

Figure S1. Ski complex components are required for successful oogenesis, related to Figure 1. (A) Egg laying assay after germline RNAi knockdown of Ski complex components

indicating a loss of fertility compared to driver control (n=2-545 eqgs). ** = p < 0.01, Tukey's posthoc test after one-way ANOVA, p < 0.001. Error bars are standard deviation (SD). (B) Quantification of the area of the germarium and egg chambers #1- #5 comparing control and tst *RNAi*, showing that egg chambers do not grow upon loss of *tst* (n=35 ovarioles). *** = p < 0.001, Student's t-test. (C) tjGAL4 soma driver control and (D) tst soma RNAi ovarioles stained with Vasa (green) and 1B1 (magenta) do not exhibit a phenotype. (E) nosGAL4 driver control and (F) tst RNAi ovarioles stained with cleaved Caspase 3 (green) and 1B1 (magenta) indicating dying egg chamber (arrow). (G) Viability assay showing there is no significant difference in eggs hatching between control and UAS-Tst,nosGal4;tst/Df (ns, p>0.05, df = 1, n=233-237 eggs, Chi-squared test). (H) WT control and (I) ski3 mutant ovarioles stained with Vasa (green) and 1B1 (magenta) indicating egg chambers that do not grow in size (vellow dashed line) and dving egg chambers in ski3 mutant (arrow). (J) qRT-PCR assaying the RNA levels of tst, ski3 and ski8 in their respective mutant background or germline RNAi normalized to control and indicating successful knockdown (n=3). * = p < 0.05, ** = p < 0.01, *** = p < 0.001, Student's t-test. Scale bars are 10µm. (**K**) Table of exosome-associated genes, their subcellular localization, and molecular function. (L) nosGAL4 driver control, (M) tst RNAi, (N) rrp6 RNAi, (O) mtr4 RNAi, (P) dis3 RNAi and (Q) rrp40 RNAi germaria stained with Vasa (green) and 1B1 (magenta) showing germarium defects and loss of germline in knockdowns of exosome-associated factors but not tst RNAi. (R) Quantification of phenotypes observed upon germline RNAi of exosome-associated factors (n=50 ovarioles). *** = p <0.001, Chi-squared test with Bonferroni correction, df=3).



Figure S2. Twister post-transcriptionally regulates a cohort of RNAs, related to Figure 2. (A) Principal Component Analysis (PCA) comparing RNA-seq data of adult WT, *tst RNAi, tst* genomic mutant (*tst*), *half pint* (*hfp*⁹), *pelota* (*pelo*¹), young WT, germline stem cell enriched

