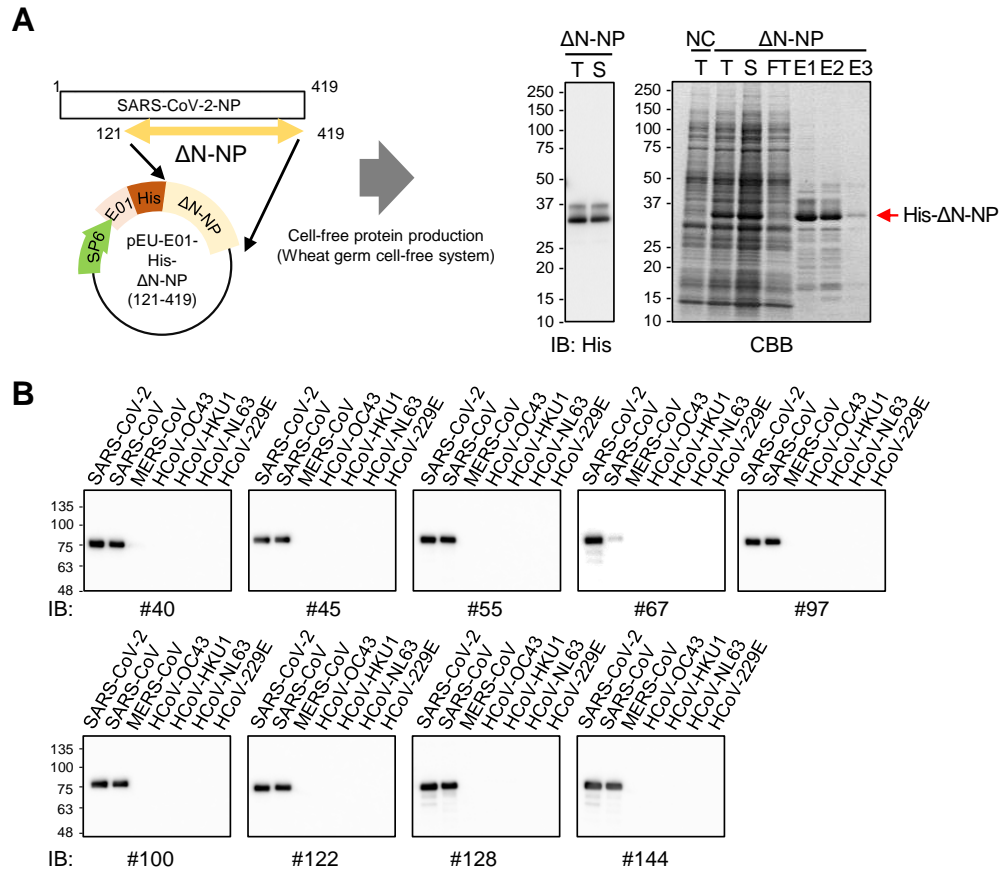


**Supplemental information**

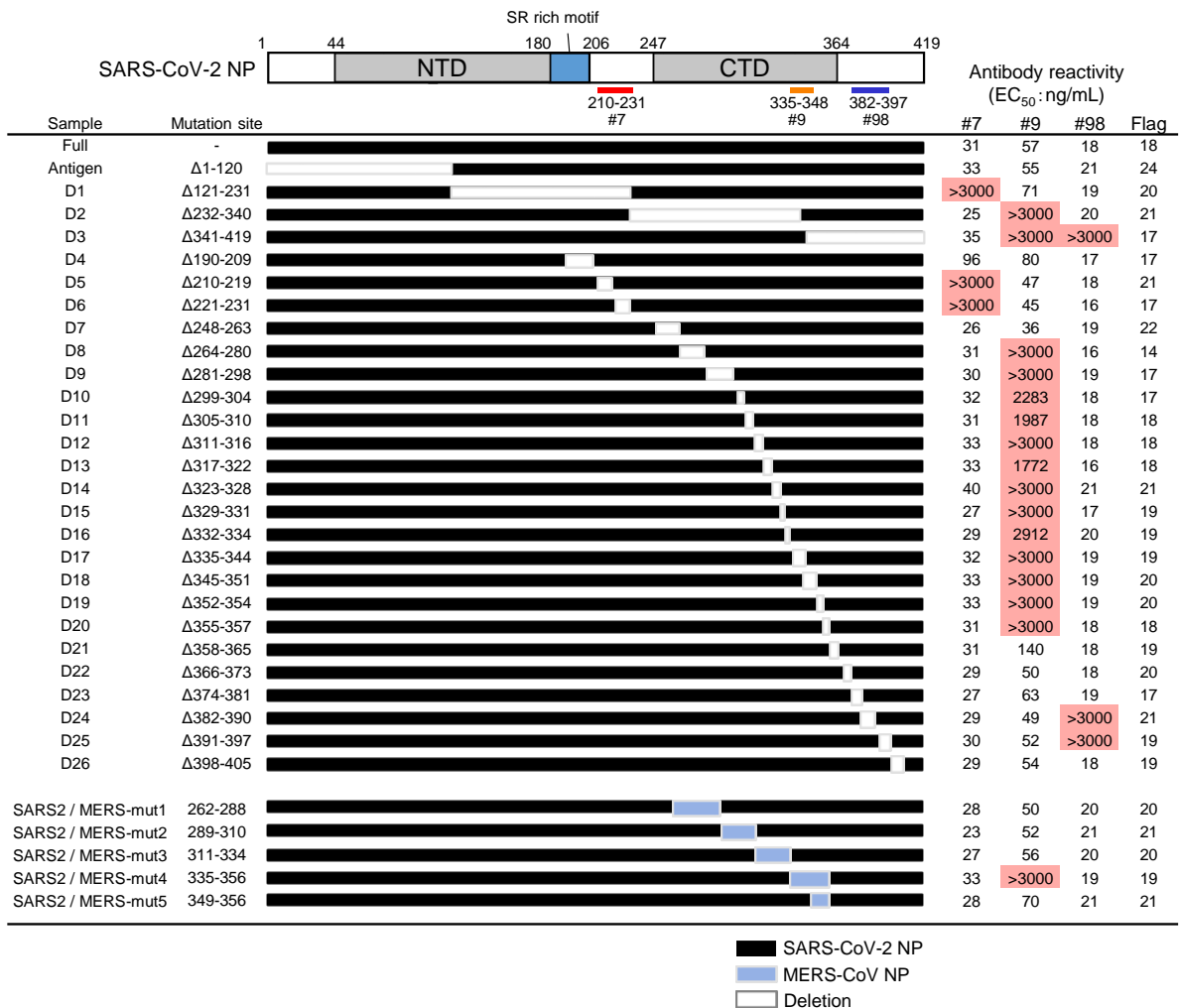
**Highly specific monoclonal antibodies  
and epitope identification against SARS-CoV-2  
nucleocapsid protein for antigen detection tests**

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**Supplementary Figure 1. (Related to Figure 1) Antigen protein production and specificity screening of developed mAbs.**

(A) Schematic representation of antigen protein production. Recombinant Histidine (His)-tagged N-terminally truncated SARS-CoV-2-NP (121-419:  $\Delta$ N-NP) was produced in a wheat germ cell-free system, and then purified using nickel-chelated Sepharose beads. Each protein fraction was analyzed by SDS-PAGE and visualized by CBB staining. Red arrow indicate the target protein. NC, Negative Control; T, Total fraction; S, Supernatant; P, Precipitate; FT, Flow-Through; E1–3, Elution fractions 1-3. (B) Flag-GST-tagged NPs derived from several human coronaviruses were produced in the wheat germ cell-free system. Reactivity of generated mAbs was validated by immunoblot analysis using the indicated antibodies.



