Pancreatic Ppy-expressing γ-cells display mixed phenotypic traits and have adaptive plasticity to engage insulin production

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Supplementary Figure 1. Inducible Ppy-YFPi lineage-tracing. a, No YFP labelling is observed in untreated two-month-old and one-year-old mice. Immunofluorescence: Ppy, red; YFP, green. Scale bars: 20µm (10µm in insets). b, Percentage of Ppy-expressing cells labelled with YFP with or without DOX treatment. No DOX (2-month-old mice): $0,0 \pm 0,0$; n=4, 1458 Ppy⁺ cells scored. No DOX (1-year-old mice): $0,0 \pm 0,0$; n=4, 696 Ppy⁺ cells scored. With DOX treatment (2-month-old mice): $84,9 \pm 0,0$; n=8, 5411 Ppy⁺YFP⁺ out of the 6389 Ppy⁺ cells scored. Error bars are s.e.m. Region of the pancreas: Ventral. Source data are provided as Source Data file (table a, b).



Supplementary Figure 2. **Validation of the anti-Ppy antibodies.** Representative islet from a 2month-old DOX-treated Ppy-YFPi mouse bearing Ppy-rtTA transgene in heterozygous (HTZ) or homozygous (HMZ) state. Ppy staining (using both anti-Ppy antibodies, red) is lost in YFP (green)-labelled cells from Ppy-rtTA HMZ mice. Any of the YFP-labelled cell in DOX-treated Ppy-rtTA HMZ mice is stained with Ppy hormone. Scale bars: 20µm (10µm in insets). Region of the pancreas: Ventral.



b

Wild-type mouse



Ppy-YFPi mouse

С



Supplementary Figure 3. γ -cell distribution in the pancreas of wild-type and Ppy-YFPi mice. a, YFP-traced γ -cells in Ppy-rtTA heterozygous (HTZ) and homozygous (HMZ) mice follow the same distribution pattern than wild-type mice, being predominantly located in the ventral region of the pancreas. Comparative immunofluorescence of ventral and dorsal pancreatic sections: Ppy, red; YFP, green; Insulin, blue. Scale bar: 40µm (section) or 20µm (islet). b-c, Quantification of Ppy⁺ (b) or YFP⁺ (c) cells per islet section in ventral and dorsal sections of wild-type and Ppy-rtTA HTZ and HMZ, respectively. WT mice (n=5 mice, 5768 and 1288 Ppy+ cells scored in ventral and dorsal regions, respectively), Ppy-rtTA HTZ (n=6 mice, 3482 and 1017 YFP+ cells scored in ventral and dorsal regions, respectively) and Ppy-rtTA HMZ (n=6 mice, 3731 and 428 YFP+ cells scored in ventral and dorsal regions, respectively). Data is presented as mean values \pm s.e.m. Two-tailed Mann-Whitney test (ns p>0.05; * p≤0.05; ** p≤0.01; *** p≤0.001); wild-type ventral versus dorsal: *P* value = 0.0079, Ppy-YFPi (HTZ) ventral versus dorsal: *P* value = 0.0022. Source data are provided as Source Data file (table c).



С





Days after DOX

+2





Supplementary Figure 4. The Ppy-YFPi model is suitable for studying the adult Ppy-cell origin in embryonic stages. a, Experimental design. b, Representative pictures of No DOX (control) and DOX E7.5-E18.5. Immunofluorescence: Ppy in red and YFP in green. c, Proportion of Ppy⁺ cells YFP-labelled. YFP-traced γ -cells are only present upon DOX administration. No DOX (n=5 mice; 0 Ppy+YFP+ out of 2515 Ppy+ cells scored) and DOX E7.5 to E18.5 (n=3 mice, 378 Ppy+YFP+ out of 492 Ppy+ cells scored). Two-tailed Mann-Whitney test, No DOX vs DOX E7.5 to E18.5 P value = 0.0179. d, Experimental protocol. DOX administration from E7.5 to E19.5 to pregnant females (T0: under DOX exposure, +1: one day after DOX removal, +2: 2 days after DOX removal). e, Relative mRNA expression of Cre recombinase normalized on Ppy expression at different DOX administration timings (no DOX, n=3 mice; T0, n=5 mice; +1, n=5 mice; +2, n=5 mice). Data are shown as fold change of normalized ct values relative to No DOX (No DOX = 1). Two-tailed Mann-Whitney test (ns p>0.05; * p≤0.05; ** p≤0.01; *** p≤0.001), No DOX versus T0: P value = 0.0357, T0 versus +2: P value = 0.0159, No DOX versus +2: P value = 0.25. f. Bihormonal YFP-labelled Ppyexpressing cells were detected at E18.5 (Ppy, red; YFP, green and Gcg, cyan). Scale bars: $20\mu m$ (10 μm in insets). Data is presented as mean values \pm s.e.m. Region of the pancreas: whole pancreas. Source data are provided as Source Data file (table d).

