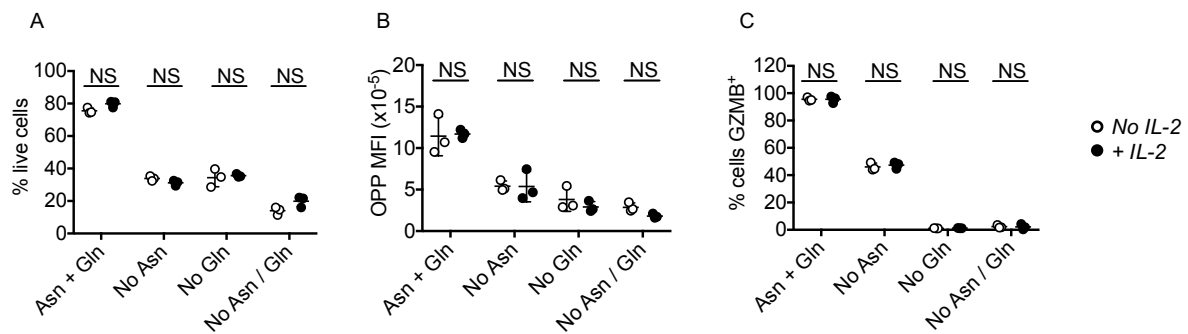


Coordination of asparagine uptake and asparagine synthetase expression modulates CD8⁺ T cell activation

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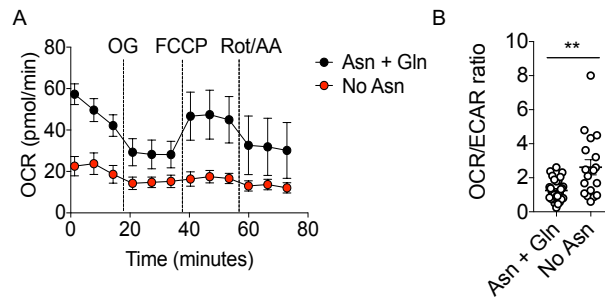
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Supplemental Figures 1-5



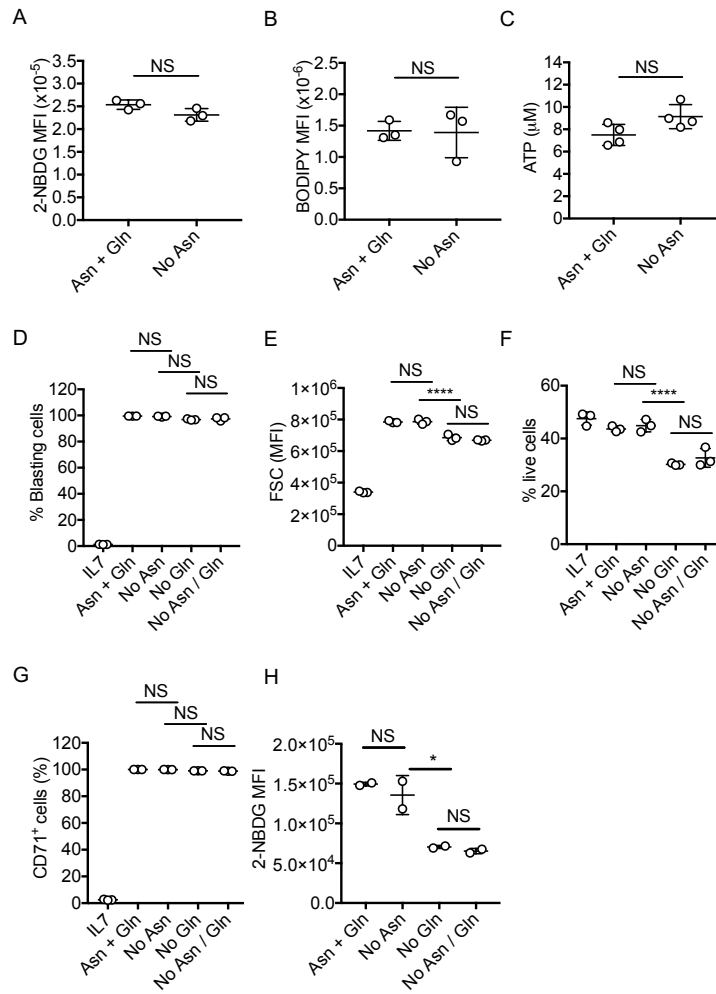
Supplemental Figure 1: Exogenous IL-2 does not rescue T cell activation in amino acid-depleted conditions

OT-1 T cells were activated for 24h (A, B) or 48h (C) in DMEM \pm Asn and/or Gln, in the presence or absence of 1ng/mL recombinant IL-2. Proportions of live cells (A) were determined by live-dead aqua dye exclusion and FACS. Levels of protein synthesis were assessed by incorporation of OPP, Click chemistry labelling and FACS (B). Intracellular granzyme B expression was assessed by FACS (C). NS – not significant as determined by 2-way ANOVA.



