

# Supplementary Information

Crystal structure of an archaeal CorB magnesium transporter

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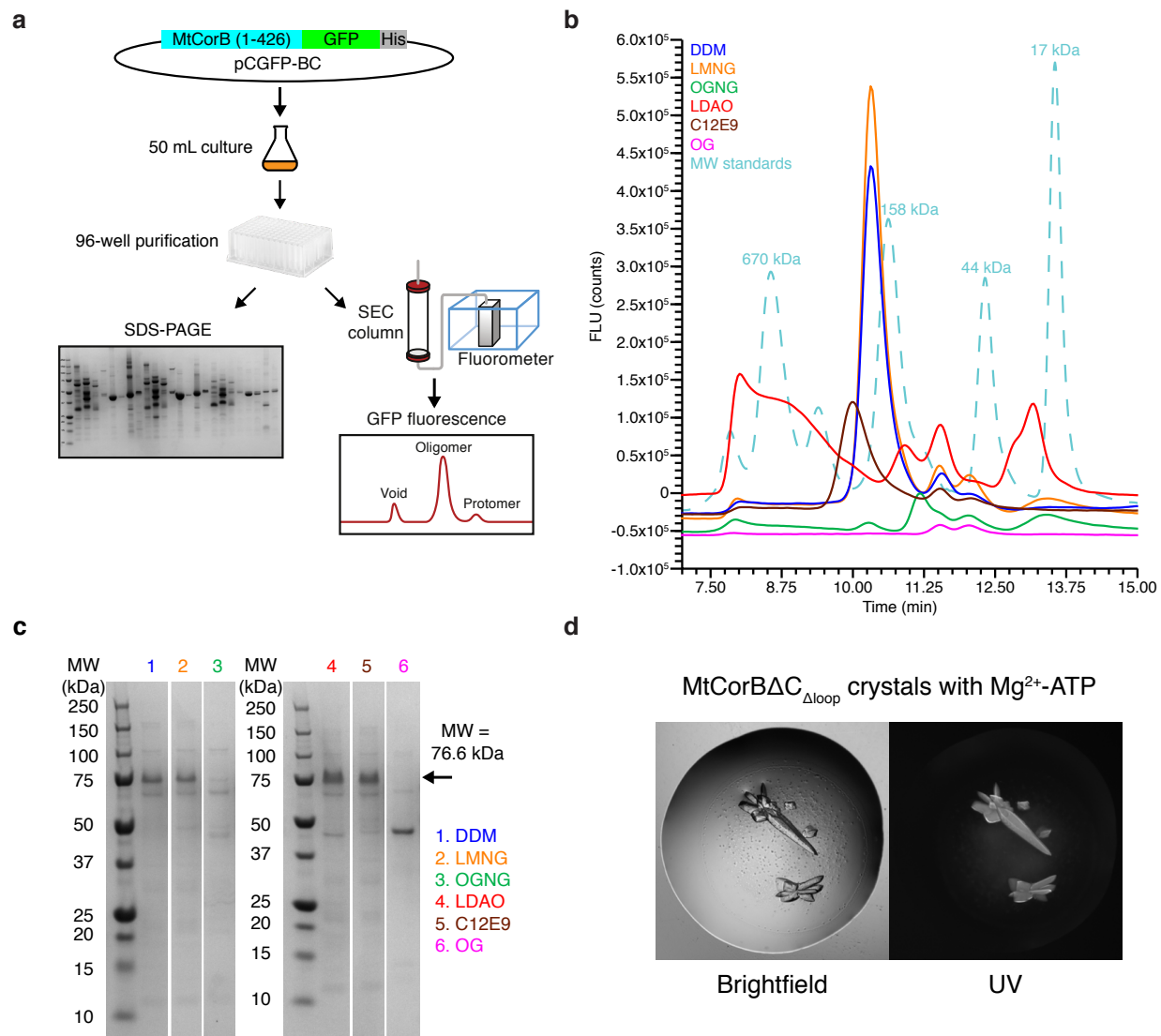
