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Supplemental information

**A GPC2 antibody-drug conjugate is
efficacious against neuroblastoma and small-cell lung
cancer via binding a conformational epitope**

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Figure S1

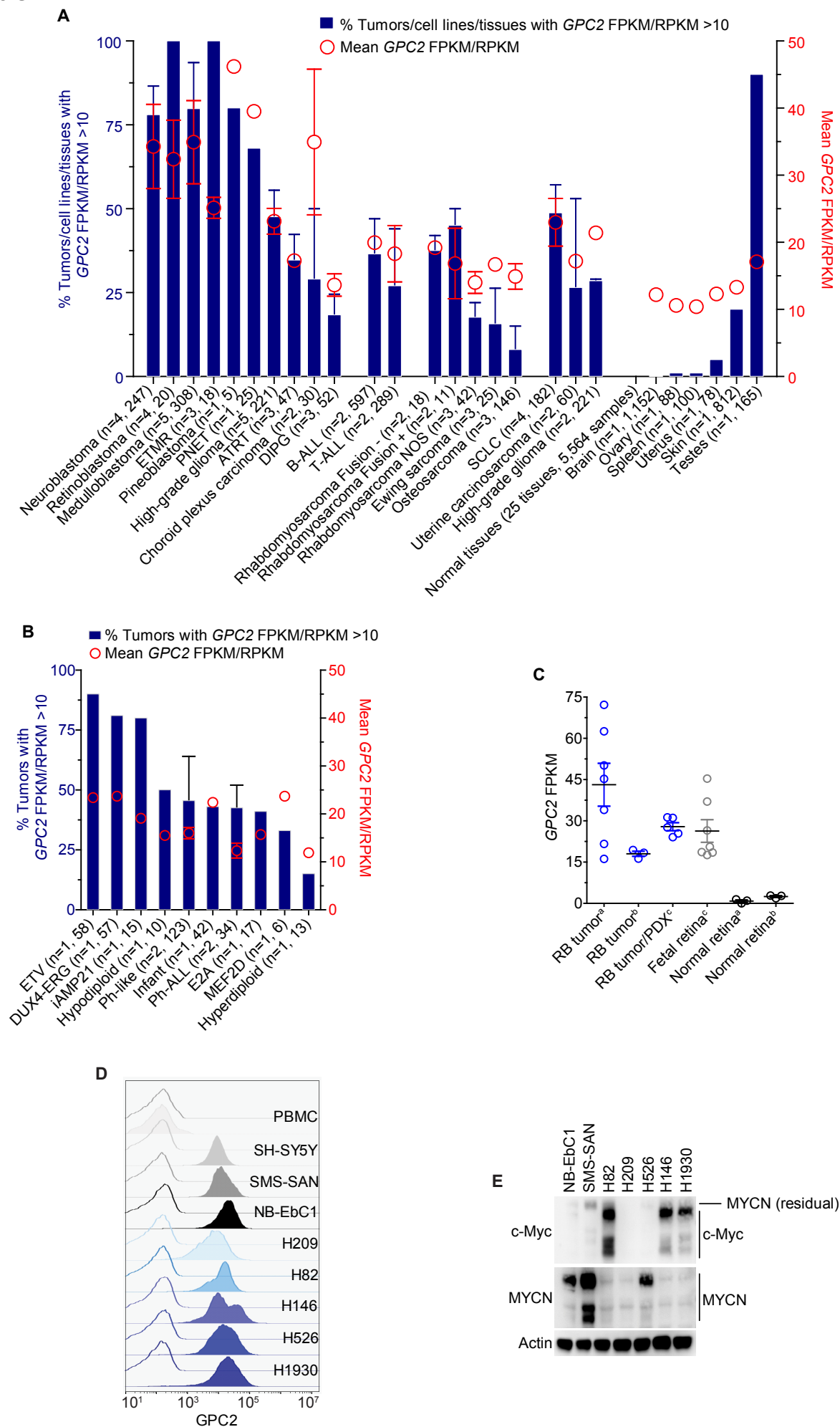


Figure S1. *GPC2* is developmentally regulated and highly expressed in other pediatric and adult cancers.

Related to Figure 1.

(A) Summary plot of *GPC2* RNA sequencing data across pediatric and adult tumor datasets. The blue bars graphed on the left y-axis represent the % of tumors/cell lines/normal tissues with *GPC2* FPKM/RPKM >10 represented as mean \pm SEM of multiple data sets per tumor histotype or normal tissue as indicated (n=1-5 datasets per tumor/cell line/tissue). The red circles graphed on the right y-axis represent the mean *GPC2* FPKM/RPKM \pm SEM of the >10 FPKM/RPKM cohort of tumors/cell lines/normal tissues in each data set. The number of data sets and total samples for each tumor histotype/normal tissue is indicated on the x-axis (n=total data sets, total tumors/cell lines/tissues).