Two-month-old wild-type mouse



b

DOX administration at one-year-old Ppy-YFPi



Supplementary Figure 5. γ -cell heterogeneity in wild-type mice persist throughout life. a, Bihormonal Ppy⁺Gcg⁺ (left; Ppy: green and Gcg: red), Ppy⁺Sst⁺ (middle; Ppy: green and Sst: red) and Ppy⁺Ins⁺ (right; Ppy: green and Ins: red) were detected in 2-month-old wild-type mice (n=5 mice). **b**, Heterogeneity in γ -cell population is maintained in one-year-old Ppy-YFPi mice (n=4 mice). Bihormonal Ppy⁺Gcg⁺ (left; Ppy: red and Gcg: cyan), Ppy⁺Sst⁺ (middle; Ppy: red and Sst: cyan) and Ppy⁺Ins⁺ (right; Ppy: red and Ins: cyan) YFP (green)-labelled could be identified. Scale bars: 20µm (10µm in insets). Region of the pancreas: Ventral. Source data are provided as Source Data file (table f).



Supplementary Figure 6. Gating strategy used for FACS sorting the YFP-labelled Ppy-expressing cells for RNA-seq analysis. Representative FACS plots for Fig. 4-5.



Supplementary Figure 7. Single-cell transcriptomics of Ppy-YFPi islets. a-d, Density plots of *Ppy* (a), *Gcg* (b), *Ins2* (c) and *Sst* (d) expression using normalized data from Ppy-YFPi islets. Red line indicates the threshold to define hormone+ cells. e, UMAP representation of all cells included in the Ppy-YFPi dataset (n=4261 cells). Red dots illustrate the endocrine population, grey dots represent all non-endocrine populations. f-j, UMAP representation of hormone positive cells: *Gcg*⁺ (orange, n=1367 cells, f), *Ins2*⁺ (green, n=548 cells, g), *Sst*⁺ (blue, n=624 cells, h) and *Ppy*⁺ (magenta, n=2626 cells, i) and *YFP*+ (fluor-green, n=1537 cells, j). Source data are provided as Source Data file (table g).





Supplementary Figure 8. Protein validation of the bihormonal Ppy-expressing cell identity. a-d, An enrichment in the production of Chga (b, d) and lapp (a, c) was observed and quantified in bihormonal Ppy+Ins+ (a, b) and Ppy+Gcg+ (c, d) in wild-type mice (n=2 mice). Scale bar: 20µm (islet) or 10µm (cell). Percentage of Chga+ (b, d; right pannel) and lapp+ (a, c; right pannel) cells in bihormonal Ppy+Ins+ and Ppy+Gcg+ compared to Ppy+Ins- and Ppy+Gcg- cells, respectively. 49 out of the 56 and 64 out of 68 Ppy+Ins+ cells expressed lapp and Chga, respectively; compared to the 50 out of 617 and 234 out of 839 Ppy+Ins- cells that express lapp and Chga, respectively. Similarly, 181 out of the 365 and 133 out of 198 Ppy+Gcg+ cells expressed lapp and Chga, respectively; compared to the 122 out of 671 and 87 out of 520 Ppy+Gcg- cells that express lapp and Chga, respectively. Ppy (a-d): red, Ins (a,b)/Gcg (c,d): grey and lapp (a,c)/ChgA (b,d): green. Region of the pancreas: Ventral. Data is presented as mean values \pm s.d. See Source data are provided as Source Data file (table m).