(GSC), cystoblast enriched (CB), and differentiating cyst enriched (Cyst), indicating that the tst mutant and tst RNAi samples cluster closely with adult WT samples. (B) Young WT ovariole stained with Vasa (green) and 1B1 (magenta) indicating the presence of only 2-3 egg chambers. Scale bars are 10µm. (C) Quantification of the latest-stage egg chambers represented in young WT and tst RNAi (n=50 ovarioles). *** = p < 0.001, Wilcoxon rank sum test. (D) Early germ cell expressed genes identified by Jevitt et al. 2020 and their fold change in tst RNA Seg samples compared to WT, only rpS10a is upregulated >4 fold. (E) Genome browser tracks of maternal RNA genes pgc and gcl indicating similar expression in WT and tst RNAi. (F) gRT-PCR assaying the pre-mRNA levels of several Tst-regulated target genes, including blanks, act42A, act57B, act87E and the non-target act5C in control and germline tst RNAi normalized to control levels and indicating no change in transcription of these genes (n=3), ns = p > 0.05. Student's t-test. (G) qRT-PCR using random hexamer primed cDNA assaying for the levels of non-target act5C and Tst-regulated genes such as act42A, act57B, act87E and blanks in control and germline tst RNAi showing that increase in Tst-regulated RNA levels are not due to poly A selection (n=3). * = p < p0.05, Student's t-test. (H) Biplot of RNA-Seg data from adult WT and tst germ line RNAi knockdown ovaries in log₂ Transcripts Per Million (TPM) highlighting upregulated RNAs (magenta) and downregulated RNAs (black) that change concordantly in both tst RNAi and tst genomic mutant ovaries; selected non-target RNAs (see methods) are in blue. (I) Violin plot of non-target genes from RNA-seq data sets that include GSCs, CBs, cysts, young WT, adult WT, unfertilized eggs (Egg), tst genomic mutant (tst), and germline tst RNAi (tst RNAi) showing that non-targets do not substantially change during the course of development. (J) Quantification of rps10a RNA expression in GSCs, CBs, cysts and egg chambers of WT control (blue) and tst RNAi (magenta) in Arbitrary Units (A.U.) (n=15-54 per cell type). *** = p < 0.001, ** = p < 0.01, Tukey's post-hoc test after two-way ANOVA, p < 0.001. (K) Quantification of dany RNA expression in GSCs, CBs, cysts and egg chambers of WT control (blue) and tst RNAi (magenta) in Arbitrary Units (A.U.) (n=13-68 per cell type). *** = p < 0.001, ** = p < 0.01, Tukey's post-hoc test after twoway ANOVA, p < 0.001. Confocal images of (L-L') nosGAL4 and (M-M') tst RNAi germaria stained for 1B1 (magenta) and Blanks protein (green and grayscale) showing expanded Blanks expression in tst RNAi cysts (orange dashed lines) and sporadic expression in early egg chambers (arrow). (N) Quantification of Blanks protein expression in GSCs, CBs, cysts and eqg chambers of WT control (gray) and tst RNAi (magenta) (n=16-92 per cell type) expressed in A.U. *** = p < 0.001, Tukey's post-hoc test after two-way ANOVA, p < 0.001. Confocal images of (**O**-O') nosGAL4 and (P-P') tst RNAi germaria stained for Vasa (magenta) and nuclear Actin (C4) antibody (green and gravscale) showing expanded nuclear Actin expression in tst RNAi cysts (orange dashed line). Scale bars are 10um, (Q) Quantification of C4 levels in GSCs, CBs, cvsts and egg chambers of control (gray) and tst RNAi (magenta) expressed in A.U. (n=36-178 per cell type). *** = p < 0.001, ** = p < 0.01, Tukey's post-hoc test after two-way ANOVA, p < 0.001.



Figure S3. Twister-regulated RNAs exhibit hallmarks of NGD, but not NMD, related to Figure 3. (A) Pie charts showing that introns occur in the 3'UTRs of Tst-regulated RNAs (magenta) and non-targets (blue) at only low frequency. (B) Metaplots showing the proportion of RNA-Seg coverage mapping to exon-intron boundaries for both Tst-regulated targets (magenta) and non-targets (blue) indicating that Tst-regulated RNAs are spliced correctly. (C) Quantification of the area of the germarium and egg chambers #1- #5 comparing control and pelo¹, showing that egg chambers do not grow upon loss of *pelo* (n=30 ovarioles). *** = p < 0.001, * = p < 0.05, Student's t-test. (D-D') WT control and (E-E') pelo¹ ovarioles stained with cleaved Caspase 3 (green) and 1B1 (magenta) indicating dying egg chamber (arrow). (F) WT control and (G) pelo¹ mutant ovarioles stained with Vasa (green) and 1B1 (magenta) indicating loss of GSC in pelo¹ (arrow). Scale bars are 10µm. (H-H') Confocal images of in situ hybridizations against the Tstregulated RNA blanks (green, grayscale) and 1B1 (magenta) in WT control showing blanks RNA expression in the undifferentiated cells and decreasing in the cyst stages and (I-I') pelo¹ ovarioles showing *blanks* mRNA expression is expanded to egg chambers. (J) Quantification of normalized log₂ polysome/input mRNA of Tst-regulated RNAs (magenta), and non-targets (blue) in CB and replicate cyst, adult WT, tst RNAi and pelo¹ samples indicating that ribosome association of Tstregulated RNAs is dynamic during development relative to non-target RNAs (p < 2.2e-16, twoway repeated-measures ANOVA; *** = p < 0.001 by estimated means post hoc tests comparing tst target RNAs normalized by non-target levels per condition). (K) Control, (L) tst RNAi and (M) pelo¹ mutant ovarioles stained for 1B1 (magenta) and Blanks (green) showing restricted Blanks protein expression restricted to undifferentiated cells in control and expanded expression in tst *RNAi* and *pelo*¹ to the cyst stages but not the egg chambers. (**N**) Quantification of Blanks protein expression in GSCs, CBs, cysts and egg chambers of WT control (gray), *tst RNAi* (light magenta) and *pelo*¹ (dark magenta) in A.U. (n=16-92 per cell type). *** = p < 0.001, Tukey's post-hoc test after one-way ANOVA, p < 0.001.



