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Supplemental information

Patient-derived iPSC-cerebral organoid modeling of the 17q11.2 microdeletion syndrome establishes *CRLF3* as a critical regulator of neurogenesis

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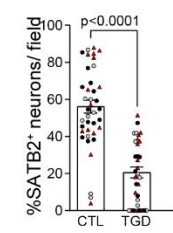
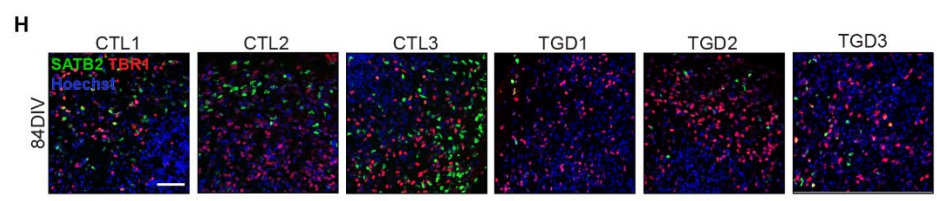
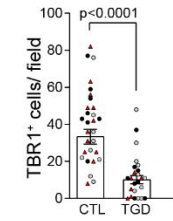
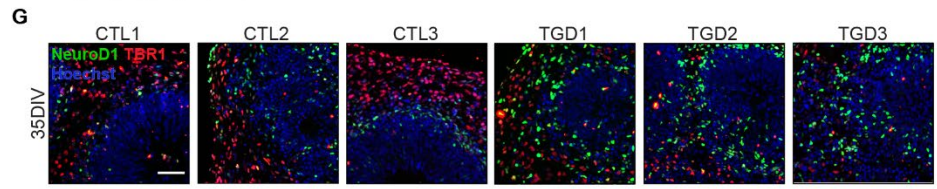
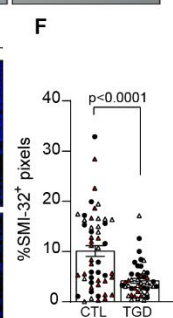
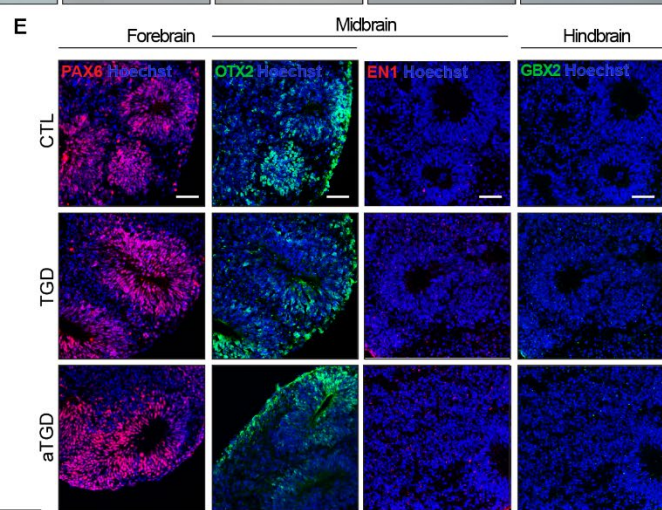
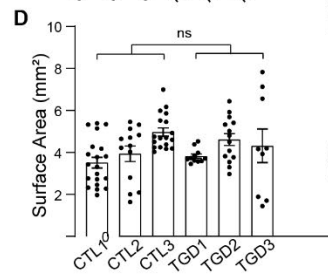
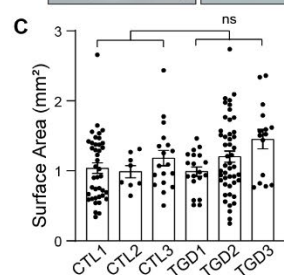
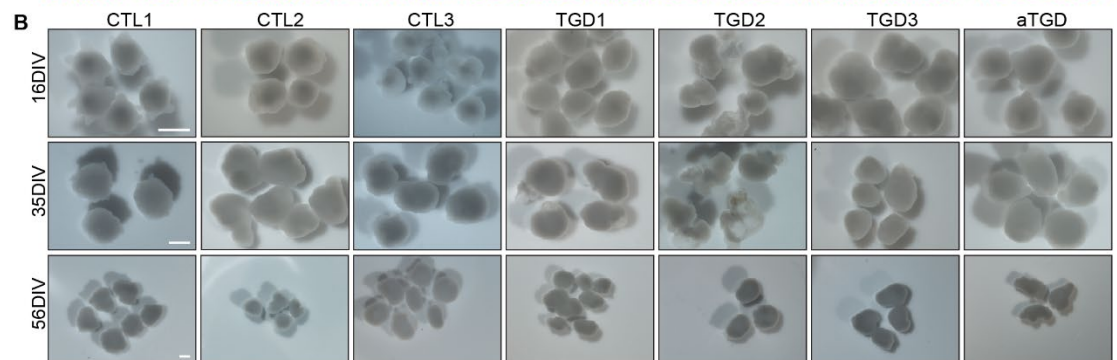
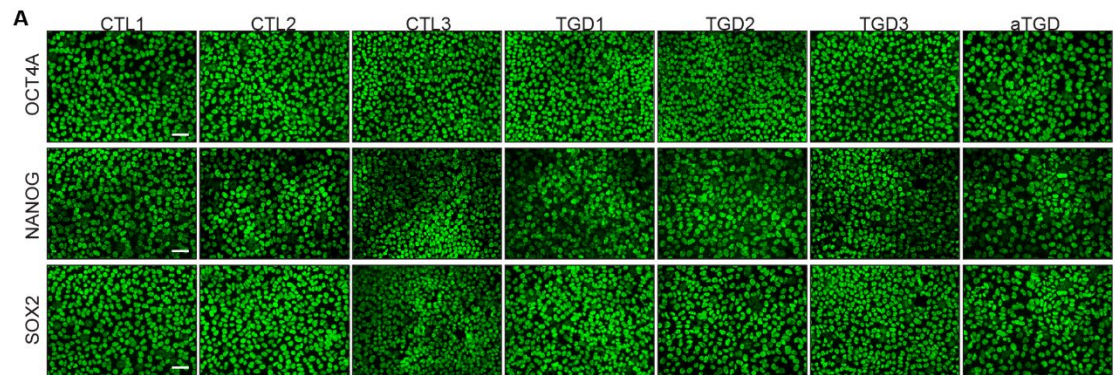


Figure S1 (related to Figure 1). Patient-derived hiPSCs and hCOs.

