

## Supporting Information for

Structures of the alkanesulfonate monooxygenase MsuD provide insight into C–S bond cleavage, substrate scope, and an unexpected role for the tetramer

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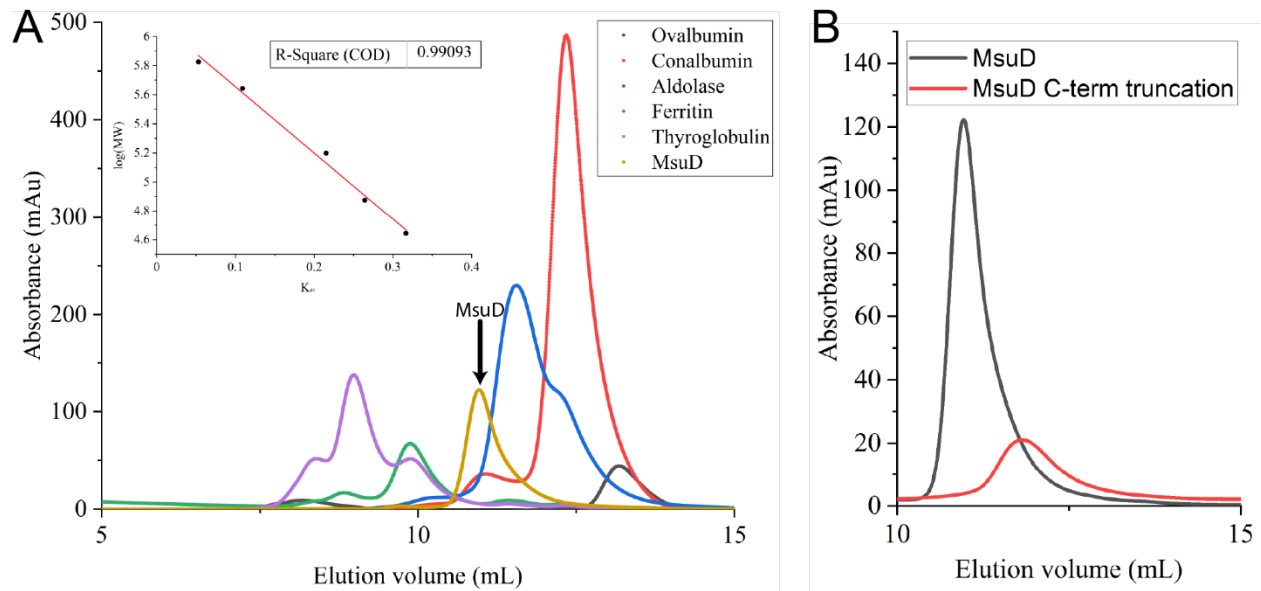
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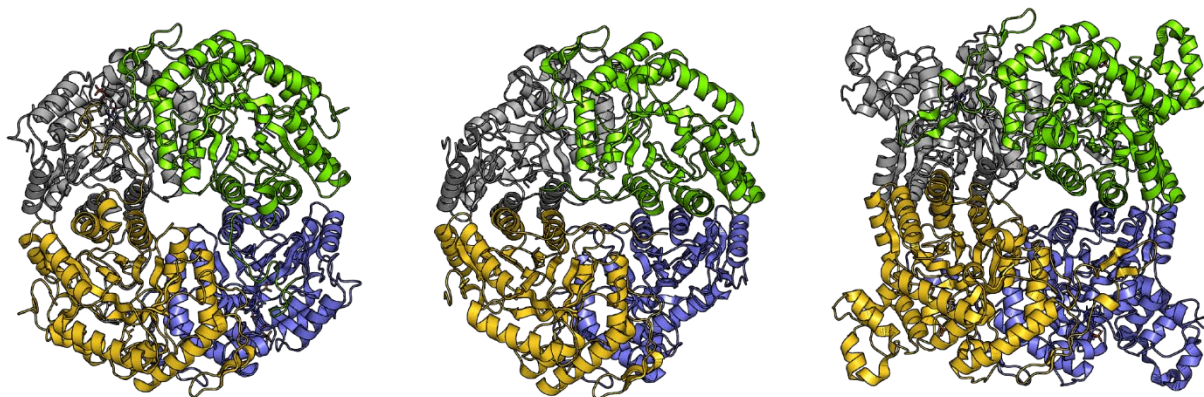
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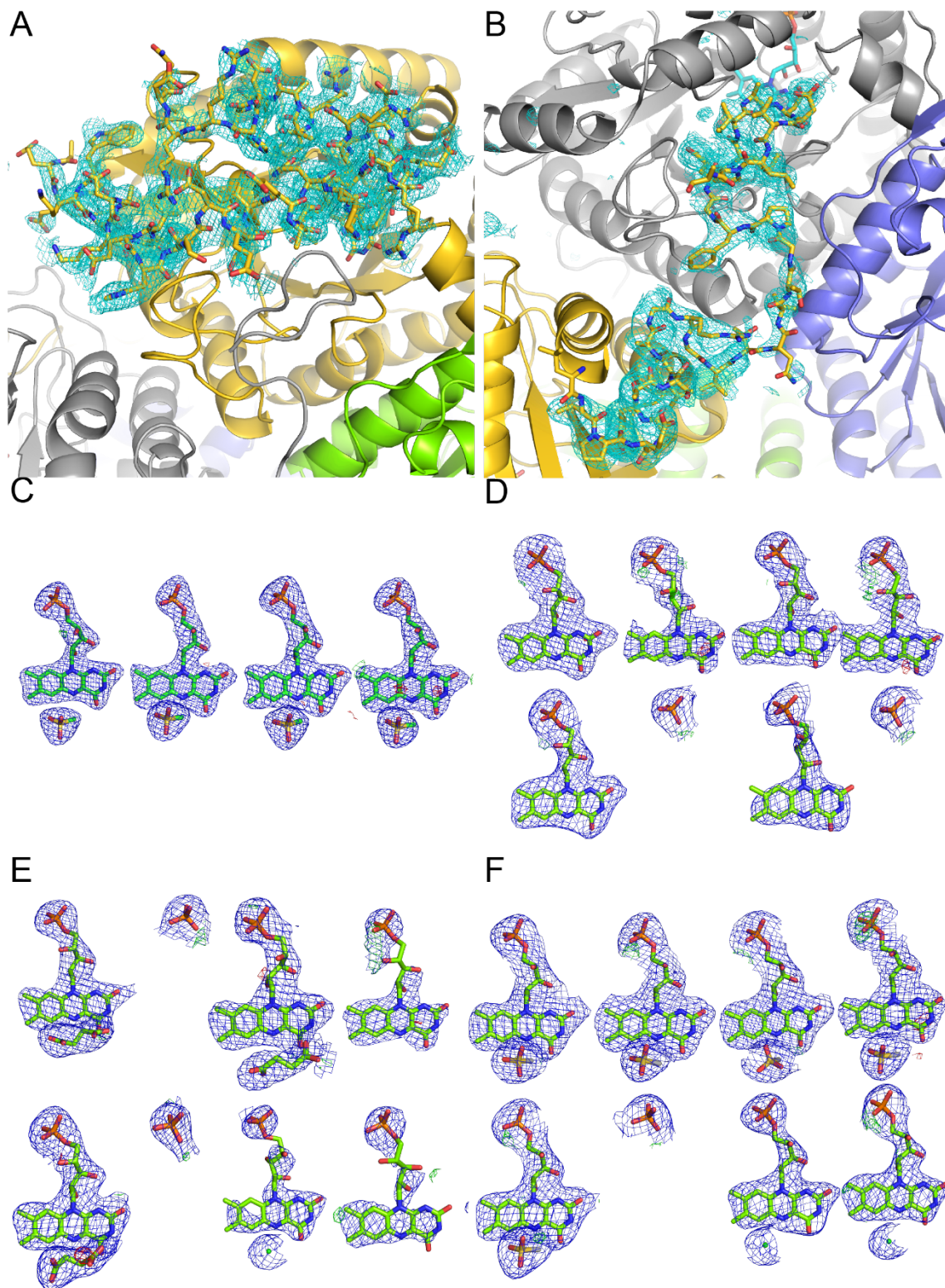
**Figure S1.** MsuD behaves as a tetramer in solution. (A) Size exclusion chromatography of MsuD on the ENrich SEC 650 column compared to Cytiva high-molecular weight protein standards reveals MsuD travels as an ~200 kDa species, most in line with a tetramer (monomer chain MW: 44.2 kDa, and homotetramer MW: 176.9 kDa). Molecular weight standards include: Ovalbumin (MW: 44 kDa), Conalbumin (MW: 75 kDa), Aldolase (MW: 158 kDa), Ferritin (MW: 440 kDa) and Thyroglobulin (MW: 669 kDa). (B) An overlay of wild type MsuD and the C-terminal truncation mutant (MsuD <sup>$\Delta$ C-16</sup>) reveal the C-terminal tail is critical for tetramerization. MsuD <sup>$\Delta$ C-16</sup> behaves as an ~113 kDa species, which corresponds to a dimer.



