Supplementary Information

SLITRK5 is a negative regulator of hedgehog signaling in osteoblasts

Jun Sun^{1†}, Dong Yeon Shin^{1, 2†}, Mark Eiseman¹, Alisha R Yallowitz¹, Na Li³, Sarfaraz Lalani¹, Zan Li¹, Michelle Cung¹, Seoyeon Bok¹, Shawon Debnath¹, Sofia Jenia Marquez¹, Tommy E. White⁴, Abdul G Khan⁴, Ivo C Lorenz⁴, Jae-Hyuck Shim⁵, Francis S. Lee⁶, Ren Xu^{3*}, Matthew B. Greenblatt^{1, 7*}

¹Department of Pathology and Laboratory Medicine Weill Cornell Medicine, New York, NY, USA

²Current Address: Research Center, LegoChem BioSciences, INC., Daejeon 34302, South Korea

³State Key Laboratory of Cellular Stress Biology, School of Medicine, Xiamen University,

Xiamen, Fujian361102, China

⁴Tri-Institutional Therapeutics Discovery Institute, New York 10021, United States

⁵Division of Rheumatology, Department of Medicine, University of Massachusetts Medical

School, 364 Plantation Street, Worcester, MA01605, USA

⁶Department of Psychiatry, Weill Cornell Medical College, New York, NY 10065, USA

⁷ Research Division, Hospital for Special Surgery, New York, NY 10021, USA

† These authors contributed equally to this work.

*Correspondence: Ren Xu 86-592- 2880577 xuren526@xmu.edu.cn

Matthew B. Greenblatt 1-212-746-1602 mag3003@med.cornell.edu



Supplementary Figure 1. a Slitrk5 mRNA level in different cells and tissues. The raw expression data were obtained from BioGPS (http://biogps.org/). Data are presented as mean±s.d., n=2 biologically independent samples. **b** Immunostaining of mouse femur sections with beta-galactosidase antibody, indicating the expression of Slitrk5 in osteoblasts.Data are representative of two independent experiments, scale bar=100 um.



Supplementary Figure 2. a Primary osteoblasts from WT and Slitrk5-/- mice were treated with the indicated doses of K252a and cultured in osteoblast differentiation medium. ALP activity was measured at day 8 of osteoblast differentiation. Data are presented as mean±s.d. n=10 biologically independent samples. **b** Slitrk5 mRNA level in SmoM2 induced medulloblastoma and adjacent tissue (GEO: GSE9299). Data are presented as mean±s.d., n=3 biologically independent samples. **c** Human osteoblast-like cells (Saos-2) infected with Slitrk5 or GFP shRNA and cultured in osteoblast differentiation medium. mRNA level of Slitrk5, Gli1, Ptch1 and Gli3 at 0, 5 and 10 days of differentiation was assessed by RT-PCR. Data are presented as mean±s.d., n=4 biologically independent samples, 2-tailed unpaired t-test.



Supplementary Figure 3. a Primary osteoblasts from WT and Slitrk5-/- mice were treated with the indicated doses of Purmorphamine and cultured in osteoblast differentiation medium. ALP activity was measured at day 8 of differentiation. Data are presented as mean±s.d. n=10 biologically independent samples. b Co-immunoprecipitation of Flag-SLITRK1 and SHH in HEK293T cells. Data are representative of two independent experiments. c Co-immunoprecipitation of Flag-SLITRK6 and SHH in HEK293T cells. Data are representative of two independent experiments. d C3H10T1/2 cells transfected with either control, Slitrk1 or Slitrk6 overexpression vectors were treated with BSA or 100ng/ml SHH in serum free medium for 48h. Gli1, Ptch1, Hhip, Slitrk1 and Slitrk6 mRNA levels were measured by RT-PCR. n=4 biologically independent samples, 2-tailed unpaired t-test. e Co-immunoprecipitation of Flag-SLITRK5 and PTCH1-YFP in C3H10T1/2 cells. Data are representative of two independent samples.