(A) Representative images of hiPSCs immunolabeled for pluripotency markers OCT4A, NANOG, and SOX2. Scale bars: 50 μ m. (B) Representative bright-field images of hCOs at 16, 35 and 56DIV. Scale bars: 1 mm. (C-D) Quantification of surface areas of hCOs at (C) 16DIV and (D) 84DIV. (E) Representative immunofluorescence images of 16DIV CTL (CTL1), TGD (TGD1) and aTGD hCOs immunolabeled for dorsal forebrain (PAX6, OTX2), midbrain (OTX2, EN1) and hindbrain (GBX2) markers. Scale bars: 50 μ m. (F) (related to **Figure 1I**) Quantitation of SMI-32⁺ immunopositive dendrites in 35DIV TGD relative to CTL hCOs. (G) Representative images of 35DIV CTL and TGD hCOs immunolabeled for early-stage immature neurons (NeuroD1) and deep-layer cortical neurons (TBR1) and quantification of the number of TBR1⁺ deep-layer neurons per image field in hCOs at 35DIV. (H) Representative images of 84DIV CTL and TGD hCOs immunolabeled for deep-layer (TBR1) and upper-layer (SATB2) neurons and quantification of %SATB2⁺ upper-layer neurons in hCOs at 84DIV. Scale bars, 100 μ m.

Independent hiPSC lines representing three different CTL or TGD lines (black, CTL1 / TGD1; white, CTL2 / TGD2; red, CTL3 / TGD3) are shown. Data are expressed as the mean \pm SEM. Each data point represents one hCO, 2-6 hCOs per experimental replicate, 3-5 experimental replicates per genotype. Statistical analysis by unpaired, two-tailed *t*-test or one-way ANOVA with Bonferroni multiple comparisons test.

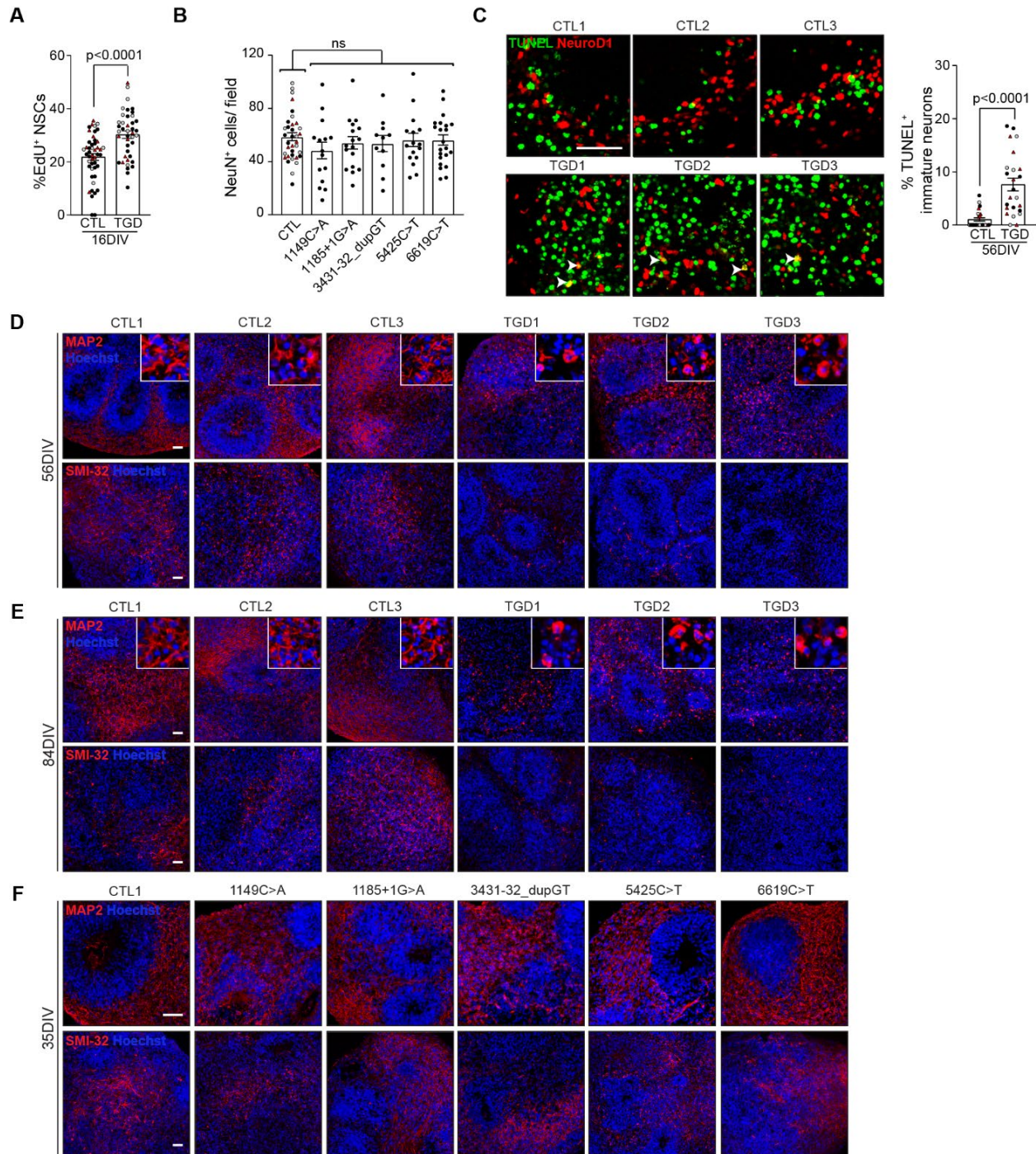
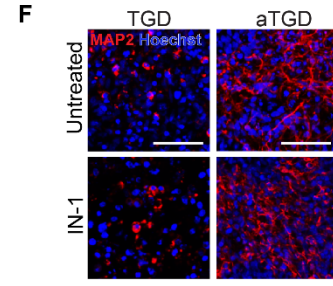
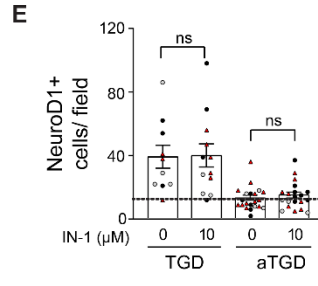
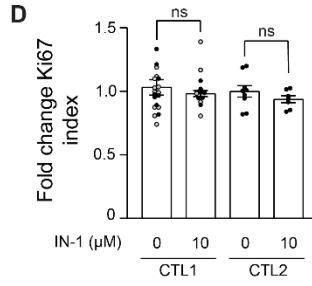
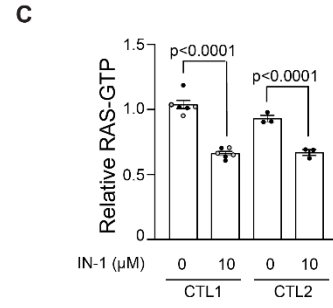
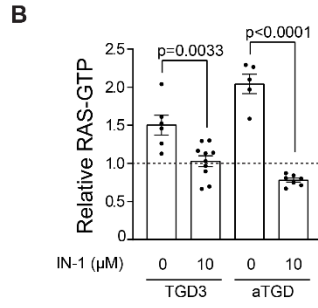
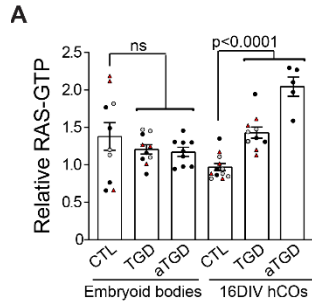