**Figure S2.** Sequence alignment of MsdD structural homologs. Homologous structures identified using the DALI server with a minimum sequence identity cut-off of 19% are displayed. The protein secondary structural elements from the MsdD structure are indicated above the MsdD sequence. Blue boxes indicate conserved and similar residues, with strictly conserved residues highlighted in red and similar residues in red text. The alignment figure was prepared using the Easy Sequencing in PostScript (ESPrpt) online program v3.0 (1). MsdD (GenID:ABA75653.1) was aligned with SsdD (GenID: CAB40391.1, seqID: 67%), RutA (GenID:AAC74097.1, seqID: 25%), PDBID: 3RAO (GenID:AAS39998.1, seqID: 32%), DmoA (GenID:ADU77278.1, seqID: 23%), LadA (GenID:ABO68832.1, seqID: 25%), RcaE (UniProt ID: A0A3B6UEK8, seqID: 23%), BdsA (GenID:BAC20180.1, seqID: 22%), and CmoJ (GenID:CAB14891.1, seqID: 24%).



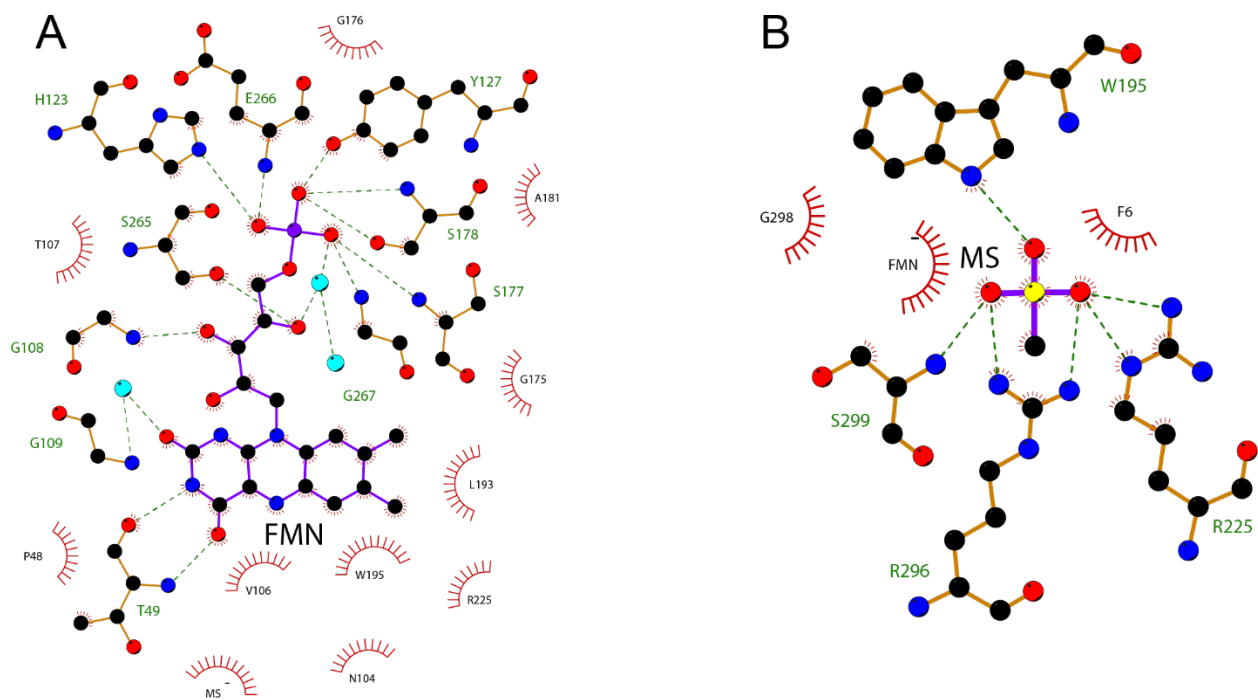
**Figure S3.** Comparison of class C flavin-dependent monooxygenases that crystallize as tetramers. Tetramers are displayed from left to right are MsuD (PDB ID 7JW9), SsuD (PDB ID 1NQK), and BdsA (PDB ID 5XKD). Protein chains are colored green, blue, yellow, and gray, respectively.



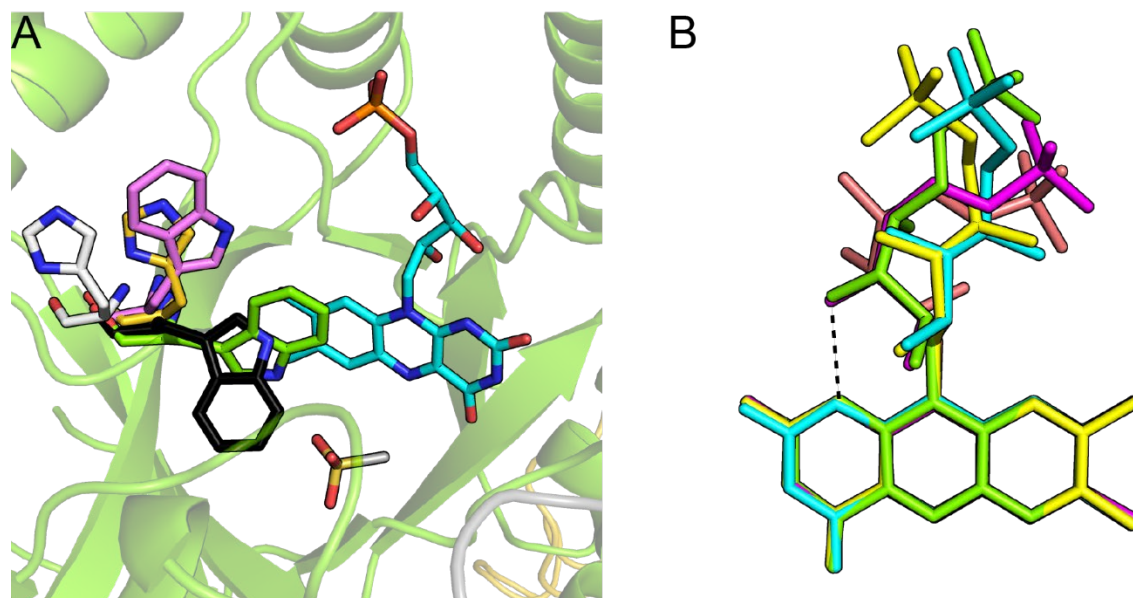
**Figure S4.** Representative polder omit electron density maps for structures of MsuD demonstrating the presence of the active site lid, protein C-terminus, and the ligands FMN, MS<sup>-</sup>,

