

Cell Reports, Volume 32

Supplemental Information

**Visual Input into the *Drosophila*
melanogaster Mushroom Body**

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A

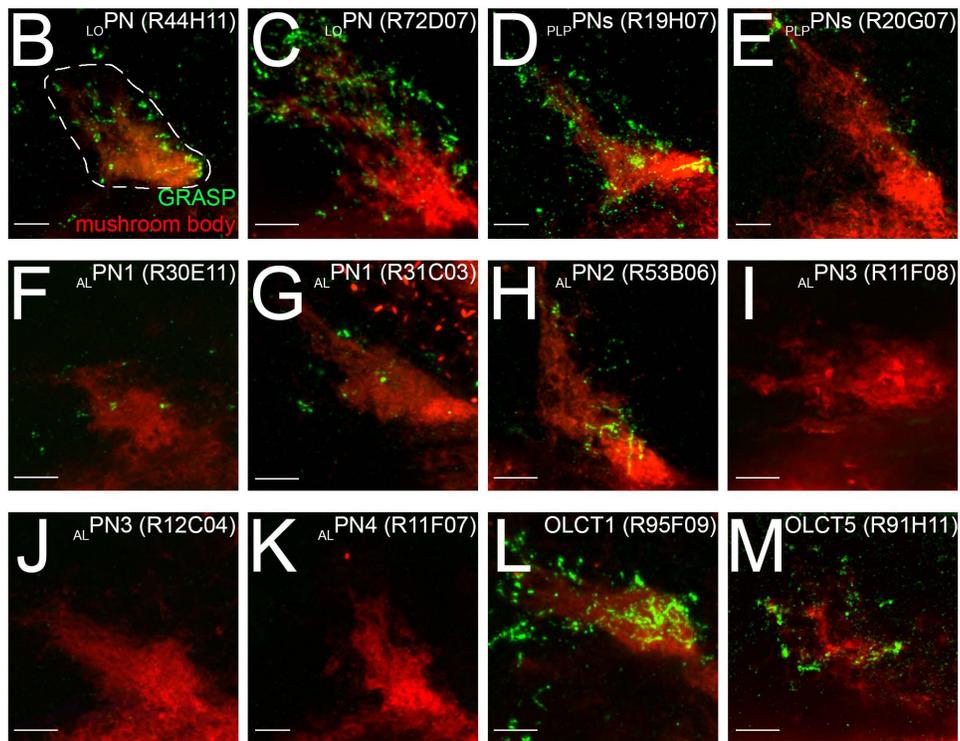
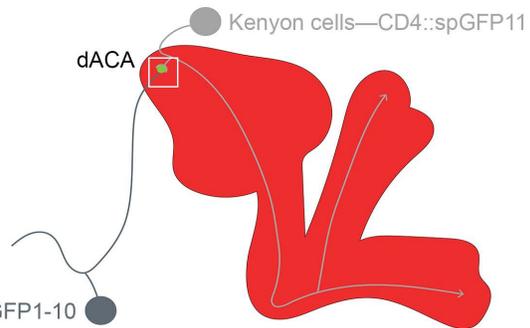


Figure S1. Identification of the transgenic lines driving expression in neurons presynaptic to the α/β_p Kenyon cells. Related to Figure 3. (A) A schematic of the *Drosophila* brain shows how the GFP Reconstitution Across Synaptic Partners (GRASP) technique was used to determine whether the selected transgenic lines drive expression in neurons that are presynaptic to the α/β_p Kenyon cells. The expression of the spGFP1-10 fragment was driven in the putative input neurons (dark grey) — using the various transgenic lines identified in the screen — and the expression of the spGFP11 fragment was driven in the mushroom body (red) using a transgenic line driving in most Kenyon cells; α/β_p Kenyon cell (light grey) can be distinguished from other Kenyon cells because they are the only Kenyon cells projecting to the dorsal accessory calyx. Reconstituted GFP molecules (here depicted by a green circle) are visible in the dorsal accessory calyx of the mushroom body (red) when the spGFP¹⁻¹⁰ fragment is expressed in synaptic partners. The dorsal accessory calyx was imaged (white square). (B-M) A strong (B-E, L-M) or weak (F-H) GRASP signal was detected in some of the selected transgenic lines; no GRASP signal was detected in other lines (I-K). The following genotypes were used in this figure: *yw/yw;UAS-syb::spGFP1-10^{unknown}*, *LEXAop-CD4::spGFP11^{unknown}/LEXAop-tdTomato^{attP5}*; *R_line-GAL4^{attP2}* (as indicated on the panel)/*MB247-LEXA^{unknown}*; . Scale bars are 5 μ m in all panels.

