

Supplemental information

**Promotion of cholangiocarcinoma growth by diverse
cancer-associated fibroblast subpopulations**

Silvia Affo, Ajay Nair, Francesco Brundu, Aashreya Ravichandra, Sonakshi Bhattacharjee, Michitaka Matsuda, LiKang Chin, Aveline Filliol, Wen Wen, Xinhua Song, Aubrianna Decker, Jeremy Worley, Jorge Matias Caviglia, Lexing Yu, Deqi Yin, Yoshinobu Saito, Thomas Savage, Rebecca G. Wells, Matthias Mack, Lars Zender, Nicholas Arpaia, Helen E. Remotti, Raul Rabadan, Peter Sims, Anne-Laure Leblond, Achim Weber, Marc-Oliver Riener, Brent R. Stockwell, Jellert Gaublonne, Josep M. Llovet, Raghu Kalluri, George K. Michalopoulos, Ekihiro Seki, Daniela Sia, Xin Chen, Andrea Califano, and Robert F. Schwabe

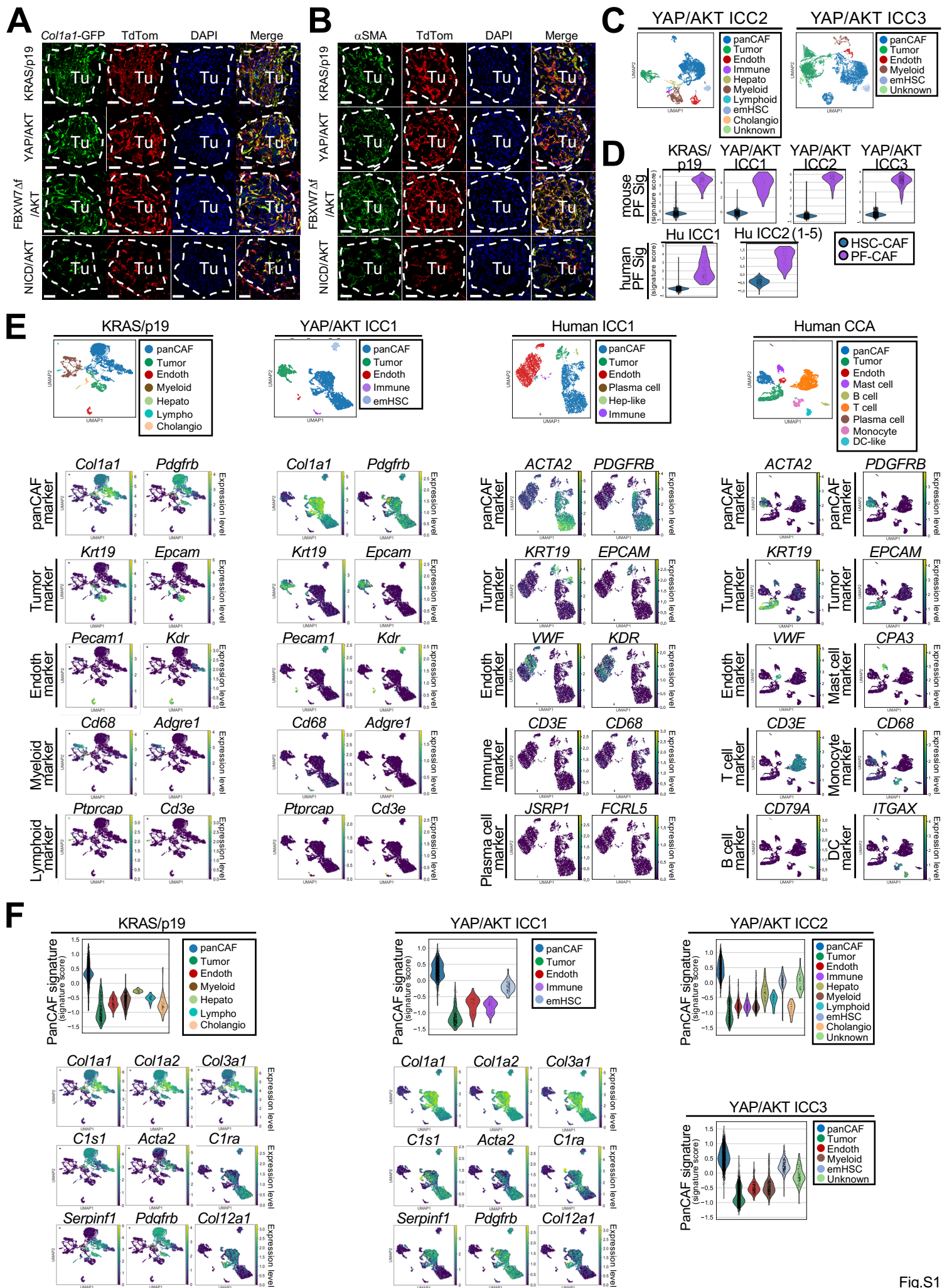


Fig.S1

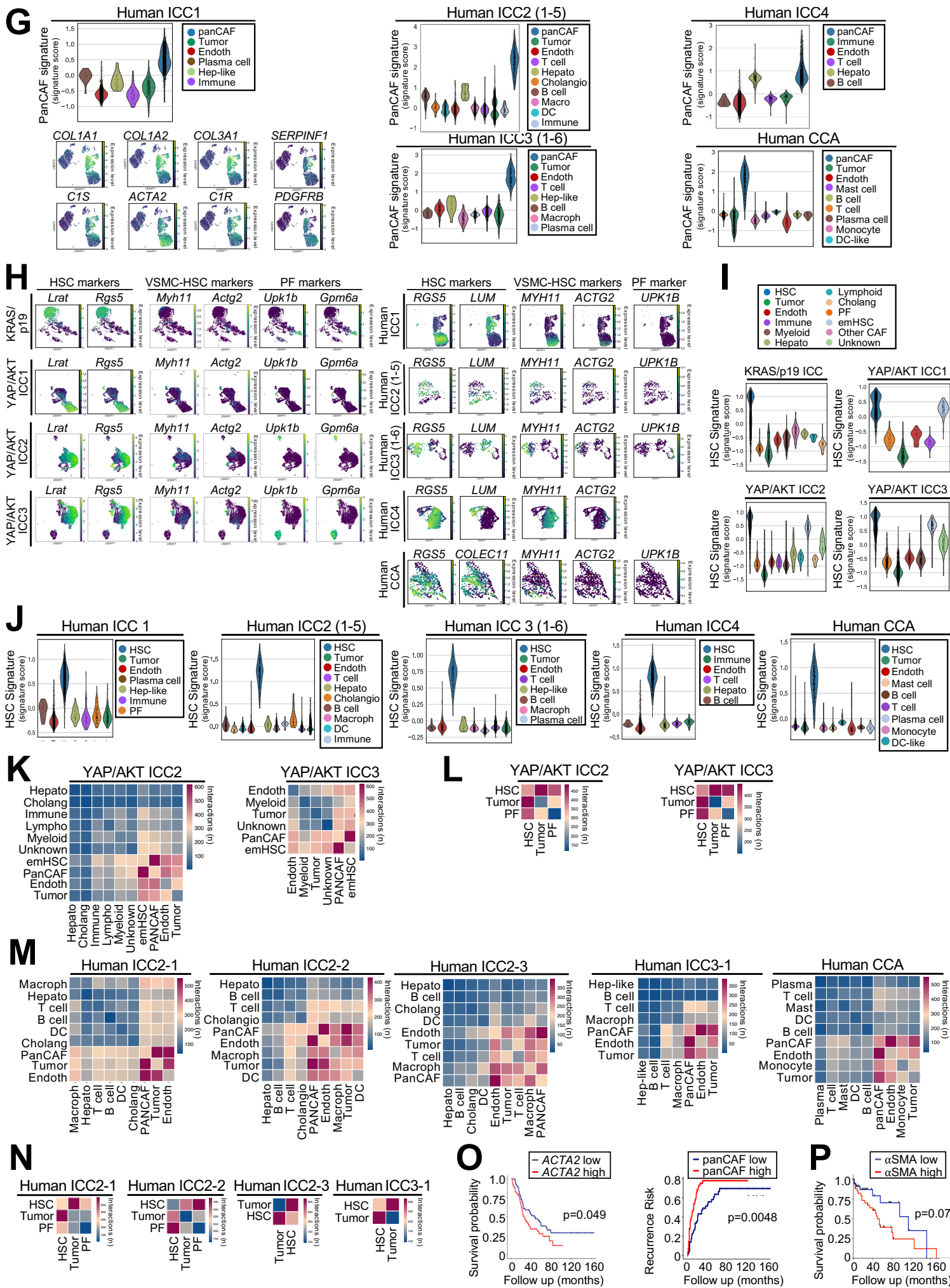


Fig.S1

Figure S1 (Related to Figure 1) | CAF origin and interactions with tumor cells in murine and human ICC. (A,B) Representative images of confocal microscopy showing co-localization of *Lrat*-Cre-induced TdTom with CAF markers *Colla1*-GFP (A) or α SMA (B) in KRAS/p19-, YAP/AKT-, FBXW7 Δ F/AKT-, NICD/AKT-induced ICC (n=3 per model) in *Lrat*-Cre⁺ TdTom⁺ *Colla1*-GFP⁺ mice. Scale bars, 50 μ m. (C) UMAPs of scRNA-seq showing all detected cell populations in the indicated samples. (D) Violin plots of scRNA-seq showing the PF signature score in the indicated samples; the width of each violin plot indicates the kernel density of the expression values. (E) UMAPs of scRNA-seq showing all detected cell populations and the normalized expression levels of indicated genes. (F,G) Violin plots showing the panCAF signature score in the indicated samples and UMAPs showing the normalized expression levels of indicated genes in (F) mouse ICC models (n=4) and in (G) human ICC (n=4) and human CCA (n=1). (H) UMAPs of scRNA-seq showing the normalized expression levels of HSC-CAF, VSMC-HSC and PF-CAF markers within panCAF in the indicated samples. (I,J) Violin plots of scRNA-seq showing the HSC signature score in (I) mouse ICC (n=4) and in (J) human ICC (n=4) and human CCA (n=1); the width of each violin plot indicates the kernel density of the expression values. (K-N) Heatmaps of scRNA-seq showing number of ligand-receptor interactions between (K,M) all cell populations and between (L,N) HSC-CAF, PF-CAF and tumor cells detected by CellphoneDB in the indicated YAP/AKT-induced ICC (n=2), human ICC (n=4) and human CCA (n=1). (O) Overall survival in 119 ICC patients with low (n=59) and high (n=60) *ACTA2* mRNA expression and recurrence risk in 113 ICC patients with low (n=56) and high (n=57) panCAF signature (Sia et al. cohort). (P) Overall survival in CCA patients with low (n=37) or high (n=40) expression of α SMA (Riener et al. TMA cohort).

