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Supplemental information

Highly specific monoclonal antibodies

and epitope identification against SARS-CoV-2

nucleocapsid protein for antigen detection tests

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Supplementary Figure 1. (Related to Figure 1) Antigen protein production and specificity screening of developed mAbs.

(A) Schematic representation of antigen protein production. Recombinant Histidine (His)-tagged N-terminally truncated SARS-CoV-2-NP (121-419: ΔN-NP) was produced in a wheat germ cell-free system, and then purified using nickel-chelated Sepharose beads. Each protein fraction was analyzed by SDS-PAGE and visualized by CBB staining. Red arrow indicate the target protein. NC, Negative Control; T, Total fraction; S, Supernatant; P, Precipitate; FT, Flow-Through; E1–3, Elution fractions 1-3. (B) Flag-GST-tagged NPs derived from several human coronaviruses were produced in the wheat germ cell-free system. Reactivity of generated mAbs was validated by immunoblot analysis using the indicated antibodies.



Supplementary Figure 2. (Related to Figure 1) Epitope mapping of mAbs.

Schematic diagram of SARS-CoV-2-NP domain architecture, mutants, and region of determined epitope of each monoclonal antibody. For epitope mapping, 26 deletion mutants (D1-D26) and 5 substitution mutants (SARS2/MERS-mut1-5) were produced as FLAG-GST-tagged proteins in the wheat germ cell-free system. NTD, N-terminal domain; LKR, flexible linker region; CTD, C-terminal domain. Reactivity of each mAb to deletion mutants was evaluated by indirect-ELISA. EC₅₀ was calculated using six different antibody concentrations, and samples with an over 5-fold increase in EC₅₀ compared to full-length NP were defined as negative (Red).



Supplementary Figure 3. (Related to Figure 1) Critical residues for desecting SARS-CoV and SARS-CoV-2 in epitope of developed mAbs.

(A) Multiple sequence alignments of NP of human coronaviruses in the epitope region using MUSCLE software. Gray shaded positions represent conserved residues among the sequences. (B) Determination of critical residues for specific detection of not SARS-CoV but SARS-CoV-2. Five point substitution mutants (SARS2/SARS1-mut1-5) were produced as FLAG-GST-tagged proteins in the wheat germ cell-free system. NTD, N-terminal domain; LKR, flexible linker region; CTD, C-terminal domain. Reactivity of each mAb to deletion mutants was evaluated by indirect-ELISA. EC_{50} was calculated using six different antibody concentrations. The samples with an over 5-fold increase in EC_{50} compared to full-length NP were defined as negative (Red).



Supplementary Figure 4. (Related to Figure 1) Shannon entropy analysis of nucleocapsid proteins and comparison of epitope sequences between prototype and current variant of SARS-CoV-2.

(A) Shannon entropy, as a quantitative measure of variation, was calculated for each amino-acid residue of SARS-CoV-2 NP derived from 8,127 strains. Binding sites of each mAb are indicated in different colors. (**B**) Amino acid sequence in the antigenic epitope of reference strain against each of the currently reported epidemic variant of SARS-CoV-2¹¹. Gray shaded positions represent conserved residues among the sequences.

Α		
#7:	210-231	aa

Number Relative ACCESSION ID 210 211 212 213 214 215 216 217 218 219 220 221 222 223 224 225 226 227 228 229 230 231 Name of strain of strains abundance R L NC_045512 8091 99.56% Μ G Ν G D Q Reference genome А G D Α L L L L L Ν L Е Α Α MT847220 BGD/BCSIR NILMRC 281/2020 3 0.04% s MT706376 USA/WI-UW-249/2020 1 0.01% Т USA/WA-S2367/2020 MT831837 0.01% V 1 F MT358640 DEU/FFM4/2020 2 0.02% V USA/MA-MGH-01121/2020 USA/CA-CZB-1484/2020 MT873396 1 0.01% v MT628169 2 0.02% . . T USA/IA-CDC-8200/2020 MT472626 2 0.02% Ĥ MT601287 BGD/BCSIR NILMRC 75/2020 13 0.16% v MT535490 USA/UT-01568/2020 2 2 0.02% MT614522 USA/WA-QDX-62/2020 0.02% С · Y MT252734 USA/WA-UW154/2020 4 0.05% . · Y MT451476 AUS/VIC680/2020 2 0.02% MT806787 USA/IN-QDX-251/2020 0.01%

