

SUPPORTING INFORMATION

Methylene Oxidation of Alkyl Sulfates by Cytochrome P450_{BM-3} and a Role for Conformational Selection in Substrate Recognition

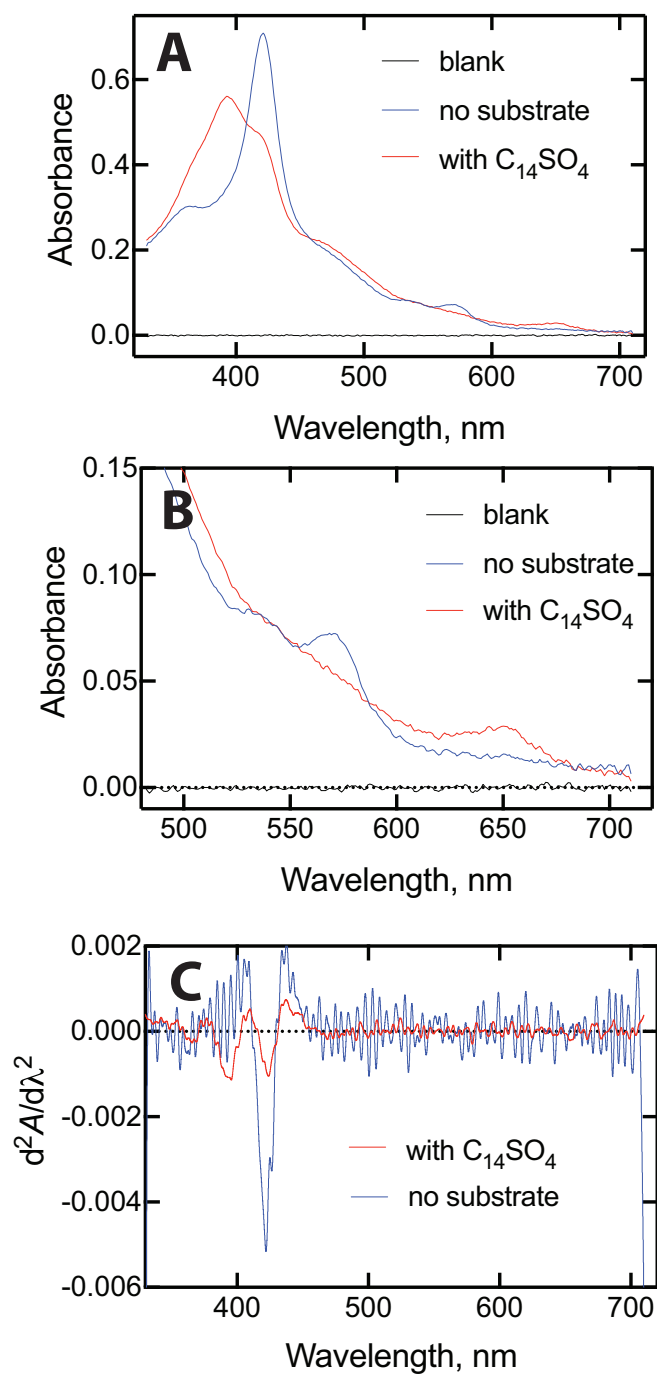
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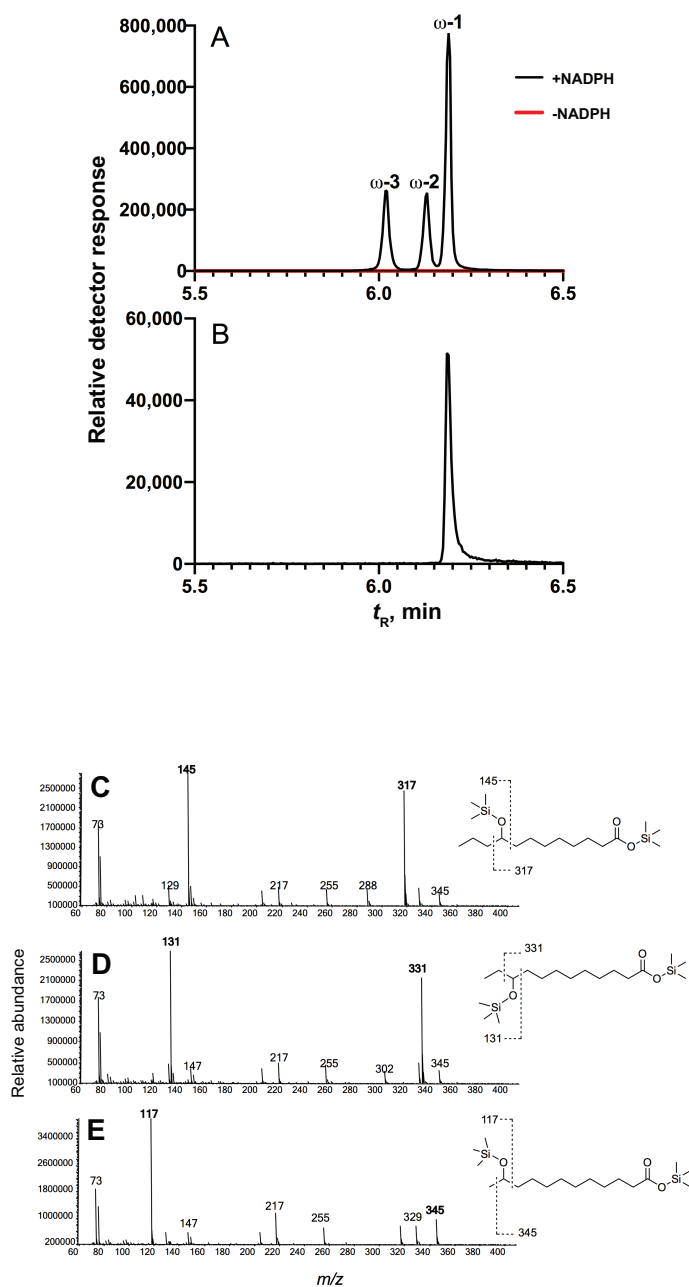
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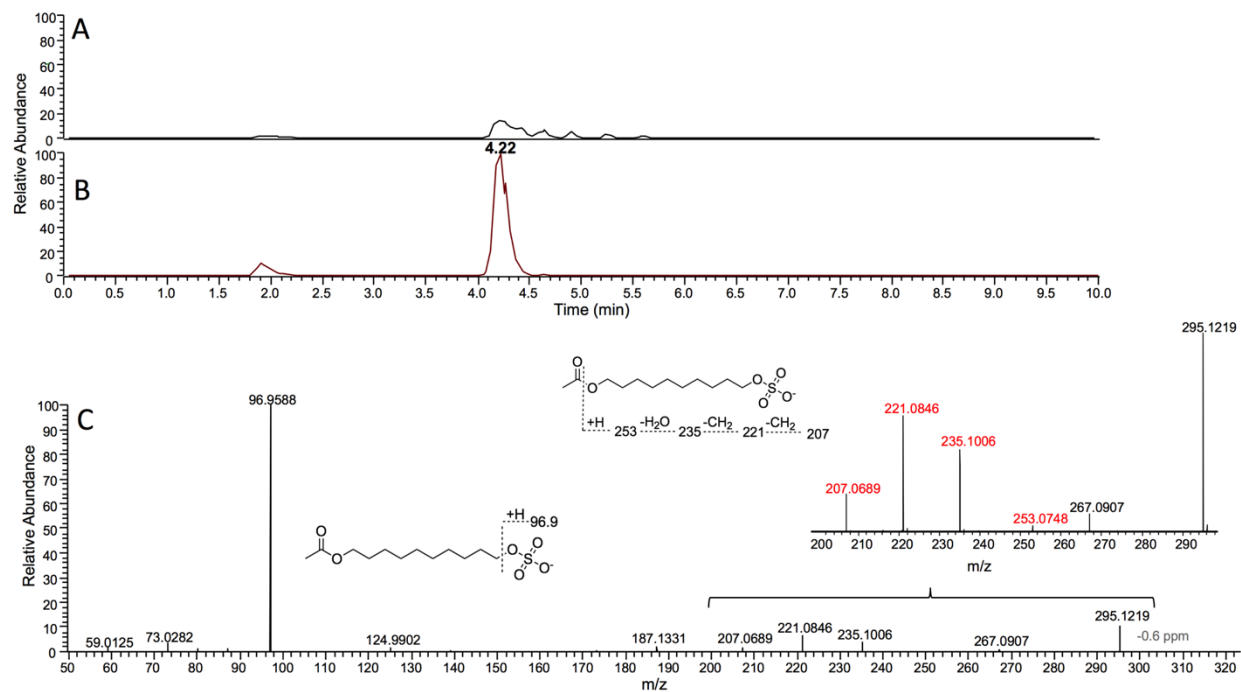
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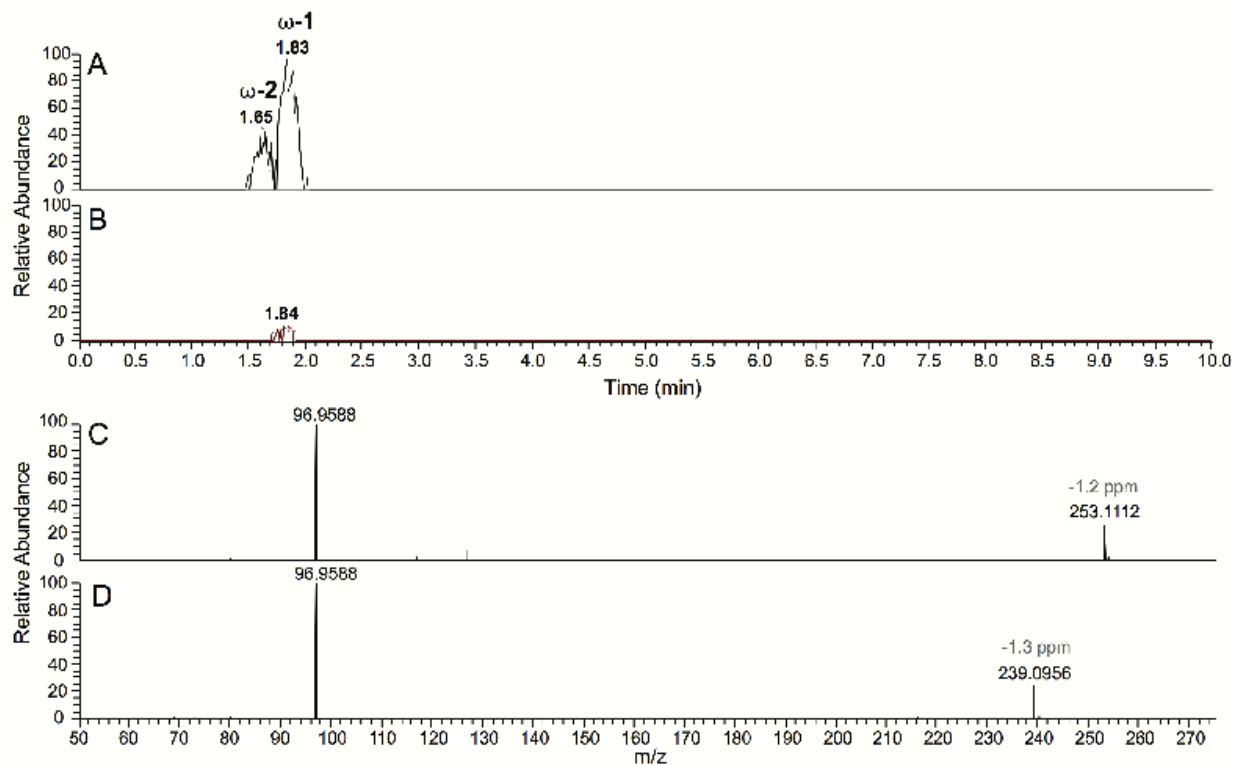
Supporting Figure S1. UV-visible spectra and second derivative spectra of P450_{BM-3} without and with tetradecyl sulfate (C₁₄SO₄). All spectra were recorded with 5 μM P450_{BM-3} in 50 mM potassium MOPS buffer (pH 7.4). **A.** Without (blue line) or with (red line) 50 μM tetradecyl sulfate (C₁₄SO₄). **B.** Part A expanded in the visible region. **C.** Second derivative spectra¹ of Part A (without substrate, blue line, filter 5 used for smoothing; with tetradecyl sulfate, red line, filter 19 used for smoothing).