(B) Summary plot of *GPC2* RNA sequencing data across pediatric ALL subtypes. The blue bars graphed on the left y-axis represent the % of ALLs with *GPC2* FPKM/RPKM >10 represented as mean \pm SEM of multiple data sets per ALL subtype as indicated. The red circles graphed on the right y-axis represent the mean *GPC2* FPKM/RPKM \pm SEM of the >10 FPKM/RPKM cohort of ALLs in each data set. The number of data sets and total samples for each ALL subtype is indicated on the x-axis (n=total data sets, total tumors).

(C) *GPC2* expression in retinoblastomas, fetal and normal retinas represented as mean \pm SEM with each circle representing an individual sample. ^aNormal retina and retinoblastoma data from GSE125903¹; ^bNormal retina and retinoblastoma data from GSE111168²; ^cNormal fetal retina and retinoblastoma tumor and PDX data from GSE87042.³

(D) Representative *GPC2* flow cytometry histograms of neuroblastoma and SCLC cell lines. SH-SY5Y, SMS-SAN and NB-EbC1 are representative neuroblastoma cell lines.

(E) c-Myc and MYCN Western blot of SCLC and neuroblastoma cell lines. SMS-SAN and NB-EbC1 are representative neuroblastoma cell lines.

ETMR, embryonal tumor with multilayered rosettes; PNET, primitive neuroectodermal tumor; ATRT, atypical teratoid rhabdoid tumor; DIPG, diffuse intrinsic pontine glioma; NOS, not-otherwise specified; SCLC, small cell lung cancer; RB, retinoblastoma; PBMC, peripheral blood mononuclear cell.

Figure S2

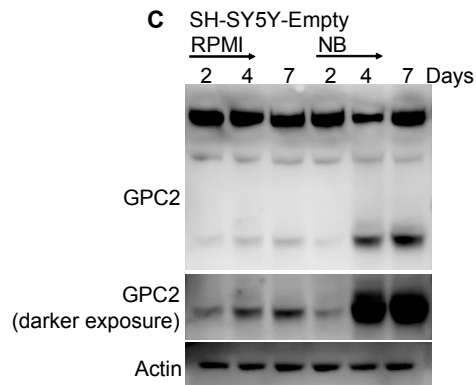
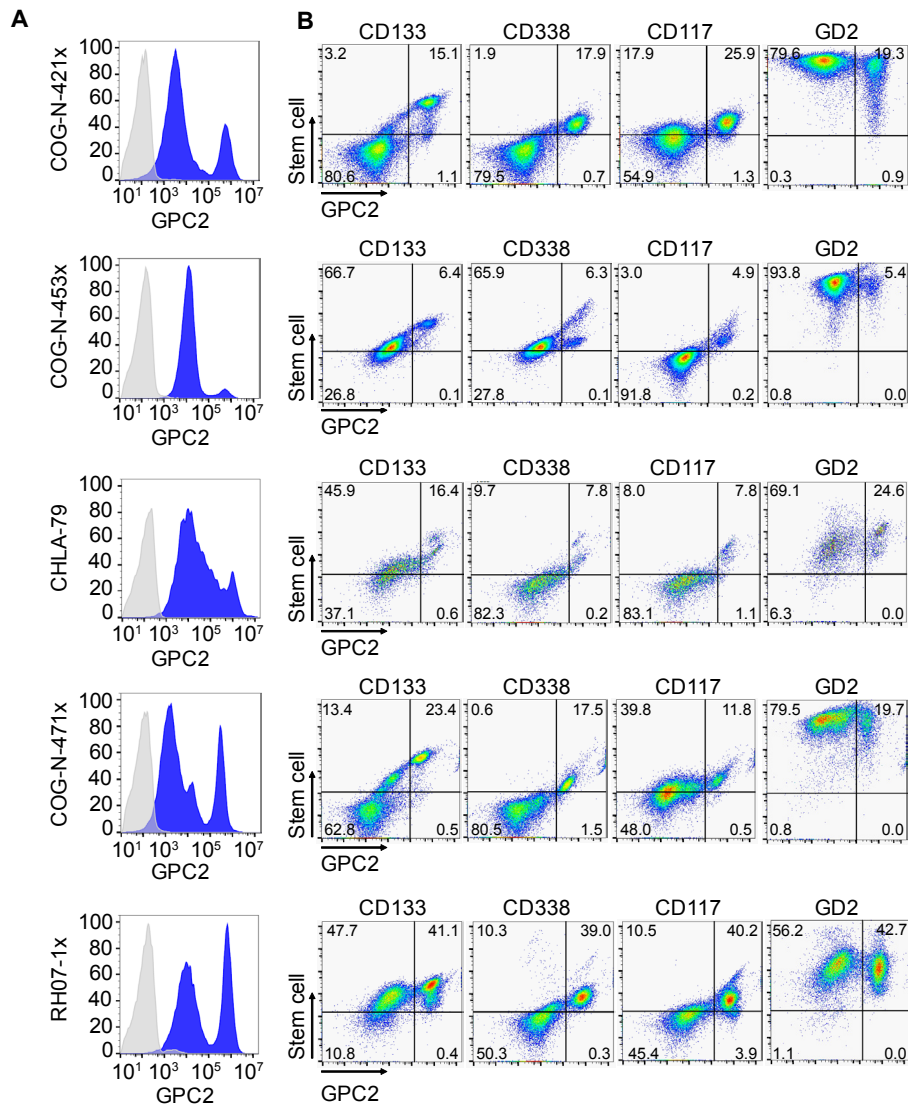