Supplementary Figure 4. a&b Representative images of TRAP staining (a) and quantification of No.Oc/B.Pm (b) of the L3 vertebrae in 7-week-old WT and Slitrk5-/- female mice. osteoclast number/bone perimeter (No.Oc./B.Pm). N=5 or 6 per group. Data are presented as mean±s.d, 2-tailed unpaired t-test, scale bar=100um. c&d Representative images of uCT (c) and quantification of bone volume/total volume (BV/TV), trabecular number per cubic millimeter (Tb.N), trabecular thickness (Tb.Th) , trabecular separation (Tb.Sp) and cortical bone thickness (Cor.Th)(d) of femurs in WT, Slitrk5+/- and Slitrk5-/- female mice at 7-week-old. N=4-7 per group. Data are presented as mean±s.d, 2-tailed unpaired t-test, scale bar=1mm.



Supplementary Table

Name	Species	shRNA primers
shSlitrk5-199	Mouse	CCGGCCTCCTGCACCTGGGTAATAACTCGAGTTATTACCCAGGTGCAGGAGGTTTTTG
shSlitrk5-200	Mouse	CCGGTGCAGAAACCATCGATTATTACTCGAGTAATAATCGATGGTTTCTGCATTTTTG
sh-control	Mouse	CCGGGCGCGATAGCGCTAATAATTTCTCGAGAAATTATTAGCGCTATCGCGCTTTTT
		qPCR primers
Slitrk5-forward	Mouse	CGCAGAACAGACTTCCTGGAAG
Slitrk5-reverse	Mouse	TCAGCCTCTCAATCCTGTTGCC
Slitrk6-forward	Mouse	AACAACCCACCATACAACGGA
Slitrk6-reverse	Mouse	TTCCTCAACTTCTTCCAAATGCT
Bsg-forward	Mouse	AGAGGACACAGGCACTTACGAG
Bsg-reverse	Mouse	GACTTCCTGGACAGAGGTTTGG
Hhip-forward	Mouse	GAGAGATCCTGTGTGGGGGCTT
Hhip-reverse	Mouse	GCTGCATTCTGAGTTGTTGGTG
Hprt-forward	Mouse	TCAGTCAACGGGGGACATAAA
Hprt-reverse	Mouse	GGGGCTGTACTGCTTAACCAG
Gli1-forward	Mouse	CCAAGCCAACTTTATGTCAGGG
Gli1-reverse	Mouse	AGCCCGCTTCTTTGTTAATTTGA
Ptch1-forward	Mouse	AAAGAACTGCGGCAAGTTTTTG
Ptch1-reverse	Mouse	CTTCTCCTATCTTCTGACGGGT
Ptch2-forward	Mouse	GGCACTCACATCCGTCAACAAC
Ptch2-reverse	Mouse	GAAGACGAGCATTACCGCTGCA
Ocn-forward	Mouse	TGCTTGTGACGAGGTATCAG
Ocn-reverse	Mouse	GTGACATCCATACTTGCAGG
Alp-forward	Mouse	CGGGACTGGTACTCGGATAA
Alp-reverse	Mouse	ATTCCACGTCGGTTCTGTTC
Sp7-forward	Mouse	TCACTTGCCTGCTCTGTTCC
Sp7-reverse	Mouse	GCGGCTGATTGGCTTCTTCT
Runx2-forward	Mouse	CCAACCGAGTCATTTAAGGCT
Runx2-reverse	Mouse	GCTCACGTCGCTCATCTTG
Gli2-forward	Mouse	ACACTGTGGAGGACTGCCTACA
Gli2-reverse	Mouse	GGCATCTCCATGCCACTGTCAT
Col1a1-forward	Mouse	CCTCAGGGTATTGCTGGACAAC
Col1a1-reverse	Mouse	CAGAAGGACCTTGTTTGCCAGG
Gli3-forward	Human	TCAGCAAGTGGCTCCTATGGTC
Gli3-reverse	Human	GCTCTGTTGTCGGCTTAGGATC
Gli1-forward	Human	AGCCTTCAGCAATGCCAGTGAC
Gli1-reverse	Human	GTCAGGACCATGCACTGTCTTG
Ptch1-forward	Human	GCTGCACTACTTCAGAGACTGG
Ptch1-reverse	Human	CACCAGGAGTTTGTAGGCAAGG
Slitrk5-forward	Human	CCTCCAGTACAATCTCATCCGC
Slitrk5-reverse	Human	TCAAGCCAGAGAAGACGCCTGA