Supplementary Information for

Skin-like mechanoresponsive self-healing ionic elastomer from supramolecular zwitterionic network

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Extended Methods

Preparation of PAAm/NaCl hydrogel. PAAm/NaCl hydrogel was prepared according to literature.^{S1} Briefly, the gel precursor was prepared by dissolving AAm and NaCl into deionized water with molar concentrations of 2.2 M and 2.74 M, respectively. MBAA (0.06 wt%) and APS (0.17 wt%) with respect to the weight of AAm monomer were then added as the cross-linker and initiator, respectively. After degassing in a vacuum chamber, TEMED (0.25 wt%, with respect to the weight of AAm monomer) was added as an accelerator. The solution was poured into a glass mold, and cured by UV light for 20 min. The obtained hydrogel was further immersed in aqueous solution with the same concentration of NaCl for more than 24 h.

Two-dimensional correlation spectroscopy (2DCOS). 2DCOS, as a mathematical method, is very suitable for the investigation of the variations of chemical groups. Its basic principle was first proposed by Noda^{S2, S3} and have been applied more and more widely to follow spectral variations under various external perturbations, such as time, temperature, pressure, concentration and other physical variables. By spreading the original spectral information along a second dimension, spectral resolution enhancement can be achieved, allowing the additional important information about molecular motions or conformational changes not readily visible in conventional analysis to be extracted.

2DCOS includes two types of correlation maps, synchronous spectrum which reflects simultaneous changes between two wavenumbers, and asynchronous spectrum which can significantly enhance the spectral resolution. The auto-peaks only appear along the diagonal in synchronous spectrum, while the cross-peaks can appear in both synchronous and asynchronous spectra. The judging rule of the sequence can be summarized as Noda's rule- that is, if the multiplication of the signs of cross-peaks (v_1 , v_2 , and assume $v_1 > v_2$) in synchronous and asynchronous spectra is positive, the change at v_1 may occur prior to v_2 , and vice versa^{S4,S5}. In Supplementary Table 2, "+" means the same signs in synchronous and asynchronous spectra, while "-" means different signs in synchronous and asynchronous spectra.



Supplementary Fig. 1. Photos of PAA/zwitterion proton-conductive elastomers indicating their good transparency.



Supplementary Fig. 2. UV-Vis transmittance spectrum of PAA/betaine elastomer film (thickness $\approx 100 \ \mu m$).



Supplementary Fig. 3. DLS size distribution of saturated betaine solution (~13.6 M). The peak at 615 nm comes from large betaine clustering aggregates.



Supplementary Fig. 4. Photos of betaine/H₂O (1:2.5), AA/betaine (1:1), AA/H₂O (1: 2.5) and AA/betaine/H₂O (1:1:2.5) mixtures. Due to the interactions among AA, betaine and water, the enhanced cosolvency of the three components is observed.



Supplementary Fig. 5. Tensile curves of the five PAA/zwitterion elastomers prepared in three batches suggest the good reproducibility of strain-stiffening response.



Supplementary Fig. 6. Initial moduli of PAA/zwitterion elastomers.



Supplementary Fig. 7. Effect of zwitterions on the enthalpic dimeric H-bonds of PAA. **a** pH values of 0.1 M PAA/zwitterion solutions by mixing PAA (with respect to monomer concentration), zwitterion, and water in the same molar ratio of 1:1:2.5. **b** Normalized ATR-FTIR spectra of PAA/zwitterion elastomers equilibrated at RH 60%.

In the cases of betaine, dimethylglycine, proline, and sarcosine, the pH values of PAA/zwitterion solutions are very close to each other, suggesting almost the same deprotonation degree of PAA. However, the relative amount of v(COOH) (H-bonded PAA) as normalized by the spectral intensity of v(COO⁻) (zwitterion) increases in the order of betaine, dimethylglycine, proline, and sarcosine, corresponding to the formation of more H-bonded crosslinks. This order is in full accordance with the increment of the initial moduli of PAA/zwitterion elastomers (Supplementary Fig. 6), which may result from the reduced zwitterion-PAA interactions in the case of tertiary and secondary amines with a lower electropositivity. The lowest initial modulus of PAA/TMAO elastomer should be caused by the highest deprotonation degree of PAA (highest pH) resulting in much less H-bonded crosslinks than the other four zwitterions.