#9. 000-040 a	#9	:	335-348	aa
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#98: 382-397 aa

ACCESSION ID	Name of strain	Number of strains	Relative abundance	335	336	337	338	339	340	341	342	343	344	345	346	347	348
NC_045512	Reference genome	8107	99.75%	G	Α	1	Κ	L	D	D	K	D	Ρ	Ν	F	Κ	D
MT873324	USA/MA-MGH-01405/2020	1	0.01%		S												
MT706363	USA/WI-UW-235/2020	1	0.01%			F					·						
MT326140	USA/WA-UW-1680/2020	2	0.02%						Ν								
MT831745	USA/WA-S2426/2020	1	0.01%		· ·									Т			
LR877181	SWE/STO-KI/2020	1	0.01%				Ν				·						
MT683418	USA/WA-UW-10138/2020	1	0.01%								Ν						
LC529905	JP/TKYE6182/2020	3	0.04%		· ·								S				
MT358737	USA/WA-UW-3994/2020	1	0.01%						G		·						
MT509498	IND/GBRC107/2020	4	0.05%								·						Υ
MT731292	MAR/RMPS02/2020	2	0.02%														н
MT135043	CHN/233/2020	1	0.01%								·	V					
MT811401	USA/SEARCH-0834-IPL/2020	2	0.02%									Y					

ACCESSION ID	Name of strain	Number of strains	Relative abundance	382	383	384	385	386	387	388	389	390	391	392	393	394	395	396	397
NC_045512	Reference genome	8079	99.41%	L	Ρ	Q	R	Q	Κ	Κ	Q	Q	Т	V	Т	L	L	Р	Α
MT811458	USA/SEARCH-0997-IPL/2020	1	0.01%							R								·	
MT252716	USA/WA-UW180/2020	2	0.02%															· .	S
MT614463	USA/CA-QDX-40/2020	1	0.01%					Κ										· .	
LC571012	JP/DP0654/2020	1	0.01%			-									-			· .	V
MT831158	USA/WA-S2249/2020	5	0.06%					н										· .	
MT252737	USA/WA-UW146/2020	2	0.02%										1					· .	
MT358637	IND/GBRC1/2020	11	0.14%			-									1			· .	
MT757020	USA/FL-BPHL-0351/2020	1	0.01%			н									-			· .	
MT451616	AUS/VIC879/2020	9	0.11%		S													· .	
MT646100	USA/MD-HP00086/2020	9	0.11%		L	-									-			· .	
MT831179	USA/WA-S2105/2020	1	0.01%	S		-									-			· .	
MT825091	IRN/COVID19-IRVSH1/2020	1	0.01%									L						· .	
MT811500	USA/SEARCH-1065-SAN/2020	1	0.01%			-				1					-			· .	
MT757121	USA/FL-BPHL-0313/2020	1	0.01%				1											· .	
MT810952	USA/SEARCH-0240-SAN/2020	1	0.01%															н	
MT434815	USA/NY-CDC-SURV0175NYC/2020) 1	0.01%									-	-	-					