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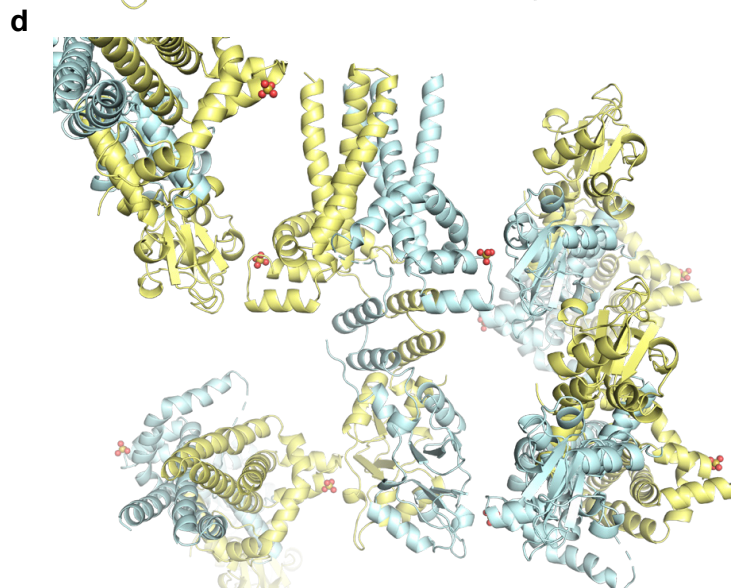
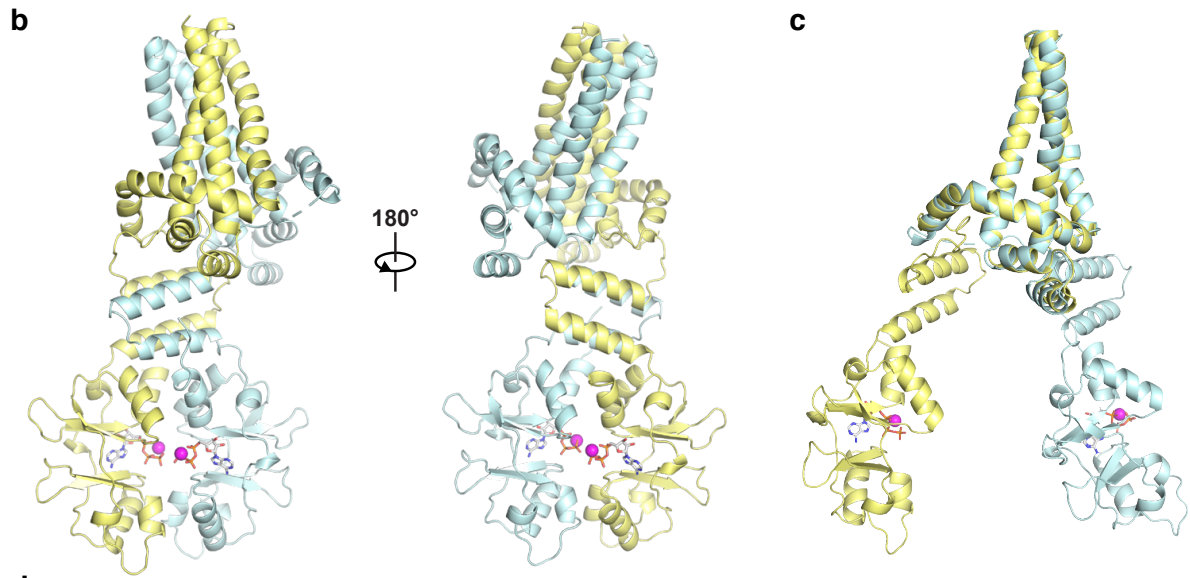
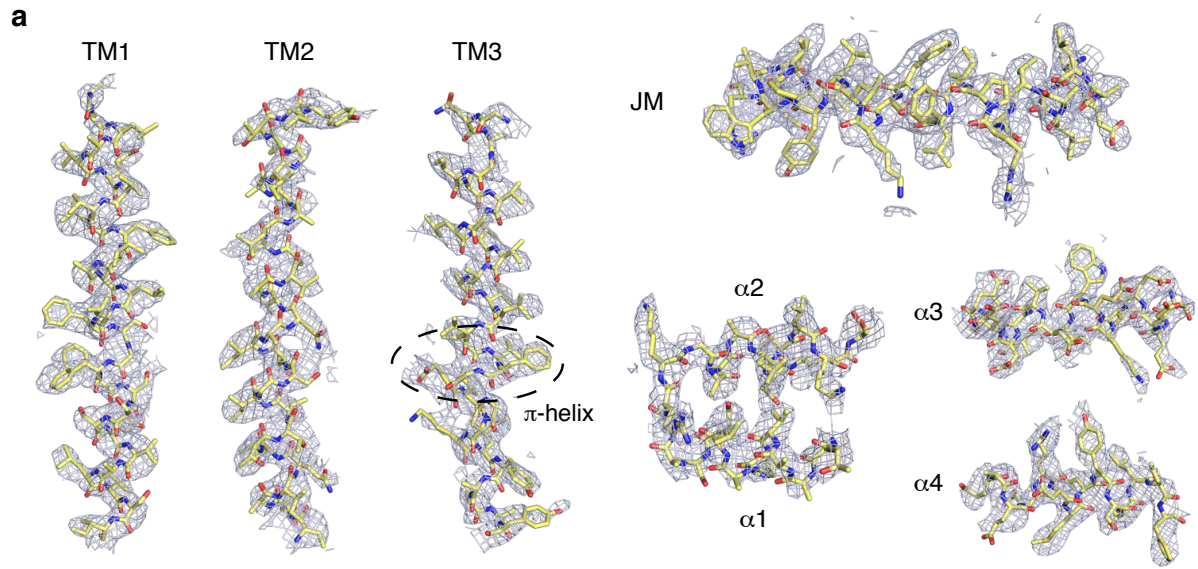