Supplementary Fig. 8. Tensile curves of PAA/betaine elastomers with different molar ratios of AA and betaine. Decreasing the ratio of betaine leads to more rigid elastomers, and the strain-stiffening effect and stretchability are also largely weakened.



Supplementary Fig. 9. Tensile curves of PAA/LiBr and PAA/CH₃COOH elastomers with different molar ratios of AA and salt/acid. The PAA/LiBr and PAA/CH₃COOH elastomers show much weakened tensile strength and stretchability compared to PAA/betaine elastomer in the same molar ratio, and the strain-stiffening effect is also largely diminished. These two sets of control samples support the role of supramolecular betaine network in the strain-stiffening behavior.



Supplementary Fig. 10. Cytotoxicity tests of PAA/betaine elastomer. Cell viability of **a**) HeLa and **b**) HepG2 cells incubated with different amounts of PAA/betaine elastomer for 12 h and 24 h, respectively. The PAA/betaine elastomer is proved to have very good biocompatibility. Data are presented as mean values \pm SD, n = 6 independent solutions.



Supplementary Fig. 11. ¹H NMR spectra of PAA ($M_w = 100,000 \text{ g mol}^{-1}$), betaine and their mixture dissolved in D₂O. The total solute concentrations are all 0.1 M.



Supplementary Fig. 12. Tensile true stress-elongation curves of PAA/betaine elastomer at different stretching rates. A higher stretching rate leads to a high modulus and more pronounced strain-stiffening effect, proving the supramolecular nature of PAA/betaine elastomer. Notably, both PAA chain relaxation and betaine chain disassociation are strongly rate-dependent. A slowing stretching rate will give enough time for betaine chain fragmentation/reorganization and PAA chain rearrangement.



Supplementary Fig. 13. Temperature-dependent wavenumber shifts of v(C-N) (betaine), $v(COO^{-})$ (betaine) and v(COOH) (H-bonded PAA).



Supplementary Fig. 14. TGA curves of the as-prepared PAA/betaine elastomer and the elastomer equilibrated at RH 60% which show the almost equal water content of 21 wt%.



Supplementary Fig. 15. Tensile true stress-elongation curves of the as-prepared PAA/betaine elastomers with different water contents.



Supplementary Fig. 16. ATR-FTIR and corresponding second derivative spectra of PAA/betaine elastomer equilibrated at different humidities. With the increase of humidity, v(COOH) (H-bonded PAA) and v(C-N) (betaine) both shift to higher wavenumbers, indicating the water-induced weakening of PAA dimeric H-bonds and betaine ionic complexes. Moreover, judging from the second derivative spectra of $v(\text{COO}^-)$, a binary change is observed indicating the conversion of ionic complexed COO⁻ at 1633 cm⁻¹ to hydrated COO⁻ at 1627 cm⁻¹ at higher humidities.



Supplementary Fig. 17. Recyclability of PAA/betaine elastomer in biologically relevant media. Photos of PAA/betaine elastomer **a**) before and **b**) after dissolving in RPMI.1640, and DMEM (high glucose) culture media, normal saline, and PBS buffer solutions, respectively. **c** Photo of the recasted PAA/betaine elastomer films. **d** Tensile curves of the recasted films at RH 60%. All the films are transparent and highly stretchable. However, since the abundant ions in the biologically relevant media strongly interrupt the physical crosslinks of PAA/betaine elastomer, strain-softening behavior is observed for all the four samples.



Supplementary Fig. 18. The recorded resistance variations of PAA/betaine elastomer adhered on human finger at different bending angles.



Supplementary Fig. 19. Temperature-dependent resistance variations and corresponding gauge factors of ionic liquids, saline solutions, PAAm/NaCl hydrogel and PAA/betaine elastomer. **a** Chemical structures of ionic liquids ([EMI][ES], [EMI][DCA], [EMI][TFSI], [EMI][BF4], [BMI][BF4], and [BMI][TFSI]). **b** Temperature-dependent resistance variations ($\Delta R/R_0$) and **c** corresponding gauge factors of ionic liquids, saline solutions (1 M KCl, 1 M NaCl), PAAm/NaCl hydrogel and PAA/betaine elastomer.



Supplementary Fig. 20. Capacitance response curve of PAA/betaine elastomer under the dynamically applied pressure of 2.45 kPa for 450 cycles.