В

#7 : 210-231 aa						#9:	335-348 a	а			#98 : 382-397 aa								
Antibody Name Mutation site (EC _{E0} :				Antibody reactivity			Mutation site		Antibody (EC _{E0} :	reactivit	iy	Name	Mutation site	Antibody reactivity (ECroping/mL)					
		#7	#9	#98	Flag			#7	#9	, #98	Flag			#7	#9	#98	Flag		
Full	-	31	57	18	18	Full	-	31	57	18	18	Full	-	31	57	18	18		
M1	A211S	28	51	17	20	M13	A336S	29	57	18	18	M25	K388R	30	52	16	18		
M2	M210I	31	54	18	20	M14	1337F	29	62	18	19	M26	A397S	28	53	16	18		
M3	M210V	30	55	18	20	M15	D340N	30	56	18	18	M27	Q386K	29	56	16	18		
M4	L230F	25	54	19	20	M16	N345T	30	63	17	19	M28	A397V	29	54	15	19		
M5	A211V	31	57	18	20	M17	K338N	30	57	18	20	M29	Q386H	28	54	17	18		
M6	A218V	40	79	19	20	M18	K342N	30	52	18	18	M30	T391I	30	52	16	19		
M7	A220T	29	58	18	20	M19	P344S	31	2658	16	19	M31	T393I	28	50	>3000	19		
M8	Q229H	27	56	19	21	M20	D340G	32	77	17	19	M32	Q384H	29	56	18	19		
M9	G212V	36	64	18	16	M21	D348Y	32	2348	16	18	M33	P383S	31	63	17	20		
M10	G212C	153	77	18	17	M22	D348H	31	1755	17	18	M34	P383L	31	58	18	21		
M11	N213Y	51	74	18	17	M23	D343V	30	>3000	17	18	M35	L382S	30	60	18	21		
M12	N228Y	56	89	17	18	M24	D343Y	31	2316	16	17	M36	Q390L	31	60	98	20		
M13	G214C	185	70	17	18							M37	K388I	30	60	19	21		
												M38	R385I	31	58	18	22		
												M39	P396H	30	58	15	20		
												M40	∆390-392	29	53	>3000	21		

Supplementary Figure 5. (Related to Figure 1) Reactivity of developed monoclonal antibodies against recombinant protein from divergent strains.

(A) Identification of amino-acid substitution in the antigenic epitope of each identical strain from 8,127 clinical strains compared with the reference strain. Dots indicate sequence identity relative to the reference strain. Gray shaded

positions represent conserved residues among the sequences. (B) Reactivity of mAbs to recombinant NP from strains possessing amino-acid substitution in the epitope regions. Forty point substitution mutants identified in (A) were produced as FLAG-GST-tagged proteins in the wheat germ cell-free system. Reactivity of each mAb to mutants was evaluated by indirect-ELISA. EC_{50} was calculated using six different antibody concentrations. The samples with an over 5-fold increase in EC_{50} compared to full-length NP were defined as negative (Red).



Supplementary Figure 6. (Related to Figure 2) Comparison of commercially available existing mAbs against SARS-CoV-2 NP.

(A) Affinity measurement of commercially available monoclonal antibodies on the octetRED96 instrument. Association and dissociation of each mAbs to full-length NP at various concentrations (50, 25, 12.5, 6.25, 3.13, 1.56 and 0.78 nM) was evaluated using AMC sensor. (B) Specificity of existing mAbs were examined by immunoblot analysis using the indicated recombinant proteins from human coronaviruses.



Supplementary Figure 7. (Related to Figure 4) Overview and procedure of LFIA.

(A) Image of LFIA Cartridge. (B) Samples were diluted with an extraction buffer (Tris buffer containing 0.1% nonionic surfactant) in the tube and one drop of the sample was added onto the sample well of LFIA cartridge. Following this, button 2 was immediately pressed to release a reducing reagent for silver amplification. After the color indicator mark turned orange (about 10 min), button 3 was pressed to release a silver-ion reagent to activate the silver amplification reaction.



Supplementary Figure 8. (Related to STAR Methods) Evaluation of clinical specimens for antigen detection assay or RT-PCR.

Clinical specimens of nasopharyngeal swabs from cases with suspected SARS-CoV-2 infection were retrieved for this study. Swabs were collected in 3 mL of viral transport medium. RNA extraction and RT-PCR using NIID-N2 set primer were carried out as an administrative inspection. The remaining clinical samples were centrifuged, and the supernatant was used for this study. For antigen detection assays, samples were diluted 2-fold (ELISA) or 3-fold (LFIA) in extraction buffer and 100 μ L (ELISA) or 1 drop (LFIA) was subjected to analysis. For RT-PCR analysis to quantify Ct value, samples were diluted to same concentration used in antigen detection assay (2-fold for ELISA, 3-fold for LFIA) with viral transport medium. RNA extraction and RT-PCR analysis were carried out using 140 μ L of diluted samples.