**Supplementary Figure 1. Detergent screening and crystallization of MtCorB.** **a**, Schematic of high-throughput screening process. **b**, Size-exclusion chromatography (SEC) profile of GFP-MtCorB purified in different detergents. The dashed line shows molecular weight (MW) standards. **c**, SDS-PAGE analysis of GFP-MtCorB purified in 6 detergents. This experiment was only performed once. **d**, MtCorB $\Delta$ loop crystals in complex with Mg<sup>2+</sup>-ATP taken with brightfield (*left*) and UV (*right*) camera.



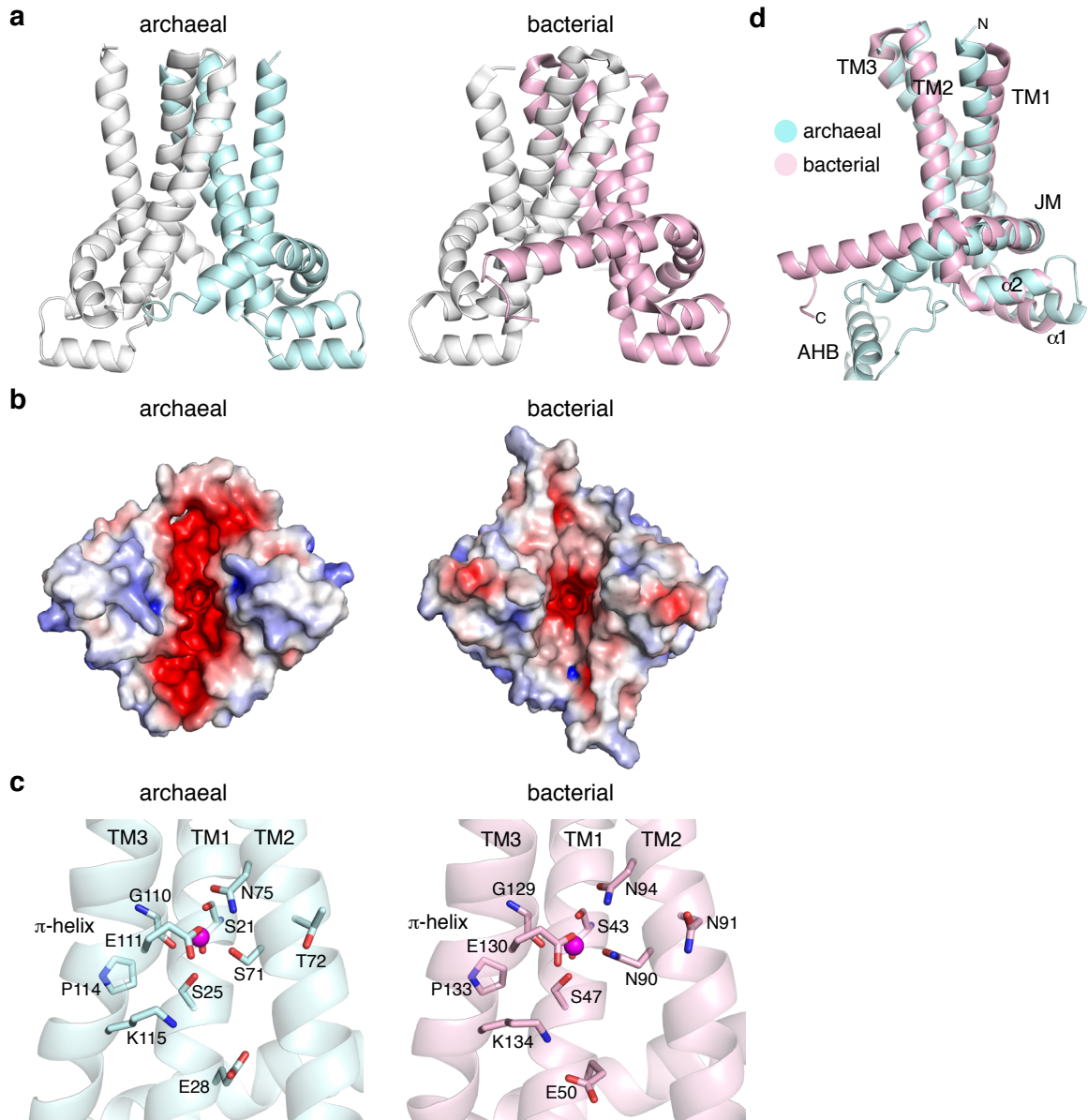
**Supplementary Figure 2. Sequence alignment of CorB/CNNM orthologs from eight representative species.** The listed CorB/CNNM orthologs and their UniProt accession numbers are: MtCorB (*Methanoculleus thermophilus*; A0A1G8XA46), TtCorB (*Tepidiphilus thermophilus*; A0A0K6IWT9), CBSDUFCH2 (*Arabidopsis thaliana*; Q84R21), MAM3 (*Saccharomyces cerevisiae*; Q12296), cnm-1 (*Caenorhabditis elegans*; A3QM97), UEX (*Drosophila melanogaster*; A0A0B7P9G0), cnm2a (*Danio rerio*; A2ATX7), and CNNM2 (*Homo sapiens*; Q9H8M5). Secondary structure corresponds to the crystal structure of MtCorB $\Delta$ C. Highlighted residues: residues involved in coordinating Mg<sup>2+</sup> ion (*magenta*), polar residues in the cavity of inward-facing conformation (*blue*), polar residues in outward-facing conformation (*green*), kink-inducing residue in JM helix (*yellow*), and ATP-binding site residues (*orange*).