Figure S2 (related to Figure 1). Neuronal differentiation defects in TGD and intragenic *NF1*-mutant hCOs.

(A) %EdU⁺ neural stem cells (NSCs) in 16DIV CTL and TGD hCOs. **(B)** Quantification of late-stage immature (NeuN⁺) neurons per image field in the SVZ of intragenic *NF1*-mutant hCOs

relative to CTL hCOs at 35DIV. **(C)** Representative images and quantification of CTL and TGD hCOs immunolabeled for TUNEL (green) and NeuroD1 (red) (co-localization indicated by white arrows) at 56DIV. **(A-C)** Independent hiPSC lines (black, CTL1 / TGD1; white, CTL2 / TGD2; red, CTL3 / TGD3) are shown. **(D-E)** Representative images of CTL and TGD hCOs immunolabeled for MAP2⁺ and SMI-32⁺ dendrites at **(D)** 56DIV and **(E)** 84DIV. **(F)** Representative control (CTL1) and intragenic *NF1*-mutant hCOs immunolabeled for dendrite-specific markers (MAP2⁺, SMI-32⁺) at 35DIV. Data are shown as the mean \pm SEM. Each data point represents one biological replicate (hCO), 2-6 biological replicates per experimental replicate, 3-5 experimental replicates per genotype. Statistical analysis by unpaired, two-tailed *t*-test or one-way ANOVA with Dunnett's multiple comparisons test. Scale bars: 50 μ m.



G

| Gene | CTL | | | TGD | | | P value | aTGD | | | P value |
|------------------|------|------|----|------|------|----|---------|------|------|----|---------|
| | Mean | SD | N | Mean | SD | N | | Mean | SD | N | |
| <i>RAB11FIP4</i> | 1.00 | 0.38 | 20 | 0.29 | 0.14 | 22 | <0.0001 | 0.56 | 0.19 | 13 | 0.0006 |

H

| Gene | CTL1 | | | CTL2 | | | CTL3 | | | F / P value |
|----------------|------|------|---|------|------|---|------|------|---|----------------|
| | Mean | SD | N | Mean | SD | N | Mean | SD | N | |
| <i>MIR193A</i> | 0.46 | 0.24 | 4 | 0.18 | 0.06 | 6 | 2.12 | 1.13 | 4 | 13.26 / 0.0012 |
| <i>MIR365B</i> | 1.70 | 1.04 | 4 | 0.48 | 0.11 | 4 | 0.58 | 0.18 | 4 | 9.98 / 0.0247 |
| <i>MIR4725</i> | 1.33 | 0.79 | 6 | 0.36 | 0.06 | 4 | 4.09 | 1.47 | 4 | 16.19 / 0.0082 |