C PPY+ vs GCG+			
Modulated Pathway (3) / State			
Role of PKR in Interferon Induction and Antiviral Response	Activated		
Aryl Hydrocarbon Receptor Signaling	Activated		
PI3K/AKT Signaling	Inhibited		

d PPY+ vs INS+				
Modulated Pathway (15) / State				
Necroptosis Signaling Pathway	Inhibited			
RhoA Signaling	Inhibited			
Regulation of Actin-based Motility by Rho	Inhibited			
Remodeling of Epithelial Adherens Junctions	Inhibited			
ILK Signaling	Inhibited			
PI3K/AKT Signaling	Inhibited			
Aryl Hydrocarbon Receptor Signaling	Inhibited			
Protein Kinase A Signaling	Inhibited			
AMPK Signaling	Inhibited			
Myc Mediated Apoptosis Signaling	Inhibited			
p53 Signaling	Inhibited			
Cardiac β-adrenergic Signaling	Inhibited			
BAG2 Signaling Pathway Inhibited				
PPARα/RXRα Activation	Activated			
PPAR Signaling	Activated			

Supplementary Figure 9. **Common transcriptomic identities between mouse and human** γ -**cells. a,** Common DEGs between PPY+ cells and each of the other endocrine cells in mouse and human. **b,** Experimental workflow to define γ -ID pathways using the mouse and human RNA-seq datasets. **c-d,** 3 and 15 differentially modulated pathways in γ -cells were identified when compared to α -(**c**) and β -cells (**d**), respectively, in both the murine and the human RNA-seq datasets. No commonly modulated pathway were identified in the Ppy cell population compared to and δ -cells (**b**) between mice and humans. The top modulated pathways are ranked based on Z-score. Activated pathways (orange), inhibited pathways (blue). Source data are provided as Source Data file (table o, p).



Supplementary Figure 10. Transcriptomic profile of human bihormonal PPY-SST and PPY-INS cells. a, Expression plot of differentially expressed genes (DEGs) between bihormonal PPY-SST (top) and PPY-INS (bottom) compared to monohormonal PPY-expressing cells in human. Each dot represents one gene. Black dots represent the genes with a P value >0.05, red dots represent the genes with P value <0.05. Gene names label the top DEGs. Grey area indicates the upregulated genes in each bihormonal population. Differential expression was calculated using a negative binomial generalized linear model in a pairwise manner between populations, taking along the number of UMIs and the number of genes as variables to regress. P-values (in panel **b** and Source Data s) are Bonferroni corrected based on the total number of genes in each dataset. **b**, Intersection of the upregulated genes in *PPY*⁺SST⁺ (top) and *PPY⁺INS⁺* (bottom) from **a**, with human δ - and β -cell ID gene lists. All bihormonal *PPY*expressing cell types share important markers with adult δ -cells (9 out of 47) and β -cells (46 out of 690). Source data are provided as Source Data file (table s).

b



Supplementary Figure 11. Ppy-expressing γ -cell ablation has no impact on body weight and glycemia. a, CRISPR-Cas9 strategy to replace the Ppy coding sequence from chromosome 11 by the DTR (diphtheria toxin receptor) sequence. LHA, left homology arm; RHA, right homology arm. **b,** PCR products of wild-type (WT), Ppy-DTR heterozygous (HTZ) and Ppy-DTR homozygous (HMZ) mice (WT band: 389bp; KI band: 223bp). No DNA corresponds to the negative control. In all subsequent experiments, only Ppy-DTR heterozygous (HTZ) were used. c, Ppy-expressing cells were efficiently ablated 15 and 3 months post-DT injection (Ppy: green; Ins: red). Region of the pancreas: Ventral. Scale bar: 20µm. No DT (n=9 mice, 3656 Ppy+ cells scored), 15 days post-ablation (n=7 mice, 2 Ppy+ cells scored) and 3 months post-ablation (n=5 mice, 7 Ppy+ cells scored). d-e, Fasting body weight curves in non-ablated Ppy-DTR controls (black) and Ppycell ablated (red) males (d) and females (e) mice. Area under de curve (AUC) of each genotype is shown in the bottom right panel. f-g, Glycemia curves in non-ablated Ppy-DTR controls (black) and Ppy-cell ablated (red) males (f) and females (g) mice. Solid line represents random fed glycemia values, dashed line represents fasting glycemia values. Body weight and glycemia values were obtained from the same mice. Males: controls, n=7 mice; ablated, n=8 mice. Females: controls, n=9 mice; ablated, n=12 mice. h-i, Intraperitoneal glucose tolerance test to six months post-ablation Ppy-DTR mice (red) compared to non-injected controls (black) males (e) and females (f) mice. Males: controls, n=7 mice; ablated, n=7 mice. Females: controls, n=8 mice; ablated, n=12 mice. Data is presented as mean values \pm s.e.m. Two-tailed Mann-Whitney test (ns p>0.05; * p \leq 0.05; ** p \leq 0.01; *** p \leq 0.001). Source data are provided as Source Data file (table v).