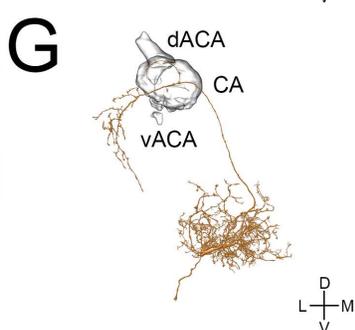
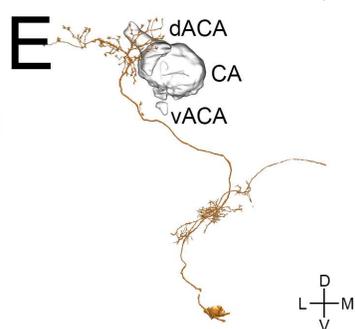
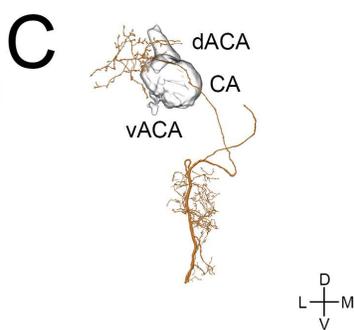
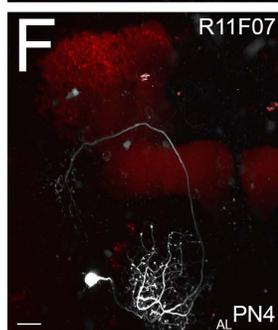
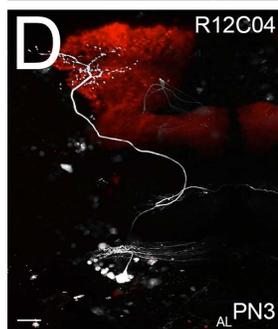
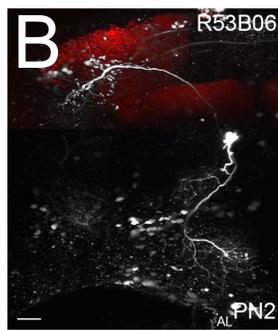
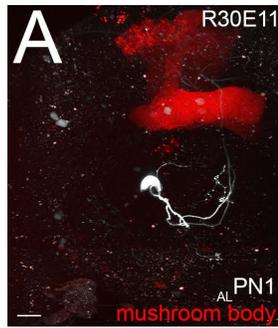


Figure S2. $_{AL}PN$ projecting from the antennal lobe to region near the dorsal accessory calyx. Related to Figure 3, Figure S1. (A, B, D, F) Four different neurons projecting from the antennal lobe to a region near the dorsal accessory calyx were identified using different transgenic lines (A: *R30E11-GAL4*, B: *R53B06-GAL4*, C: *R12C04-GAL4* and D: *R11F071-GAL4*); all the photo-labeled neurons project from the antennal lobe but each neuron has a distinct morphology. (C, E, G) Three of the four $_{AL}PN$ -like neurons were identified in the hemibrain connectome. (A) $_{AL}PN1$ extends its dendrites in the posterior antennal lobe and projects its axons in the superior lateral protocerebrum. (B-C) $_{AL}PN2$ extends its dendrites into the column region of the posterior antennal lobe, a region known to be activated by high humidity, as well as in the sub-esophageal ganglion, a gustatory processing center, and projects its axons in the superior lateral protocerebrum. (D-E) $_{AL}PN3$ extends its dendrites into the arm region of the posterior antennal lobe, a region known to be activated by low humidity and projects its axons in the superior lateral protocerebrum. (F-G) $_{AL}PN4$ extends its dendrites broadly in the anterior antennal lobe, an olfactory processing center, and projects its axons in the posterior lateral protocerebrum. The images in (C), (E) and (G) were taken directly from the Neuprint platform. The neuropil domains are defined by Neuprint platform. The following genotypes were used in this figure: *yw/yw;MB247-DsRed^{unknown},UAS-C3PA-GFP^{unknown}/UAS-C3PA-GFP^{attP40};UAS-C3PA-GFP^{attP2},UAS-C3PA-GFP^{VK00005},UAS-C3PA-GFP^{VK00027}/R_line-GAL4^{attP2}* (as indicated on the panel);. Scale bars are 20 μ m.