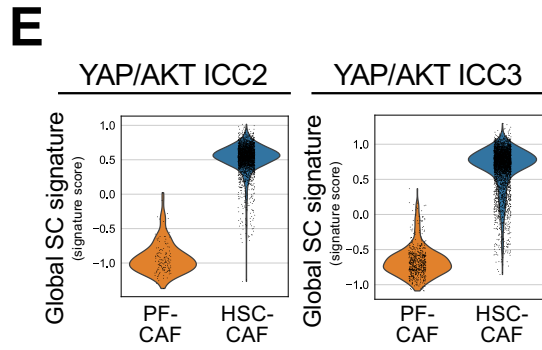
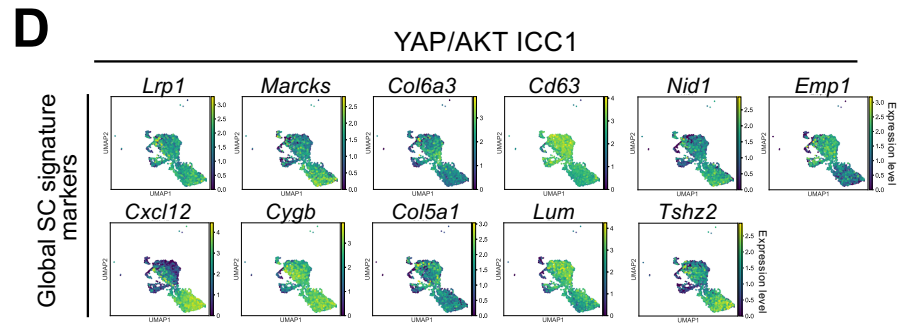
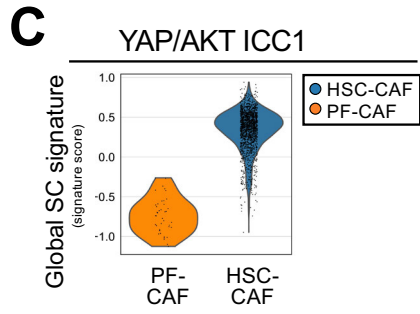
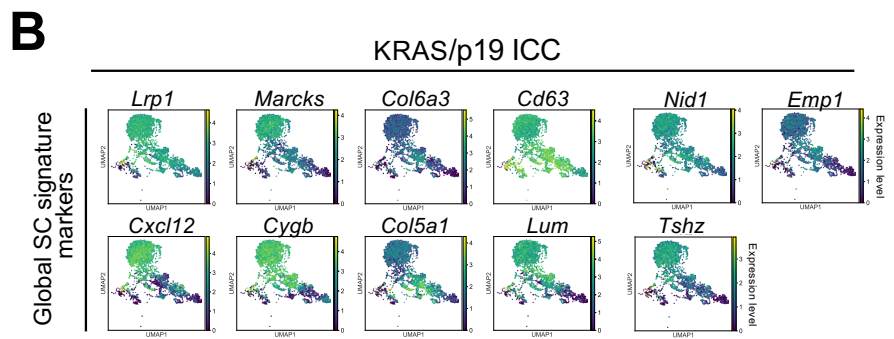
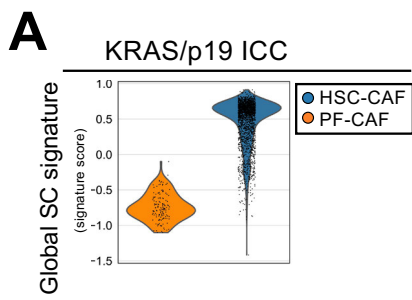
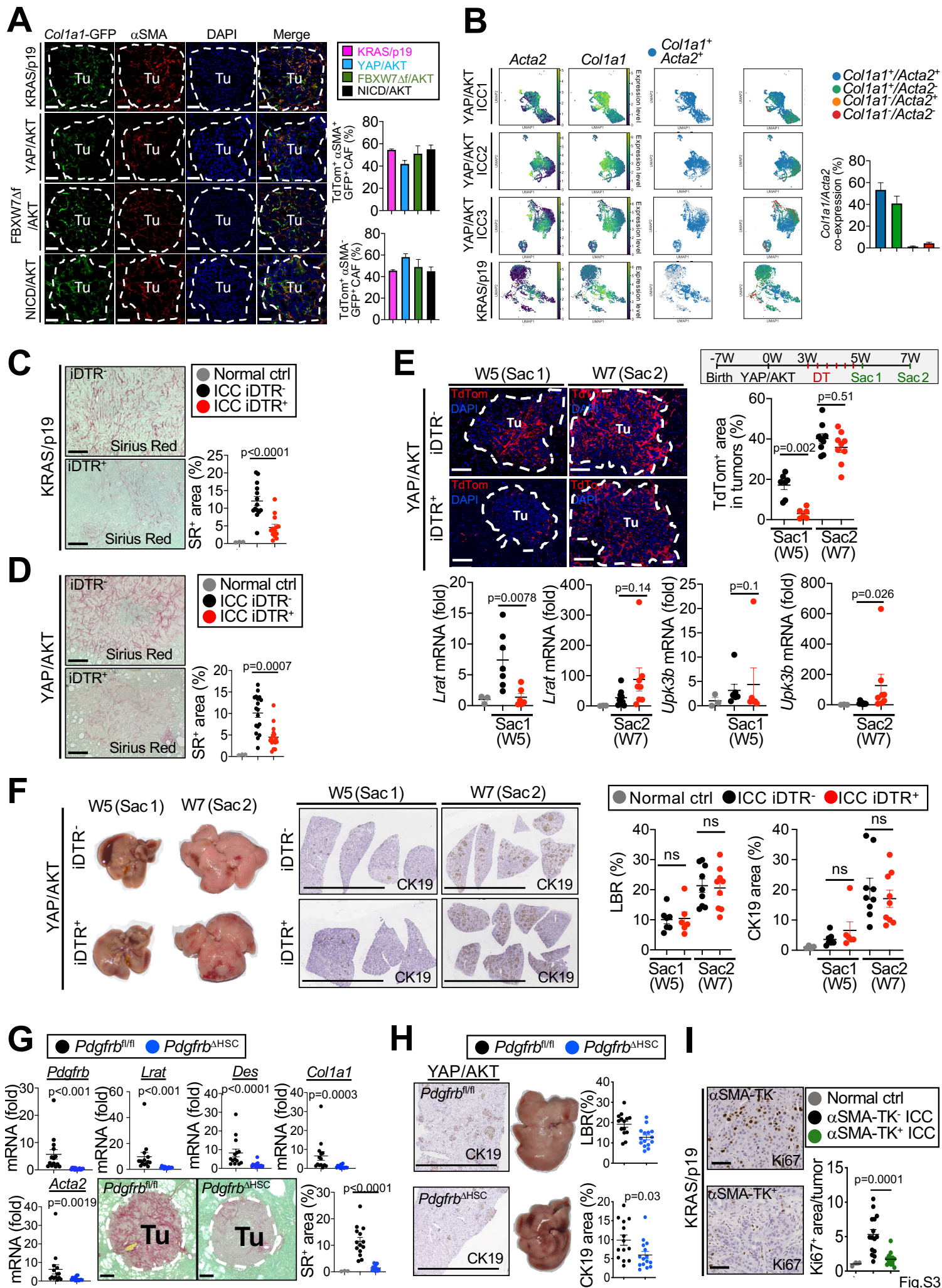


Figure S2 (Related to Figure 2) | Comparison of CAF from ICC and PDAC by global stellate cell and portal fibroblast signatures by single cell RNA-sequencing. (A) Violin plot showing the global SC signature in KRAS/p19-induced ICC (n=1). The width of each violin plot indicates the kernel density of the expression values. (B) UMAPs showing the normalized expression levels of the global stellate cell (global SC) signature markers in KRAS/p19 ICC. (C,E) Violin plot showing the global SC signature relative to randomly selected genes in YAP/AKT-induced ICC (n=3). The width of each violin plot indicates the kernel density of the expression values. (D) UMAPs showing the normalized expression levels of the global stellate cell (global SC) signature markers in YAP/AKT ICC (n=1).