Figure S2. GPC2 is a putative neuroblastoma stem cell marker. Related to Figure 2.

(A, B) GPC2 flow cytometry histograms (A) and GPC2 vs. CD133, CD338, CD117 and GD2 plots (B) of neuroblastoma PDXs (COG-N-421x, COG-N-453x, COG-N-471x and RH07-1x) and cell line xenograft (CHLA-79).

(C) GPC2 Western blot of SH-SY5Y-Empty^{RPMI} vs. SH-SY5Y-Empty^{NB} cells.

Figure S3

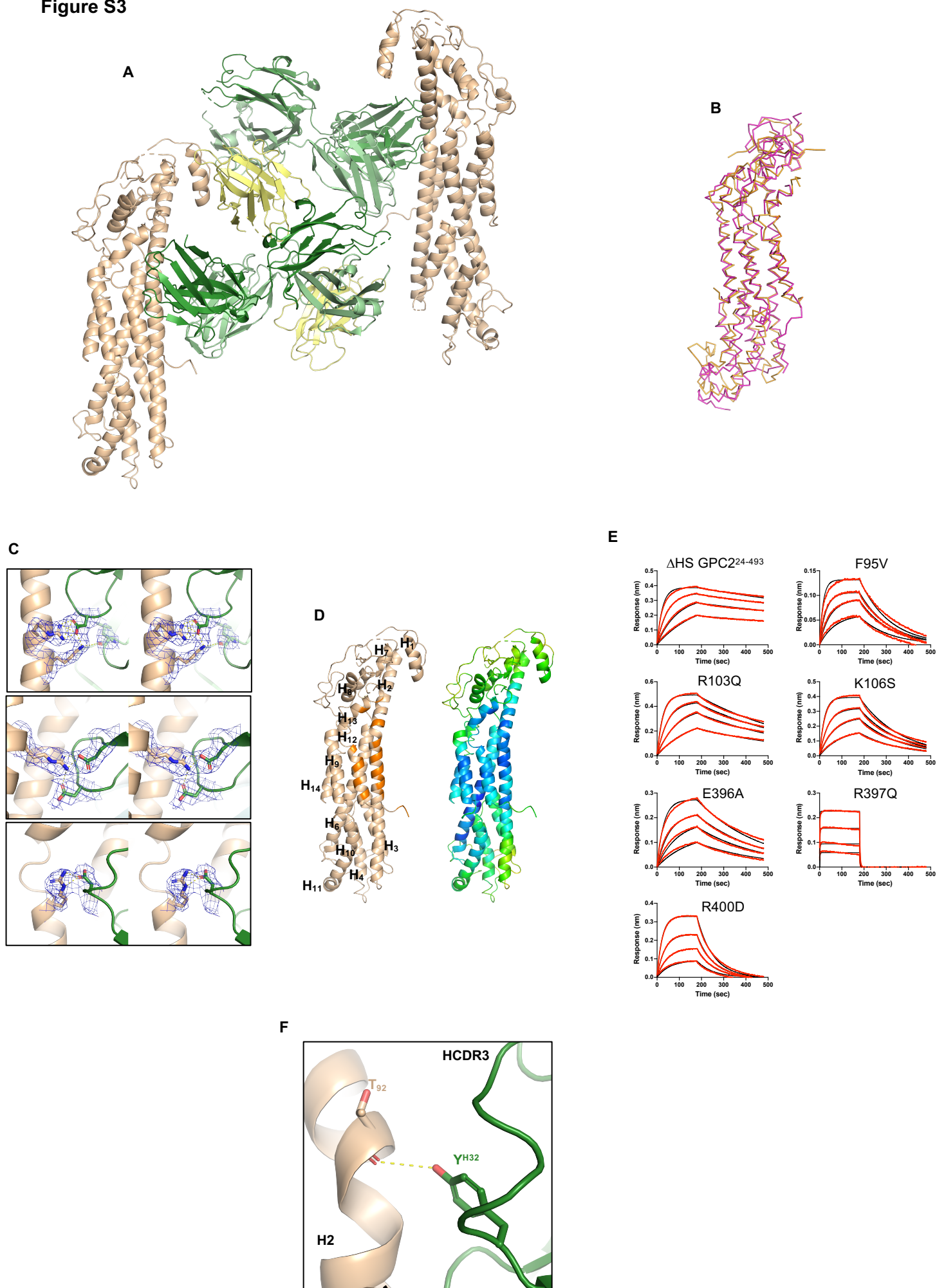


Figure S3. The D3-GPC2-Fab binds a conformational GPC2 protein epitope. Related to Figure 3.