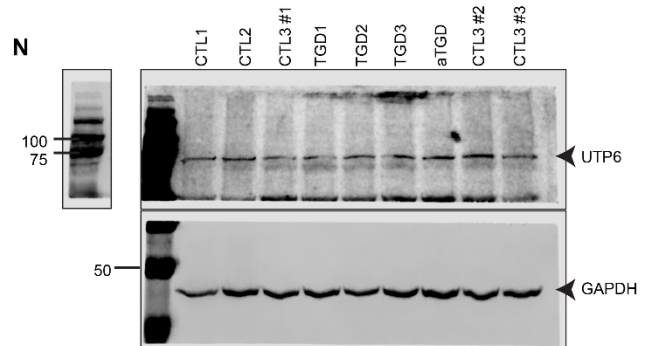
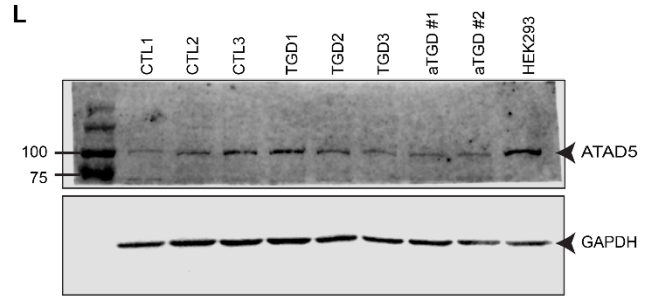
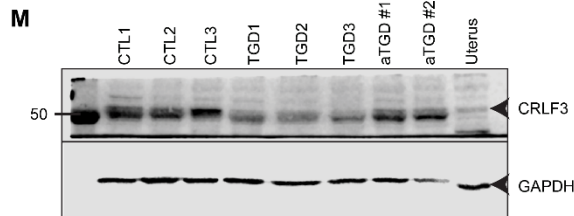
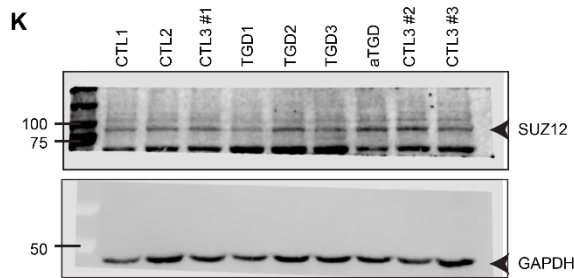
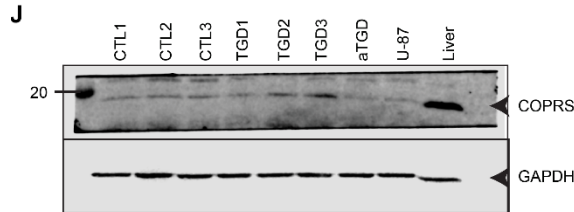
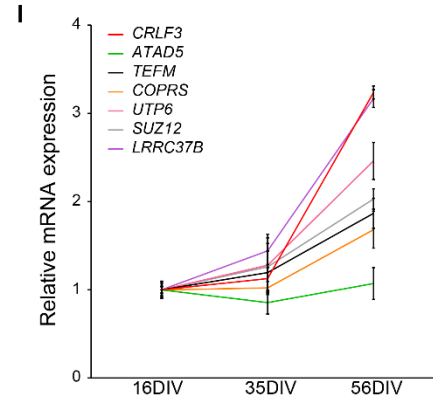


Figure S3 (related to Figure 2 and 3). RAS activity and differential gene expression analysis of TGD and CTL hCOs.

(A) RAS activation in CTL and TGD 8DIV embryoid bodies and 16DIV hCOs. (B-C) Reduced RAS activity in (B) TGD3 and aTGD 16DIV hCOs and (C) CTL1 and CTL2 hCOs following 10 μ M pan-RAS-IN-1 (IN-1) treatment. The mean CTL hCO RAS activity was assigned a value of 1 (dotted line). (A-C) Each data point represents an independent experimental replicate consisting of 20 pooled embryoid bodies or 4 pooled hCOs. Statistical analysis by unpaired, two-tailed *t*-test or one-way ANOVA with Dunnett's multiple comparisons. (D) Quantification of NSC proliferation (fold change in %Ki67⁺ NSCs) in control hCOs at 16DIV with or without IN-1 treatment. Each data point represents one hCO, 2-6 hCOs per experimental replicate, 3-5 experimental replicates per genotype. Statistical analysis by unpaired, two-tailed *t*-test. (E) Number of early-stage immature (NeuroD1⁺) neurons per image field in the SVZ of 16DIV TGD3 and aTGD hCOs with and without IN-1 treatment. Each data point represents one hCO, 3-12 hCOs per clone. Statistical analysis by unpaired, two-tailed *t*-test comparing TGD3 and aTGD hCOs with control values (indicated by dotted line). (A-E) All data are shown as the mean \pm SEM. Independent (A, E) hiPSC lines (black, CTL1 / TGD1 / aTGD1; white, CTL2 / TGD2 / aTGD2; red, CTL3 / TGD3, aTGD-3), or (C-D) independent hiPSC clones (black, clone 1; white, clone 2; red, clone 3) are shown. (F) Representative images of 16DIV TGD3 and aTGD hCOs with and without RAS-IN-1 treatment immunolabeled for MAP2⁺ dendrites. Scale bars: 50 μ m. (G) mRNA expression of *RAB11FIP4* in 56DIV hCOs showing gene deletion status in TGD1-3 and aTGD. Statistical analysis by unpaired, two-tailed *t*-test. (H) RT-qPCR analysis of microRNA gene expression in CTL hCOs at the time point of highest expression (16DIV). Statistical analysis by one-way ANOVA; *F*-ratio / *P* values reported. *MIR4733* was not expressed in CTL hCOs. Each mRNA expression data point represents one biological replicate (hCO), 2-3 hCOs per experimental replicate. (I) Time course analysis of mRNA expression in 16, 35 and 56DIV CTL hCOs for 7 protein-coding genes included in differential gene expression

analysis, illustrating highest transcript expression levels for 6 of the 7 genes at 56DIV. *ATAD5* had no change in expression over time. Each time point represents 2 independent experimental replicates of CTL1 hCOs with each experimental replicate containing 2 biological replicates (hCOs). Data are shown as the mean \pm SEM. (**J-N**) Representative unprocessed western blots of CTL and TGD protein expression including (**J**) COPRS, (**K**) SUZ12, (**L**) ATAD5, (**M**) CRLF3 and (**N**) UTP6.