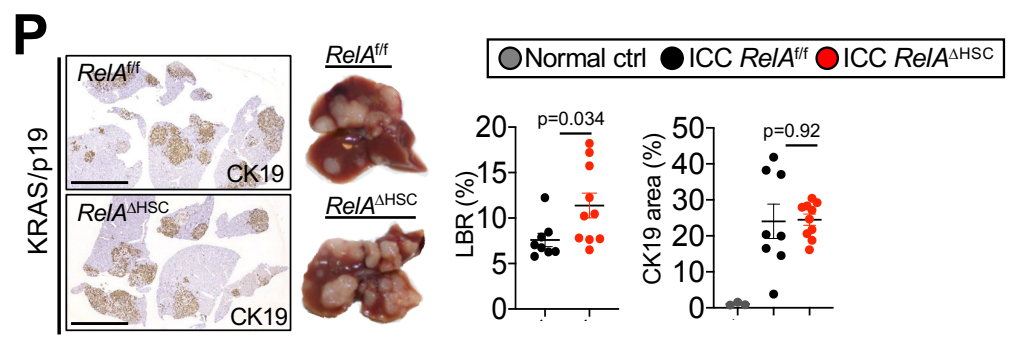
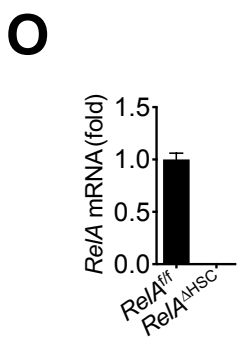
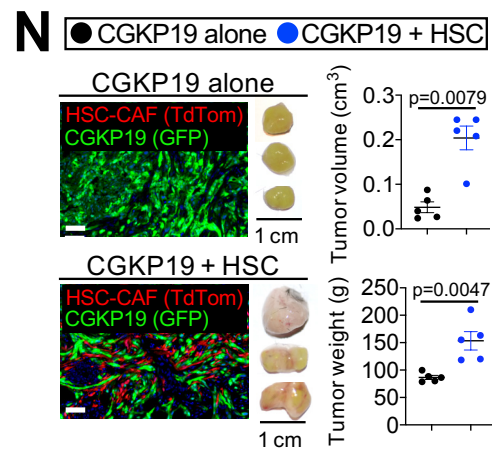
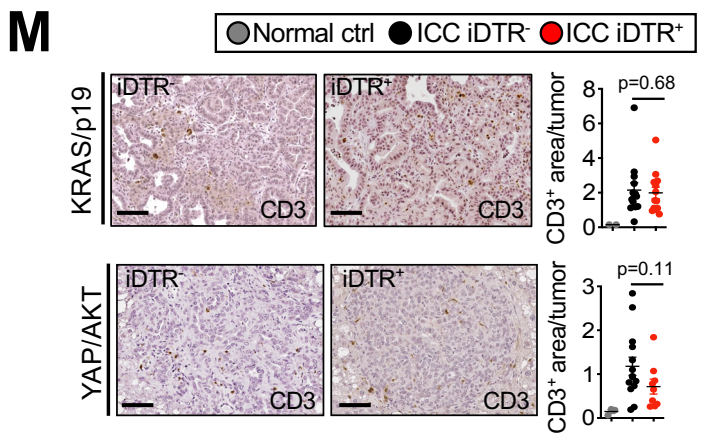
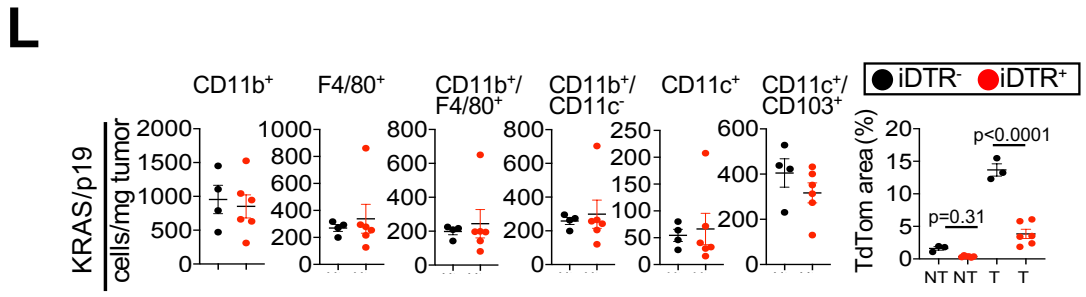
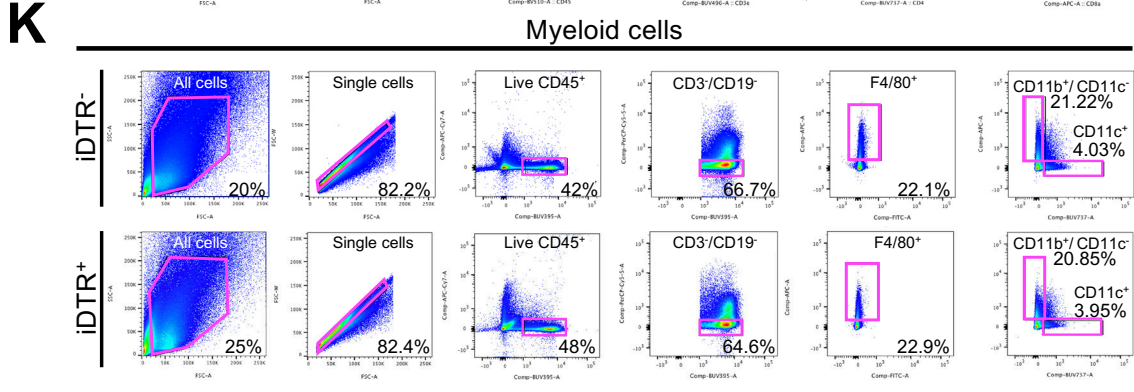
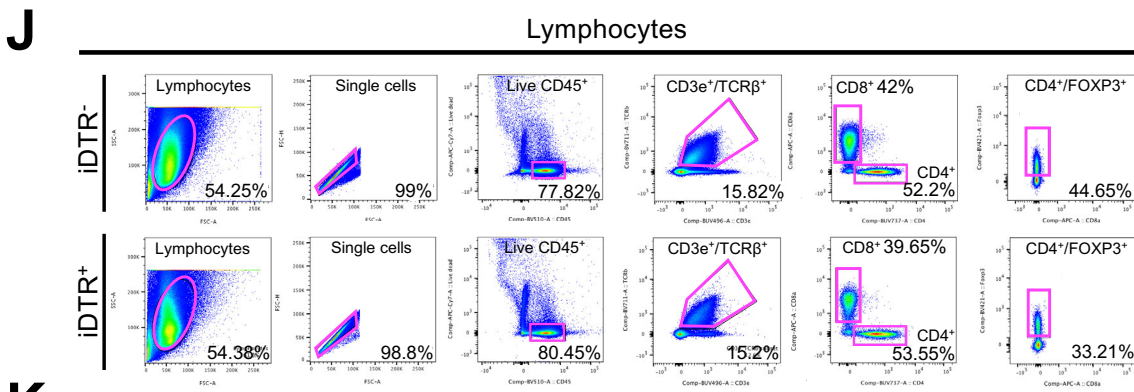


Fig.S3

Figure S3 (Related to Figure 3) | CAF promote ICC. (A) Representative images of confocal microscopy and quantifications showing co-localization of *Colla1*-GFP and α SMA in KRAS/p19-, YAP/AKT-, FBXW7 Δ F/AKT-, NICD/AKT-induced ICC (n=3 per model) in *Lrat*-Cre⁺ TdTom⁺ *Colla1*-GFP⁺ mice. Scale bars, 50 μ m. (B) UMAPs of scRNA-seq showing the normalized expression levels of indicated genes and populations and quantifications of *Colla1* and *Acta2* in KRAS/p19- (n=1) and YAP/AKT-induced ICC (n=3). (C,D) HSC-derived CAF were depleted by injecting diphtheria toxin 6 times (3x/week) in *Lrat*-Cre⁺, TdTom⁺, iDTR⁺ or *Lrat*-Cre⁺, TdTom⁺, iDTR⁻ littermates from week 5 to 7 after tumor induction (late depletion) and HSC depletion was confirmed by sirius red staining in (C) KRAS/p19 (n=13-15 mice/group) and (D) YAP/AKT-induced ICC (n=11-16 iDTR⁻ mice/group). Scale bar 100 μ m. (E-F) For early HSC-derived CAF depletion, *Lrat*-Cre⁺ TdTom⁺ iDTR⁺ mice and *Lrat*-Cre⁺ TdTom⁺ iDTR⁻ littermate controls were injected 6 times with DT (3x/week) starting 3 weeks after the plasmid injection. (E) HSC-derived CAF depletion and repopulation by HSC-CAF or PF-CAF were determined by measuring the TdTom⁺ area and *Lrat* (HSC marker) as well as *Upk3b* (PF marker) mRNA expression in YAP/AKT-induced ICC after the last DT injection (sac1, n=6-7 mice per group) or 2 weeks later (sac2, n=9 mice per group); (F) representative images of whole livers, liver body ratio (LBR), IHC and CK19⁺ quantifications from above groups. Scale bars 1 cm. (G-H) *Pdgfr β* , *Lrat*, *Des*, *Colla1*, *Acta2* mRNA expression and representative images of sirius red and quantifications in *Pdgfr β* ^{f/f} (n=14) and *Pdgfr β* ^{Δ HSC} (n=15) mice in YAP/AKT-induced ICC (G). Scale bar 100 μ m. Representative images of livers, CK19 IHC as well as quantification of LBR and CK19⁺ area from above groups (H). Scale bars 1 cm. (I) Representative micrographs and quantifications of Ki67 IHC from KRAS/p19-induced ICC in α SMA-TK mice (n=15 mice/group). Scale bars 100 μ m. (J,K) Representative gating strategy in the tumors of KRAS/p19-induced ICC, iDTR⁻ (n=4) and iDTR⁺ (n=6) mice for (J) lymphoid and (K) myeloid subsets. (L) Flow cytometric analysis of CD11b⁺, F4/80⁺, CD11b⁺/F4/80⁺, CD11b⁺/CD11c⁻, CD11c⁺, CD11c⁺/CD103⁺ subsets in the tumors from KRAS/p19-induced ICC as well as confirmation of depletion via TdTom⁺ area in *Lrat*-Cre⁺TdTom⁺ iDTR⁻ (n=4) and *Lrat*-Cre⁺TdTom⁺ iDTR⁺ (n=6) mice. (M) Representative micrographs and quantifications of CD3 IHC from KRAS/p19-induced ICC (n=14 iDTR⁻ mice, n=13 iDTR⁺ mice) and YAP/AKT-induced ICC (n=14 iDTR⁻ mice, n=9 iDTR⁺ mice). Scale bars 100 μ m. (N) Subcutaneous co-injections of CGKP19 KRAS/p19 ICC-derived cells alone or with HSC in *Rag2*KO mice. (O) *RelA* mRNA expression in HSC from *RelA*^{f/f} (n=2) and *RelA* ^{Δ HSC} (n=2). (P) Representative images of CK19 IHC, livers, LBR and CK19⁺ area from *RelA*^{f/f} (n=8) and *RelA* ^{Δ HSC} (n=10) from KRAS/p19-induced ICC. For panels A-I, L-P data are shown as mean \pm SEM, and statistical significance was determined using two-sided unpaired T-test (G,H,L,N) or Mann-Whitney (F,H,L,N,P) for groups of two; and by one-way ANOVA followed by Sidak's posthoc test (C,E,F,M,P) or Kruskal-Wallis test with Dunn's posthoc test (A,B,D,G,I) for groups of three.

