOVA in Prism 7. Error bars, standard deviation, n=3; *p<0.05; **p<0.01.

Figure S11. B3GLCT N-GT-like domain adopts a GT-A fold. (A) Protein backbone alignment between B3GLCT N-GT-like domain (tan), B3GLCT C-GT model (green), MFNG (PDB 2J0B, blue) and B3GNT2 (PDB 7JHN, purple). Structure alignment was visualized with Chimera. **(B)** The DxD motif, G-loop, catalytic aspartic acid, and C-His for B3GLCT N-GT-like domain model (tan), C-GT domain model (green) are aligned with MFNG (PDB 2J0B, blue), B3GNT2 (PDB 7JHN, purple). DxD motifs, G-loops, and catalytic aspartic acid of all four domains also overlayed, whereas C-His of the N-GT domain is missing.

Figure S1 HUMAN MOUSE	10 20 30 40 MRPPACWWLLAPPALLALLTCSLAFGLASEDTKKEVK.QSQDLEKSGI MRPPALLALFSCSAAFALMSEEIKEKVT.PSQDLRQSSL
ZEBRAFISH FRUITFLY STARFISH	MLSIKCQNGYSVTQYFFLWSLVAFITAKGDEVENPAKVHGTDKPSSG. MPTRICVLFGLLCLASLCCGSNKPG. MAASIVDPYFPRDLTVAEPNL
HUMAN MOUSE	50, 60, 70, 80, 90, SRKNDIDLKGIVFVIQSOSNSFHAKRAEQLKKSILKOAADITOE.LPS.VLLLH PGRHDIDLKEIVFVIOSOSNSFHAKRAEOLKKNTIKOAANITOD.LPR.VLLLH
ZEBRAFISH FRUITFLY STARFISH	SQLDLREVVFVIQSQRNSFHVRQAEKLRKDLLLQTQTLQESPPVILLLE EVLLVIACPPHPQQARSDCLALSHNVLEQQRALELAGIFPEDFVLKVH EGHHATTGEDLVFIIRSQQASPHDQWAKQVKADIEQQYRIINLGNPQVFLLH
HUMAN	00 110 120 130 140 150 OlakoegawiilpilpisytYsrnsswiffceee <mark>tr</mark> i.Oirk <mark>u</mark> let <mark>u</mark> rrydeskew
MOUSE ZEBRAFISH FRUITFLY STARFISH	QLAKQEGAWTILPLPHFSVITYSKNSAWIFFCEEETRU.QIPTLDTTRRYDPSKEW TISDNEGDWSILPLPRISSQFGKNSSWIMFLEEDTRV.KLQKTHEVIKKFDRRKEW VMHELFNSWTMLDALPHLRAQARVLGARTEWIIWCQHNTRVSSIRGLEQTRRQNPRELA QQYEFSGVWTILPVLPKLNEDFGQ.AKWLFFCEEQTRV.SVHGLQVTKYNPQQEW
	N-GT-like Domain
HUMAN	160 170 180 190 200 210 FlGkalhDeeatiihhyafsenPtvekyPdfaAgwalsipLynkTtkri, ksesiks
MOUSE ZEBRAFISH FRUITFLY STARFISH	FLGKALYDEESTIIHHYAFSENPTVFKYPDFAAGWALSIPLVNKLAKRLKSEALKS FLGKPLHDEESTIIHHYAFSENPTAFEYPDFSAGWALSIPLINRLASKIEEEPLKS FYGHALYDAEATIIHHFSNYKDPORFPYPMLSAGVVFTGALLRRLADLVAPSGONITVHS FLGRALODRAASVIHHYRFHDDPSKFSYPDFEAGWLISTGUIRGUADRWETEEHRM
LITERAN	
HOMAN MOUSE ZEBRAFISH FRUITFLY	DFTIDLKHEIALYIWDKGGGPALTPVPEFGTEDVDPF. DFTIDLKHEIALYIWDKGGGPALTPVPEFGTEDVDPR. DFTIDLKHEVALYIWEDGKGPRITGVPELCTL. AEHNKRAQS DFSIDASHELARFIFDNVSPDPHISTPISGGILLKSASYIGSTPTSVPNRKLPC.
STARFISH	
	25 <u>0</u> 26 <u>0</u> 27 <u>0</u> 280
HUMAN MOUSE ZEBRAFISH FRUITFLY	CATTFHSFLPLCRKPVKKKDIFVAVKTCKKFHGDRIPIVK CVTTFHSFLPLCGVPVKKEDIFVAVKTCKKFHADRIPIVK CATTVSSHSPLCGEPVKIENIFVAVKTCKKFHSDRVPVVK LLHAQPEEPLTLGQRRNGCHTTGSHYFAIKTCAKFHKERIPIIE
STARFISH	KALNKAIRDGPQEEDG <mark>CVSA</mark> HPKGL <mark>P</mark> TCAEAVLKED <mark>I</mark> LF <mark>AVKTCKKFH</mark> KD RVP IVQ
:	290 300 310 320 330 340
HUMAN MOUSE ZEBRAFISH FRUITFLY	QTWESQASLIEYYSDYTENSIPTVDLGIPNTDRGHCGKTFAILERFLNRSQDKTAWLV KTWAAQASLIEYYSDYAETAIPTVDLGIPNTDRGHCGKTFAILERFLNHSUNKISWLV KTWGKQASLLEYYSDYADPSIPTINLGVPNTERGHCGKTFAILRFLSSHVPRTDWLL RTWAADARNRRYYSDVADVGIPAIGTGIPVOTGHCAKTMAILOLSLKDIGKOLDIRWLM
STARFISH	QTWGKHISHIVFVSDVQDDTIPTIASGVPNTERGHCGKLFAIFEMFNSK.PEŸLKYSWLV
	350 360 370 380 390
HUMAN MOUSE ZEBRAFISH EBULTELY	IVDDDTLISISRLOHLLSCYDSGEPVFLGERYGYG.LGTG.GYSYITGG IVDDDTLISISRLRHLLSCYDSSDPVFLGERYGYG.LGTG.GYSYVTGG IVDDDTLISIPRLOALLSCYESSEPLCLGERYGYG.LGGG.GYSYITGG
STARFISH	VADDDTILSVARLRALLSCYNAKKLVFLGERYGYGHLKLGWGYDYLTGG
	400 410 420 430 440 450
HUMAN MOUSE ZEBRAFISH	GGMVFSREAVRRLLASKCRCYSNDAPDDMVLGMCFSGLGIPVTHSPLFHQARPVDYPKDY GGMVFSREAIRRLLVSSCRCYSNDAPDDMVLGMCFSGLGVPVTHSPLFHQARPVDYPKDY GGMLFSREAVVQLLSSCCNCYSNDAPDDMVLGMCLNSLRVPVTHSPLFHQARPEDYARDF
FROITFLY STARFISH	GGNIFSRAGIQQLLATGCKCNKDEDPDDMILGYCLQALGVEATHVAGMHQAREODYAGEL GGMIFSRAGIQQLLATGCKCNKDEDPDDMIFGMCTKRHDMPITHSPLFHQARESDYAVGY
HUMAN MOUSE ZEBRAFISH	460, 470, 480, 490, LSHQVPISFHKHWNIDPVKVYFTWLAPSDEDKARQETQKGFREEL LAHQIPVSFHKHWHIDPVKVYLTWLAPSEEDQATQETQKDPREEL LSHQTPISFHKHWNIDPIAVFNKWI
FRUITFLY STARFISH	LQLHAPLTFHKFWNTDPEHTYRRWLGGSMVNRSAPLAAHKEQPAAGPLHMTMGRHSLAAG LQHQTPVSFHKHLNADPLAVYRDWFKHADEVDSRTHGAQGEREEL