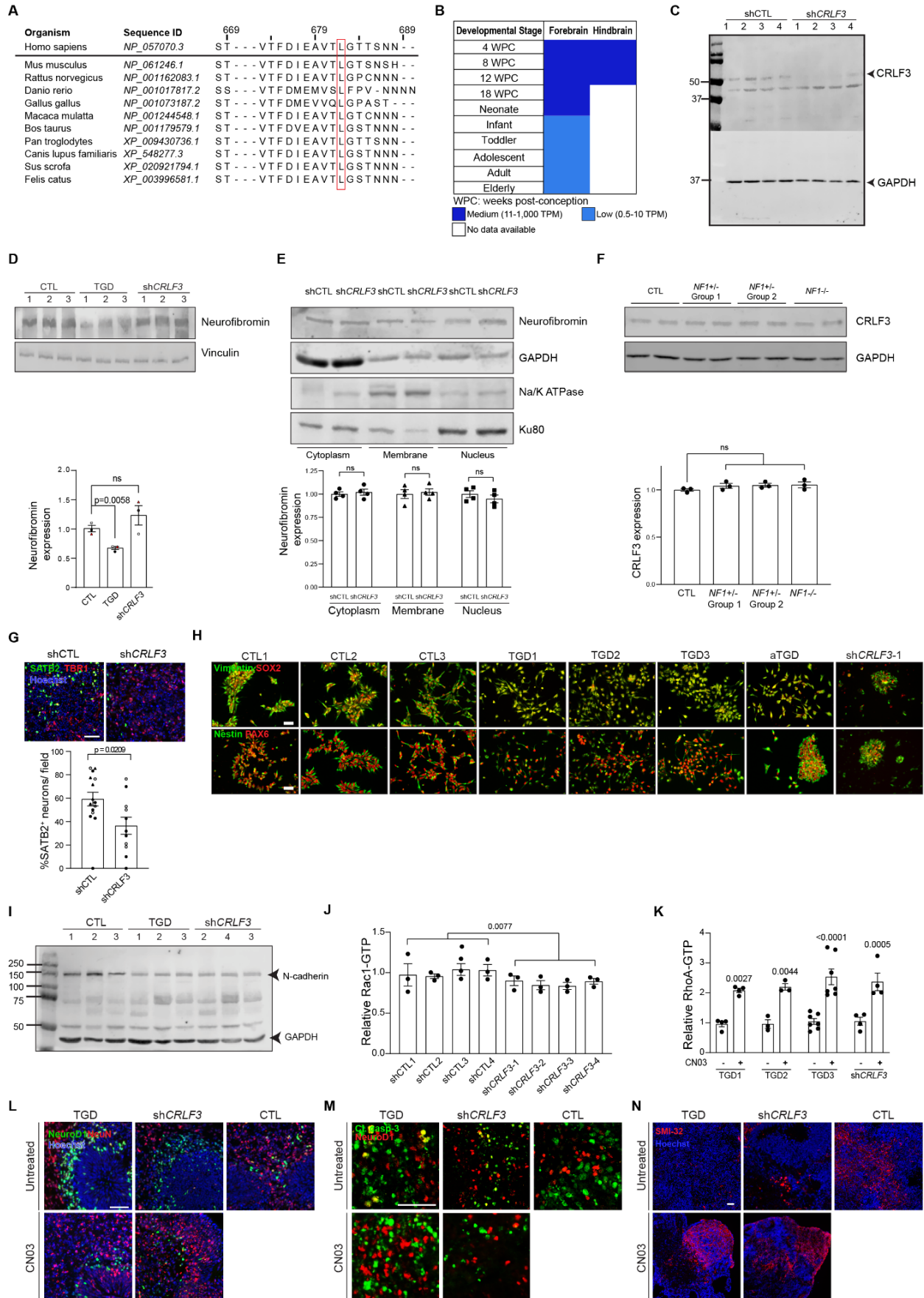


Figure S4 (related to Figures 3 and 4). *CRLF3* sequence conservation, developmental expression, and downstream signaling.

(A) Amino acid sequence alignments revealed 92.8% conservation in p.Leu389 between human and 303 vertebrate *CRLF3* orthologs. Ten representative orthologs from NCBI's Eukaryotic Genome Annotation pipeline are shown, with p.Leu389 outlined in red. (B) Heat map of *CRLF3* mRNA expression levels in the human forebrain and hindbrain at different developmental stages, as reported by the Expression Atlas: Human RNA-seq time-series of the development of seven major organs. TPM: transcripts per million. (C) Uncropped western immunoblot from **Figure 4A**. (D) Neurofibromin relative expression in CTL, TGD, and sh*CRLF3* hiPSC-derived NSCs. Independent hiPSC lines (black, CTL1 / TGD1 / sh*CRLF3*-1; white, CTL2 / TGD2 / sh*CRLF3*-2; red, CTL3 / TGD3 / sh*CRLF3*-3) are shown. Statistical analysis by unpaired, two-tailed *t*-test. (E) Immunoblots and quantitation of neurofibromin expression in different subcellular fractions (cytoplasm, membrane, nucleus) in shCTL and sh*CRLF3* NPCs. GAPDH (cytoplasm), Na/K ATPase (membrane) and human-specific Ku80 (nucleus) were used as loading controls. (F) Immunoblot and quantitation of *CRLF3* expression in NPCs harboring *NF1* point mutations, either conferring <30% reduced (Group 1), or >70% reduced (Group 2) neurofibromin levels, NPCs harboring homozygous null *NF1* mutations (*NF1*^{-/-}), or non-mutant controls. GAPDH was used as a loading control. (E-F) Data are expressed as the mean ± SEM. Statistical analysis by (E) unpaired, two-tailed *t*-test or (F) one-way ANOVA with Bonferroni post-test correction. ns, not significant. (G) Representative images of 84DIV shCTL and sh*CRLF3* hCOs immunolabeled for deep-layer (TBR1) and upper-layer (SATB2) neurons and quantification of %SATB2⁺ upper-layer neurons in hCOs at 84DIV. Scale bar: 100 μm. (H) hiPSC-derived NSCs immunolabeled for NSC markers SOX2, Vimentin, Nestin and PAX6. Scale bar: 50μm. (I) Unprocessed western immunoblot from **Figure 4H**. (J) Rac1 activity levels in shCTL and sh*CRLF3* NSCs. Each data point represents individual NSC sample. Statistical analysis by unpaired, two-tailed *t*-test. (K) RhoA activity in 2DIV TGD and sh*CRLF3* hCOs with