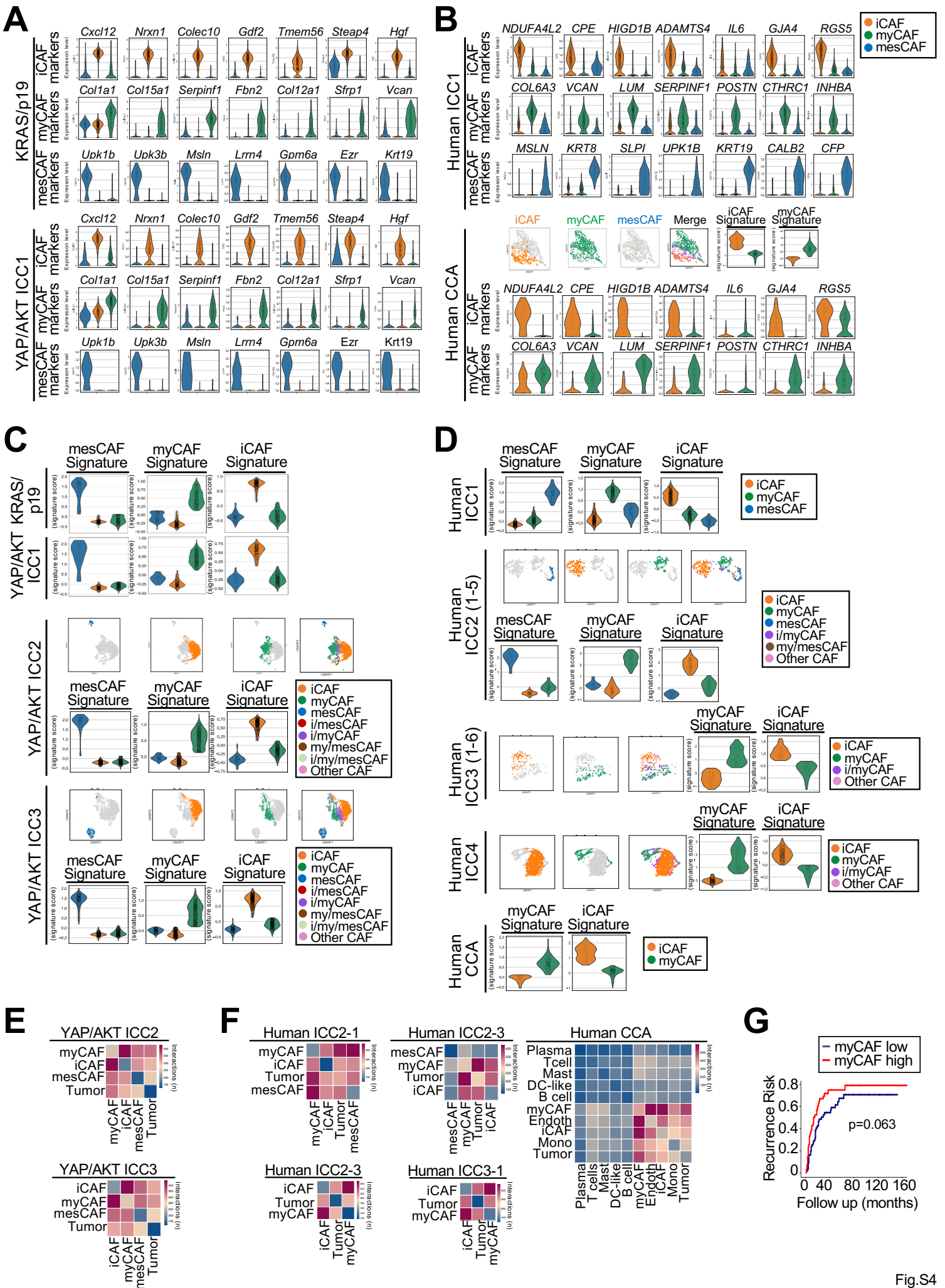


Fig.S4

Figure S4 (Related to Figure 4) | Single cell RNA-sequencing of CAF subpopulations in murine and human ICC. (A,B) Violin plots showing the normalized expression levels of the top genes from iCAF, myCAF and mesCAF signatures in (A) KRAS/p19- , YAP/AKT-induced ICC and (B) human ICC and CCA in one representative sample each. (C,D) Violin plots showing the iCAF, myCAF and mesCAF signature scores in the indicated (C) KRAS/p19- induced ICC, YAP/AKT-induced ICC, (D) human ICC and human CCA samples. The width of each violin plot indicates the kernel density of the expression values. (E,F) Heatmaps showing number of ligand–receptor interactions between iCAF, myCAF, mesCAF and tumor cells in the indicated (E) YAP/AKT-induced ICC, (F) human ICC and human CCA samples, analyzed by CellPhone DB. (G) Recurrence risk in 113 ICC patients with low (n=56) and high (n=57) myCAF signature (Sia et al. cohort).

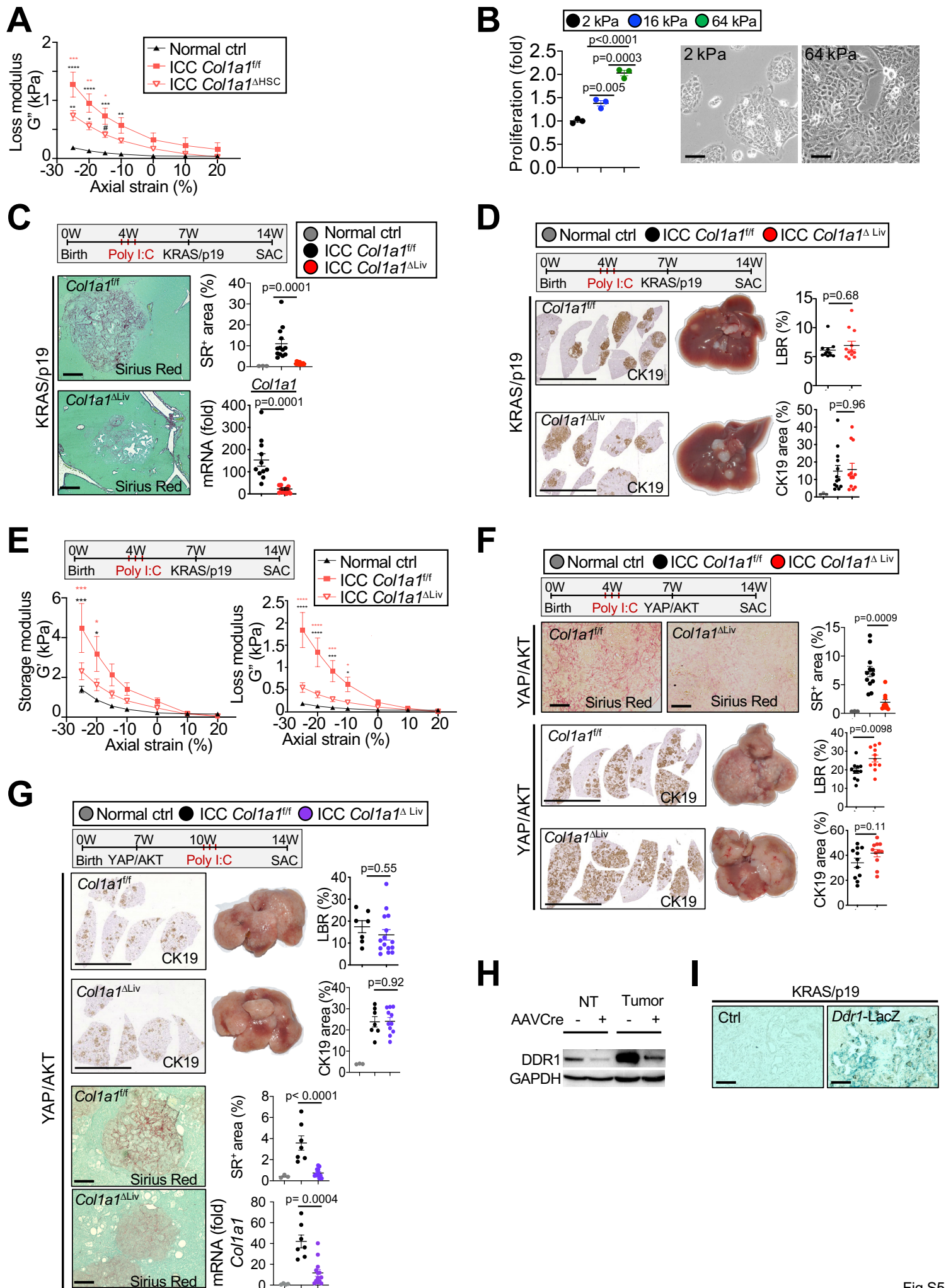


Figure S5 (Related to Figure 5) | CAF promote tumor growth independently of type I collagen. (A) Loss modulus G'' (a measure of viscosity), was determined for tumors from $Collal^{f/f}$ (n=3) and $Collal^{\Delta HSC}$ (n=4) in a KRAS/p19-induced ICC and in control livers (n=2) by shear rheometry. Curves are mean \pm SEM. Using 2-way ANOVA: * $p \leq 0.05$, ** $p \leq 0.01$, **** $p \leq 0.0001$, # $0.05 < p \leq 0.10$ vs ctrl (black) or vs $Collal^{\Delta HSC}$ (red). (B) Proliferation assessed by PicoGreen in HuCCT-1 plated on 2kPa, 16 kPa and 64 kPa plates (n=3 each group) and representative pictures of HuCCT-1 plated on 2kPa and 64 kPa plates. (C-E) Mx1-Cre activation for the deletion of *Collal* was induced by 3 i.p. injections of poly(I:C) before tumor induction starting when mice were 4 weeks old. (C) Representative images and quantification of sirius red staining and tumor *Collal* mRNA expression in $Collal^{f/f}$ (n=13) and $Collal^{\Delta Liv}$ (n=12) in KRAS/p19-induced ICC. Scale bars 100 μ m. (D) Representative images of CK19 IHC, livers, LBR and CK19⁺ area from KRAS/p19-induced ICC for $Collal^{f/f}$ (n= 13) and $Collal^{\Delta Liv}$ (n=12) mice. Scale bars 1 cm. (E) Storage modulus G' (a measure of elasticity) and loss modulus G'' (a measure of viscosity), were determined for tumors from $Collal^{f/f}$ (n=3) and $Collal^{\Delta Liv}$ (n=6) in KRAS/p19 ICC or control liver (n=2) by shear rheometry. Curves are mean \pm SEM. Using 2-way ANOVA: * $p \leq 0.05$, ** $p \leq 0.01$, **** $p \leq 0.0001$, # $0.05 < p \leq 0.10$ vs ctrl (black) or vs $Collal^{\Delta Liv}$ (red). (F) Representative images and quantifications of sirius red staining, livers, LBR and the CK19⁺ area in $Collal^{f/f}$ (n=11) and $Collal^{\Delta Liv}$ (n=11) mice from YAP/AKT-induced ICC. Scale bars 1 cm. (G) Mx1-Cre activation for the deletion of *Collal* was induced by 3 i.p. injections of poly(I:C) starting 3 weeks after tumor induction. Representative images of sirius red staining and livers, LBR, *Collal* mRNA expression and the CK19 and sirius red-positive area in $Collal^{f/f}$ (n=7) and $Collal^{\Delta Liv}$ (n=11) mice from YAP/AKT-induced ICC. Scale bars 1 cm. (H) DDR1 western blot in *Ddr1*^{f/f} NT (n=1) and T (n=1) and (I) representative XGAL staining in *Ddr1*-LacZ reporter (n=1) and wt littermate (n=1) mice in KRAS/p19-induced ICC. Data are shown as mean \pm SEM, and statistical significance was determined by two-sided unpaired T-test (D,F) or Mann-Whitney (E) for groups of two; and by 2-way ANOVA (A,E) or one-way ANOVA (B,G) or Kruskal-Wallis test with Dunn's posthoc test (C,D) for groups of three.