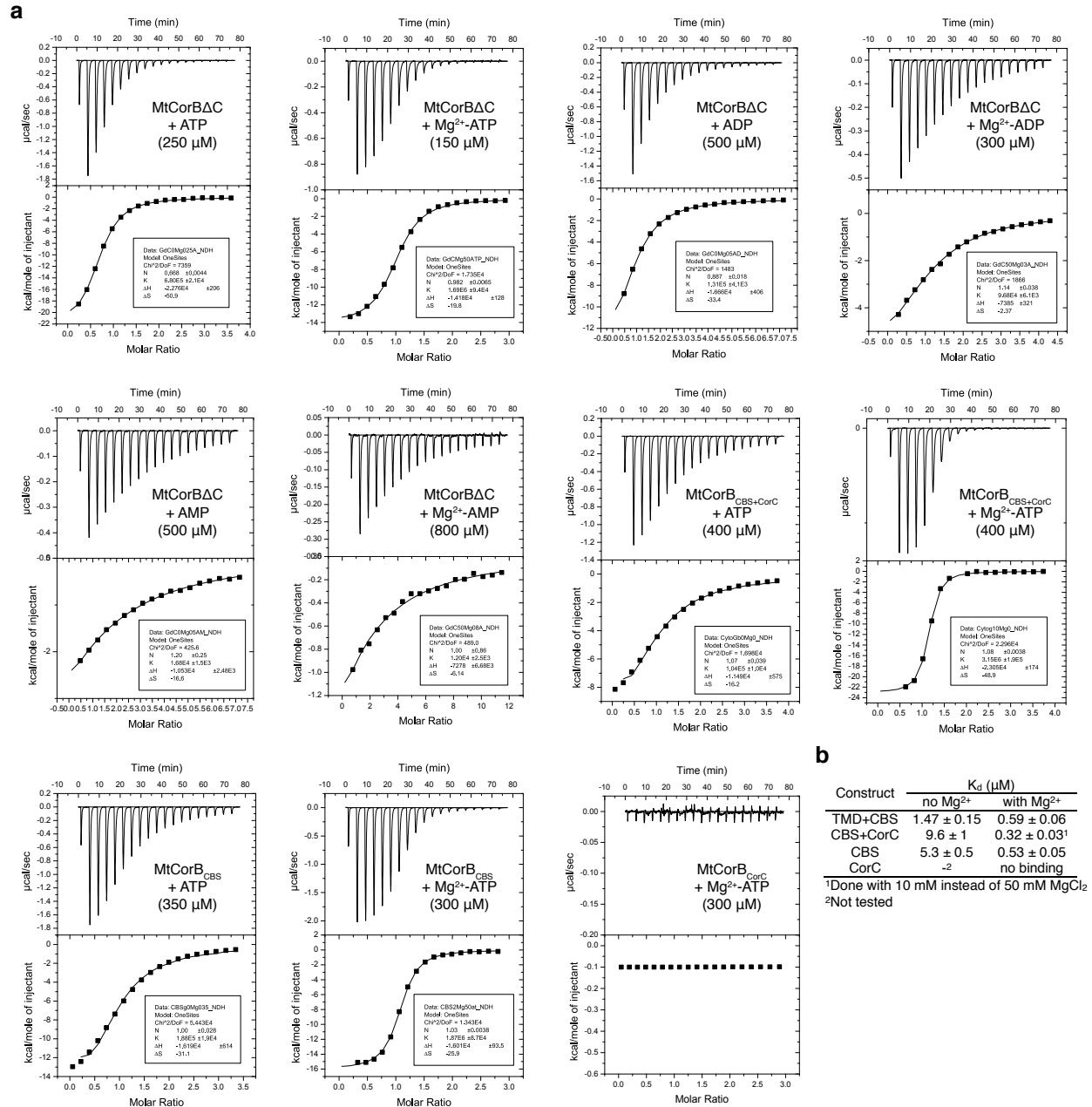


**Supplementary Figure 3. Overall structure of MtCorB $\Delta$ C bound to Mg<sup>2+</sup>ATP.**

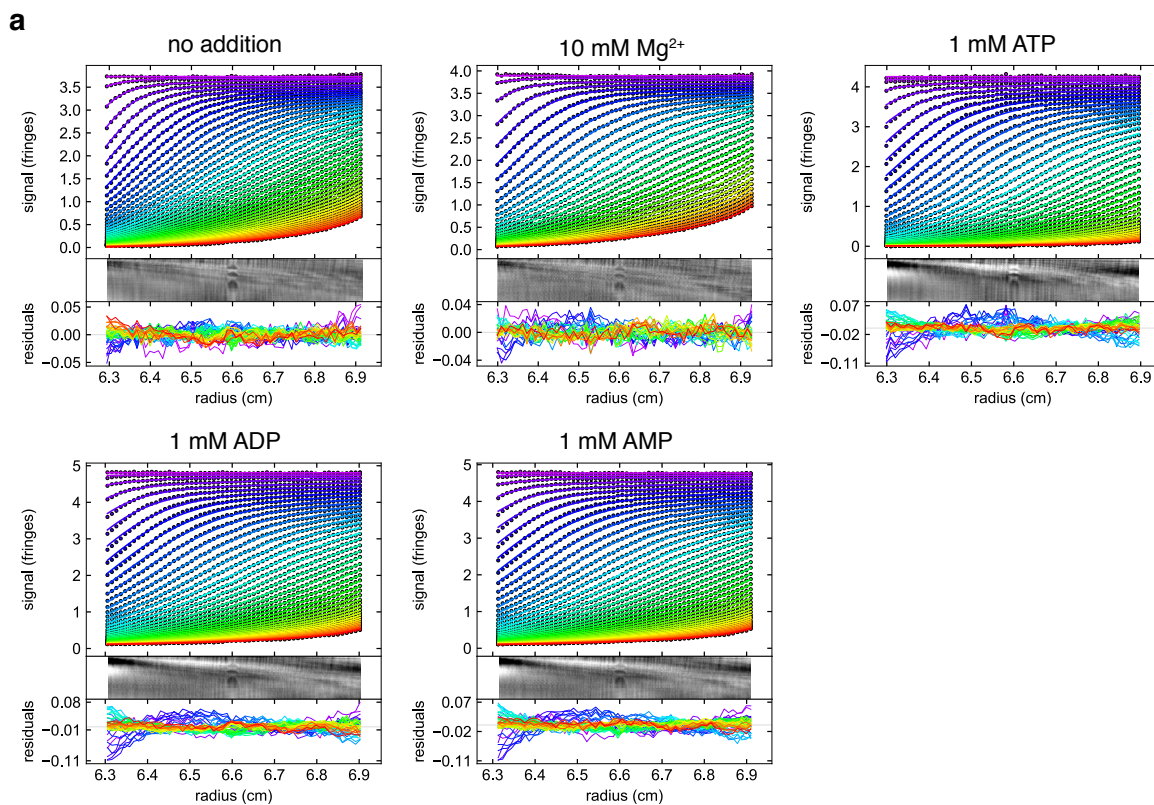
**a**, Representative simulated annealing 2F<sub>o</sub>-F<sub>c</sub> composite omit map for TMD and AHB, contoured at 1.0  $\sigma$ . **b**, Front and rear view of the MtCorB $\Delta$ C homodimer showing asymmetry between TMD and cytosolic domains. **c**, Overlay of the TMD of the two protomers. **d**, Crystal packing is assisted with a sulfate molecule.