(A) Overview of the co-crystal structure that shows two NCS-related copies of the D3-GPC2-Fab/ Δ HHS GPC2²⁴⁻⁴⁹³ complex in the asymmetric unit. GPC2 is shown in wheat, the D3-GPC2-Fab in green and the V_{HH} domain utilized to aid in crystallization in yellow.

(B) Superposition of the GPC1 crystal structure (PDB ID: 4ACR, pink) on the GPC2 structure (orange), shown as C α trace.

(C) Stereo images of D3-GPC2-Fab residues (green) interacting with GPC2 residues (wheat). The 2F_o-F_c map (blue mesh) is rendered at a 1.0 σ contour level. Interactions: GPC2-R103 and -K106 with Fab_{HC}-D99 and Fab_{LC}-L91 (top); GPC2-R397 with Fab_{HC}-E95 and -D99 (middle); GPC2-R400 with Fab_{HC}-D31 (bottom).

(D) GPC2 with helices numbered and D3-GPC2-Fab epitope traced in orange (left) and GPC2 colored based on B-factors (right). The color spectrum ranges from B-factors of 30 Å^2 (blue) to 150 Å^2 (orange), with colder colors indicative of regions of overall lower flexibility, such as the D3-GPC2-Fab epitope.

(E) Representative kinetics plots showing the binding of D3-GPC2-Fab to the Δ HHS GPC2²⁴⁻⁴⁹³ protein and its respective point mutants. The data (red) is fitted with a 1:1 model (black).

(F) The Ser/Thr92 interaction (wheat) with heavy chain residues of D3-GPC2-Fab (green).

