## Supplemental data for

## TYRO3 induces anti-PD-1/PD-L1 therapy resistance by limiting innate immunity and tumoral ferroptosis

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## Supplemental Figure 1. TYRO3 expression is correlated with anti-PD-1/PD-L1 resistance.

(A) Relative expression of the indicated kinases in 4T1-P and 4T1-R tumor cells by RTK kinase array. (B) Overall survival in melanoma patients who received PD-1 antibody therapy with high and low *EPHB2*, *FLT3*, and *TRKA* mRNA expression. (C) Overall survival in melanoma patients who received PD-1 antibody therapy with high and low *AXL/MERTK* mRNA expression. (D) Overall survival with high and low mRNA *TYRO3* expression in breast cancer, neuroblastoma, bladder cancer, and melanoma patients. (E) Relationship between overall survival and cytotoxic T lymphocyte levels in breast cancer patients with high and low mRNA *TYRO3* expression. (F) IHC staining of p-TYRO3 in lung cancer patients who received anti-PD-1/PD-L1 therapy. Resistant cases n = 12. Non-resistant cases n = 17. \*p = 0.0356, two-way unpaired t-test. Scale bars: left 50µm, right 25µm.



Supplemental Figure 2. Cell proliferation assay and pathways downstream of TYRO3. (A) Cell proliferation of *Tyro3*-OE and *Tyro3*<sup>-/-</sup> cells compared with their respective control cells, n = 3. NS p = 0.63, ns p = 0.72, two-way ANOVA. (B) Carboxyfluorescein succinimidyl ester (CFSE) staining of 4T-1P, *Tyro3*-OE, 4T1-R, and *Tyro3*<sup>-/-</sup> cells. Samples were stained on day 1 and collected on day 3 followed by flow cytometric analysis.



Supplemental Figure 3. TYRO3 favors anti-inflammatory tumor microenvironment. (A) Heat map showing normalized marker expression of each immune cell cluster from mass cytometric analysis. (B) Relative MFI of PD-L1 in tumor cells from anti-PD-1 treated 4T1-P and *Tyro3*-OE tumors, ns p = 0.96, two-way unpaired t-test. (C) Relative MFI of Caspase-3 in tumor cells from anti-PD-1 treated 4T1-P and *Tyro3*-OE tumors. Ns p = 0.356, two-way unpaired t-test.

(**D**) Relative mRNA expression of M1/M2 macrophage marker, *HLADRA1* \*p = 0.02, *MRC1* \*\*p = 0.001, *ARG1* \*p = 0.011, *IL10* \*p = 0.046, n = 2, two-tailed unpaired t-test. (**E**) Relative MFI of CD11c and CD206 in bone marrow–derived macrophages (BMDMs) cultured in the conditioned medium (CM) from 4T-1P or *Tyro3*-OE tumor cells, n = 3. CD11c \*\*p = 0.0072, CD206 \*\*p = 0.0078, and (**F**) in BMDMs cultured in the CM from 4T1-R/*Tyro3*<sup>-/-</sup> cells, n = 3. CD11c, ns p = 0.123, CD206 \*\*p = 0.0073. Two-tailed unpaired t-test. (**G**) Relative *VEGF* mRNA expression in 4T1-P and *Tyro3*-OE cells, n = 3. \*\*\*p = 0.0003, and in (**H**) BT549 and *TYRO3*-OE cells, n = 3. \*\*\*p = 0.0008. Two-tailed unpaired t-test. (**I**) Relative mRNA expression of M1/M2 macrophage marker, *CXCL10* \*\*p = 0.002, *CXCL10* plus axitinib ns p = 0.39, *HLADRA1* \*p = 0.019, *HLADRA1* plus axitinib ns p = 0.5, *IL6* \*p = 0.003, *ARG1* plus axitinib \*p = 0.047, *IL10* \*p = 0.02, *IL10* plus axitinib ns p = 0.6; n = 3, two-tailed unpaired t-test.