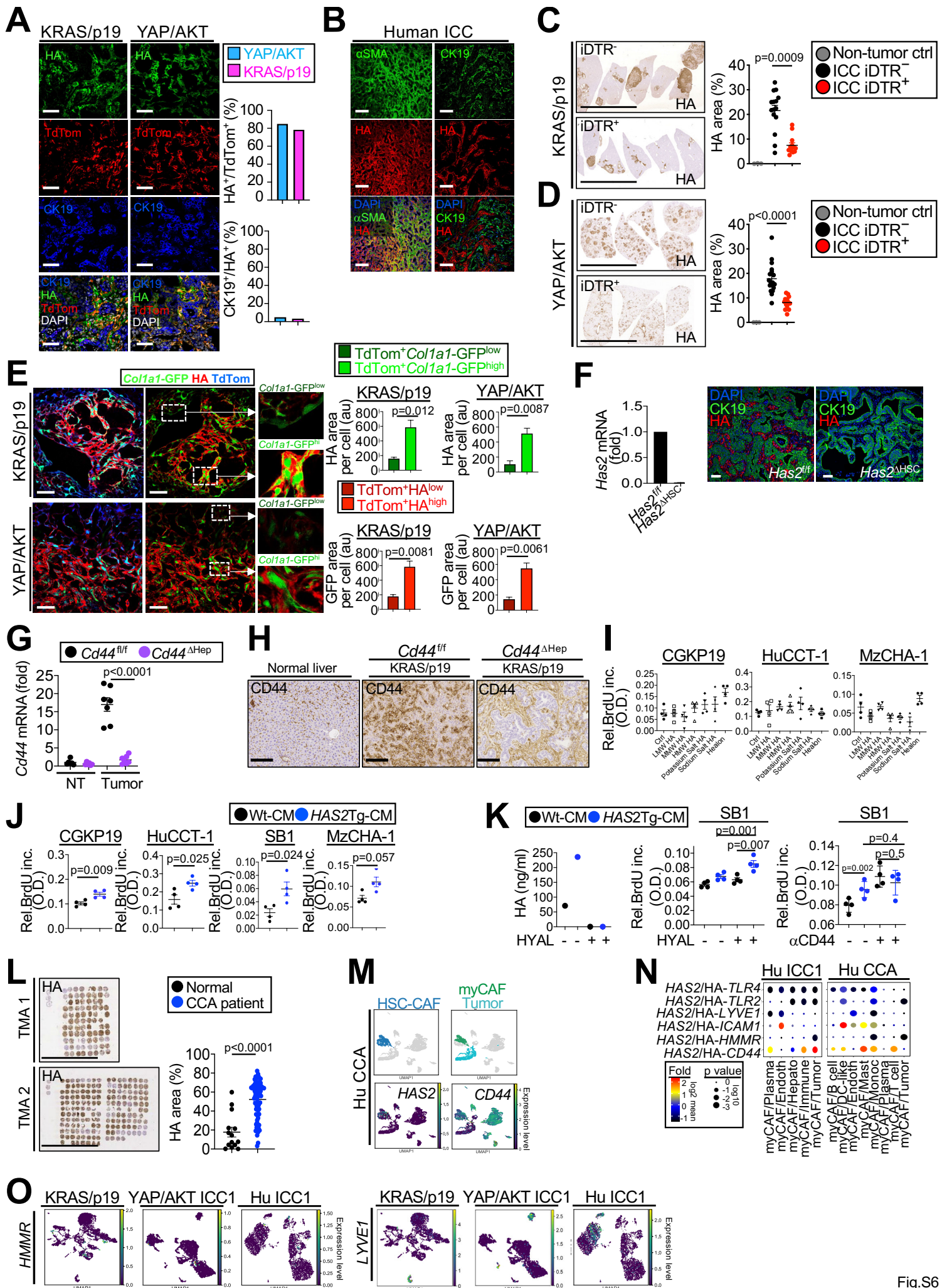


Fig.S6

Figure S6 (Related to Figure 6) | myCAF-derived HAS2 sustain tumor growth in ICC. (A) Representative micrograph and quantifications of confocal microscopy show co-localization of *Lrat*-Cre-induced TdTom (HSC derived-CAF) with HA and absent co-localization with CK19 in KRAS/p19- (n=1) and YAP/AKT- (n=1) ICC. (B) Confocal microscopy in human ICC (n=1) showed co-localization of HA with α SMA but not with CK19. Scale bars, 50 μ m. (C,D) Representative micrographs of HA IHC and quantification in CAF-depleted mice from (C) KRAS/p19-ICC (n=15 iDTR⁻, n=13 iDTR⁺ mice) and (D) YAP/AKT-induced ICC (n=16 iDTR⁻, n=11, iDTR⁺ mice). Scale bars 1 cm. (E) Representative micrograph and quantifications of confocal microscopy show high expression of myCAF HA in cells with high expression of myCAF marker *Colla1*-GFP and low expression of myCAF HA in cells with low expression of myCAF marker *Colla1*-GFP, in KRAS/p19- and YAP/AKT-induced ICC (n=3 tumors per group). Scale bars, 50 μ m. (F) *Has2* mRNA expression in HSC isolated from *Has2*^{f/f} (n=1) and *Has2*^{ΔHSC} (n=2) and representative confocal microscopy showing absent HA in *Has2*^{ΔHSC} mice. Scale bars, 50 μ m. (G) *Cd44* mRNA expression in tumor (T) (n=6 per group) and non-tumor (NT) (n=7 per group) tissue from *Cd44*^{f/f} and *Cd44*^{ΔHep} mice. (H) Representative micrographs of CD44 IHC in normal liver and tumor area in KRAS/p19-induced ICC in *Cd44*^{f/f} and *Cd44*^{ΔHep} mice. (I) BrdU incorporation assay in mouse (CGKP19) and human (HuCCT1 and MzCHA) tumor cells treated with different types of HA. (J) BrdU incorporation assay in human and mouse tumor cells treated with conditioned medium (CM) from *HAS2*-Tg and littermate wt mice (n=4/group). (K) HA content in the CM from *HAS2*-Tg and littermate control wt mice by ELISA in presence or absence of hyaluronidase (HYAL) (n=1 per condition, left panel) and BrdU incorporation assay in SB1 cells treated with CM from *HAS2*-Tg or littermate wt mice in the presence or absence of HYAL or after pre-treatment with CD44 antibody or IgG control (n=4 per condition, middle and right panel). (L) Representative micrographs and quantifications of HA-stained area of human CCA TMAs (n=19 ICC and n=58 ECC) and normal liver (n=14). Scale bars 1 cm. (M) UMAPs showing the normalized expression levels of *HAS2* and *CD44* in human CCA (n=1). (N) Representative CellphoneDB heatmap showing interactions of *HAS2*-expressing myCAF with HA-receptor expressing cells in human ICC and in CCA. (O) UMAPs showing the normalized expression levels of *HMMR* and *LYVE1* in KRAS/p19-, YAP/AKT-induced ICC and in human ICC. For panels C-E, G, I-L; data are shown as mean \pm SEM, statistical significance was determined by two-sided unpaired T-test (J) or Mann-Whitney (J, L) groups of two; and by one-way ANOVA followed by Sidak's posthoc test (D, E, K, G, I) or Kruskal-Wallis test with Dunn's posthoc test (C, I) groups of three.