and succinate. (A) and (B) Polder omit electron density for the best ordered lid region and C-terminus (chain A) from ternary-MsuD (cocrystal) are displayed. Protein chains A-D are colored green, blue, yellow, and gray, respectively. Polder mFo-DFc omit maps were calculated for segments of five residues at a time and are contoured at  $3\sigma$  level, carved to 2 Å about the lids. Polder omit maps for ligands (FMN,  $MS^-$ , and succinate) are shown from chains A/B/C/D on top and E/F/G/H on the bottom of the MsuD tetramer in (C) ternary-MsuD (cocrystal), (D) binary-soak MsuD, (E) binary-titrated MsuD, and (F) ternary-soak MsuD. Omit maps for ligands are at  $5\sigma$  level and Fo-Fc electron density maps at  $\pm 3\sigma$  level, colored green for positive peaks and red for negative peaks, carved up to 2 Å about the ligands.

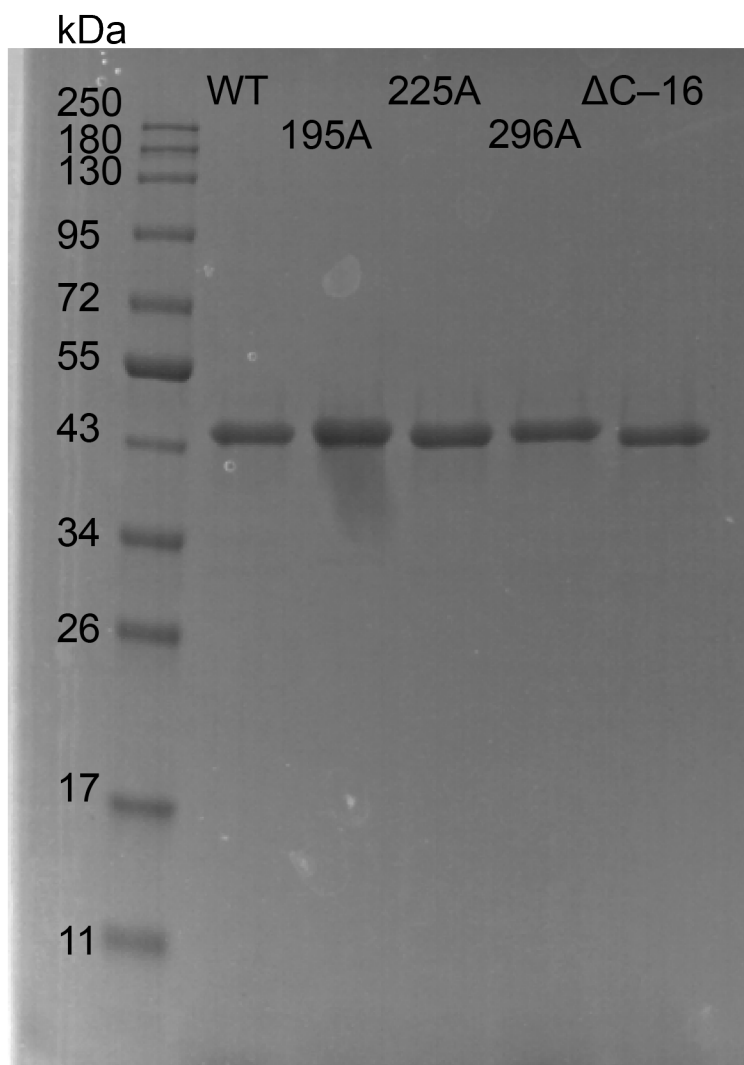




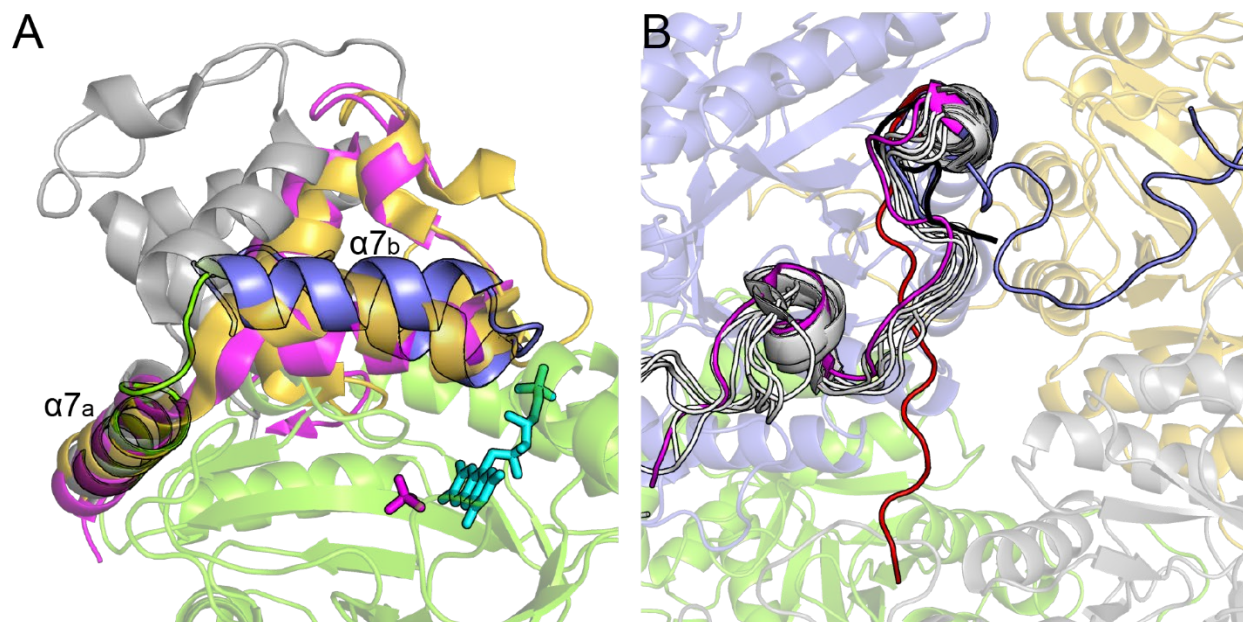
