

Figure S1. Effect of GPx1 on TNF- α -induced apoptosis (Related to Fig. 1).

(a) Expression of RIPK1 and RIPK3 protein in various cancer cell types. hRIPK and mRIPK present human and mouse RIPK protein, respectively. Actin is a loading control. (b) FACS analysis of the siRNA-transfected cancer cells untreated or treated with T/C for 6 h. PI, propidium iodide. (c) TUNEL staining of the T/C-treated HeLa cell. Arrows indicate the apoptotic cells. (d) Apoptotic cell death in the T/C-treated MCF10A, normal mammary epithelial cells, transfected with control or GPx1 siRNA. Note that MCF10A was not susceptible for TNF- α -induced apoptosis. (e) HeLa cells were transfected with three different sequences of siRNAs specific to GPx1. The knockdown level of GPx1 protein was verified by immunoblotting. The transfected HeLa cells were treated with T/C for 6 h. According to the result, the siRNA #3 is mainly used for further experiments. (f and g) TNF- α -induced necroptosis in the RIPK3-positive cancer cells. HeLa cells (g) were transfected with pcDNA vector encoding human RIPK3 tagged with Flag eiptope and treated with T/C/Z. ZVAD (Z) was pretreated to induced caspase-independent necroptosis in cancer cells.

Data in the graph are means ± SD of percent of apoptotic cells (n = 3, *P<0.001, **P<0.0001). Control siRNA is specific to firefly luciferase. N.S., not significant.



Figure S2. Effect of GPx1 on TNF- α -induced apoptosis in the Smac mimetic (Related to Fig. 1).

HeLa and MDA-MB-231 cells were transfected with control and GPx1 siRNAs and stimulated with TNF- α -in the presence of a Smac mimetic, BV6 (B) for 6 h. ZVAD (Z) was also pretreated to inhibit the apoptosis. Note that BV6 alone slightly induced apoptosis. Data in the graph are means ± SD of percent of apoptotic cells (n = 3, *P<0.05, **P<0.005, #P<0.001). Control siRNA is specific to firefly luciferase.



Figure S3. Level of lipid hydroperoxide in the GPx1-depleted MDA-MB-231 cells (Related to Fig. 2).

The level of lipid hydroperoxide (LOOH) was measured using a LOOH-specific fluorescent probe, C11-BODIPY^{581/591}, in the MDA-MB-231 cells (**a**). Cells were transfected with either control or Gpx1 siRNA (siCon or siGpx1) for 48 h and treated with C11-BODIPY (1 μ M). A ferroptosis inducer (Erastin) was used in parallel for control experiment. The GPx depletion was verified by immunoblotting (**b**). Data in the graph are means ± SD of the relative rations of green versus red fluorescence intensities averaged from 40 cells (*n* = 3). Representative images are shown. N.S., not significant.

Figure S4. Lee et al



Figure S4. GPx1 differentially regulates MAPK activation in the MDA-MB-231 cells (Related to Figure 3). (a) The siRNA-transfected MDA-MB-231 cells were treated with T/C and lysed for immunoblotting. The immune bands for the phosphorylated MAPKs were quantified and normalized by the intensity of corresponding MAPK bands. Data in the graph are means \pm SD of fold change of band intensities (*n* = 3, **P*<0.05, ***P*<0.001). An immunoblot against α -tubulin is the loading control.

(b and c) The siRNA-transfected MDA-MB-231 (b) and HeLa (c) cells were pretreated with MAPK inhibitors (10 µM each) for 30 min. Apoptosis was measured after T/C treatment for 6 h. Data in the graph are means ± SD of percent of apoptotic cells (n = 3, *P<0.01). SP600125 (SP) and JNK inhibitor VIII (JNK,-VIII) are specific JNK inhibitors, while SB203880 (SB) is specific p38 inhibitor. N.S., not significant.