and without 1 $\mu\text{g}/\text{mL}$ CN03 RhoA activator (CN03) treatment for 24 hours. Each data point represents 6 pooled hCOs. Statistical analysis by two-way ANOVA with Sidak's multiple comparison test performed comparing untreated with treated hCOs. All data are shown as the mean \pm SEM and the *P* values are shown above each bar. **(L-N)** Representative images of **(L)** NeuroD1⁺ (green)/ NeuN⁺ (red) neurons, **(M)** cleaved caspase-3⁺ apoptotic immature neurons and **(N)** SMI-32⁺ dendrites in 35DIV CTL, TGD and *shCRLF3* hCOs with and without CN03 treatment. Scale bars: 50 μm .

Table S1 (related to Figure 1). Patient-derived CTL1-3, TGD1-3 and aTGD (atypical TGD) hiPSC lines and isogenic hiPSC lines CRISPR/Cas9-engineered to harbor NF1 patient *NF1* gene mutations.

| Genotype | Sex | Age (years) | Specimen source | No. clones |
|----------------------|--------------------|-------------|-----------------|------------|
| CTL1 ^a | Male | Fetal | Skin biopsy | 2 |
| CTL2 ^b | Male | 27 | Skin biopsy | 1 |
| CTL3 ^c | Male | 41 | Skin biopsy | 1 |
| TGD1 | Male | 44 | Skin biopsy | 2 |
| TGD2 | Male | 6 | Urine | 2 |
| TGD3 | Male | 11 | Blood | 3 |
| aTGD | Female | 16 | Blood | 3 |
| NF1 patient mutation | Protein level | | Mutation Type | No. clones |
| c.1149C>A | p.Cys383X | | Nonsense | 2 |
| c.1185+1G>A | p.Asn355_Lys395del | | Splice site | 2 |
| c.3431-32_dupGT | p.Thr1145Val_FS | | Frameshift | 2 |
| c.5425C>T | p.Arg1809Cys | | Missense | 2 |
| c.6619C>T | p.Gln2207X | | Nonsense | 1 |

^aBJFF.6 commercially available

^bDr. Matthew B. Harms (WUSM)

^cDr. Fumihiko Urano (WUSM)

Table S2 (related to Figure 3). Human genomic DNA whole-exome sequencing.

| Patient ID | SRS-2 | Age (years) | Sex | <i>CRLF3</i>-mutation | <i>NF1</i>-mutation |
|-------------------|--------------|--------------------|------------|------------------------------|----------------------------|
| OtB3317 | 81 | 10 | M | c.1166T>C | c.5305C>T |
| OtC6610 | 48 | 11 | F | | c.3137_3138delCA |
| OtB3335 | 64 | 11 | M | | c.1756_1759delACTA |
| OtB3325 | 45 | 11 | F | | c.3888T>G |
| OtC6607 | 70 | 11 | F | | c.3449C>T |
| OtB3313 | 98 | 13 | M | c.1166T>C | c.7255_7256delCT |
| OtC6614 | 48 | 13 | M | | c.2965G>T |
| OtC6612 | 50 | 13 | M | c.1166T>C | c.910C>T |
| OtB3333 | 91 | 13 | M | c.1166T>C | c.204+1G>T |
| OtB3326 | 54 | 15 | F | c.1166T>C | c.2125T>C |
| OtB3321 | 88 | 15 | M | | c.6855C>A |
| OtB3312 | 98 | 15 | M | c.1166T>C | c.4514delG |
| OtC6619 | 46 | 16 | F | | c.4006C>T |
| OtC6615 | 76 | 16 | M | | c.205-19T>A |
| OtB3319 | 74 | 16 | F | c.1166T>C | c.4985G>A |
| OtB3323 | 56 | 17 | M | | c.1885G>A |
| OtB3336 | 46 | 18 | M | | c.3520C>T |

Table S3 (related to Figure 4). Differentially expressed gene list filtered for non-significant genes in the comparison of TGD vs sh*CRLF3* samples.