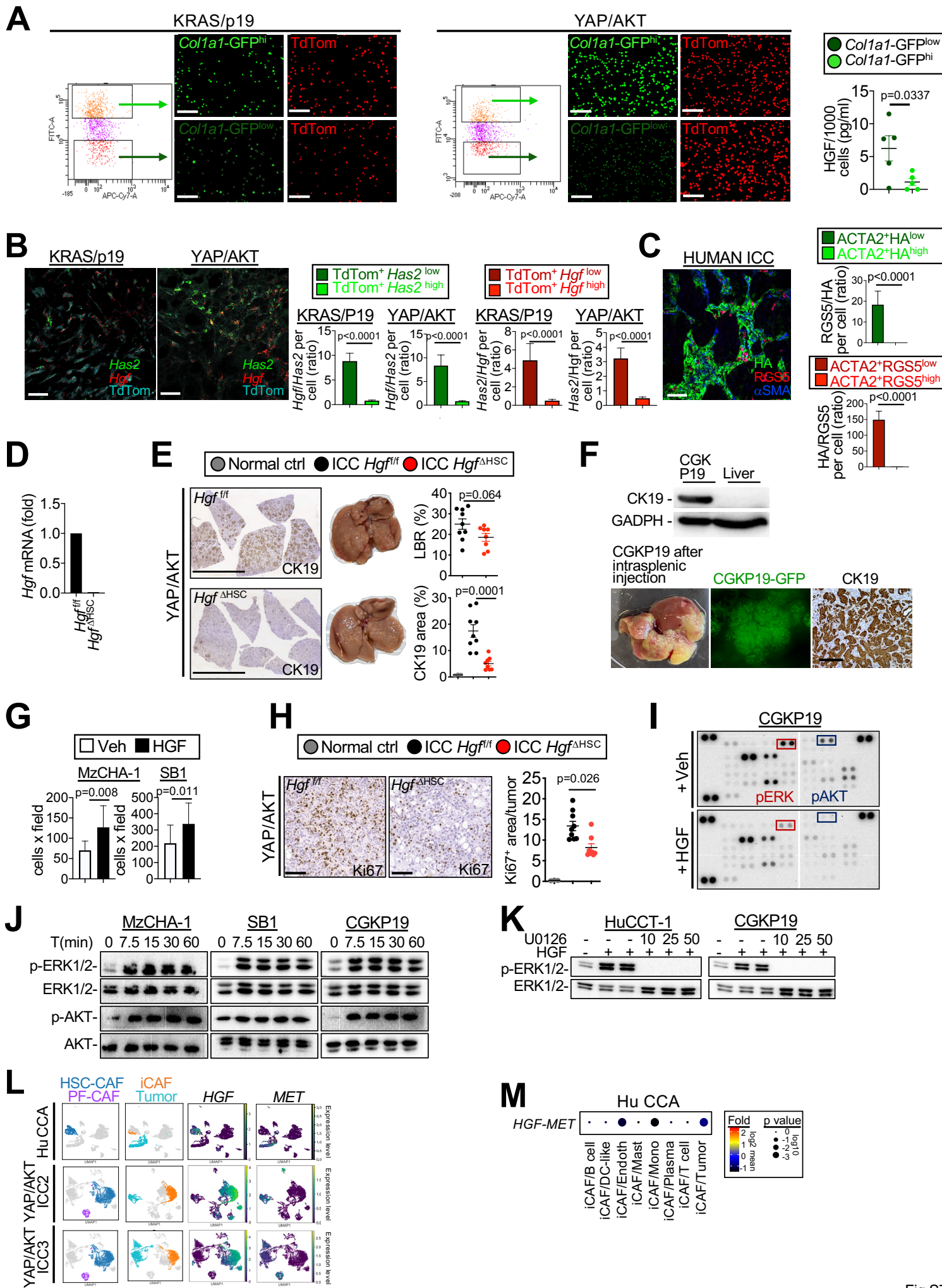


Fig.S7

Figure S7 (Related to Figure 7) | iCAF-derived HGF promotes ICC growth. (A) Representative gating strategy and pictures showing the *Lrat-Cre⁺TdTom⁺Colla1-GFP^{high}* (myCAF) and *Lrat-Cre⁺TdTom⁺Colla1-GFP^{low}* (iCAF) cells sorted and freshly plated from tumors of KRAS/p19- (n=3) and YAP/AKT-induced ICC (n=2); and ELISA showing that *Lrat-Cre⁺TdTom⁺Colla1-GFP^{low}* iCAF produce more HGF compared to the *Lrat-Cre⁺TdTom⁺Colla1-GFP^{high}* myCAF 24 hours after plating. Scale bars, 100 μ m. (B) Confocal microscopy representative images of RNAscope showing *Has2* and *Hgf* in *Lrat-Cre⁺TdTom⁺* mice in KRAS/p19- and YAP/AKT-induced ICC. Scale bars, 50 μ m. (C) Representative fluorescent confocal microscopy image of iCAF marker RGS5 and myCAF marker HA and quantifications in human ICC (n=1). Scale bars, 50 μ m. (D) *Hgf* mRNA expression in HSC from *Hgf^{f/f}* (n=1) and *Hgf ^{Δ HSC}* (n=2). (E) Representative images of CK19 immunohistochemistry, livers, LBR and CK19⁺ area from *Hgf^{f/f}* (n=9) and *Hgf ^{Δ HSC}* (n=8) livers in YAP/AKT-induced ICC. Scale bars 1 cm. (F) CK19 western blot in the CGKP19 cell line (n=1) and whole liver (n=1) and representative picture of liver, CGKP19 native fluorescence and CK19 immunohistochemistry in CGKP19 spleen-injected mouse. Scale bar 100 μ m. (G) Human MzCHA-1 and murine SB1 cell lines were treated with recombinant human or mouse HGF (25 ng/ml, n=5) or vehicle (n=5) and cell number was determined after 48h. (H) Representative micrographs of Ki67 IHC in *Hgf^{f/f}* and *Hgf ^{Δ HSC}* in YAP/AKT-induced ICC. Scale bars 100 μ m. (I) CGKP19 cells were treated with mouse HGF, followed by phospho-kinase array. (J,K) Human (MzCHA1) and murine (SB1 and CGKP19) CCA cell lines were treated with (J) recombinant HGF (K) in the presence of MEK1/2 inhibitor U0126 or vehicle and were tested by western blot for phosphorylated and total ERK1/2. (L) UMAPs showing the normalized expression levels of *HGF* and *MET* in human CCA (n=1) and in YAP/AKT-induced ICC (n=2). (M) CellphoneDB showing interaction of *HGF*-expressing iCAF with *MET*-expressing cells in human CCA (n=1). For panels A-C,E,G,H; data are shown as mean \pm SEM, statistical significance was determined by two-sided unpaired T-test (A,E,G) or Mann-Whitney (B,C) groups of two; and by one-way ANOVA followed by Sidak's posthoc test (E) or Kruskal-Wallis test with Dunn's posthoc test (H) groups of three.

Table S1 (Related to Figure 1 and Figure S1). Description of mouse and human scRNA sequencing samples used for CAF analysis

MOUSE SAMPLES scRNA-seq (GSE154170)

Sample ID	ICC model	Enrichment	HSC-CAF PF-CAF% analysis	iCAF, myCAF, mesCAF% analysis	CellphoneDB analysis	Total CAF number	HSC-CAF number	PF-CAF number	Non-PF, non-HSC other CAF number	iCAF number	myCAF number	mesCAF number	multiCAF number	Non- i/my/ mesCAF other CAF number
YAP/AKT ICC1	YAP/AKT plasmids HDTV1*	100% Col1a1-GFP+	✓	✓	✓	1830	1788	42	none	790	835	41	95	69
YAP/AKT ICC2	YAP/AKT plasmids HDTV1*	70% Col1a1-GFP+ and 30% unpurified tumor suspension	✓	✓	✓	3256	3117	139	none	1747	1026	137	282	64
YAP/AKT ICC3	YAP/AKT plasmids HDTV1*	100% Col1a1-GFP+	✓	✓	✓	4683	4076	607	none	2183	1262	540	563	135
KRAS/p19	KRAS/p19 plasmids HDTV1*	70% Col1a1-GFP+ and 30% unpurified tumor suspension	✓	✓	✓	2662	2324	149	189	1350	686	204	367	55

*HDTV1= Hydrodynamic tail vein injection

HUMAN SAMPLES scRNA-seq

Sample ID	Source paper	Source Sample ID	HSC-CAF PF-CAF% analysis	iCAF, myCAF, mesCAF% analysis	CellphoneDB analysis	Total CAF number	HSC-CAF number	PF-CAF number	Non-PF, non-HSC other CAF number	iCAF number	myCAF number	mesCAF number	multiCAF number	Non- i/my/ mesCAF other CAF number
Human ICC1	Zhang et al., 2020	GSE142784, GSM4240155 ICC24S	✓	✓	✓	2574	2397	177	0	1413	781	144	49	187
Human ICC2-1 *	Zhang et al., 2020	GSE138709, GSM4116585	✓	✓	✓	124	74	50	0	42	21	43	8	10
Human ICC2-2 *	Zhang et al., 2020	GSE138709, GSM4116584	✓	✓	✓	218	199	19	0	98	78	3	34	5
Human ICC2-3	Zhang et al., 2020	GSE138709, GSM4116583	✓	✓	✓	42	42	0	0	24	12	0	6	0
Human ICC2-4	Zhang et al., 2020	GSE138709, GSM4116581				14	14	0	0	2	11	0	0	1
Human ICC2-5	Zhang et al., 2020	GSE138709, GSM4116580				11	11	0	0	6	0	0	0	5
Human ICC3-1	Ma et al., 2019	GSE125449 Set1, S11_P06_LCP29	✓	✓	✓	150	150	0	0	62	37	0	31	20
Human ICC3-2	Ma et al., 2019	GSE125449 Set1, S09_P04_LCP25				24	24	0	0	3	15	0	1	5
Human ICC3-3	Ma et al., 2019	GSE125449 Set1, S08_P03_LCP26				13	13	0	0	2	8	0	1	2
Human ICC3-4	Ma et al., 2019	GSE125449 Set1, S12_P07_LCP30				16	16	0	0	2	10	0	2	2
Human ICC3-5	Ma et al., 2019	GSE125449 Set1, S20_P12_LCP35				37	37	0	0	8	17	0	5	7
Human ICC3-6	Ma et al., 2019	GSE125449 Set1, S19_P11_LCP39				12	12	0	0	1	8	0	3	0
Human ICC4	Zhang et al., 2020	GSE142784, GSM4240156 ICC32S	✓	✓		1228	1228	0	0	905	172	0	72	79