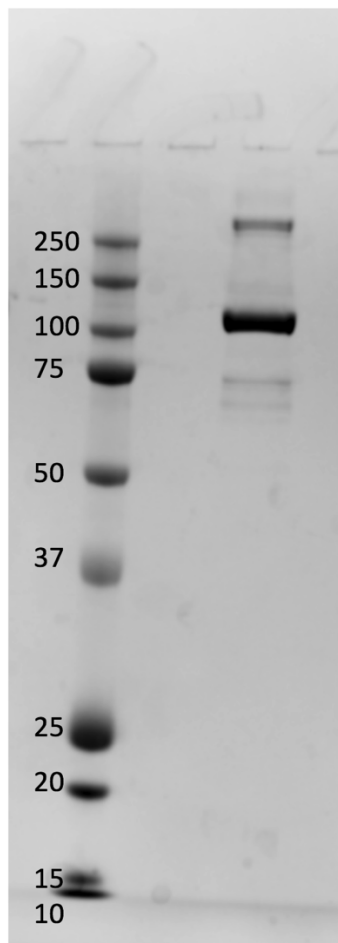
Supporting Figure S2. Identification of lauric acid oxidation products formed by P450_{BM-3} by GC-MS. **A.** Ion chromatogram resulting from incubation of lauric acid with P450_{BM-3} and NADPH, following derivatization to trimethylsilyl products. The GC-MS traces (*m/z* 345, *vide infra*) are shown for the reaction in the presence (black trace) and absence (red trace) of NADPH. **B.** Ion chromatogram for authentic standard 11-hydroxy lauric acid, following derivatization to trimethylsilyl derivative. **C-E.** mass fragmentation spectra of reaction products (from Part A). **C,** ω -3 product; **D,** ω -2 product; **E,** ω -1 product.