| Gene symbol | P value (TGD vs. sh<i>CRLF3</i>) | FDR step up (TGD vs. sh<i>CRLF3</i>) | Fold change (TGD vs. sh<i>CRLF3</i>) |
|--------------------|---|---|---|
| <i>KCP</i> | 0.1073 | 0.1689 | 2.99 |
| <i>SPN</i> | 0.2332 | 0.3201 | 2.90 |
| <i>THSD7A</i> | 0.0109 | 0.0248 | 2.84 |
| <i>MMP23B</i> | 0.0177 | 0.0373 | 2.67 |
| <i>ACOT11</i> | 0.0456 | 0.0826 | 2.25 |
| <i>ASCL1</i> | 0.9106 | 0.9346 | 1.82 |
| <i>DACT1</i> | 0.0700 | 0.1184 | 1.75 |
| <i>LDHAP4</i> | 0.7063 | 0.7715 | 1.29 |
| <i>ADGRE5</i> | 0.7043 | 0.7696 | 1.25 |
| <i>RUBCNL</i> | 0.8635 | 0.8992 | 1.20 |
| <i>NEFM</i> | 0.5160 | 0.6030 | 1.13 |
| <i>EPB41L4A</i> | 0.9420 | 0.9570 | -1.09 |
| <i>TENM2</i> | 0.2461 | 0.3342 | -1.27 |
| <i>MDGA2</i> | 0.1567 | 0.2308 | -1.27 |
| <i>CAMK4</i> | 0.5252 | 0.6119 | -1.31 |
| <i>SORBS2</i> | 0.0812 | 0.1341 | -1.64 |
| <i>ATCAY</i> | 0.1408 | 0.2112 | -1.69 |
| <i>PTX3</i> | 0.0732 | 0.1229 | -1.72 |
| <i>MANEAL</i> | 0.0455 | 0.0825 | -1.78 |
| <i>ITGB8</i> | 0.0134 | 0.0296 | -1.86 |
| <i>DCLK2</i> | 0.0091 | 0.0214 | -1.91 |
| <i>SYT5</i> | 0.0236 | 0.0474 | -2.25 |
| <i>SYP</i> | 0.0102 | 0.0234 | -2.25 |
| <i>RASGRP1</i> | 0.0241 | 0.0481 | -2.58 |
| <i>MSI1</i> | 0.0047 | 0.0121 | -2.58 |
| <i>CRABP1</i> | 0.0052 | 0.0134 | -2.82 |
| <i>FCHO1</i> | 0.0056 | 0.0142 | -2.85 |
| <i>ECEL1</i> | 0.0114 | 0.0257 | -2.90 |
| <i>PLEKHA7</i> | 0.0097 | 0.0225 | -3.02 |
| <i>ULBP1</i> | 0.0051 | 0.0131 | -3.43 |
| <i>MMRN1</i> | 0.0103 | 0.0237 | -3.89 |

Table S4 (related to Figures 1-4). Summary of experimental samples, replicates and statistical tests used.