Figure S4

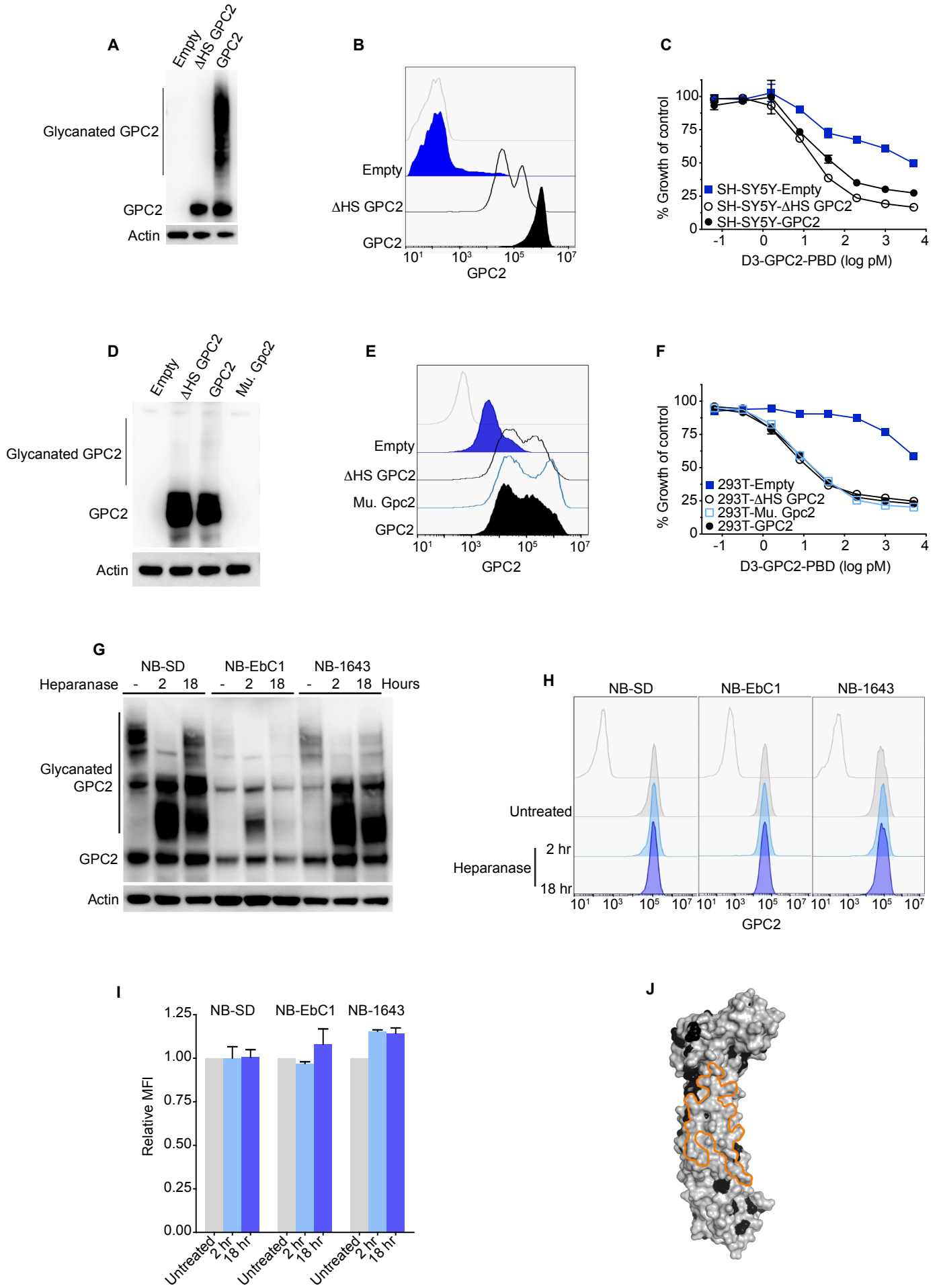


Figure S4. GPC2-associated heparan sulfate chains do not significantly alter D3-GPC2 antibody binding.

Related to Figure 3.

(A-C) GPC2 Western blot (A), D3-GPC2-IgG1 flow cytometry histograms (B) and relative growth plots 4 days after treatment with D3-GPC2-PBD (C) of SH-SY5Y-Empty, -GPC2 and - Δ HS GPC2 isogenic cells.

(D-F) GPC2 Western blot (D), D3-GPC2-IgG1 flow cytometry histograms (E) and relative growth plots 4 days after treatment with D3-GPC2-PBD (F) of HEK293T-Empty, -murine Gpc2, -human GPC2 and - Δ HS GPC2 transfected cells. GPC2 antibody used in Western blot in D does not recognize murine Gpc2.

(G) GPC2 Western blot of neuroblastoma cells after treatment with recombinant heparanase.

(H) Representative D3-GPC2-IgG1 flow cytometry histograms of heparanase treated neuroblastoma cells.

(I) Summary of relative GPC2 MFIs of heparanase treated neuroblastoma cells from H.

(J) Human and mouse GPC2 sequence conservation mapped on human GPC2 crystal structure, with variable amino acid residues in black and the D3-GPC2-Fab epitope residues outlined in orange.

Data in I is represented as mean \pm SEM of two independent replicates. Data in C and F is represented as mean \pm SEM from a representative experiment done in technical triplicate with each experiment being done two independent times.

