Supplementary information

Enrichment of NPC1-deficient cells with the lipid LBPA stimulates autophagy, improves lysosomal function, and reduces cholesterol storage

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Supplemental Figures



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Fig. S1. Related to Fig. 1. Phospholipid composition of NPC1 mutant fibroblasts treated with PG.

A, *r*epresentative filipin images and quantification of filipin intensity in parental (WT) and PG or PC treated CRISPR-Cas9 edited NPC1 KO HeLa cells. Western blot demonstrates stable NPC1 gene deletion. In Western Blot samples were analyzed in biological replicates. For each condition one band was cropped and bands for WT and NPC1 KO were spliced. Splicing is indicated with a vertical line. In images, bar=30µm. Data from 2 independent experiments. N=100-200 cells/condition. *B*, LC-MS analysis of NPC1-mutant fibroblasts treated with 100µM PG for 12, 24 and 48h. Bar graph indicate fold-change of average relative mol% of all lipids measured, normalized to untreated CTR. N=2. *C*, LC-MS analysis of NPC1-mutant fibroblasts treated with 100µM PC for 24h. Bar graph indicate fold-change of average relative mol% of all lipids measured, normalized to CTR. *D*, LC-MS analysis of cardiolipin in NPC1-deficient fibroblasts treated with 100µM PC for 24 and 48h relative to untreated CTR. *E*, Cell viability assay in NPC1 deficient cells treated with PG. All graphed data show mean ±SD. *p<0.05, **:p<0.01, ***:p<0.001 compared to untreated cells (CTR) in two-tailed Student's test.

Nomenclature abbreviations are: PA, phosphatidic acid; PC, phosphatidylcholine; PCe, ether phosphatidylcholine; PE, phosphatidylethanolamine; PEp, plasmalogen phosphatidylethanolamine; PG, phosphatidylglycerol; PI, phosphatidylinositol; PS, phosphatidylserine; LPC, lysophosphatidylcholine; LPCe, ether lysophosphatidylcholine; LPE, lysophosphatidylethanolamine; LPEp, plasmalogen lysophosphatidylethanolamine; LPI, lysophosphatidylinositol; LBPA lysobisphosphatidic acid; AcylPG acyl phosphatidylglycerol, NAPE N-acyl phosphatydylethanonamine, NAPS N-acyl-phosphatidylserine, NSer Nacylserine



Fig. S2. Related to Fig. 2 and 3. PG treatment does not alter early autophagic markers in NPC1-deficient fibroblasts

A, Western Blot analysis and quantification of LC3-II and p62 in control and PG-treated NPC1 KO HeLa cells at 48h post-treatment. Data from 3-5 independent experiments. On the upper panel samples were analyzed in biological replicates. For each condition one band was cropped and bands for CTR and PG were spliced. Splicing is indicated with a vertical line. *B*, Representative images and quantification of LC3-positive puncta in NPC1 fibroblasts at 48h. Scale bar, 20µm. N>60 cells/condition. *C*, Representative confocal images and quantification of LC3 staining in control and PG-treated NPC1 KO HeLa cells at 48h post-treatment. *D*, Representative images and quantification of LC3 puncta in neural stem cells treated with 50µM PC or PG liposomes. *E*, Western Blot analysis and quantification of autophagy markers in NPC1-deficient fibroblasts. Splicing is indicated with a vertical line. *F*, Time course of labeled with LAMP2 LY distribution in NPC1 deficient fibroblasts treated with PG. Insets illustrate the size of LY. N-nuclei. Scale bar, 20µm. All graphed data show mean ±SD. ***:p<0.001 compared to untreated cells (CTR) in two-tailed t-test. Error bars indicate SD.



Fig. S3. Related to Fig. 4. ASM inhibition reverses the effects of PG/LBPA administration in NPC1-knockout cells.

A, ASM activity in parental and NPC1 KO HeLa cells treated with anionic phospholipids for 24 and 48h: PG (100μM), PC (100μM) and PC:LBPA (1:1 mol:mol) (100μM:100μM). ***p<0.001 vs NPC1 KO control. ##p<0.01 vs parental cell line (WT HeLa). B, efficient inhibition of ASM in AML treated NPC1 KO HeLa cells without and with 100µM PG treatment. AML concentration was 5µM. AML was added alone or together with PG for 48h. C, Representative epifluorescent images of filipin staining in NPC1 KO HeLa cells treated with DMSO (48h), AML alone (24 and 48h), PG alone (48h) or in combination of PG (48h) and AML (24 and 48h) and quantification. Filipin intensity was quantified in >50 cells in each group and data represented as % of DMSO control. Scale bar, 30µm. Statistically significant changes are indicated. ***p<0.001 vs DMSO control^{. ###}p<0.001 vs PG alone in two-tailed Student's test. AML concentration 5µM. D, representative confocal images and image quantification of LC3-II in NPC1 KO HeLa cells treated with DMSO, AML, PG, or PG+AML at indicated time points as in C. LC3-II-positive puncta were quantified by intracellular staining using a-LC3 antibody and represented as # per cell. Individual dots represent average # per cell/image. Average cell # in image is 8 cells. Scale bar is 20 µm. ***:p<0.001 compared to DMSO control, #:p<0.05, in two-tailed Student's test. Error bars indicate SD. AML concentration 5uM. PG 100uM. E, Western blot and quantification of LC3-II and p62 autophagy markers in NPC1-deficient fibroblasts treated with DMSO, AML, PG, PG+AML as in C and D. β-actin was used as loading control. AML 5µM, PG 100µM. F. Acid ceramidase activity in NPC1 deficient fibroblasts treated with 100µM PG and 500nM or 1µM of the acid ceramidase inhibitor Bodipy-Soclac. Inhibitor was added alone or together with PG for 48h. G. Filipin intensity quantification in NPC1-deficient fibroblasts treated with PG (100uM) and indicated concentrations of Bodipy-Soclac. Bodipy-Soclac was added alone or together with PG for 48h. H, Cell viability assay for 48h treatment of WT and NPC1 fibroblasts with Bodipy-Soclac in concentration range 100nM-5 μ M. All graphical data expressed as mean \pm SD.



Fig. S4. Related to Figure 6. Cell viability and filipin imaging following treatment with PC:PG. *A*, cell viability following treatment with PC:PG at 48h. Cell viability was determined using Cell Titer-Glo assay. The concentration of total lipid is presented: e.g. 200μ M PC:PG (1:1 mol:mol) (100μ M PC+ 100μ M PG). *B*, *r*epresentative images and filipin quantification in PC:PG treated NPC1 fibroblasts at 24 and 48h. Scale bar, 30μ m. Bar graph represents data from 2 independent experiments. ***:p<0.001 compared to untreated cells (CTR) in two-tailed t-test. 200μ M PC:PG (1:1 mol:mol) (100μ M PC+ 100μ M PG). All graphed data show mean ±SD.