**Figure S5.** Analysis of residues and surrounding environment of (A) FMN and (B) MS<sup>-</sup> in chain A of ternary-MsuD, showing interactions within hydrogen bonding distance and residues within van der Waals contact distances. Figure generated using LigPlot+ (2).



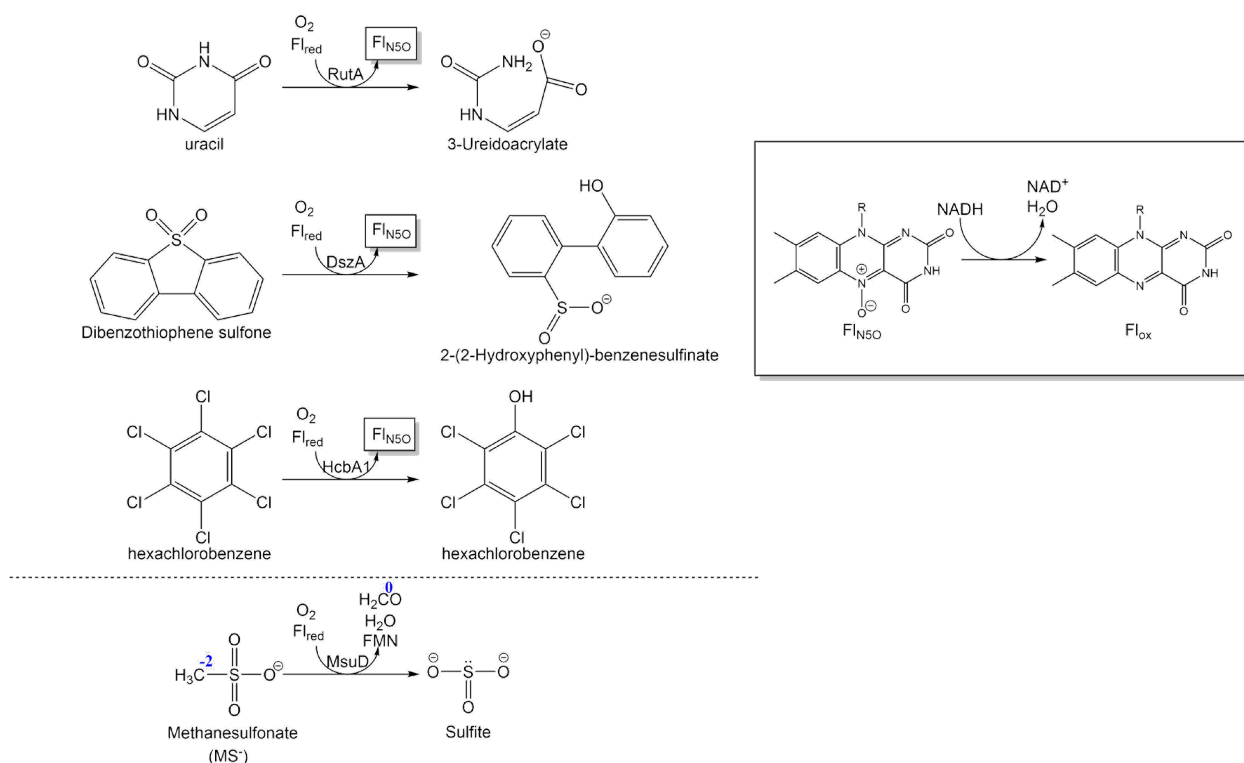
**Figure S6.** (A) Overlay of unliganded MsuD (black), ternary-MsuD (green), SsuD (pink), CmoJ (white), and PDBID: 3RAO (orange), demonstrating the position of aromatic residues analogous to W195 in the absence of FMN. FMN is displayed with cyan carbons from the ternary-MsuD cocrystal structure. (B) Overlay of FMN from different homolog structures demonstrating differences in ribityl tail positioning. The isoalloxazine rings of FMN were superimposed for MsuD (green, PDB ID 7JW9) with RutA (cyan, PDB ID 5WAN), LadA (purple, PDB ID 3B9O), RcaE (yellow, PDB ID 5W4Y) and BdsA (pink, PDB ID 5XKD).