Figure S5

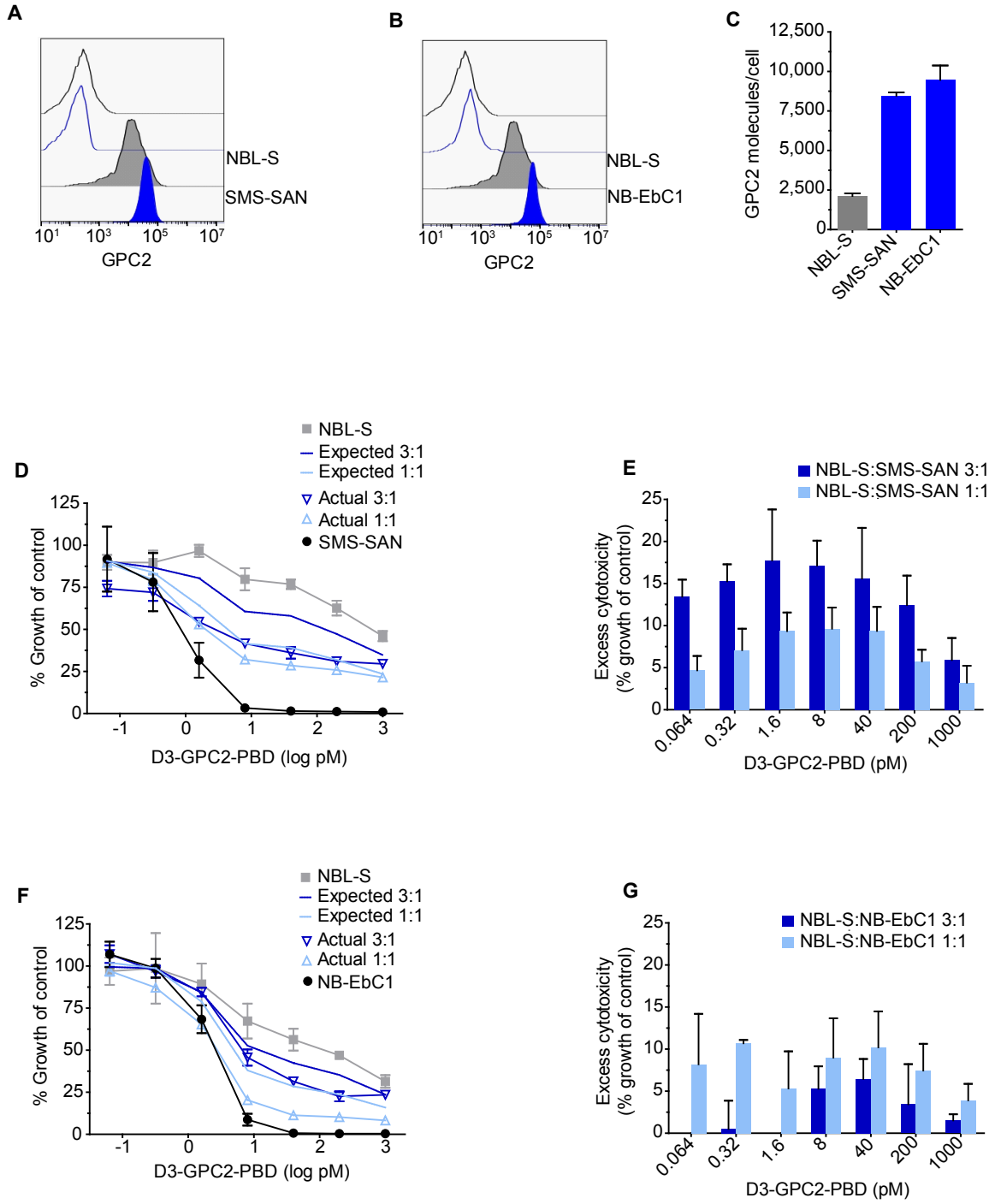


Figure S5. The D3-GPC2-PBD ADC induces bystander cell killing in paired high/low-GPC2 expressing neuroblastoma cell lines. Related to Figure 4.

(A, B) Representative GPC2 flow cytometry histograms of NBL-S and SMS-SAN (A) and NBL-S and NB-EbC1 (B) neuroblastoma cells.

(C) GPC2 molecules/cell for NBL-S, NB-EbC1 and SMS-SAN neuroblastoma cells.

(D) Representative actual and expected relative growth plots of indicated ratios of NBL-S/SMS-SAN co-incubated cells 6 days after treatment with the D3-GPC2-PBD ADC.

(E) Summary plot of excess cytotoxicity observed in co-incubated NBL-S/SMS-SAN cells.

(F) Representative actual and expected relative growth plots of indicated ratios of NBL-S/NB-EbC1 co-incubated cells 6 days after treatment with the D3-GPC2-PBD ADC.

(G) Summary plot of excess cytotoxicity observed in co-incubated NBL-S/NB-EbC1 cells.

Data in C, E and G is represented as mean \pm SEM of 2-3 independent replicates. Data in D and F is represented as mean \pm SEM from a representative experiment done in technical duplicate with each experiment being done at least two independent times.