Supplemental Figure 4. TYRO3 inhibits tumor cell ferroptosis. (A) Relative mRNA expression of *Slc3a2*, *Fth1* and *Gpx4* in *Tyro3*-OE and *Tyro3*<sup>-/-</sup> cells, n = 3. *Slc3a2*: 4T1-P vs. *Tyro3*-OE \*p = 0.03, 4T1-R vs *Tyro3*<sup>-/-</sup>, \*\*\*p = 0.0002; *Fth1*: 4T1-P vs. *Tyro3*-OE, \*\*p = 0.003; *Gpx4*: 4T1-P vs. *Tyro3*-OE \*p = 0.014, 4T1-R vs *Tyro3*<sup>-/-</sup>, \*\*p = 0.006, two-tailed unpaired t-test. (B) Co-expression of *SLC3A2* and *TYRO3* in breast cancer patients, Pearson = 0.22, \*\*\*\*p = 2.98e<sup>-</sup>

<sup>12</sup>. (C) IHC staining of SLC3A2 and TYRO3 in melanoma tissue array, n = 77. Pearson Chi-square, \*\*\*p = 0.000273. Scale bars: left 200 $\mu$ m, right 20 $\mu$ m. (**D**) MFI of Granzyme B in CD8<sup>+</sup> T cells in anti-PD-1 treated 4T1-P and *Tyro3*-OE tumors, n = 3. Ns p = 0.75, two-tailed unpaired t-test. (E) MDA concentration in 4T1-P and Tyro3-OE cells, cells were treated by 10uM erastin for 24 hours. MDA content was normalized to protein concentration, n = 3. \*\*\*p = 0.0001, two-tailed unpaired t-test. (F) MDA concentration in 4T1-R and  $Tyro3^{-/-}$  cells, n = 3. \*\*\*p = 0.00013, two-tailed unpaired t-test. (G) Dual-luciferase reporter assay of the ARE-reporter. Cells were transfected with TYRO3 overexpression plasmid, with or without AKT dominant-negative (AKT-DN) plasmid, n = 3. \*\*\*\*p < 0.0001, ns p = 0.16, two-tailed unpaired t-test. (H) Relative lipid ROS in PY8119 cells that were primed with or without 200 nM Pros1 for 24h then treated with 2µM erastin and/or  $5\mu$ M fer-1 for 8h, n = 3. \*\*\*p = 0.0006, two-tailed unpaired t-test. (I) Effect of LDC1267 on MDA concentration in 4T1-R cells. Cells were treated by  $5\mu$ M LDC1267 for 24h, n = 3. \*\*p = 0.002, two-tailed unpaired t-test. (J) Western blot analysis of TYRO3 expression in B16F10 and Tyro3<sup>-/-</sup> cells; and in (K) PY8119 and Tvro3<sup>-/-</sup> cells. TUBULIN was used as a loading control. (L) Relative lipid ROS in B16F10 or *Tyro3<sup>-/-</sup>* cells treated with 5 $\mu$ M LDC1267 for 12h, n = 3. \*\*\*\*p < 0.0001, ns p = 0.67; two-tailed unpaired t-test. (M) Relative lipid ROS in PY8119 or  $Tyro3^{-/-}$  cells treated with  $5\mu$ M LDC1267 for 12h, n = 3. \*\*\*\*p < 0.0001, ns p = 0.08; two-tailed unpaired t-test. (N) The percentage of 7-AAD<sup>+</sup> cells in B16F10 cells or  $Tyro3^{-/-}$  cells treated with 5µM LDC1267 for 24h, n = 3. \*\*\*\*p < 0.0001, ns p = 0.33; two-tailed unpaired t-test. (**O**) The percentage of 7-AAD<sup>+</sup> cells in PY8119 cells or *Tyro3<sup>-/-</sup>* cells treated with  $5\mu$ M LDC1267 for 24h, n = 3. \*\*\*\*p < 0.0001, ns p = 0.85; two-tailed unpaired t-test.