**Figure S7.** Recombinantly expressed MsuD variants were purified and analyzed by SDS-PAGE. Approximately 1  $\mu$ g samples of fully purified MsuD including WT and variants were loaded onto a BioRad AnyKD Protein Gel and stained with SimplyBlue SafeStain. The MW sizes of the NEB Protein Ladder are labeled.



**Figure S8.** Structural superimpositions of MsuD homologs reveal similarities and differences in their lids and C-termini. (A) An overlay of several lids from MsuD homologs is displayed. Protein overlays are displayed as ribbons. MsuD is colored green with FMN and MS<sup>-</sup> in cyan and magenta sticks, respectively. A variable number of helices are present in the lid region of different homologs, and an overlay demonstrates how the first helix is consistently placed. The homologs shown are RcaE (orange), LadA (magenta) and CmoJ (gray). (B) A comparison of the C-termini in MsuD and its homologs is displayed. The C-termini of the three tetrameric monooxygenase structures are displayed for MsuD (blue), SsuD (red), and BdsA (magenta), and the dimeric homologs (DmoA, EmoA, LadA, RcaE, CmoJ, and PDB ID 3SDO) are shown in white. The C-terminus in MsuD is in a conformation not previously observed.



**Figure S9.** The proposed reactions of RutA, DszA, and HcbA1 demonstrate differences in substrate oxidation state with MsuD. Recent work by Matthew's et al have proposed the use of an N5-(hydro)peroxy ( $\text{Fl}_{\text{N5OO}}$ ) intermediate in the reactions of three class C two-component flavin-dependent monooxygenases, RutA, DszA, and HcbA1, and a flavin N5-oxide is formed in the reaction cycle (3). Considering that the starting substrates of RutA, DszA, and HcbA1 are uracil, dibenzothiophene sulfone, and hexachlorobenzene, respectively, these substrates are in a more oxidized state at the carbon center proposed for attack by the  $\text{Fl}_{\text{N5OO}}$  than an alkanesulfonate substrate for MsuD. Therefore, the mechanisms of RutA, DszA, and HcbA1 would generate  $\text{Fl}_{\text{N5O}}$  and must invoke an NADH molecule for the final water reduction step to regenerate oxidized flavin. Oxidation states for carbon in the MsuD reaction are shown.

## Supporting Tables.

**Table S1.** Genome and gene information for *msu* and *sfn* genes from *P. fluorescens* P0-1, *P. aeruginosa* PAO1, and *P. putida* KT2440.