* Different tumors from same patient

<i>Acot7</i>	<i>Bpgm</i>	<i>Col4a5</i>	<i>Dap</i>	<i>Frem1</i>	<i>Impdh1</i>	<i>Mdk</i>	<i>Olfml2b</i>	<i>Plpp5</i>	<i>Scpep1</i>	<i>Speccl</i>	<i>Tubb3</i>
<i>Acta2</i>	<i>Capg</i>	<i>Col5a2</i>	<i>Dbp</i>	<i>Fst</i>	<i>Inhba</i>	<i>Medag</i>	<i>P4ha2</i>	<i>Pmepa1</i>	<i>Sac1</i>	<i>Spp1</i>	<i>Tyms</i>
<i>Actg2</i>	<i>Cd59a</i>	<i>Col5a3</i>	<i>Ddah1</i>	<i>Fstl1</i>	<i>Im2a</i>	<i>Meox1</i>	<i>Pafah1b3</i>	<i>Podxl2</i>	<i>Serf1</i>	<i>Srpx</i>	<i>Uck2</i>
<i>Adam19</i>	<i>Cd9</i>	<i>Col6a2</i>	<i>Dpys13</i>	<i>Fstl3</i>	<i>Kctd11</i>	<i>Mfap2</i>	<i>Palld</i>	<i>Prrx1</i>	<i>Serpine2</i>	<i>Ste2</i>	<i>Vcan</i>
<i>Adcy7</i>	<i>Cd93</i>	<i>Col7a1</i>	<i>Ebf1</i>	<i>Fxyd5</i>	<i>Kdelr3</i>	<i>Mfap4</i>	<i>Pcbp4</i>	<i>Prss35</i>	<i>Serpinf1</i>	<i>Stk17b</i>	<i>Vegfa</i>
<i>Ahna2</i>	<i>Cda</i>	<i>Col8a1</i>	<i>Egln3</i>	<i>Fxyd6</i>	<i>Kij26b</i>	<i>Mgll</i>	<i>Pdgfr1</i>	<i>Ptgfrn</i>	<i>Sesn3</i>	<i>Sulfl</i>	<i>Vmp1</i>
<i>Ak1</i>	<i>Cdh11</i>	<i>Colec12</i>	<i>Emp1</i>	<i>Fzd1</i>	<i>Kremen1</i>	<i>Mgp</i>	<i>Pdlim3</i>	<i>Pik7</i>	<i>Sfrp1</i>	<i>Tfpi2</i>	<i>Wisp1</i>
<i>Akr1b8</i>	<i>Cdkn2a</i>	<i>Comtd1</i>	<i>Eno1</i>	<i>Gas7</i>	<i>Lama4</i>	<i>Mical2</i>	<i>Pdzrn3</i>	<i>Ptn</i>	<i>Sh3bgr13</i>	<i>Tgfb3</i>	<i>Wnt11</i>
<i>Aldh1a2</i>	<i>Cdkn2b</i>	<i>Cop2</i>	<i>Enpp1</i>	<i>Gm15867</i>	<i>Lef1</i>	<i>Minos1</i>	<i>Pgam1</i>	<i>Ptprb</i>	<i>Sh3pxd2b</i>	<i>Thy1</i>	<i>Zswin7</i>
<i>Ank</i>	<i>Cercam</i>	<i>Col1</i>	<i>Env4</i>	<i>Gng11</i>	<i>Lefly1</i>	<i>Mmp14</i>	<i>Pgf</i>	<i>Rab7b</i>	<i>Shisa4</i>	<i>Thyn1</i>	
<i>Ankrd37</i>	<i>Ch25h</i>	<i>Cox4i2</i>	<i>Evl</i>	<i>Gpar1</i>	<i>Lgals1</i>	<i>Mmp2</i>	<i>Pgk1</i>	<i>Rap2b</i>	<i>Siva1</i>	<i>Timp1</i>	
<i>Anpep</i>	<i>Chst11</i>	<i>Cpxm2</i>	<i>Fam198b</i>	<i>Gpc1</i>	<i>Lgi2</i>	<i>Mtch1</i>	<i>Piezo2</i>	<i>Rapgef4</i>	<i>Slc16a3</i>	<i>Tm4sf1</i>	
<i>Aoc1</i>	<i>Chst2</i>	<i>Crjfl</i>	<i>Fam20a</i>	<i>Gpm6b</i>	<i>Lhfp12</i>	<i>Nbl1</i>	<i>Pik3ip1</i>	<i>Rgs16</i>	<i>Slc27a1</i>	<i>Tmem2</i>	
<i>Apol9a</i>	<i>Clecl1a</i>	<i>Csrp2</i>	<i>Fam20c</i>	<i>Gpr153</i>	<i>Lsp1</i>	<i>Ncam1</i>	<i>Pkn</i>	<i>Rgs19</i>	<i>Slc39a14</i>	<i>Tmem45a</i>	
<i>Aprt</i>	<i>Clmp</i>	<i>Ctcr1</i>	<i>Fam69b</i>	<i>Hlf0</i>	<i>Ltbp2</i>	<i>Ndufa412</i>	<i>Pla1a</i>	<i>Rnf149</i>	<i>Slc7a2</i>	<i>Tnc</i>	
MOUSE mesCAF											
<i>Adgrd1</i>	<i>Cd2ap</i>	<i>Flr2</i>	<i>Igfbp6</i>	<i>Mmd</i>	<i>Rbbp8</i>	<i>Slc9a3r1</i>	<i>Tmem98</i>				
<i>Angptl7</i>	<i>Celf2</i>	<i>Fth1</i>	<i>Il18</i>	<i>Msln</i>	<i>Rspo1</i>	<i>Sipi</i>	<i>Tnfrsf12a</i>				
<i>Anxa3</i>	<i>Cpe</i>	<i>Gas6</i>	<i>Krt18</i>	<i>Ndr1</i>	<i>S100a1</i>	<i>Smpd3</i>	<i>Tnfrsf12a</i>				
<i>Atp1b1</i>	<i>Cxadr</i>	<i>Gm20186</i>	<i>Krt7</i>	<i>Ndufa4</i>	<i>Saa3</i>	<i>Snrpg</i>	<i>Trf</i>				
<i>Bcd1</i>	<i>Dab2</i>	<i>Gm8186</i>	<i>Krt8</i>	<i>Nkain4</i>	<i>Sbsn</i>	<i>Spint2</i>	<i>Ucp2</i>				
<i>C3</i>	<i>Ddr1</i>	<i>Gpc3</i>	<i>Ldhb</i>	<i>Pdgfc</i>	<i>Serpinh6b</i>	<i>Simn1</i>	<i>Upk1b</i>				
<i>Cd151</i>	<i>Ezr</i>	<i>Gpm6a</i>	<i>Lgals3</i>	<i>Pdpn</i>	<i>Shb</i>	<i>Tcea3</i>	<i>Upk3b</i>				
<i>Cd200</i>	<i>Fbxo33</i>	<i>Hspb1</i>	<i>Lrrn4</i>	<i>Rab11a</i>	<i>Slc39a8</i>	<i>Tmem151a</i>	<i>W1</i>				
HUMAN iCAF											
<i>ADAMTS4</i>	<i>AGT</i>	<i>APOE</i>	<i>ARHGDI1B</i>	<i>CCL19</i>	<i>CCL21</i>	<i>COLEC11</i>	<i>CPE</i>	<i>GEM</i>	<i>GJA4</i>		
<i>GPX3</i>	<i>HIGD1B</i>	<i>IL6</i>	<i>ISYNA1</i>	<i>LHFP</i>	<i>MAP1B</i>	<i>MT1A</i>	<i>NDUFA4L2</i>	<i>PKD4</i>	<i>RGS5</i>		
HUMAN myCAF											
<i>APOD</i>	<i>CCL11</i>	<i>COL1A1</i>	<i>COL1A2</i>	<i>COL3A1</i>	<i>COL5A1</i>	<i>COL6A3</i>	<i>CTGF</i>	<i>CTHRC1</i>	<i>CYP11B1</i>	<i>FNI</i>	
<i>INHBA</i>	<i>ISLR</i>	<i>LUM</i>	<i>MMP14</i>	<i>POSTN</i>	<i>PTGDS</i>	<i>SERPINF1</i>	<i>SFRP2</i>	<i>SPON2</i>	<i>VCAN</i>		
HUMAN mesCAF											
<i>ANXA1</i>	<i>ANXA2</i>	<i>BDKRB1</i>	<i>C19orf33</i>	<i>C3</i>	<i>CALB2</i>	<i>CCDC80</i>	<i>CFB</i>	<i>CRABP2</i>			
<i>CXCL1</i>	<i>CXCL6</i>	<i>EFEMP1</i>	<i>EGFL6</i>	<i>EMP3</i>	<i>EZR</i>	<i>HMOX1</i>	<i>HP</i>	<i>HSPA6</i>			
<i>IFI27</i>	<i>IGFBP6</i>	<i>ITLN1</i>	<i>KRT18</i>	<i>KRT19</i>	<i>KRT8</i>	<i>LINC01133</i>	<i>LOX</i>	<i>MT1E</i>			
<i>MT1G</i>	<i>MT1X</i>	<i>MXRA5</i>	<i>PDPN</i>	<i>PLA2G2A</i>	<i>PRG4</i>	<i>PRSS23</i>	<i>PTGIS</i>	<i>RP11-572C15.6</i>			
<i>S100A10</i>	<i>S100A16</i>	<i>S100A6</i>	<i>SAA1</i>	<i>SAA2</i>	<i>SERPINE2</i>	<i>SH3BGL3</i>	<i>SLC12A8</i>	<i>SLPI</i>	<i>TM4SF1</i>		

epithelial cell proliferation	GO:0050673	429	21354	1	81	6	38	5	0.005504332	59	9	0.26239298	94	9
ovulation cycle	GO:0042698	84	21354	0.003359416	90	6	34	1	1	50	3	0.268790672	64	4
Drug metabolism - other enzymes	KEGG:00983	88	8477	0.766693576	1	1	12	3	0.105508027	22	3	0.312597491	32	3
response to organic substance	GO:0010033	3434	21354	0.054186759	96	32	41	18	0.02613305	85	30	0.339735166	87	28
retinoic acid binding	GO:0001972	16	21091	0.412292076	1	1	1	1	0.006497501	7	2	0.412292076	1	1
response to vitamin D	GO:0033280	34	21354	1	73	2	33	2	0.028210783	91	4	0.463704759	72	3
enzyme linked receptor protein signaling pathway	GO:0007167	918	21354	1	50	8	45	8	0.030949112	96	15	0.615961375	85	12
establishment of planar polarity	GO:0001736	63	21354	1	-1	-1	46	2	0.023886084	99	5	0.675713007	44	3
establishment of tissue polarity	GO:0007164	64	21354	1	-1	-1	46	2	0.025815633	99	5	0.707927442	44	3
organic acid metabolic process	GO:0006082	948	21354	1	100	12	59	13	1	73	10	0.711685349	50	9
response to nutrient	GO:0007584	191	21354	1	73	4	35	5	0.038299754	91	7	0.820203001	38	4
retinoid binding	GO:0005501	34	21091	0.876120661	1	1	35	2	6.39148E-05	73	5	0.876120661	1	1
cellular response to organic substance	GO:0071310	2461	21354	0.074678934	51	17	41	16	0.142191115	85	23	0.88319428	72	19
response to vitamin	GO:0033273	105	21354	1	73	2	35	3	0.012639842	91	6	0.998213458	72	4
isoprenoid binding	GO:0019840	39	21091	1	1	1	1	1	0.000130485	73	5	1	1	1
oxidoreductase activity, acting on paired donors, with incorporation or reduction of molecular oxygen	GO:0016705	221	21091	0.930457494	82	5	87	7	1	73	2	1	80	4
hyaluronan synthase activity	GO:0050501	4	21091	0.024150363	58	2	34	1	1	44	1	1	26	1
monooxygenase activity	GO:0004497	160	21091	1	29	2	87	6	1	73	2	1	76	3
cell morphogenesis involved in differentiation	GO:0000904	767	21354	1	76	7	94	10	0.00096158	87	15	1	98	11
midgut development	GO:0007494	12	21354	1	19	1	51	3	1	34	1	1	97	2
cellular response to growth factor stimulus	GO:0071363	671	21354	1	94	8	34	6	0.029504316	85	12	1	71	8
cellular response to endogenous stimulus	GO:0071495	1329	21354	0.315544868	48	11	38	10	0.034857925	85	17	1	64	11
cell development	GO:0048468	2361	21354	1	90	17	86	14	0.035493224	87	24	1	98	21
rhythmic process	GO:0048511	347	21354	0.035959652	90	9	83	2	1	82	5	1	64	4
monocarboxylic acid metabolic process	GO:0032787	616	21354	1	100	9	95	12	1	73	8	1	50	7
response to extracellular stimulus	GO:0009991	566	21354	1	6	2	72	10	1	55	7	1	38	6
digestive tract development	GO:0048565	138	21354	1	19	1	51	5	1	68	4	1	38	3
response to growth factor	GO:0070848	698	21354	1	58	6	34	6	0.043605217	85	12	1	71	8
extracellular polysaccharide metabolic process	GO:0046379	3	21354	0.049348422	58	2	34	1	1	44	1	1	26	1
extracellular polysaccharide biosynthetic process	GO:0045226	3	21354	0.049348422	58	2	34	1	1	44	1	1	26	1
Factor: GLI; motif: NSTGGGTGGTCY	TF:M10254	9881	21859	1	94	53	90	61	0.022650143	82	56	1	90	55
Factor: Smad2; motif: NCTGYCTGN	TF:M10366	6664	21859	1	82	37	88	45	1	95	40	1	45	22