### Supplementary Figure 2. (Related to Figure 1) Epitope mapping of mAbs.

Schematic diagram of SARS-CoV-2-NP domain architecture, mutants, and region of determined epitope of each monoclonal antibody. For epitope mapping, 26 deletion mutants (D1-D26) and 5 substitution mutants (SARS2/MERS-mut1-5) were produced as FLAG-GST-tagged proteins in the wheat germ cell-free system. NTD, N-terminal domain; LKR, flexible linker region; CTD, C-terminal domain. Reactivity of each mAb to deletion mutants was evaluated by indirect-ELISA. EC<sub>50</sub> was calculated using six different antibody concentrations, and samples with an over 5-fold increase in EC<sub>50</sub> compared to full-length NP were defined as negative (Red).

**A**

#7 : 210-231 aa

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SARS-CoV-2 210 M A G N G G D A A - - - - - L A L L L L D R L N Q L E 231
SARS-CoV 211 M A S G G G E T A - - - - - L A L L L L D R L N Q L E 232
MERS-CoV 206 I G A V G G D - - - - - L L Y L D L L N R L Q 223
HCoV-OC43 216 - A N S G N R T P - - - - - T S G V T P D M A D Q I A 237
HCoV-HKU1 215 - S N S N F R H S - - - - - D S I V K P D M A D E I A 236
HCoV-NL63 193 L K N L G F D - - - - - N Q S K S P S 206
HCoV-229E 190 L K S L G F D K P Q E K D K K S A K T G T P K P S R N Q S P A 220
  
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#9 : 335-348 aa

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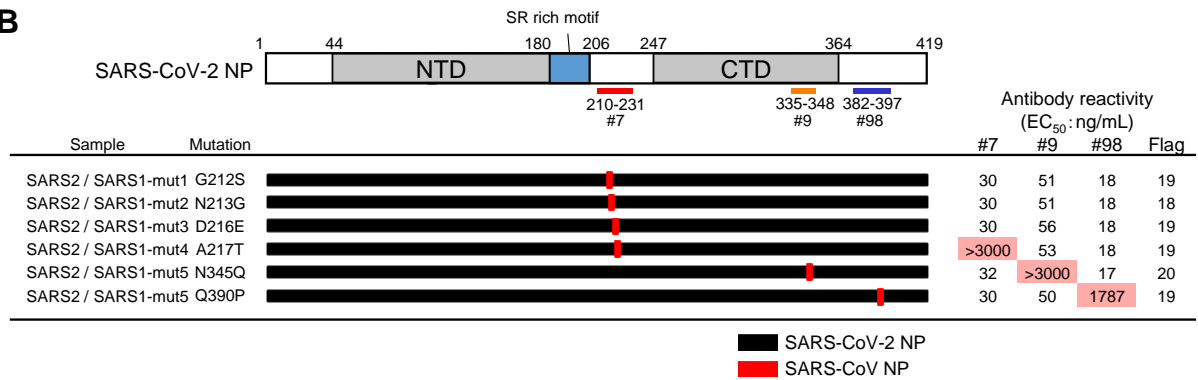
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SARS-CoV 336 G A I K L D D K D P Q F K D 349
MERS-CoV 333 G A I K L D P K N P N Y N K 346
HCoV-OC43 355 G A I R F D S T L S G F E T 368
HCoV-HKU1 350 G S I R F D S T L P G F E T 363
HCoV-NL63 308 Y K M L V A K D N K N L P K 321
HCoV-229E 323 T R V T V P K D H P H L G K 336
  