Sequence Name	Protein Name	Length	locus_tag	old locus_tag	gene product annotation
P. aeruginosa PAO1 NC_002516	SfnR1	1131	PA2354		transcriptional regulator
	MsuC	1185	PA2355		FMNH <sub>2</sub> -dependent monooxygenase
	MsuD	1146	PA2356		methanesulfonate monooxygenase
	MsuE	561	PA2357		FMN reductase MsuE
	SfnR2	1086	PA2359		transcriptional regulator
	SfnG	1095	PA3954		hypothetical protein
P. fluorescens Pf0-1 NC_007492	MsuE	564	PFL01_RS19665	PfI01_3915	FMN reductase
	MsuD	1146	PFL01_RS19670	PfI01_3916	FMNH <sub>2</sub> -dependent alkanesulfonate monooxygenase
	MsuC	1188	PFL01_RS19675	PfI01_3917	acyl-CoA dehydrogenase family protein
	SfnR	1104	PFL01_RS19680	PfI01_3918	sigma-54-dependent Fis family transcriptional regulator
	SfnG	1095	PFL01_RS14520	PfI01_2879	dimethyl sulfone monooxygenase SfnG
P. putida NC_002947	SfnR	1104	PP_RS14410	PP_2771	sigma-54-dependent Fis family transcriptional regulator
	MsuC	1188	PP_RS14415	PP_2772	acyl-CoA dehydrogenase family protein
	hypothetical protein CDS	159	PP_RS14420	PP_2773	hypothetical protein
	SfnB	1233	PP_RS14335	PP_2755	SfnB family sulfur acquisition oxidoreductase
	hypothetical protein CDS	387	PP_RS14340	PP_2756	hypothetical protein
	sugar ABC transporter substrate-binding protein CDS	972	PP_RS14345	PP_2757	sugar ABC transporter substrate-binding protein
	sugar ABC transporter substrate-binding protein CDS	1029	PP_RS14350	PP_2758	sugar ABC transporter substrate-binding protein
	sugar ABC transporter ATP-binding protein CDS	1539	PP_RS14355	PP_2759	sugar ABC transporter ATP-binding protein
	ABC transporter permease CDS	978	PP_RS14360	PP_2760	ABC transporter permease
	ABC transporter permease CDS	987	PP_RS14365	PP_2761	ABC transporter permease
	SfnA	1239	PP_RS14370	PP_2762	acyl-CoA dehydrogenase family protein
	SfnF	561	PP_RS14375	PP_2764	FMN reductase
	SfnG	1092	PP_RS14380	PP_2765	dimethyl sulfone monooxygenase SfnG

**Table S2.** Structural alignment results for MsuD from the DALI server (4), containing matches above or equal to 19% sequence identity. With the exceptions of RcaE, 1LUC, and 5LXE, these proteins are predicted to be class C flavin-dependent monooxygenases.

Chain	RMSD	% id	Number of aligned C $\alpha$	Name	Description	References
1NQK	1.3	69	328	SsuD	Alkanesulfonate monooxygenase	(5)
3RAO	2.0	32	320	N/A	Luciferase-like monooxygenase	(6)
5WAN	2.1	25	312	RutA	Pyrimidine monooxygenase	(7)
5W4Z	2.1	23	312	RcaE	Riboflavin lyase	(8)
3B9O	2.5	25	328	LadA	Long-chain alkane monooxygenase	(9)
5XKD	2.6	22	336	BdsA	Dibenzothiophene monooxygenase	(10)
3SDO	2.3	21	248	N/A	Nitrilotriacetate monooxygenase	(11)
6ASL	2.7	24	248	CmoJ	Cysteine salvage pathway	(8)
5DQP	2.8	19	320	EmoA	Ethylenediaminetetraacetic acid monooxygenase	(12)
6AK1	2.8	23	328	DmoA	Dimethylsulfide monooxygenase	(13)
1LUC	2.7	19	296	N/A	Bacterial luciferase	(14)
3FGC	2.7	18	304	N/A	Bacterial luciferase	(15)
5LXE	2.4	21	288	Rh-fgd1	Glucose-6-phosphate dehydrogenase	(16)

**Table S3.** Summary of ligand occupancies and lid regions within MsuD crystal structures. The active site lid consists of residues D250–L282. In structures with an ordered C-terminus, the final four residues are unable to be built. For soaking experiments, FMN binding appears to be strongest in chains A/C and E/G of the two MsuD tetramers, whereas FMN binds strongly in all molecules of the cocrystal structure (highlighted yellow).

		Ternary cocrystal				Unliganded				Binary-soak				Ternary-soak				Binary-titrated			
Chain		FMN	MS <sup>-</sup>	Lid	C-terminus	FMN	MS <sup>-</sup>	Lid	C-terminus	FMN	MS <sup>-</sup>	Lid	C-terminus	FMN	MS <sup>-</sup>	Lid	C-terminus	FMN	MS <sup>-</sup>	Lid	C-terminus
Tetramer One	A	F <sup>a</sup>	F	O <sup>b</sup>	O	-	-	D	D	F	-	O	D	NF	P	O	D	NF	- <sup>f</sup>	O	D
	B	F	F	O	O	-	-	D	D	P	-	D	D	P	P	O	D	- <sup>c</sup>	-	D	O
	C	F	F	O	O	-	-	D	D	F	-	O	D	NF	P	O	D	NF	- <sup>f</sup>	O	D
	D	F	F	O	O	-	-	D	D	NF	-	D	D	P	P	O <sup>g</sup>	D	P	-	D	O
Tetramer Two	E					-	-	D	D	F	- <sup>f</sup>	O	D	NF	P	O	D	NF	- <sup>f</sup>	O	D
	F					-	-	D	D	- <sup>c</sup>	-	D	D	- <sup>c</sup>	-	D	D	- <sup>c</sup>	-	D	D
	G					-	-	D	D	P	-	D	D	P	- <sup>e</sup>	D <sup>h</sup>	D	P	-	D	D
	H					-	-	D	D	- <sup>c</sup>	-	O <sup>d</sup>	O	P	- <sup>e</sup>	D	O	P	-	D	O

<sup>a</sup> Due to the moderate resolution of liganded structures, occupancies of ligand are reported as fully bound (F, 100%), nearly fully bound (NF, 90% - 99%), or partially bound (P, <90%).

