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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	\square The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
\boxtimes	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	A description of all covariates tested
\boxtimes	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
\boxtimes	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

No software is used in data collection

Data analysis

Using liftover code to convert reference genome from NCBI37/mm9 to GRCm38/mm10

Using TopDom code to identify the TAD boundaries in populated Hi-C data $\,$

The code of Si-C method is available on https://github.com/TheMengLab/Si-C/tree/master/Si-C_code RMSD calculation https://github.com/TheMengLab/Si-C/tree/master/analysis/structure_analysis/analysis/align/rmsd

Separation score calculation https://github.com/TheMengLab/Si-C/tree/master/analysis/structure_analysis/analysis/analysis/align/sepscore_gyr

Identifying domain boundaries in single cells https://github.com/TheMengLab/Si-C/tree/master/analysis/structure_analysis/analysis/align/

sepscore_gyr/boundary_chr

 $Identify\ chromosome\ compartment\ https://github.com/TheMengLab/Si-C/tree/master/analysis/compartment$

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about <u>availability of data</u>

Dual use research of concern

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

	tures by Si-C are presented at http TheMengLab/Nuc_3D_structure_C	s://github.com/TheMengLab/Si-C_3D_structure. 3D structures generated by Nucdynamics are presented at ell1.	
Field-spe	ecific reporting		
Please select the o	one below that is the best fit for	your research. If you are not sure, read the appropriate sections before making your selection.	
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For a reference copy of	the document with all sections, see <u>nat</u>	ure.com/documents/nr-reporting-summary-flat.pdf	
Life scier	nces study des	ign	
All studies must di	sclose on these points even wh	en the disclosure is negative.	
Sample size	8 cell samples are used in this work because only single-cell Hi-C data of these 8 cells are deposited in Laue's work.		
Data exclusions	No data is excluded in our analysis.		
Replication	For each cell, we constructed 20 3D structure replicas for different resolution. All of these structure replicas satisfy >99% constraints derived from Hi-C data. The chromosome structures exhibited by different replicas are highly consistent.		
Randomization	From single-cell Hi-C data of each cell, we calculated 20 structure replicas with initial structure randomly generated using self-avoidance random walk algorithm.		
Blinding	The investigators were blinded to group allocation while data obtaining and analysis		
<u> </u>	<u> </u>	materials, systems and methods s of materials, experimental systems and methods used in many studies. Here, indicate whether each material,	
		are not sure if a list item applies to your research, read the appropriate section before selecting a response.	
	perimental systems	Methods	
n/a Involved in the	·	n/a Involved in the study	
Antibodies		ChIP-seq	
Eukaryotic cell lines		Flow cytometry	
Palaeontology and archaeology		MRI-based neuroimaging	
	Animals and other organisms		
	Human research participants		
Clinical da	ta		