**Supplementary Figure 4. Structural comparisons of the TMD of archaeal (this work) and bacterial (PDB: 7CFF) CorB proteins. a**, TMD homodimerizes in both archaeal and bacterial CorB proteins. **b**, Electrostatic surface potential analysis ( $\pm 5 \text{ kT e}^{-1}$ ) shows conservation of negatively charged cavity in both archaeal and bacterial CorB proteins from an intracellular view. **c**, Conservation of the polar residues in the  $\text{Mg}^{2+}$  binding site and  $\pi$ -helical turn of archaeal and bacterial CorB proteins. **d**, Structural overlay of the TMD of archaeal and bacterial CorB proteins showing distinct differences in the length and orientation of the TM1 and JM helices.



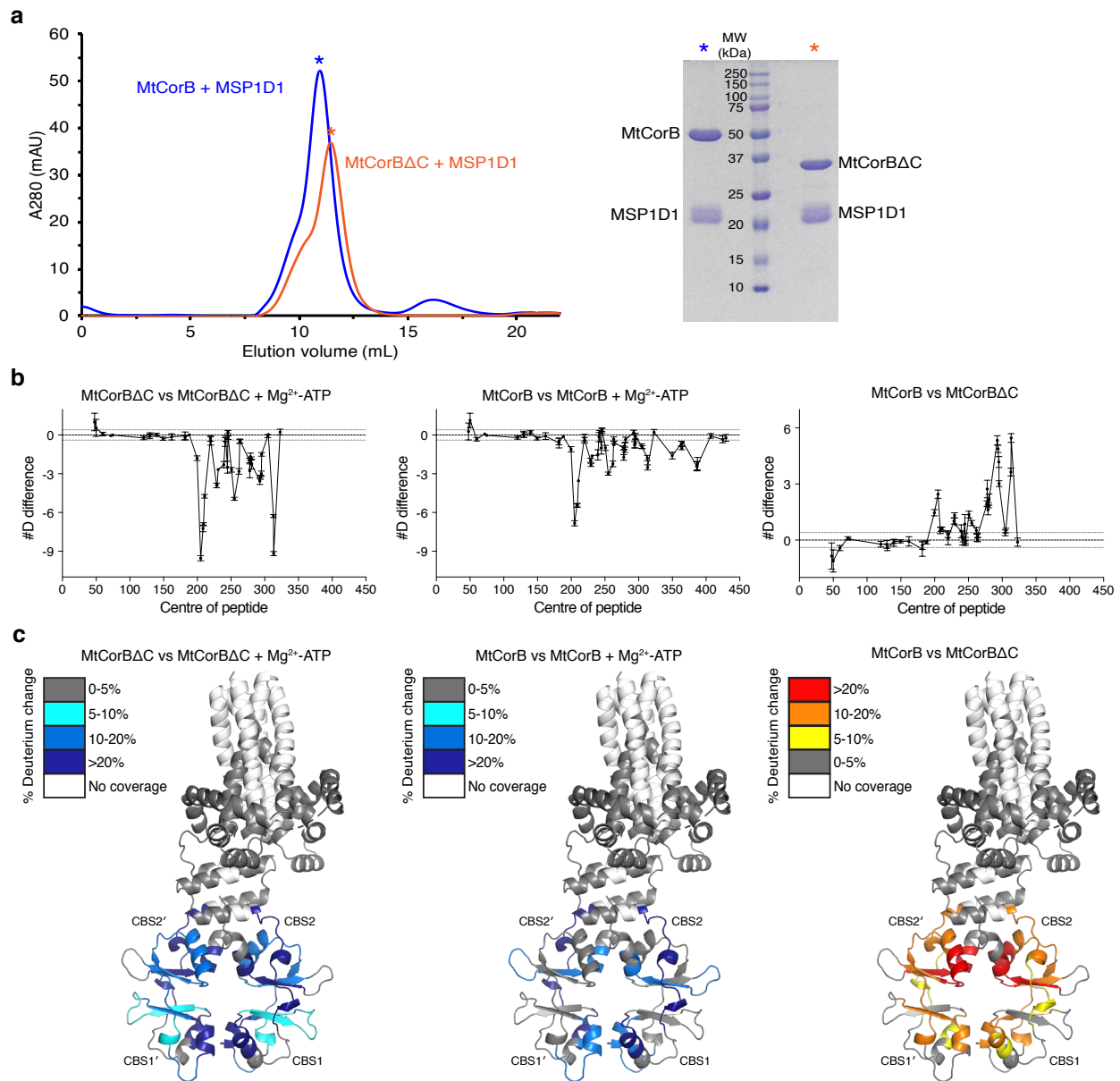
**Supplementary Figure 5. Raw ITC thermograms. a**, ITC thermograms showing various MtCorB constructs binding to adenosine nucleotides (concentrations indicated in parentheses) in absence or presence of 50 mM  $\text{MgCl}_2$ . **b**, Table summarizing the affinities of different MtCorB constructs binding to ATP in presence or absence of 50 mM  $\text{MgCl}_2$ .