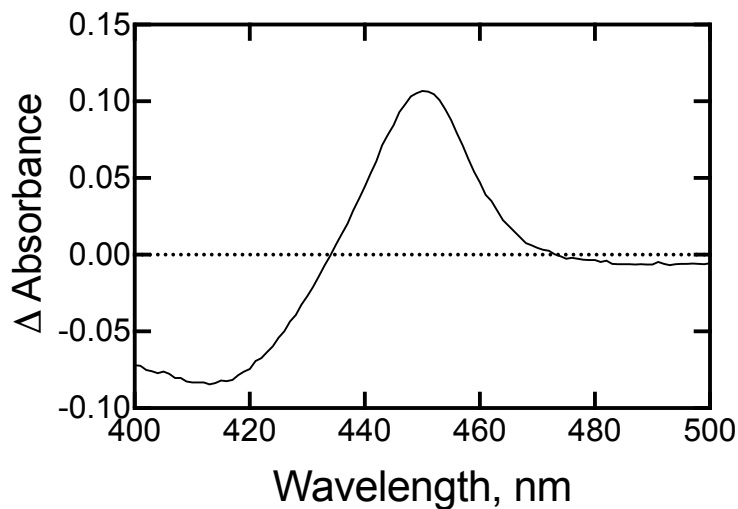
Supporting Figure S3. LC-HRMS of P450_{BM-3} reaction products of dodecyl sulfate following Baeyer-Villiger derivatization to esters. **A.** Dodecyl sulfate incubated with NADPH but without P450_{BM-3} for 2 minutes. **B.** Dodecyl sulfate incubated with NADPH and P450_{BM-3}. Both reactions were followed by oxidation of alcohol products using Jones reagent² and then Baeyer-Villiger ketone oxidation³ to form esters (Scheme 2). Extracted ion chromatograms for *m/z* 295.1221 are shown in Parts A and B, corresponding to product ester derivatives. **C.** HRMS (ESI) mass fragmentation spectrum of the ester formed from incubation of dodecyl sulfate with P450_{BM-3} and NADPH followed by the chemical oxidation steps.



Supporting Figure S4. LC-HRMS of P450_{BM-3} reaction products of dodecyl sulfate following Baeyer-Villiger derivatization to esters and base hydrolysis. **A.** Dodecyl sulfate incubated with NADPH and P450_{BM-3} for 2 minutes. **B.** Dodecyl sulfate incubated with P450_{BM-3} in the absence of NADPH. Both reactions were followed by oxidation of alcohol products using Jones reagent,² Baeyer-Villiger ketone oxidation³ to form esters, and then NaOH treatment (Scheme 2). Extracted ion chromatograms for *m/z* 253.1115 and 239.0959 (combined) are shown in both Parts A and B, corresponding to the alcohol products detected after base hydrolysis. **C, D.** HRMS (ESI) mass fragmentation spectra of the alcohols formed from incubation of tetradecyl sulfate with P450_{BM-3} and NADPH followed by the two chemical oxidation steps and base hydrolysis: **C,** fragmentation spectrum of the peak eluted at *t_R* 1.83 min peak; **D,** fragmentation spectrum of the peak eluted at *t_R* 1.65 min peak..



Supporting Figure S5. Purity of P450_{BM-3} preparation determined using SDS-polyacrylamide gel electrophoresis (7.5%, w/v). The molecular weight standard markers (labeled as kDa) are on the left and the P450_{BM-3} sample (10 pmol) is in the right lane. The higher band in the P450_{BM-3} sample (~ 260 kDa) is presumed to be a dimer of the enzyme.



Supporting Figure S6. Fe^{2+} -CO complex vs. Fe^{2+} difference spectrum of P450_{BM-3} preparation (1.25 μM) used for this study.

References

1. O'Haver, T. C.; Green, G. L., Numerical Error Analysis of Derivative Spectrometry for the Quantitative Analysis of Mixtures, *Anal. Chem.* **1976**, *48*, 312-318.
2. Bowden, K.; Heilbron, I. M.; Jones, E. R. H.; Weedon, B. C. L., 13. Researches on Acetylenic compounds. Part I. The Preparation of Acetylenic Ketones by Oxidation of Acetylenic Carbinols and Glycols, *J. Chem. Soc.* **1946**, 39-45.
3. Olah, G. A.; Wang, Q.; Trivedi, N. J.; Prakash, G. K. S., Baeyer-Villiger Oxidation of Ketones to Esters with Sodium Percarbonate/Trifluoroacetic Acid, *Synthesis-Stuttgart* **1991**, 739-740.