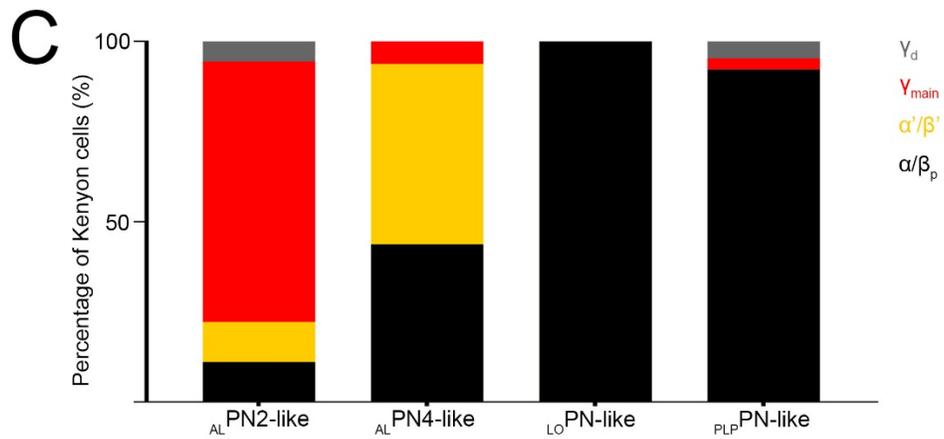
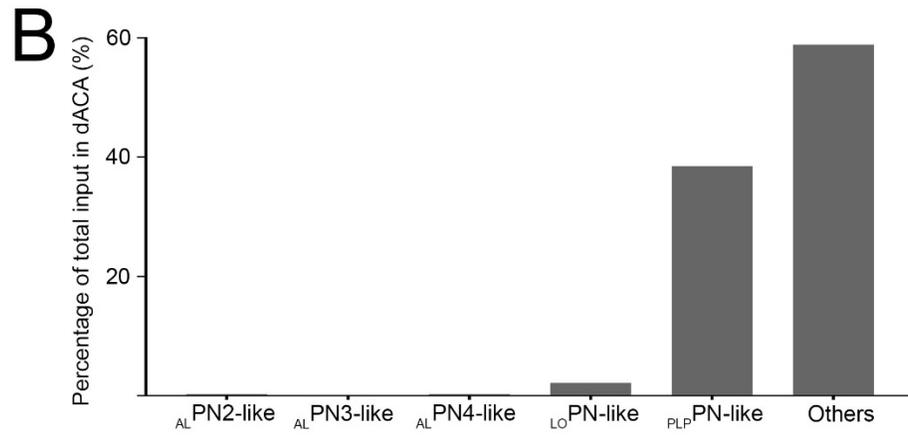
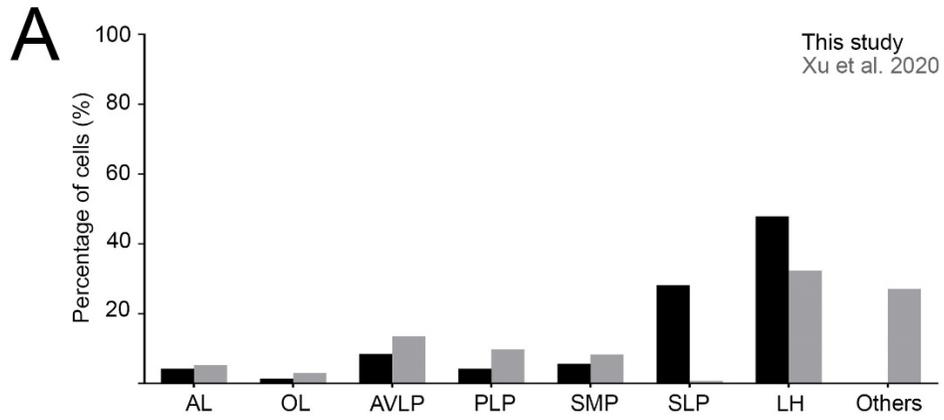