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#98 : 382-397 aa

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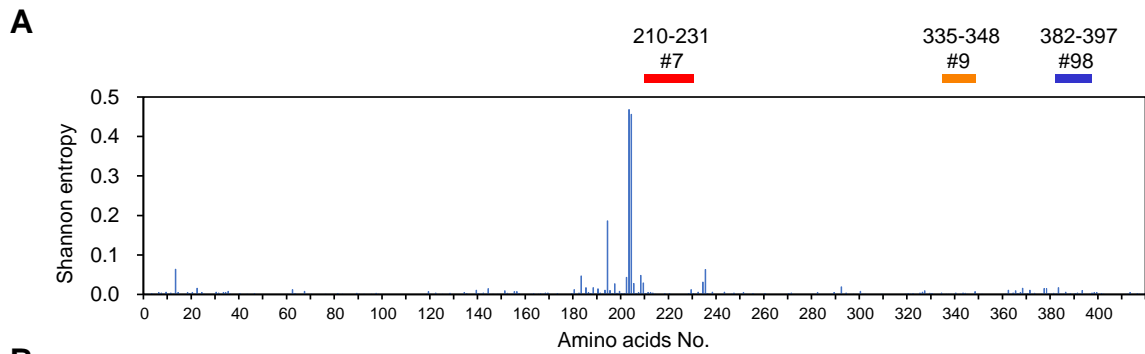
SARS-CoV-2 382 L P Q R Q K K Q - - - - Q T V T L L P A 397
SARS-CoV 383 L P Q R Q K K Q - - - - P T V T L L P A 398
MERS-CoV 382 P P K E Q R V Q - - - - G S I T - - - - 393
HCoV-OC43 412 A V P K S R V Q Q N K S R E L T - - - - 427
HCoV-HKU1 410 S A G T Q H I S - - - - N D F T - - - - 421
HCoV-NL63 354 S I P E S - - - - - K P L A - - - - 362
HCoV-229E 367 S P A T A - - - - - E P V R - - - - 375
  
```

**B**



**Supplementary Figure 3. (Related to Figure 1) Critical residues for desecting SARS-CoV and SARS-CoV-2 in epitope of developed mAbs.**

(A) Multiple sequence alignments of NP of human coronaviruses in the epitope region using MUSCLE software. Gray shaded positions represent conserved residues among the sequences. (B) Determination of critical residues for specific detection of not SARS-CoV but SARS-CoV-2. Five point substitution mutants (SARS2/SARS1-mut1-5) were produced as FLAG-GST-tagged proteins in the wheat germ cell-free system. NTD, N-terminal domain; LKR, flexible linker region; CTD, C-terminal domain. Reactivity of each mAb to deletion mutants was evaluated by indirect-ELISA. EC<sub>50</sub> was calculated using six different antibody concentrations. The samples with an over 5-fold increase in EC<sub>50</sub> compared to full-length NP were defined as negative (Red).



**B**

**#7 : 210-231 aa**

PANGO lineage of SARS-CoV-2	Amino acid sequence of epitope
A (Prototype)	210 M A G N G G D A A L A L L L L D R L N Q L E 231