γPdx1 OE + NO DT γPdx1 OE + DT YFP Ins Pdx1 YFP Pdx1 d 1 ** 2 ** ** ** ** ** 50



40·

30·

20-

10-

0

С







Supplementary Figure 12. Synergistic y-cell reprogramming by inducing Pdx1 expression and **ablating** β -cells. **a**, Transgenes required for γ -cell lineage tracing and ectopic Pdx1 expression combined with β -cell ablation. **b**, YFP-labelled γ -cells express Pdx1 in control and DT-treated mice. Approximately, half of the γ -cells expressing Pdx1 engage insulin production after β -cell ablation. The experiment was performed once with mice treated asynchronously depending on their availability. Scale bar: 20µm (islet) or 10µm (cell). Insulin: red; YFP: green; Pdx1: white. c, Percentage of YFP⁺ γ -cells expressing insulin in γ Pdx1 after β -cell loss. Two-tailed Mann-Whitney test (ns p>0.05; * p≤0.05; ** p≤0.01; *** p≤0.001), DT versus γPdx1OE: P value = 0.0016, γPdx1OE versus DT+γPdx1OE: P value = 0.0079, DT versus DT+γPdx1OE: P value = 0.0016. d, Percentage of Insulin+ cells labelled with YFP in γ Pdx1 after β -cell loss. Two-tailed Mann-Whitney test, DT versus yPdx1OE: P value = 0.0016, yPdx1OE versus DT+yPdx1OE: P value = 0.0079, DT versus DT+yPdx1OE: P value = 0.0016. DT, n=8 mice, 209 Ins+YFP+ out of 5477 YFP+ cells scored (this condition is taken for reference from Fig. 8e-f); γ Pdx1, n=5 mice, 233 Ins+YFP+ out of 2301 YFP+ cells scored; yPdx1+DT, n=5 mice, 652 Ins+YFP+ out of 1784 YFP+ cells scored. Region of the pancreas: Ventral. Data is presented as mean values \pm s.e.m. Source data are provided as Source Data file (table x).

Supplementary Table 1

List of primer sequences for qPCR

Gene	Primer sequence forward	Primer sequence reverse
B-actin	5' AAGGCCAACCGTGAAAAGAT 3'	5' GTGGTACGACCAGAGGGATAC 3'
Gapdh	5' TCCATGACAACTTTGGCATTG 3'	5' CAGTCTTCTGGGTGGCAGTGA 3'
Рру	5' GAGCTGAAGGCAGGGAGC 3'	5' GTTTGCAAGGGAGCAGGTTG 3'
Cre	5' GCCGCGCGAGATATGG 3'	5' GCCACCAGCTTGCATGATC 3'
GFP	5' CTACCCCGACCACATGAAGC 3'	5' GTAGTTGCCGTCGTCCTTGA 3'
Руу	5' CGCAGCTCTGTTCTCCAAC 3'	5' CAAACCTTCTGGCCGAGACC 3'

Supplementary Table 2

List of antibodies used for immunofluorescence

Primary antibodies	Dilution	Company
guinea pig anti-Pdx1	1/750	C.W. Wright
guinea pig anti-porcine insulin	1/400	DAKO (A0564)
rabbit anti-insulin	1/3000	Molecular Probes (701265)
mouse anti-glucagon	1/1000	Sigma (G2654)
rabbit anti-glucagon	1/200	DAKO (A0565)
mouse anti-somatostatin	1/200	BCBC Ab1985
rabbit anti-somatostatin	1/200	DAKO (A0566)
goat anti-somatostatin*	1/200	Santa Cruz Biotechnology (sc-55565)*
rabbit anti-GFP	1/400	Molecular Probes (A11122)
chicken anti-GFP	1/500	Abcam (ab13970)
mouse anti-Ppy	1/200	Y. Fujitani
mouse anti-Ppy	1/1000	R&D Biosystems (MAB62971)
mouse anti-Pyy	1/1000	Abcam (ab112474)
rabbit anti-Chga	1/200	Abcam (ab68271)
rabbit anti-lapp	1/500	Abcam (ab254259)

* Out of stock

Secondary antibodies	Dilution	Company
Alexa 568	1/500	Molecular Probes
Alexa 488	1/500	Molecular Probes
Alexa 647	1/500	Molecular Probes
Alexa 405	1/500	Molecular Probes
СуЗ	1/500	Southern Biotech
Cy5	1/500	Southern Biotech
FITC	1/500	Southern Biotech
TRITC	1/500	Southern Biotech