

## Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

### Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size ( $n$ ) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided  
*Only common tests should be described solely by name; describe more complex techniques in the Methods section.*
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g.  $F$ ,  $t$ ,  $r$ ) with confidence intervals, effect sizes, degrees of freedom and  $P$  value noted  
*Give  $P$  values as exact values whenever suitable.*
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's  $d$ , Pearson's  $r$ ), indicating how they were calculated

*Our web collection on [statistics for biologists](#) contains articles on many of the points above.*

### Software and code

Policy information about [availability of computer code](#)

Data collection

Data analysis

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research [guidelines for submitting code & software](#) for further information.

### Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

## Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences  Behavioural & social sciences  Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

## Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

|                 |  |
|-----------------|--|
| Sample size     | Sample size for the discovery dataset was determined by the available data, with statistical analysis plan pre-planned. No pre-determined power calculations were performed. Large publicly available datasets were utilised for validation. |
| Data exclusions | No data were excluded from analyses.   |
| Replication     | Data was independently verified using independent publicly available datasets (as referenced in the manuscript) and reproducible.  |
| Randomization   | Allocation was not random - multivariate analysis was performed as reported.   |
| Blinding        | Authors were blinded to pnsTING status of samples for scoring of immune markers and analysis of gene expression data.  |

## Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

### Materials & experimental systems

| n/a                                 | Involvement in the study                                  |
|-------------------------------------|---|
| <input type="checkbox"/>            | <input checked="" type="checkbox"/> Antibodies            |
| <input type="checkbox"/>            | <input checked="" type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology    |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Animals and other organisms      |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants      |
| <input type="checkbox"/>            | <input checked="" type="checkbox"/> Clinical data         |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern     |

### Methods

| n/a                                 | Involvement in the study                        |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq               |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry         |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

## Antibodies

### Antibodies used

STING - 19851-1-AP ProteinTech  
 ER - NCL-L-ER-6F11, Leica  
 HER2 - NCL-L-CB11, Leica  
 Ki-67 - 790-4286, Roche  
 CK - M3515, Dako  
 CD3 - 790-4341, Roche  
 CD4 - 790-4423, Roche  
 CD8 - M7103, Dako  
 CD20 - M0755, Dako  
 CD68, PA0273, Leica  
 CD45RO - NCL-L-UCLH1, Leica  
 CD163 - PA0090, Leica  
 FoxP3 - LS-C210349, LSBio  
 ICOS - 89601, Cell Signalling  
 mTOR - 2976, Cell Signalling  
 IDO1 - 86630, Cell Signaling  
 PL-L1 - 790-4905, Roche  
 PD-L1, M4420, Spring Bioscience  
 TIM3 - 45208, Cell Signaling

See Supplementary table 1 for full list of antibodies and IHC protocols used in this study.

### Validation

Antibodies have been validated using control tissues (tonsil, spleen, whole face tumour sections) and optimised in this manner.

Validation

Antibodies are either diagnostic grade or validated by the manufacturer, in addition to quality control steps performed in house under the CRUK Accelerator programme.

## Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)

MDA-MB-436 EV - originally obtained from ATCC with sh-empty vector described in Parkes et al, JNCI, 2017.

Authentication

Authenticated as BRCA mutant previously (Parkes et al, JNCI, 2017)

Mycoplasma contamination

All cell lines tested negative for mycoplasma contamination and are tested on a regular basis.

Commonly misidentified lines  
(See [ICLAC](#) register)

Not relevant

## Clinical data

Policy information about [clinical studies](#)

All manuscripts should comply with the ICMJE [guidelines for publication of clinical research](#) and a completed [CONSORT checklist](#) must be included with all submissions.

Clinical trial registration

*Provide the trial registration number from ClinicalTrials.gov or an equivalent agency.*

Study protocol

*Note where the full trial protocol can be accessed OR if not available, explain why.*

Data collection

*Describe the settings and locales of data collection, noting the time periods of recruitment and data collection.*

Outcomes

*Describe how you pre-defined primary and secondary outcome measures and how you assessed these measures.*