Figure S3. Identification of the projection neurons connecting to the α/β_p Kenyon cells in the dorsal accessory calyx in the connectome. Related to Figure 2, Figure 5. (A-C) The 60 α/β_p Kenyon cells reconstructed in the *Drosophila* hemibrain connectome receive input from 133 neurons in the dorsal accessory calyx. (A) These input neurons can be divided into seven clusters based on the location of their cell bodies: the antennal lobe (AL), optic lobe (OL), anterior ventral lateral protocerebrum (AVLP), lateral horn (LH), superior lateral protocerebrum (SLP), superior medial protocerebrum (SMP) and posterior lateral protocerebrum (PLP) clusters; neurons with somata located outside these clusters are defined as “others”. The percentage of neurons found in each cluster (grey) is similar to the one measured using the *en masse* photo-labeling technique reported in this study (black). (B) Among these 133 input neurons, neurons morphologically similar to $_{AL}PNs$, $_{LO}PN$ and $_{PLP}PNs$ were identified; the percentage of input α/β_p Kenyon cells receive in the dorsal accessory calyx from these different types of neuron varies across types. (C) $_{AL}PN$ -like, $_{LO}PN$ -like and $_{PLP}PN$ -like neurons connect to different types of Kenyon cell; whereas $_{AL}PNs$ connect mostly to olfactory Kenyon cells (namely γ_{main} (red) and α'/β' (yellow) Kenyon cells), $_{LO}PN$ -like and $_{PLP}PN$ -like neurons connect mostly to α/β_p Kenyon cells (black). Very few connections to γ_d Kenyon cells (grey) are found.

Type of neuron	Location of cell body	Number of neurons	Driver line(s)	Connections to α/β_p Kenyon cells	
				GRASP	Dye/photo-labeling technique
^{LO} PN* (OCLT1)	PLP cluster	1	R44H11	strong signal	4.1% ($n = 27$)
			R72D07	strong signal	3.0% ($n = 27$)
			R95F09	strong signal	n/A
^{PLP} PN	LH cluster	13	R19H07	strong signal	All: 14.1% ($n = 24$) Single: 1.9% ($n = 32$)
			R20G07	weak signal	
^{AL} PN1	AL cluster	1	R30E11	weak signal	0 ($n = 26$)
			R31C03	weak signal	
^{AL} PN2	AL cluster	1	R53B06	weak signal	0 ($n = 30$)
^{AL} PN3	AL cluster	1	R11F08	no signal	0 ($n = 26$)
			R12C04	no signal	
^{AL} PN4	AL cluster	1	R11F07	no signal	0 ($n = 25$)

Table S1. Neurons projecting to the dorsal accessory calyx. Related to Figure 5, Figure S1.

The neurons identified in the screen were divided into three types — ^{LO}PN, ^{PLP}PN and ^{AL}PN — based on the location of their dendrites — lobula (LO), posterior lateral protocerebrum (PLP) and antennal lobe (AL). The ^{AL}PN type was further divided into four subtypes — ^{AL}PN1, ^{AL}PN2, ^{AL}PN3 and ^{AL}PN4 — based on the fact that each one of these neurons were identified using different transgenic lines. The asterisk demarks the only neuron identified by our study that is similar to a neuron described in the Yagi et al. (2016) study.

Type of neuron (this study)	Type of neuron (Xu et al., 2020)	Neuprint ID number	Number of α/β_p Kenyon cells connected
LOPN	PVL02b_pct (1)	419908058	9 (2.14%)
PLPN (13)	PDL20y_pct (5)	479969106	20 (4.99%)
		5813019543	10 (2.17%)
		450597298	6 (1.88%)
		389536926	13 (4.73%)
		5813009790	2 (0.32%)
	PDL23g_pct (4)	5813011738	19 (6.91%)
		480310599	3 (0.26%)
		454335501	12 (4.12%)
		295461570	2 (0.42%)
	PDL20ob_pct (2)	5813011717	8 (2.63%)
		511634742	3 (0.23%)
	PDL20z_pct (2)	512023157	22 (6.61%)
		480997558	12 (3.21%)
ALPN2	VP multi adPN mALT	5813063239	2 (0.26%)
ALPN4	olfactory multi lvPN mALT	1037293275	4 (0.26%)
N/A	PVL10a_pct	479935033	43 (15.14%)

Table S2. Input neurons of the dorsal accessory calyx as identified in the *Drosophila melanogaster* hemibrain connectome. Related to Figure 6, Figure S3. The neurons connecting to α/β_p Kenyon cells in the dorsal accessory calyx — and their corresponding neurons, as described in this study — are listed; the numbers in parentheses indicate the number of neurons reported for each type. The identity number can be used to visualize each of these neurons using the Neuprint platform. Also listed are the number of α/β_p Kenyon cells connected to a given input neuron, as reported in Neuprint v1.0.1, as well as the percentage of input these neurons provide to the α/β_p Kenyon cells in parentheses.