Figure S6

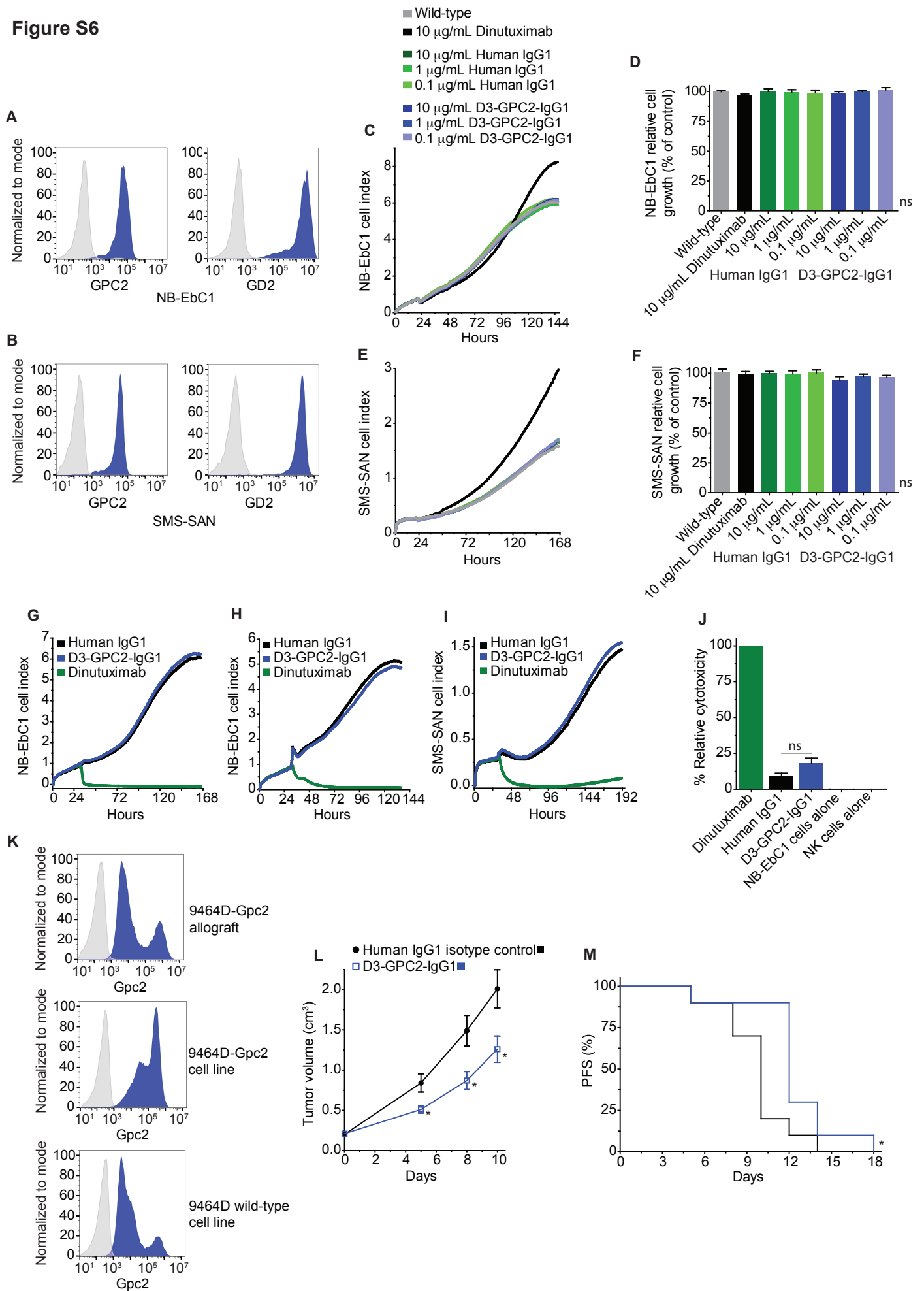


Figure S6. The D3-GPC2-IgG1 antibody alone does not significantly inhibit neuroblastoma cell growth or induce ADCC. Related to Figures 3 and 4.

(A) NB-EbC1 GPC2 (left) and GD2 (right) flow cytometry histograms.

(B) SMS-SAN GPC2 (left) and GD2 (right) flow cytometry histograms.

(C, D) NB-EbC1 cell growth after treatment with D3-GPC2-IgG1 as measured by RT-CES (C) or CellTiter-Glo® assay (D).

(E, F) SMS-SAN cell growth after treatment with D3-GPC2-IgG1 as measured by RT-CES (E) or CellTiter-Glo® assay (F).

(G, H) NB-EbC1 cell growth after treatment with D3-GPC2-IgG1 and co-incubation with human NK cells (G) or PBMCs (H).

(I) SMS-SAN cell growth after treatment with D3-GPC2-IgG1 and co-incubation with human NK cells.

(J) NB-EbC1 relative cytotoxicity after treatment with D3-GPC2-IgG1 and co-incubation with human NK cells as measured by an LDH-Glo™ cytotoxicity assay.

(K) Murine neuroblastoma cell line 9464D, isogenic cell line 9464D-Gpc2 and 9464D-Gpc2 allograft Gpc2 flow cytometry histograms.

(L, M) Tumor growth of 9464D-Gpc2 allografts treated with D3-GPC2-IgG1 (n=10 mice/cohort; represented as mean ± SEM; L) and corresponding progression-free survival (PFS; M).

Data in C-J is represented as either mean or mean ± SEM from representative experiments done in at least technical duplicate at least two independent times. ns, not significant; *, $p < 0.05$.