B.1.1.7 (501Y.V1)	210 M A G N G G D A A L A L L L L D R L N Q L E 231
B.1.351 (501Y.V2)	210 M A G N G G D A A L A L L L L D R L N Q L E 231
B.1.427	210 M A G N G G D A A L A L L L L D R L N Q L E 231
B.1.429	210 M A G N G G D A A L A L L L L D R L N Q L E 231
B.1.525	210 M A G N G G D A A L A L L L L D R L N Q L E 231
B.1.526	210 M A G N G G D A A L A L L L L D R L N Q L E 231
P.1 (501Y.V3)	210 M A G N G G D A A L A L L L L D R L N Q L E 231
P.2	210 M A G N G G D A A L A L L L L D R L N Q L E 231

**#9 : 335-348 aa**

PANGO lineage of SARS-CoV-2	Amino acid sequence of epitope
A (Prototype)	335 G A I K L D D K D P N F K D 348
B.1.1.7 (501Y.V1)	335 G A I K L D D K D P N F K D 348
B.1.351 (501Y.V2)	335 G A I K L D D K D P N F K D 348
B.1.427	335 G A I K L D D K D P N F K D 348
B.1.429	335 G A I K L D D K D P N F K D 348
B.1.525	335 G A I K L D D K D P N F K D 348
B.1.526	335 G A I K L D D K D P N F K D 348
P.1 (501Y.V3)	335 G A I K L D D K D P N F K D 348
P.2	335 G A I K L D D K D P N F K D 348

**#98 : 382-397 aa**

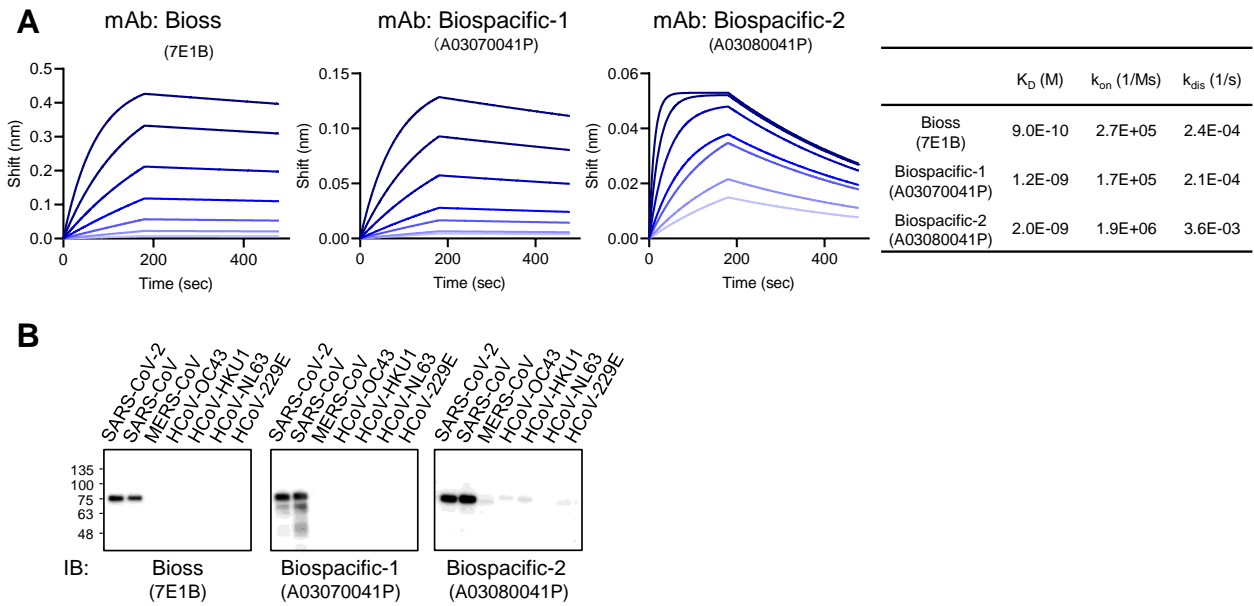
PANGO lineage of SARS-CoV-2	Amino acid sequence of epitope
A (Prototype)	382 L P Q R Q K K Q Q T V T L L P A 397
B.1.1.7 (501Y.V1)	382 L P Q R Q K K Q Q T V T L L P A 397
B.1.351 (501Y.V2)	382 L P Q R Q K K Q Q T V T L L P A 397
B.1.427	382 L P Q R Q K K Q Q T V T L L P A 397
B.1.429	382 L P Q R Q K K Q Q T V T L L P A 397
B.1.525	382 L P Q R Q K K Q Q T V T L L P A 397
B.1.526	382 L P Q R Q K K Q Q T V T L L P A 397
P.1 (501Y.V3)	382 L P Q R Q K K Q Q T V T L L P A 397
P.2	382 L P Q R Q K K Q Q T V T L L P A 397