cell differentiation	GO:0030154	0.038207701	1.4178491	4248	34	18	21257	LEF1,PTN,HAS2,MEOX1,CDKN2A,MEGF10,SERPINF1,COL8A1,CDKN2B,FBN2,ROR2,COL7A1,ETV4,SFRP1,WNT11,TMEFF1,CSPG4,FST
gland morphogenesis	GO:0022612	0.038280124	1.4170267	139	25	4	21257	PTN,CDKN2A,ETV4,SFRP1
negative regulation of multicellular organismal process	GO:0051241	0.040857245	1.3887309	1326	33	10	21257	LEF1,PTN,CDKN2A,SERPINF1,CDKN2B,ETV4,SFRP1,WNT11,ADCY7,CSPG4
epithelial to mesenchymal transition	GO:0001837	0.042519125	1.3714157	137	26	4	21257	LEF1,HAS2,SFRP1,WNT11
negative regulation of biological process	GO:0048519	0.047486952	1.3234257	5477	26	17	21257	NOTUM,DKK2,LEF1,PTN,CRLF1,ITIH5,CDKN2A,SERPINF1,NKD2,CDKN2B,FBN2,ROR2,COL7A1,GFRA2,ETV4,SFRP1,WNT11
cellular response to platelet-derived growth factor stimulus	GO:0036120	0.048107092	1.3177909	28	6	2	21257	PTN,HAS2
collagen-containing extracellular matrix	GO:0062023	1.87E-11	10.727259	339	43	13	21187	PTN,ITIH5,COL15A1,SERPINF1,COL8A1,FBN2,COL7A1,SFRP1,TMEFF1,CSPG4,S100A6,MFAP4,PCOLCE2
extracellular matrix	GO:0031012	3.25E-11	10.487735	452	43	14	21187	PTN,ITIH5,COL15A1,SERPINF1,COL8A1,FBN2,COL7A1,SFRP1,WNT11,TMEFF1,CSPG4,S100A6,MFAP4,PCOLCE2
extracellular region	GO:0005576	5.76E-07	6.2396066	2713	44	22	21187	NOTUM,DKK2,PTN,CRLF1,ITIH5,COL15A1,SERPINF1,AMN,COL8A1,FBN2,COL7A1,SFRP1,WNT11,PRSS35,TMEFF1,CSPG4,FST,S100A6,PLA1A,MFAP4,PCOLCE2,TNFRSF9
basement membrane	GO:0005604	2.18E-06	5.6623636	104	32	6	21187	PTN,COL15A1,SERPINF1,COL8A1,COL7A1,TMEFF1
extracellular space	GO:0005615	0.003622364	2.4410079	1885	44	14	21187	DKK2,PTN,CRLF1,COL15A1,SERPINF1,AMN,COL8A1,COL7A1,SFRP1,WNT11,FST,PLA1A,MFAP4,TNFRSF9
collagen trimer	GO:0005581	0.022181242	1.6540141	88	22	3	21187	COL15A1,COL8A1,COL7A1
microfibril	GO:0001527	0.039580835	1.4025151	11	39	2	21187	FBN2,MFAP4
Wnt signaling pathway	KEGG:04310	5.64E-05	4.248439	152	26	7	6926	NOTUM,DKK2,LEF1,NKD2,ROR2,SFRP1,WNT11
Cushing syndrome	KEGG:04934	0.017493086	1.7571336	146	29	5	6926	LEF1,CDKN2A,CDKN2B,WNT11,ADCY7
Collagen chain trimerization	C:R-MMU-8948	0.026137604	1.5827342	38	22	3	8671	COL15A1,COL8A1,COL7A1
Collagen biosynthesis and modifying enzymes	C:R-MMU-1650	0.045313253	1.3437748	59	43	4	8671	COL15A1,COL8A1,COL7A1,PCOLCE2
Factor: AP-2; motif: SNNCCNCAGGCN; match class: 1	TF:M00915_1	0.010591738	1.9750328	2395	48	17	21846	NOTUM,LEF1,LRRC55,CRLF1,SERPINF1,NKD2,CDKN2B,FBN2,ROR2,ADCY7,ACTG2,TMEFF1,FST,MFAP4,VCAN,CPXM2,DOK5
Factor: Smad4; motif: NCAGACAN; match class: 0	TF:M07368_0	0.010657189	1.9723573	5290	40	23	21846	DKK2,LRRC55,HAS2,CRLF1,MEOX1,CDKN2A,MEGF10,SERPINF1,COL8A1,NKD2,COL7A1,ETV4,SFRP1,WNT11,D630003M21RIK,PRSS35,ADCY7,ACTG2,TMEFF1,CSPG4,S100A6,MFAP4,LSP1
Factor: Smad4; motif: NCAGACAN	TF:M07368	0.010657189	1.9723573	5290	40	23	21846	DKK2,LRRC55,HAS2,CRLF1,MEOX1,CDKN2A,MEGF10,SERPINF1,COL8A1,NKD2,COL7A1,ETV4,SFRP1,WNT11,D630003M21RIK,PRSS35,ADCY7,ACTG2,TMEFF1,CSPG4,S100A6,MFAP4,LSP1
Factor: Sp3; motif: ASMCTGGGSRGGG; match class: 1	TF:M00665_1	0.018213747	1.7396007	1757	41	13	21846	LEF1,CRLF1,MEOX1,MEGF10,FBN2,INSC,COL7A1,ETV4,WNT11,CSPG4,FST,NREP,VCAN
Factor: FKLf; motif: BGGGNGGVMd; match class: 1	TF:M01837_1	0.026764655	1.5724384	985	27	8	21846	DKK2,LEF1,HAS2,NKD2,ROR2,GFRA2,ETV4,D630003M21RIK
Factor: WT1; motif: SMCNCCNSC	TF:M01118	0.032278059	1.4910926	8133	27	21	21846	NOTUM,DKK2,LEF1,HAS2,CRLF1,MEOX1,ITIH5,CDKN2A,MEGF10,AMN,NKD2,CDKN2B,FBN2,INSC,ROR2,COL7A1,GFRA2,ETV4,SFRP1,WNT11,D630003M21RIK
Factor: WT1; motif: SMCNCCNSC; match class: 0	TF:M01118_0	0.032278059	1.4910926	8133	27	21	21846	NOTUM,DKK2,LEF1,HAS2,CRLF1,MEOX1,ITIH5,CDKN2A,MEGF10,AMN,NKD2,CDKN2B,FBN2,INSC,ROR2,COL7A1,GFRA2,ETV4,SFRP1,WNT11,D630003M21RIK

Table S6 (Related to Figure 6). Combined top DGE core matrisome filtered CAF YAP/AKT and KRAS/p19 versus quiescent HSC.

Gene	baseMean	log2FoldChange YAP/AKT	lfcSE	stat	pvalue	padj	baseMean	log2FoldChange KRAS/p19	lfcSE	stat	pvalue	padj	Sum log2Fold Kras/p19 and YAP/AKT	
Mgp	9960.9976	13.3806	0.6823	-19.6120	1.22E-85	1.11E-82	4142.7631	12.1766	0.5451	-22.3367	1.63E-110	1.36E-107	25.5572	Core Matrisome
Col6a5	2969.9244	13.1437	1.1259	-11.6744	1.72E-31	1.79E-29	967.4053	11.6372	0.9047	-12.8630	7.27E-38	6.65E-36	24.7809	Core Matrisome
Spp1	15673.0196	12.0905	0.5016	-24.1062	2.15E-128	9.40E-125	15253.7126	12.1529	0.4242	-28.6461	1.79E-180	4.34E-177	24.2434	Core Matrisome
Col6a6	963.6615	12.6791	1.3962	-9.0814	1.07E-19	4.95E-18	231.4441	10.6974	1.6731	-6.3936	1.62E-10	2.27E-09	23.3765	Core Matrisome
Coll5a1	15353.4502	11.2533	0.8004	-14.0593	6.76E-45	1.45E-42	12457.9640	11.1875	0.8002	-13.9810	2.04E-44	2.28E-42	22.4408	Core Matrisome
Mfap4	2391.5150	11.4487	1.1358	-10.0801	6.76E-24	4.32E-22	1234.1193	10.5493	1.2482	-8.4515	2.88E-17	7.83E-16	21.9980	Core Matrisome
Fbn2	9533.2964	11.5753	0.4199	-27.5667	2.79E-167	2.03E-163	1643.1586	9.1123	0.3408	-26.7375	1.72E-157	3.76E-154	20.6877	Core Matrisome
Cilp	1370.9420	10.3084	1.0201	-10.1053	5.23E-24	3.39E-22	1237.1727	10.2419	0.8896	-11.5124	1.14E-30	7.41E-29	20.5503	Core Matrisome
Dmbt1	289.6714	11.0294	2.0308	-5.4310	5.60E-08	6.41E-07	94.1498	9.4647	1.1964	-7.9113	2.55E-15	5.89E-14	20.4941	Core Matrisome
Igfbp5	5785.2698	10.7093	0.4993	-21.4477	4.79E-102	1.16E-98	2744.1844	9.7364	0.3274	-29.7352	2.70E-194	7.34E-191	20.4457	Core Matrisome
Lamc3	434.4838	10.7764	1.4269	-7.5524	4.27E-14	1.14E-12	176.7730	9.5884	1.0171	-9.4270	4.22E-21	1.47E-19	20.3648	Core Matrisome
Has2	240.0764	10.0202	1.0867	-9.2209	2.95E-20	1.44E-18	266.4770	10.2429	1.0519	-9.7375	2.09E-22	8.13E-21	20.2631	Core Matrisome
Igfbp6	243.2684	10.0338	1.1285	-8.8914	6.03E-19	2.58E-17	250.1232	10.1387	1.0424	-9.7263	2.33E-22	9.04E-21	20.1726	Core Matrisome
Coll10a1	48.5075	9.1512	1.1911	-7.6831	1.55E-14	4.41E-13	91.6773	10.1460	1.0657	-9.5208	1.72E-21	6.18E-20	19.2973	Core Matrisome
Col28a1	67.8336	9.6355	1.1450	-8.4152	3.92E-17	1.43E-15	62.1539	9.5822	1.1508	-8.3268	8.31E-17	2.20E-15	19.2177	Core Matrisome
Npnt	33.5049	8.6128	1.1403	-7.5534	4.24E-14	1.13E-12	58.0432	9.4710	1.1570	-8.1861	2.70E-16	6.80E-15	18.0838	Core Matrisome
Thbs4	327.9637	9.2556	1.2470	-7.4220	1.15E-13	2.93E-12	197.6863	8.5902	1.3166	-6.5247	6.82E-11	9.90E-10	17.8458	Core Matrisome
Smoc1	231.6377	8.6320	0.8131	-10.6162	2.50E-26	1.87E-24	147.9011	8.0631	0.6929	-11.6363	2.69E-31	1.83E-29	16.6951	Core Matrisome
Mfap5	41.7127	6.7429	1.0069	-6.6968	2.13E-11	4.14E-10	332.4165	9.7752	0.8407	-11.6270	3.00E-31	2.03E-29	16.5181	Core Matrisome