**b**

Condition	Sedimentation coefficient (S)		Estimated MW (kDa)		RMSD	Theoretical MW (kDa)	
	Peak 1	Peak 2	Peak 1	Peak 2		Monomer	Dimer
no addition	1.84	-	16.2	-	0.0049		
10 mM Mg <sup>2+</sup>	1.78	-	16.0	-	0.0042		
1 mM ATP	-	2.49	-	32.0	0.0045	14.5	29.0
1 mM ADP	-	2.40	-	30.4	0.0044		
1 mM AMP	-	2.44	-	30.9	0.0050		

**Supplementary Figure 6. Summary of SV-AUC results.** **a**, Sedimentation velocity AUC profiles of MtCorB<sub>CBS</sub> in presence of various ligands. Interference of the sample are plotted against the radial position in the cell. One in every 75 scans is plotted. **b**, Summary of experimental sedimentation coefficients and estimated molecular weights.



### Supplementary Figure 7. Reconstitution of MtCorB into MSP1D1 nanodiscs and HDX-MS

**analyses. a**, SEC profile and SDS-PAGE analysis of MtCorB and MtCorB $\Delta$ C reconstituted in

MSP1D1 nanodiscs. The experiment was repeated twice with similar results. **b**, HDX-MS

analysis of three sets of experiments. The sum of the # of deuterons protected from across all

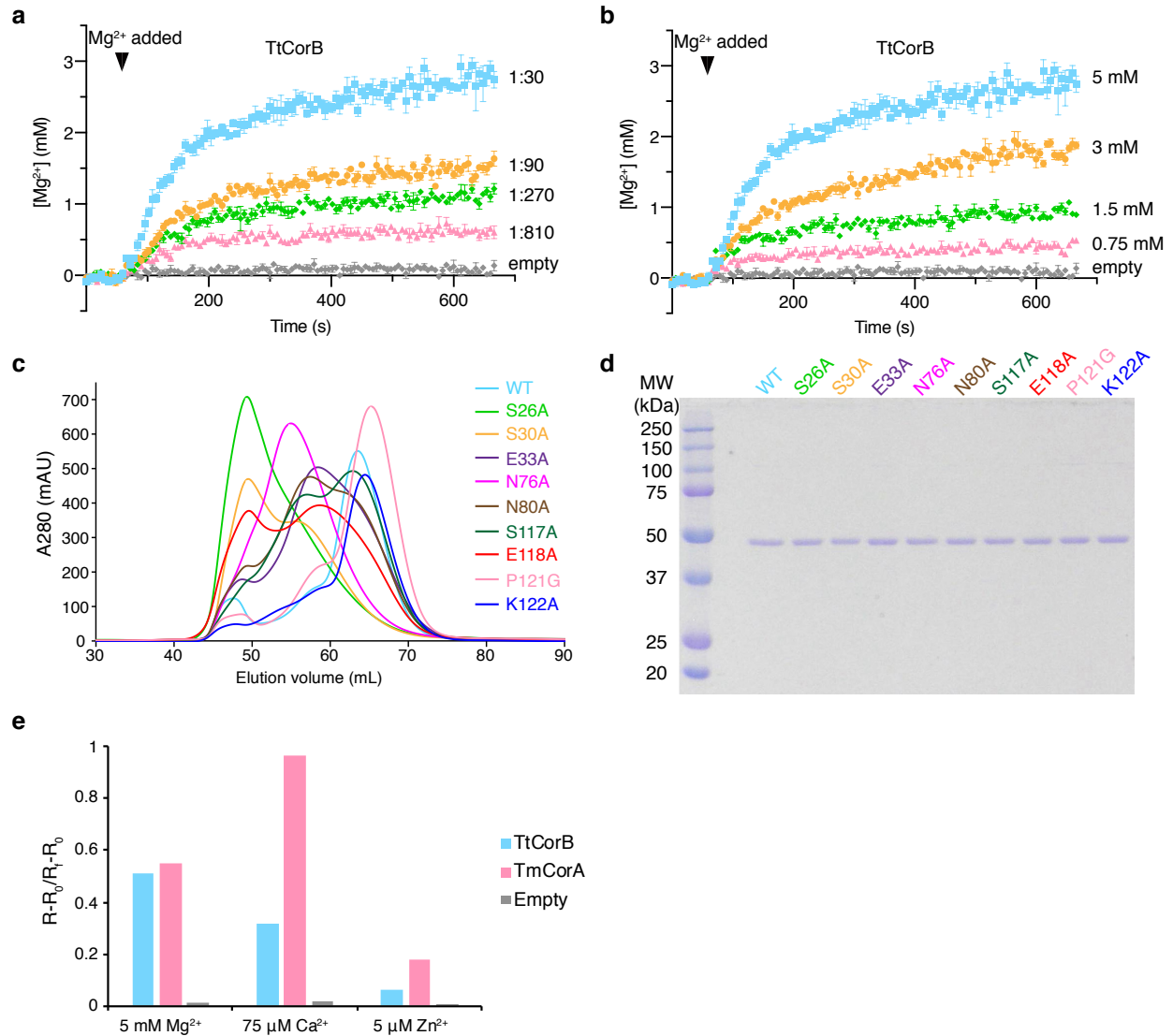
timepoints is shown. Each point represents a single peptide, with them being graphed on the x-

