Supporting Information

Design of a stable cyclic peptide analgesic derived from sunflower seeds that targets the κ -opioid receptor for the treatment of chronic abdominal pain

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Figure S1. (*A*) Concentration-dependent displacement of [³H]-diprenorphine (1 nM; DPN) by helianorphin-12 and 14-18 in HEK293 cells stably expressing mouse KOR (n=3). To determine specific binding, nonspecific binding was subtracted from total binding. (*B*) cAMP inhibition was measured following mouse KOR activation by helianorphin-12 (n=4). Data are normalized to 100% and are mean \pm SD.



Figure S2. Stability of helianorphin-19 was determined by UPLC-MS after incubation in 100% simulated gastric fluid (SGF) (n=3).



Figure S3. Quality control of helianorphin-19 was determined by (*A*) RP-HPLC and (*B*) MALDI mass spectrometry. The chromatogram at 215 nm indicates a purity >95% and the monoisotopic mass signal (1790.0 m/z) represents oxidized cyclic peptide.



Figure S4. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-2.



Figure S5. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-3.



Figure S6. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-4.



Figure S7. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-5.



Figure S8. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-6.



Figure S9. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-7.



Figure S10. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-8.



Figure S11. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-9.



Figure S12. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-10.



Figure S13. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-11.



Figure S14. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-12.



Figure S15. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-13.



Figure S16. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-14.



Figure S17. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-15.



Figure S18. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-16.











Figure S21. Full one-dimensional ¹H NMR spectrum (0-10 ppm) of helianorphin-19.



Figure S22. Full one-dimensional ¹³C NMR spectrum (0-200 ppm) of helianorphin-19.

Table S1. Analytical	data of synthesized peptides.
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Peptide	HPLC purity (%)*	HPLC retention times (min)	Calculated mass (m/z)	Observed mass (m/z) [#]
SFTI-1	100.0	36.9	1513.8	1513.6
helianorphin-1	99.8	33.4	2362.3	2362.0
helianorphin-2	99.7	37.0	2526.5	2526.1
helianorphin-3	99.9	38.5	1739.9	1739.6
helianorphin-4	96.0	37.6	1904.0	1903.8
helianorphin-5	99.3	10.5	1583.8	1583.8
helianorphin-6	99.3	7.6	1571.7	1571.7
helianorphin-7	100.0	10.1	1512.7	1512.7
helianorphin-8	99.5	9.2	1623.8	1623.8
helianorphin-9	98.3	8.8	1651.8	1651.9
helianorphin-10	94.2	10.0	1577.8	1577.8
helianorphin-11	93.5	10.2	1464.7	1464.7
helianorphin-12	100.0	9.8	1575.8	1575.8
helianorphin-13	100.0	10.5	1603.8	1603.8
helianorphin-14	98.9	7.2	2362.3	2362.2
helianorphin-15	99.5	7.8	2501.4	2501.4
helianorphin-16	100.0	9.6	2608.5	2608.5
helianorphin-17	100.0	9.4	2608.5	2608.9
helianorphin-18	99.1	7.3	1891.1	1891.4
helianorphin-19	99.9	20.0	1790.0	1790.0

*Purity of SFTI-1 and helianorphin 1-4 and 19 (calculated by automatic peak integration from 5-45 min) was determined by RP-HPLC using a Phenomenex Jupiter C₁₈ column (5 μ M, 300 Å, 150 x 2 mm) and a linear gradient of 5-65% solvent B in 60 min at a flow rate of 1 mL/min. Purity of helianorphin 5-18 was assessed by RP-UPLC using a Phenomenex Luna Omega column (1.6 μ m C₁₈ 100 Å, 50 x 2.1 mm) and a linear gradient of 1-61% of solvent B in 15 min at a flowrate of 0.6 mL/min was applied. Purity was calculated by automatic peak integration from 3-15 min.

[#]Monoisotopic masses of peptides obtained by ESI- and/or MALDI-MS are shown.