В



TSR2-8

GFP

B3GLCT TSR2-8

GFP

FigureS6 A

В











B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	MS-VGRRRIKLLGILM MRPPACWWLLAPPALLALLTCSLAFGLASEDTKKEVKQSQDLEKSGISRKNDIDLKGIVF 	15 60 0
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	MANVFIYFIMEVSKSSSQEKNGKGEVIIPKEKFWKIS VIQSQSNSFHAKRAEQLKKSILKQAADLTQELPSVLLLHQLAKQEGAWTILPLLPHFSVT	52 120 0
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	TPPEAYWNREQEKLNRQYNPILSMLTNQTGEAGRLSNISHLNYCE YSRNSSWIFFCEEETRIQIPKLLETLRRYDPSKEWFLGKALHDEEATIIHHYAFSENPTV	97 180 0
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	NLPDRFKD- FKYPDFAAGWALSIPLVNKLTKRLKSESLKSDFTIDLKHEIALYIWDKGGGPPLTPVPEF MHCRLFRGMAGALFTLL	118 240 17
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	FLLYLRCRNYSLLIDQPDKCAKKPFLLLAIKSL-TPHFARRQAIRES CTNDVDFYCATTFHSFLPLCRKPVKKKDIFVAVKTCKKFHGDRIPIVKQT CVGLLSLRYHSSLSQRMIQGALRLNQRNPGPLELQLGDIFIAVKTTWAFHRSRLDLLLDT ::: * * : :::*:*: * * : ::	164 290 77
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	WGQESNAGNQTVVRVFLLGQTPPEDNHPDLSDMLKFESEKHQDILMWNYRDTFFNLS WESQASLIEYYSDYTENSIPTVDLGIPNTDRGHCGKTFA WVSRIRQQTFIFTDSPDERLQERLGPHLVVTNCSAEHSHPALSCK * : : . * ::::	221 329 122
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	LKEVLFLRWVSTSCPDTEFVFKG DDD VFVNTHHILNYLNSLSKTKAKDLFIGDVIHNAGP ILERFLNRSQDKTAWLVIV DDD TLISISRLQHLLSCYDSGEPVFLGERYGY MAAEFDAFLVSGLRWFCHV DDD NYVNPKALLQLLKTFPQDRDVYVGKPSLNRPI : .: : *** :. : : * : ::*.	281 380 176
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	HRDKKLKYYIPEVVYSGLYPPYAGGGGGFLYSGHLALRLYHITDQVHLYPIDDV GLGTGGYSYITGGGGGMVFSREAVRRLLASKCRC-YSNDAPDDM HASELQSKNRTKLVRFWFATGGAGFCINRQLALKMVPWASGSHFVDTSALIRLPDDC : :**.*: :: **	334 422 233
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	YTGMCLQ-KLGLVPEKHKGFRTFDIEEKNKNNICSYVDLML VLGMCF-SGLGIPVTHSPLFHQARPVDYPKDYLSHQVPISFHKHWNIDPVKVYFT-WL TVGYIIECKLGGRLQPSPLFHSHLETLQLLGAAQLPEQVTLSYGVFEGKLNVIKLP * : ** *:: *: .	374 478 289
B3GNT2_HUMAN B3GLCT_HUMAN MFNG_MOUSE	VHSRKPQEMIDIWSQLQSAHLKC 397 APSDEDKARQETQKGFREEL 498 GPFSHEEDPSRFRSLHCLLYPDTPWCPLLAAP 321 :: .:	