| Relative to main Figure | Readout | Sample type | Assay | Quantitation | Independent CTL lines used | Independent mutant lines used | Min. times experiment performed | Min. biological replicates/line or clone | Statistical test used |
|-------------------------|-------------------|-------------------|------------------|--------------------|-------------------------------------|-------------------------------------|---------------------------------|---|--|
| Figure 1 | Ki67 | hCOs | IF: Figure 1B | Figure 1D | 3 CTL | 1 aTGD | 3 | 7 | unpaired, 2-tailed t-test |
| | NeuroN1 | hCOs | IF: Figure 1C | Figure 1E | 3 CTL | 4 (TGD1, TGD2, TGD3, aTGD) | 3 | 8 | unpaired, 2-tailed t-test |
| | Cleaved Caspase-3 | hCOs | IF: Figure 1C | Figure 1F | n/a | 2 (TGD3, aTGD) | 3 | 7 | unpaired, 2-tailed t-test |
| | MAP2 | hCOs | IF: Figure 1H | Figure 1G | 3 CTL | 1 aTGD | 3 | 10 | unpaired, 2-tailed t-test |
| | SMI-32 | hCOs | IF: Figure 1I | n/a | 3 CTL cell lines (CTL1, CTL2, CTL3) | 3 TGD cell lines (TGD1, TGD2, TGD3) | 3 | 9 | n/a |
| | SMI-312 | hCOs | IF: Figure 1J | Figure 1F | 3 CTL | 1 aTGD | 3 | 11 | unpaired, 2-tailed t-test |
| | SMI-32 | 2D neurons | IF: Figure 1J | Figure 1J | 3 CTL | 1 aTGD | 3 | 9 | n/a |
| | TBR1 | hCOs | IF: Figure 1G | Figure 1I | 3 CTL | 1 aTGD | 3 | 6 | 1-way ANOVA with Bonferroni correction |
| | SATB2 | hCOs | IF: Figure 1H | Figure 1H | 3 CTL | 1 aTGD | 3 | 6 | unpaired, 2-tailed t-test |
| | Ki67 | hCOs | IF: Figure 2B | Figure 2B | 3 CTL | 3 TGD | 3 | 10 | unpaired, 2-tailed t-test |
| | RAS-GTP | hCOs | ELISA: Figure 2C | Figure 2C | 3 CTL | 4 (TGD1, TGD2, TGD3, aTGD) | 3 | 7 | unpaired, 2-tailed t-test |
| Ki67 | hCOs + IN-1 | IF: Figure 2D | Figure 2D | n/a | 2 (TGD3, aTGD) | 3 | 12 | 1-way ANOVA with Bonferroni correction | |
| NeuroD1 | hCOs | IF: Figure 2E | n/a | 3 CTL | 1 aTGD | 3 | 9 | n/a | |
| NeuN | hCOs | IF: Figure 2F | Figure 2E | 3 CTL | 1 aTGD | 3 | 9 | unpaired, 2-tailed t-test | |
| Cleaved Caspase-3 | hCOs | IF: Figure 2F | Figure 2F | 3 CTL | 1 aTGD | 3 | 8 | unpaired, 2-tailed t-test | |
| MAP2 | hCOs | IF: Figure 2G | n/a | 3 CTL | 1 aTGD | 3 | 9 | n/a | |
| SMI-32 | hCOs | IF: Figure 2G | n/a | 3 CTL | 1 aTGD | 3 | 9 | n/a | |
| SMI-312 | hCOs | IF: Figure 2G | n/a | 3 CTL | 1 aTGD | 3 | 9 | n/a | |
| EDU | hCOs | n/a | Figure 2A | 3 CTL | 3 TGD | 3 | 8 | n/a | |
| NeuN | hCOs | n/a | Figure 2B | 3 CTL | 5 intragenic cell lines | 3 | 15 | unpaired, 2-tailed t-test | |
| TUNEL | hCOs | IF: Figure 2C | Figure 2C | 3 CTL | 3 TGD | 3 | 7 | 1-way ANOVA with Bonferroni correction | |
| RAS-GTP | EBs | ELISA: Figure 3A | Figure 3A | 3 CTL | 4 (TGD1, TGD2, TGD3, aTGD) | 3 | 10 | unpaired, 2-tailed t-test | |
| RAS-GTP | hCOs | ELISA: Figure 3A | Figure 3A | 3 CTL | 4 (TGD1, TGD2, TGD3, aTGD) | 3 | 12 | 1-way ANOVA with Dunnett's multiple comparison | |
| RAS-GTP | hCOs + IN-1 | ELISA: Figure 3B | Figure 3B | n/a | 2 (TGD3, aTGD) | 3 | 9 | 1-way ANOVA with Dunnett's multiple comparison | |
| RAS-GTP | hCOs + IN-1 | ELISA: Figure 3C | Figure 3C | 2 CTL (CTL1, CTL2) | n/a | 3 | 12 | unpaired, 2-tailed t-test | |
| Ki67 | hCOs + IN-1 | n/a | Figure 3D | 2 CTL (CTL1, CTL2) | n/a | 3 | 6 | unpaired, 2-tailed t-test | |
| NeuroD1 | hCOs + IN-1 | n/a | Figure 3E | n/a | 2 (TGD3, aTGD) | 3 | 3 | unpaired, 2-tailed t-test | |
| CRF3 | hCOs | WB: Figure 3C | Figure 3C | 3 CTL | 4 (TGD1, TGD2, TGD3, aTGD) | 4 | 4 | unpaired, 2-tailed t-test | |
| Figure 3 | SRS-2 Score | patient DNA | SRS-2 analysis | Figure 3E | 10 patients without CRF3 mutation | 7 patients with CRF3 mutation | n/a | 7 | unpaired, 2-tailed t-test |
| | mRNA Expression | hCOs | mRNA expression | Figure S3I | 1 CTL (CTL1) | n/a | 2 | 4 | n/a |
| | CRF3 | shCRF3 iPSCs | WB: Figure 4A | Figure 4A | 4 shCTL | 4 shCRF3 | 3 | 4 | unpaired, 2-tailed t-test |
| | Ki67 | shCRF3 hCOs | n/a | Figure 4B | 4 shCTL | 4 shCRF3 | 3 | 9 | unpaired, 2-tailed t-test |
| | NeuroD1 | shCRF3 hCOs | IF: Figure 4C | Figure 4C | 4 shCTL | 4 shCRF3 | 3 | 4 | unpaired, 2-tailed t-test |
| | NeuN | shCRF3 hCOs | IF: Figure 4C | Figure 4C | 4 shCTL | 4 shCRF3 | 3 | 4 | unpaired, 2-tailed t-test |
| | Cleaved Caspase-3 | shCRF3 hCOs | IF: Figure 4D | Figure 4D | 4 shCTL | 4 shCRF3 | 3 | 3 | unpaired, 2-tailed t-test |
| | SMI-32 | shCRF3 hCOs | IF: Figure 4E | Figure 4E | 4 shCTL | 4 shCRF3 | 3 | 3 | unpaired, 2-tailed t-test |
| | N-cadherin | NSCs | WB: Figure 4H | Figure 4H | 3 CTL | 6 (3 TGD; 3 shCRF3) | 3 | 3 | 1-way ANOVA with Bonferroni correction |
| | Rac1-GTP | NSCs | ELISA: Figure 4I | Figure 4I | 3 CTL | 3 TGD | 3 | 3 | unpaired, 2-tailed t-test |
| | RhoA-GTP | NSCs | ELISA: Figure 4J | Figure 4J | 3 CTL | 3 TGD | 3 | 3 | unpaired, 2-tailed t-test |
| RhoA-GTP | shCRF3 NSCs | ELISA: Figure 4K | Figure 4K | 4 shCTL | 4 shCRF3 | 3 | 3 | unpaired, 2-tailed t-test | |
| NeuroD1 | hCOs + CN03 | IF: Figure S4L | Figure 4L | 3 CTL | 3 TGD; 2 shCRF3 | 3 | 5 | unpaired, 2-tailed t-test | |
| NeuN | hCOs + CN03 | IF: Figure S4L | Figure 4M | 3 CTL | 3 TGD; 2 shCRF3 | 3 | 4 | 1-way ANOVA with Tukey multiple comparison | |
| Cleaved Caspase-3 | hCOs + CN03 | IF: Figure S4M | Figure 4N | 3 CTL | 3 TGD; 2 shCRF3 | 3 | 3 | 1-way ANOVA with Tukey multiple comparison | |
| SMI-32 | hCOs + CN03 | IF: Figure S4N | Figure 4O | 3 CTL | 3 TGD; 2 shCRF3 | 3 | 3 | 1-way ANOVA with Tukey multiple comparison | |
| Neurofibromin | NSCs | WB: Figure S4E | Figure S4E | 3 shCTL | 3 shCRF3 | 3 | 3 | unpaired, 2-tailed t-test | |
| Neurofibromin | NSCs | WB: Figure S4E | Figure S4E | 3 shCTL | 3 shCRF3 | 3 | 3 | unpaired, 2-tailed t-test | |
| CRF3 | hCOs | WB: Figure S4F | Figure S4F | 2 CTL | 4 (NF1+, 2 NF1-) | 3 | 3 | 1-way ANOVA with Bonferroni correction | |
| SATB2 | shCRF3 hCOs | IF: Figure S4G | Figure S4G | 3 shCTL | 2 shCRF3 | 3 | 5 | unpaired, 2-tailed t-test | |
| Rac1-GTP | shCRF3 NSCs | ELISA: Figure S4J | Figure S4H | 4 shCTL | 4 shCRF3 | 3 | 3 | unpaired, 2-tailed t-test | |
| RhoA-GTP | NSCs + CN03 | ELISA: Figure S4K | Figure S4I | n/a | 3 TGD; 2 shCRF3 | 3 | 3 | 2-way ANOVA with Sidak's multiple comparison test | |