# nature research

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### **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

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Fora	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	$oxed{x}$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	🗴 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided  Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
x	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
x	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated
	Our web collection on <b>statistics for biologists</b> contains articles on many of the points above.

#### Software and code

Policy information about availability of computer code

Data collection

RNA sequencing was performed using the Illumina HiSeq2500 platform with 2x100 read pairs.

Data analysis

The details of data analysis and the software/code used are provided in the relevant subsections of the Methods section. Also, the code for analyses and parameters are available at https://github.com/hemberg-lab/yeastDrop-Seq.

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

#### Data

Policy information about <u>availability of data</u>

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The accession number for the raw sequencing data corresponding to the four treatment conditions is GEO: GSE165686.

## Life sciences study design

		<u> </u>			
All studies must disc	lose on these	points even when the disclosure is negative.			
Sample size	We targeted a sample size based on the original (mammalian cell optimized) Drop-Seq technique (Macosko E. et al, Cell, 2015).				
Data exclusions	No data were excluded.				
	Due to the inherent nature of stochasticity associated with gene expression, there were cell-to-cell differences (as expected) in the analyzed RNA transcript numbers even for cells grown in the same growth condition.				
Randomization	N/A (no random	andomization)			
Blinding	N/A (no blinding	'A (no blinding)			
Reporting for specific materials, systems and methods  We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.					
Materials & experimental systems Methods					
n/a Involved in the	· · · · · · · · · · · · · · · · · · ·				
Antibodies	,	ChIP-seq			
<b>x</b> Eukaryotic c	ell lines	Flow cytometry			
× Palaeontolo	ogy and archaeology				
Animals and	nd other organisms				
	Human research participants				
Clinical data					
Dual use research of concern					
Eukaryotic cell lines					
Policy information about cell lines					
Cell line source(s)	Saccharomyces cerevisiae (Acar Lab)				
Authentication	Validated its appropriate auxotrophy on plates lacking the appropriate aminoacids				
Mycoplasma contam	mination N/A				
Commonly misider (See <u>ICLAC</u> register)					