**Supplementary Figure 4. (Related to Figure 1) Shannon entropy analysis of nucleocapsid proteins and comparison of epitope sequences between prototype and current variant of SARS-CoV-2.**

(A) Shannon entropy, as a quantitative measure of variation, was calculated for each amino-acid residue of SARS-CoV-2 NP derived from 8,127 strains. Binding sites of each mAb are indicated in different colors. (B) Amino acid sequence in the antigenic epitope of reference strain against each of the currently reported epidemic variant of SARS-CoV-2<sup>11</sup>. Gray shaded positions represent conserved residues among the sequences.



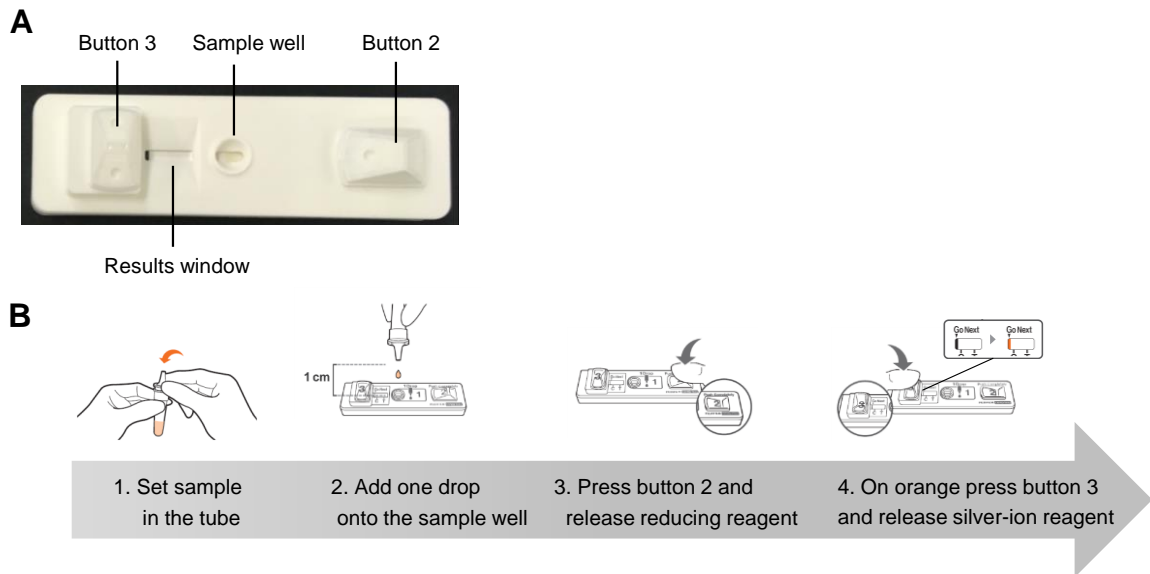
positions represent conserved residues among the sequences. (B) Reactivity of mAbs to recombinant NP from strains possessing amino-acid substitution in the epitope regions. Forty point substitution mutants identified in (A) were produced as FLAG-GST-tagged proteins in the wheat germ cell-free system. Reactivity of each mAb to mutants was evaluated by indirect-ELISA.  $EC_{50}$  was calculated using six different antibody concentrations. The samples with an over 5-fold increase in  $EC_{50}$  compared to full-length NP were defined as negative (Red).



**Supplementary Figure 6. (Related to Figure 2) Comparison of commercially available existing mAbs against SARS-CoV-2 NP.**

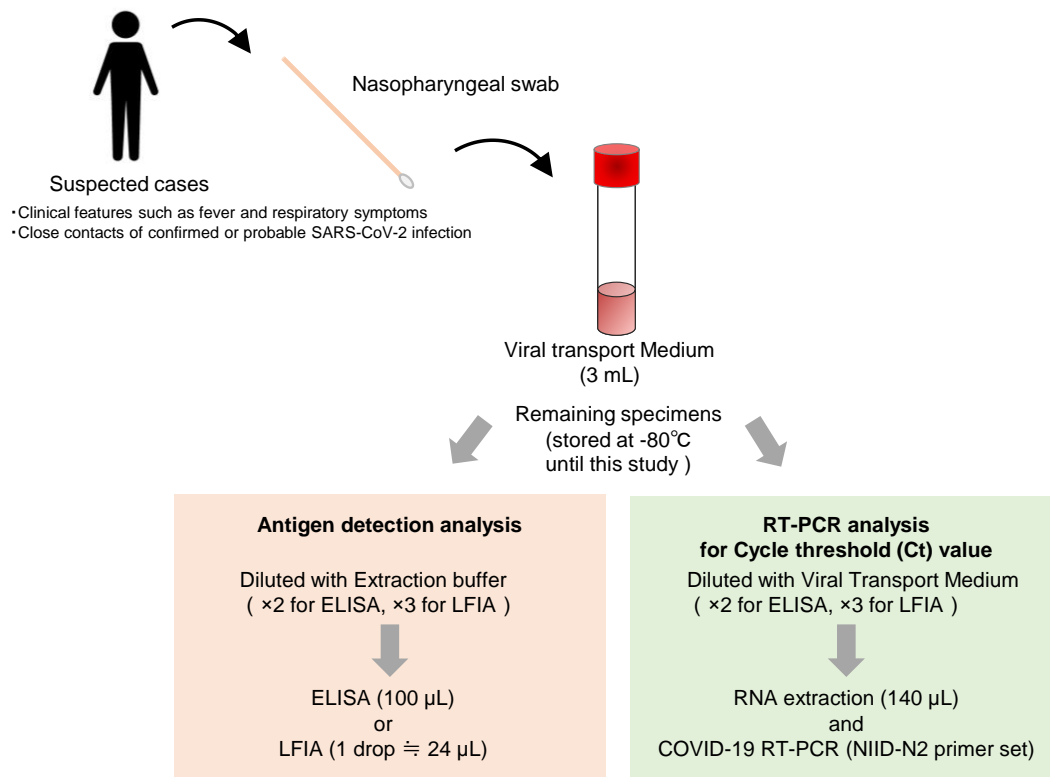
(A) Affinity measurement of commercially available monoclonal antibodies on the octetRED96 instrument. Association and dissociation of each mAbs to full-length NP at various concentrations (50, 25, 12.5, 6.25, 3.13, 1.56 and 0.78 nM) was evaluated using AMC sensor. (B) Specificity of existing mAbs were examined by immunoblot analysis using the indicated recombinant proteins from human coronaviruses.