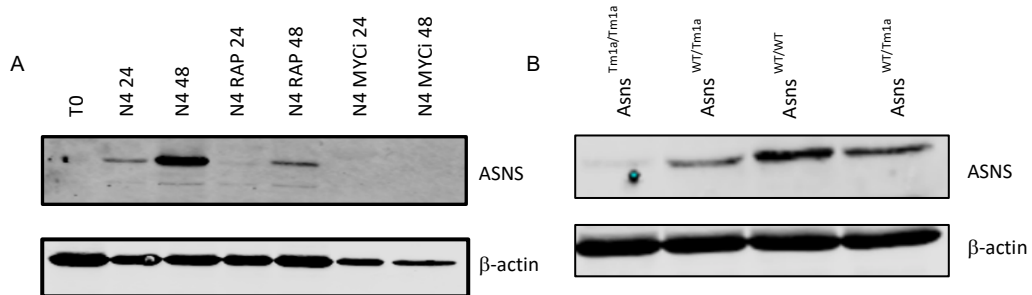
Supplemental Figure 2: Reduced mitochondrial function in Asn-deprived CD8⁺ T cells

OT-I T cells were activated with SIINFEKL peptide for 24h in DMEM ± Asn. (A) Oxygen consumption rates (OCR) were measured using the Seahorse XFe96 analyser, using a Mitostress™ kit. OG – oligomycin, FCCP- Carbonyl cyanide-4-(trifluoromethoxy)phenylhydrazone. Rot/AA – rotenone/antimycin A. Values are mean OCR readings from replicate wells (n=6) ± SD. (B) Ratio for the basal OCR/extracellular acidification rates (ECAR) rates for cells stimulated in the presence or absence of Asn (dots represent technical replicates, n=30). ** - p<0.01 by Student’s *t*-test. Data represent one of two repeated experiments.



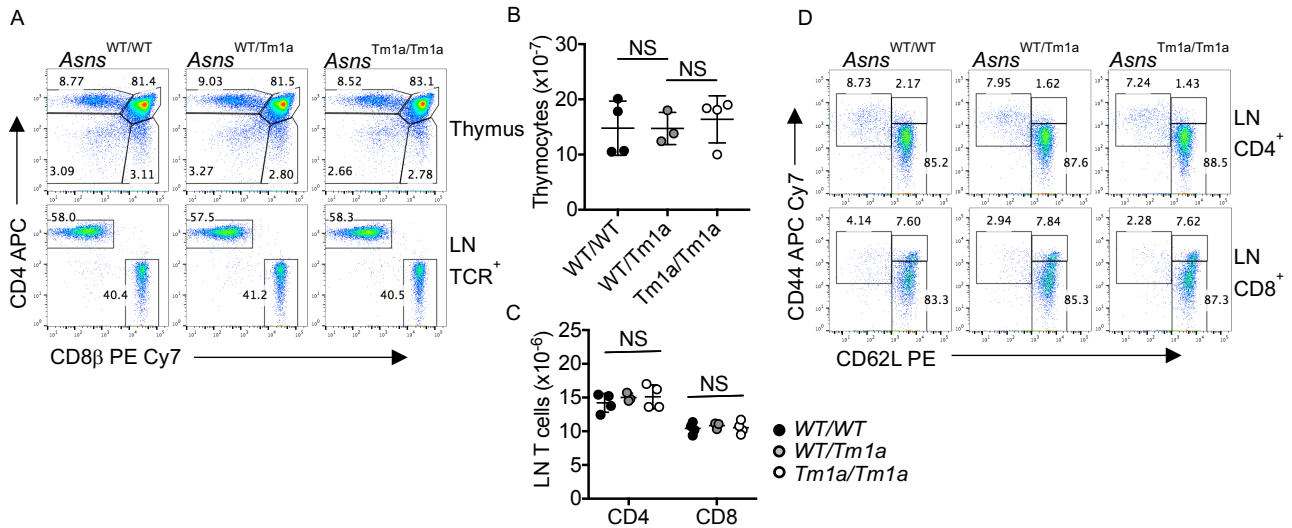
Supplemental Figure 3: T cells lose dependence upon extracellular Asn upon prolonged activation

OT-1 T cells were activated for 72h with SIINFEKL peptide in DMEM \pm Asn (A-C). Uptake of fluorescent 2-NB-d-glucose (A) and BODIPYTM-C16 (B) was assessed by FACS. Data shown are mean fluorescence intensities (MFI). (C) Cellular ATP levels were assessed using a luminescent assay. (D-H) OT-I T cells were stimulated with SIINFEKL peptide in complete IMDM for 24h, then were switched to DMEM \pm Asn \pm Gln for 24h. Proportions of blasting cells (A), cell size (FSC) (B), viability (C), CD71 expression (D) and 2-NBDG uptake (E) by OT-I T cells after 48h of stimulation were assessed by flow cytometry. Data are representative of 3 repeated experiments, dots represent technical replicates (for part C n=4, otherwise n=3). NS – not significant, * p<0.05, **** p<0.001 as assessed by Mann-Whitney test (A-C) or by one-way ANOVA with Tukey's multiple test (D-H).



Supplemental Figure 4: Replicate western blotting experiments for ASNS expression

A) OT-1 T cells were activated for 0-48h with SIINFEKL (N4) peptide in the presence of rapamycin (RAP) or Myc inhibitor (MYCi). B) Levels of ASNS in activated *Asns*^{Tm1a} gene-trap T-lymph node T cells. Data show western blots of ASNS in cell lysates; β actin serves as a loading control.



Supplemental Figure 5: T cell development and homeostasis are unimpeded in *Asns*^{Tm1a} mice

(A) Thymocyte and lymph node (LN) cell populations from littermate *Asns*^{Tm1a} mice were analysed by flow cytometry. Total thymocytes (B) and LN CD4 and CD8 T cells (C) were enumerated from 3-4 age-matched male mice of each genotype. (D) Proportions of naïve, effector and memory cells within gated CD4⁺ and CD8⁺ T cell populations were analysed by analysis of CD44 and CD62L expression by flow cytometry. FACS dotplots are representative of 5-6 mice of each genotype. Values on dotplots represent % cells within the defined gates. NS – not significant, as determined by one-way ANOVA