axis according to its central residue. The data points represent mean  $\pm$  SD (n = 3 independent

measurements). **c**, Mapping of deuteration onto the structure of MtCorB $\Delta$ C. Regions that showed



significant decreases or increases in exchange (defined as >5%, 0.4 Da, and a two-tailed Student's *t*-test  $p < 0.01$ ) are colored in blue or red respectively. The source data are provided as a Source Data file.



**Supplementary Figure 8. Liposome mag-fura-2 Mg<sup>2+</sup> transport assay of TtCorB proteins.**

**a**, TtCorB-incorporated proteoliposome (protein/lipid ratios of 1:30 to 1:810) shows protein-dependent Mg<sup>2+</sup> transport. The data points represent mean ± SEM (n = 3 independent measurements).

**b**, The TtCorB-mediated Mg<sup>2+</sup> transport depends on external MgCl<sub>2</sub> concentration. The data points represent mean ± SEM (n = 3 independent measurements).

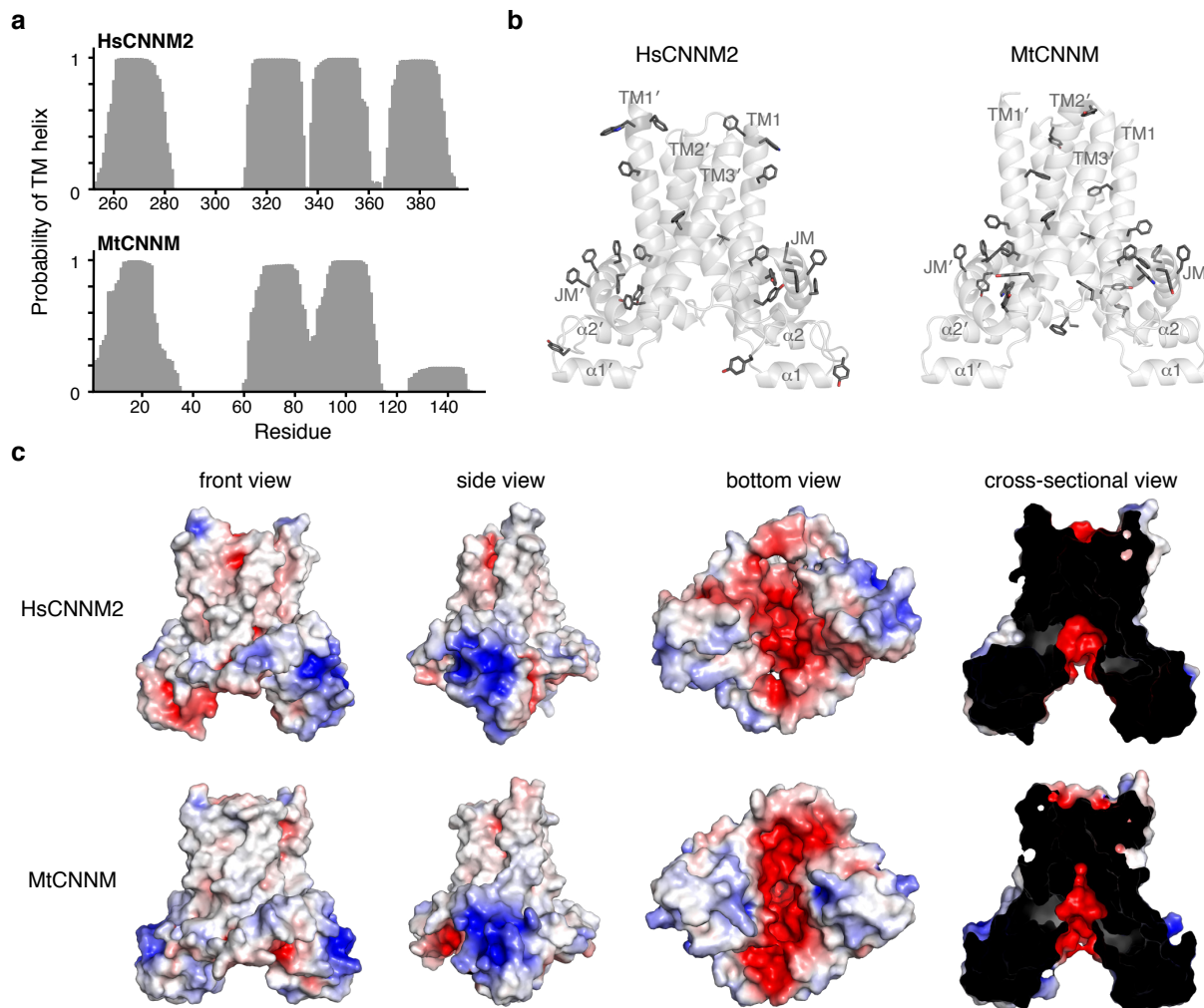
**c**, SEC profiles of TtCorB mutants. The experiment was repeated twice with similar results.