**Supplementary Figure 7. (Related to Figure 4) Overview and procedure of LFIA.**

(A) Image of LFIA Cartridge. (B) Samples were diluted with an extraction buffer (Tris buffer containing 0.1% non-ionic surfactant) in the tube and one drop of the sample was added onto the sample well of LFIA cartridge. Following this, button 2 was immediately pressed to release a reducing reagent for silver amplification. After the color indicator mark turned orange (about 10 min), button 3 was pressed to release a silver-ion reagent to activate the silver amplification reaction.



**Supplementary Figure 8. (Related to STAR Methods) Evaluation of clinical specimens for antigen detection assay or RT-PCR.**

Clinical specimens of nasopharyngeal swabs from cases with suspected SARS-CoV-2 infection were retrieved for this study. Swabs were collected in 3 mL of viral transport medium. RNA extraction and RT-PCR using NIID-N2 set primer were carried out as an administrative inspection. The remaining clinical samples were centrifuged, and the supernatant was used for this study. For antigen detection assays, samples were diluted 2-fold (ELISA) or 3-fold (LFIA) in extraction buffer and 100 µL (ELISA) or 1 drop (LFIA) was subjected to analysis. For RT-PCR analysis to quantify Ct value, samples were diluted to same concentration used in antigen detection assay (2-fold for ELISA, 3-fold for LFIA) with viral transport medium. RNA extraction and RT-PCR analysis were carried out using 140 µL of diluted samples.