<sup>b</sup> O= ordered; D= disordered

<sup>c</sup> Phosphate has been placed in these chains as a substitute for the phosphate head of FMN as the rest of the FMN density is not present

<sup>d</sup> The lid of this chain was only partially built up to R262, denoting a potential alternate conformation

<sup>e</sup> A chloride ion was placed in these chains of the ternary soak

<sup>f</sup> Succinate was placed in these chains of the binary structures; no MS<sup>-</sup> was used in the experiment

<sup>g</sup> lacks electron density for R278 and R279

<sup>h</sup> contains electron density for a7<sub>c</sub> only



**Table S4.** The ribityl tail from different FMN-bound structures of the structural homologs of MsdD demonstrate altered torsion angles. The torsion angles of the ribityl tail range from nearly all 180° in the fully extended state of CmoJ to a more compact state within BdsA.

Structure	PDB ID	Torsion angle of ribityl tail (°)			
		N10-C1'-C2'-C3'	C1'-C2'-C3'-C4'	C2'-C3'-C4'-C5'	C3'-C4'-C5'-O5'
MsdD	7JW9	-131.8	-77.4	70.7	-161.1
RutA	5WAN	178.6	-53.1	-179.9	60.8
BdsA	5XKD	124.4	67.7	42.5	-84.0
RcaE	5W4Y	-174.8	-58.3	-178.5	56.6
LadA	3B9O	-125.7	-84.2	4.7	-91.0
CmoJ	6ASL	-179.1	179.6	-177.8	-171.8
LuxA	3FGC	169.0	-74.3	-99.3	-31.4

**Table S5.** Comparison of activity of alkanesulfonate monooxygenases from different organisms and genes with different sulfonate group containing substrates.

			Substrates					
			Methanesulfonate	Octanesulfonate	Pentanesulfonate	HEPES	PIPES	MOPS
Organism and Gene	MsuD <i>P. fluorescens</i>	% relative activity	86 ± 7	100 ± 10	62 ± 5	12 ± 4	14 ± 7	19 ± 7
	MsuD <i>P. aeruginosa</i> <sup>a</sup>	% relative activity	100	N/A	13.0	9.0	31.0	17.2
	SsuD <i>E. coli</i> <sup>b</sup>	% relative activity <sup>c</sup>	1.5	100	87.3	22.9	63.1	78.6

<sup>a</sup> (17)

<sup>b</sup> (18)

<sup>c</sup> Octanesulfonate activity was scaled from 46.3% of 100% (4.1  $\mu\text{mol min}^{-1} \text{mg}^{-1}$ ) 1,3-Dioxo-2-isoindolineethanesulfonic acid activity for comparison of substrates tested in this work. Other substrates were similarly scaled.

**Table S6.** The *msuD* gene of *P. fluorescens* Pf0-1 was amplified by PCR using the forward and reverse DNA primers shown. The site for restriction enzyme NheI is underlined in the forward primers and the site for restriction enzyme HindIII is underlined in the reverse primers. Mutagenesis was done with forward and reverse designed primers.

Name	Sequence
Pfl01_msuD_fw (Pfl01_3916_fw)	5'- GAT ATA <u>GCT AGC</u> ATG GAT GTT TTC TGG TTC CTG CCG ACT CAC GGC GAC GG -3'
Pfl01_msuD_rv (Pfl01_3916_rv)	5'- GAT ATA <u>AAG CTT</u> TCA GGC GTT GGC TTT GGC GGG CAG TAC ATC GTT GGC G -3'
MsuD-W195A-F	5'- C GTG TAC CTG ACG <u>GCA</u> GGC GAA CCA CCG GC -3'
MsuD-W195A-R	5'- TCG ACC TGC TCG GCG GCC -3'
MsuD-R225A-F	5'- G TTC GGC ATC <u>GCG</u> CTG CAC GTG ATC GTG -3'
MsuD-R225A-R	5'- TTC ACT TTG CGC CCG TGA -3'
MsuD-R296A-F	5'- C GGT CTG GTG <u>GCA</u> GGC GGT TCC GGC ACT G -3'
MsuD-R296A-R	5'- ACG CCG GCC CAC AGG TTG -3'
MsuD-Stop-F	5'- TC ACC GGG CCA <u>TGA GAC</u> GAA ATG ATC GCC AAC G -3'
MsuD-Stop-R	5'- GGT TGG TGA CGC CGC GTC -3'

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