Figure S4. Twister-regulated RNAs are bound by Hfp and do not exhibit suboptimal codon usage, related to Figure 4. (A) Codon Adaptation Index (CAI) comparison for Tst-regulated RNAs (magenta) versus non-targets (blue) indicating a higher CAI for Tst-regulated RNAs

(Wilcoxon rank sum test); and all genes (gray). (B) Table of the Actin paralog genes indicating that they have similar CAI values. (C) Schematic of the act5C::GFP, (C') act57B::GFP and (C'') act57B PT Mutant::GFP reporters that are under the control of a germline promoter (pgc) and a neutral 5'UTR (nos), and 3'UTR (K10). (D) Quantification of hfp⁹ oogenesis defect phenotypes compared to control (n=50 ovarioles, p<0.001, Chi-squared test with Bonferroni correction, df=3). (E) Quantification of the area of the germarium and egg chambers #1-#5 comparing control and hfp^9 , showing that eqg chambers do not grow upon loss of hfp (n=25 ovarioles). *** = p < 0.001, ** = p < 0.01, Student's t-test. (**F-F**') WT control and (**G-G**') hfp^9 ovarioles stained with cleaved Caspase 3 (green) and 1B1 (magenta) indicating dying egg chamber (arrow). (H-H') Germaria stained for Hfp (green and grayscale) and DAPI (magenta) indicating both cytoplasmic and nuclear Hfp expression during early oogenesis. Scale bar is 10um. (I) Western blot analysis of a subcellular fractionation probing input, cytoplasm and nucleus fractions for the cytoplasmic marker Orb, the nuclear marker Fibrillarin, and Hfp. Hfp is present in both the nucleus and cytoplasmic fractions. (J) Hfp-HA RIP and gRT-PCR analyses indicating an enrichment of the Tstregulated targets act57B, rps10a, and abd-a levels compared to non-targets such as pgc and act5C levels in Hfp RIP samples compared to input (n=3). *** = p < 0.001, ** = p < 0.01, * = p < 0.05, Student's t-test. (K) 3-Way Venn diagram of Tst-regulated RNAs, Pelo-regulated RNAs and Hfp-regulated RNAs indicating that Tst, Pelo and Hfp co-regulate 59% (122/207) of their target RNAs (p<9.84x10E-169, Hypergeometric test). (L) Metaplot of the proportion of RNA-seq coverage mapping to exon-intron boundaries in *hfp*⁹ mutant and control (heterozygous) RNA-seq data sets for Tst-regulated RNAs (magenta) indicating correct splicing. (M) Co-Immunoprecipitation and Western blot of Pelo-HA probed for Hfp and Tst-GFP, indicating that these 3 proteins interact in vivo. IgG lane shows Mouse IgG heavy chain signal indicated by (*).



Figure S5. Some Twister-regulated RNAs are required in the undifferentiated stages for germline maintenance and are detrimental when expressed after differentiation, related to Figure 5. (A) Schematic of a screen to test the requirement of Tst-regulated genes in the germline. Using UAS-Dcr2;nosGAL4 that also carries a bamGFP (differentiaton marker) reporter select RNAs were depleted in the germline. F1 ovaries were dissected and phenotypes were assessed by 1B1, Vasa and GFP staining. (B) nosGAL4 driver control, (C) tsf1 RNAi, (D) lsp1y RNAi, and (E) CG15067 RNAi stained with Vasa (blue), 1B1 (magenta), and BamGFP (green) each exhibiting either a differentiation defect or complete loss of germ line. (F) nosGAL4 driver control ovariole stained with Vasa (green) and 1B1 (magenta) and (G) germline UAS-Act57B overexpression showing egg chamber defects (arrow). (H) Quantification of oogenesis defect phenotypes observed upon germline UAS-Act57B overexpression (n=50 ovarioles, p < 0.001. Chi-squared test with Bonferroni correction, df=1). (I) nosGAL4:bamGFP driver control ovariole, (J) >tst RNAi;bamGFP, (K) >ski3 RNAi;bamGFP, and (L) >ski8 RNAi;bamGFP ovarioles stained with 1B1 (magenta) and GFP (green) show correct BamGFP expression. (M) nosGAL4 driver control and (N) >tst RNAi ovarioles stained for EdU (green) and 1B1 (magenta) showing proper endocycling. Scale bars are 10µm.