**d**, SDS-PAGE analysis of proteoliposomes reconstituted with TtCorB mutants (protein/lipid ratios of 1:30).

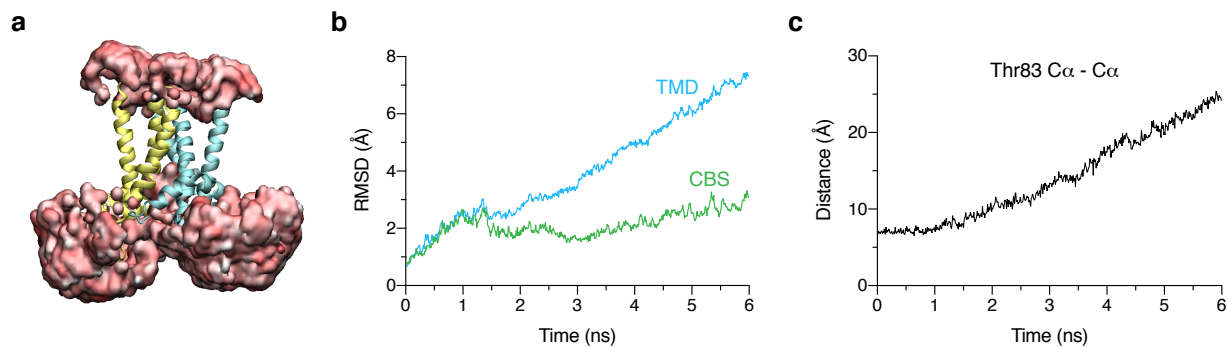
**e**, Comparison of ion transport by TtCorB and TmCorA proteoliposomes 10 min after



addition of the indicated ions. The relative change in fluorescence is plotted with  $R =$  ratio of fluorescence with excitation at 330 nm /369 nm,  $R_0 =$  fluorescence ratio before addition of cations (final concentrations are shown), and  $R_f =$  fluorescence ratio after lysing liposomes with Triton X-100.



**Supplementary Figure 9. Analysis of HsCNNM2 model structure.** **a**, Prediction of transmembrane helices in human CNNM2 and archaeal MtCorB with TMHMM server. HsCNNM2 is predicted to have four TM helices. **b**, Clustering of aromatic residues at the phospholipid-solvent interface and JM helix. **c**, Comparison of the electrostatic surfaces of human and archaeal TMD ( $\pm 5 \text{ kT e}^{-1}$ ).



**Supplementary Figure 10. Molecular dynamics simulations of MtCorB. a,** Hydration analysis of TMD showing an inward-facing, solvent-exposed cavity. **b,** Structural deviations of TMD and CBS-pair domain during the targeted MD simulation of inward-facing to outward-facing conformation. **c,** Opening of the dimerization interface as shown by increased C $\alpha$  distances between Thr83 of both protomers.

**Supplementary Table 1. Mutagenic primers used in the study.**

Construct	Forward Primer	Reverse Primer
MtCorB Δ323-426	catgctggaagaactggttctcgagcaccaccaccacc accactg	cagtggagggtgggtggtgctcgagaaccagtctcca gcatg
MtCorB Δ259-262	gatgtgttctctgcgcagacctccgctacc	ggtagcggaggctgcgcagagaacacatc
MtCorB R235L	gaaaccggcttctcttattcccggttaccac	gtggtaaaccgggataagagagaagccggttc
TtCorB S26A	gttctgctgctgaccgccggttcgctccatg	catggacgcgaaaccggcggcagcagcagaac
TtCorB S30A	gaccagcggttcgcggccatgtctgaaaccg	cggttcagacatggccgcgaaaccgctggtc
TtCorB E33A	gtttcgcgtccatgtctgcaaccgttatgatggcagc	gctccatcataacggttcagacatggacgcgaaac
TtCorB N76A	gagcgttatcctgctggtgccaacgcggttaacgttg	ccaacgttaaccgctggcaaccagcaggataacgctc
TtCorB N80A	ctggtaacaacgcggttgccgtggtgctgcgacc	ggtcgcagcaccaacggcaaccgctgttaaccag
TtCorB S117A	cttctgatcctggtgttcgccgaaatcaccccgaaag	ctttcggggtgatttcggcgaacaccaggatcaggaag
TtCorB E118A	gatcctggtgttcagcgcgaatcaccccgaaagtatc	gataactttcggggtgattgcgctgaacaccaggatc
TtCorB P121G	gtgttcagcgaatcacccgggaaagtatcggagcgcg	cgcgctccgataactttccgggtgatttcgctgaacac
TtCorB K122A	cagcgaatcaccccggcagttatcggagcgcgttac	gtaacgcgctccgataactgccggggtgatttcgctg