Temperature/°C	τ*	β
20	353.35	0.438
30	128.53	0.514
40	63.77	0.543
50	36.21	0.530

Supplementary Table 1. Fitting parameters (τ^* and β) in the simulation of iso-strain–stress relaxation results (Fig. 2d).

Supplementary Table 2. Final results of the multiplication of the signs of each cross-peak in 2DCOS synchronous and asynchronous spectra of PAA/betaine elastomer (Fig. 2f).

1392	-	-	-	-	
1402	+	-	+		
1639	+	-			
1701	+				
1728					
	1728	1701	1639	1402	1392

According to Noda's rule, the final specific order for PAA/betaine elastomer during heating is given as follows: $1392 \rightarrow 1728 \rightarrow 1639 \rightarrow 1402 \rightarrow 1701 \text{ cm}^{-1}$ (\rightarrow means prior to or earlier than), i.e. v(C-N) (betaine-betaine complex) $\rightarrow v(\text{COOH})$ (free PAA) $\rightarrow v(\text{COO}^{-})$ (betaine) $\rightarrow v(\text{C-N})$ (betaine-PAA complex) $\rightarrow v(\text{C=O})$ (H-bonded PAA).

Supplementary Table 3. Extended comparison of the overall performance between this work and previously reported polyzwitterion-based hydrogels and ionic skin materials.

Ionic skin	Mechano	Self-	Max.	Elastic	Transparency	Air	Anti-	Adhesion	Ref.
materials	response	healing	strain	recovery	(%)	stability	freezing		
			(%)	ratio (%)			(°C)		
PMAEDS/CNF	Softening	None	920	N.A.	High	Poor	N.A.	Yes	[S6]
hydrogel									
PDMAPS/silk	Softening	24 h	6000	N.A.	High	Good	N.A.	Yes	[S7]
fibroin/CaCl ₂									
hydrogel									
PAA-co-	Softening	2 h,	>10000	N.A.	90	Poor	N.A.	Yes	[S8]
DMAPS/NaCl		100%							
hydrogel									
PDA-clay-PSBMA	Softening	24 h,	900	93.8	Low	Poor	N.A.	Yes	[S9]
hydrogel		80%							
pCB/pSB ZEN	Softening	None	169.9	N.A.	High	Poor	N.A.	N.A.	[S10]
hydrogel									
Dopamine-triggered	N.A.	Fast	>350	N.A.	~90	N.A.	N.A.	Yes	[S11]
PSBMA hydrogel									
PAAm/PF127	Softening	Yes	2716	~95	83.8	Poor	N.A.	N.A.	[S12]
hydrogel									
HPC/PVA DN	Softening	None	975	~90	Opaque	Poor	N.A.	N.A.	[S13]
hydrogel									
PAA-MEA	Softening	None	868	~100	~100	Good	-90	N.A.	[S14]
organohydrogel									
PVA-CNF	Softening	None	660	Good	90	Good	-70	N.A.	[S15]
organohydrogel									
Cellulose-IL	Softening	15 min,	~105	N.A.	>90	Good	N.A.	Yes	[S16]
dynamic gel		~100%							
PolyTA ionogel	Softening	6 h,	>10000	99%	95	Good	N.A.	Yes	[S17]
		~100%							
PEA DN ionogel	Softening	None	>5000	~94	95	Good	-70	Yes	[S18]
PVDF-HFP ionogel	Softening	24 h,	~2000	~80	98	Good	N.A.	N.A.	[S19]
		99.1%							
PIL/TFSI elastomer	Softening	2 h,	540	95	Opaque	Good	N.A.	Yes	[S20]
		~100							
PAAm-IL gel	Softening	Yes	900	N.A.	92	N.A.	-20	N.A.	[S21]
PAAm/ChCl	Softening	72 h,	450	N.A.	95	Good	-23	N.A.	[S22]
elastomer		94							
PAA/betaine	Stiffening	12 h,	1600	97.9	99.7	Good	-40	Yes	This
elastomer		100%							work

Note: PMAEDS = PDMAPS = PSBMA. None of self-healing property refers to chemically or nanocrystal-crosslinked gels that are considered to be unable to heal themselves once damaged. Air stability means the stability of the material for maintaining stable ionic conduction as exposed in air. For gels, air stability is closed related to their ability of solvent retention in air.

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