Figure S7

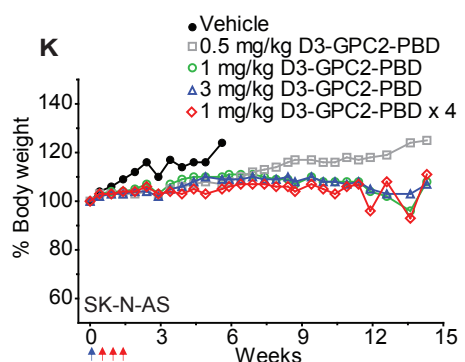
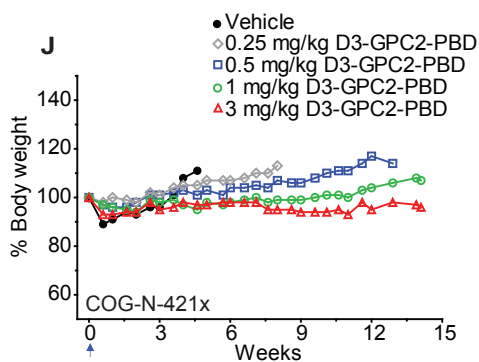
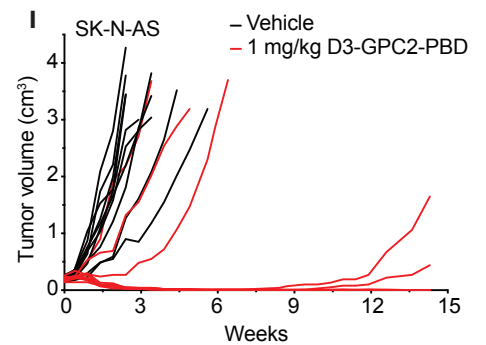
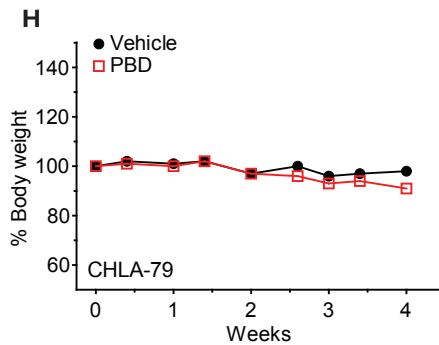
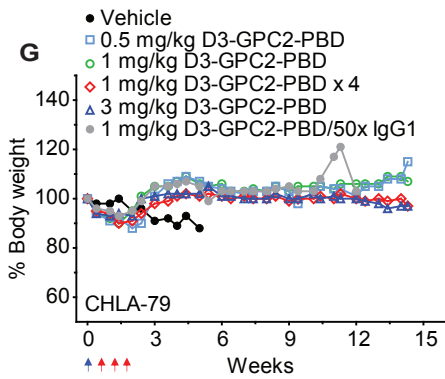
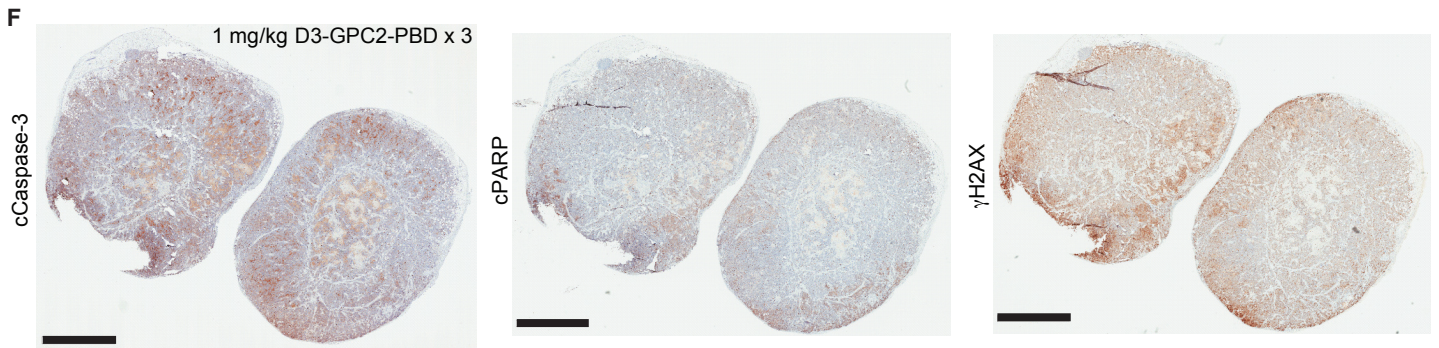
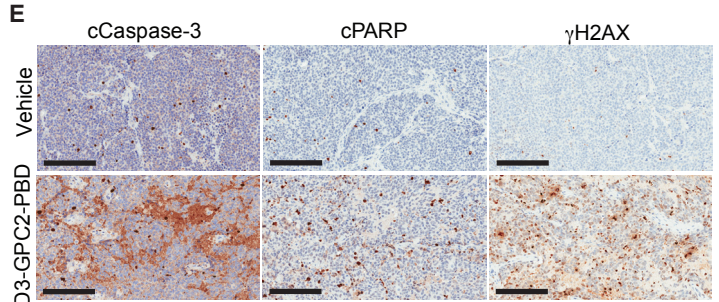
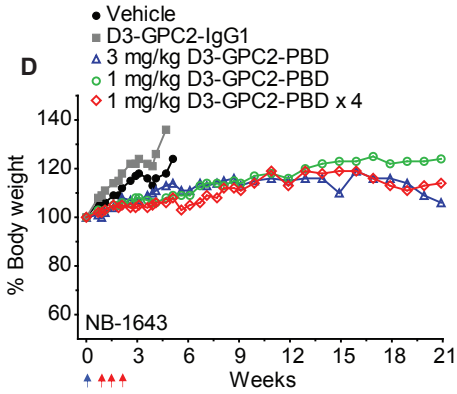
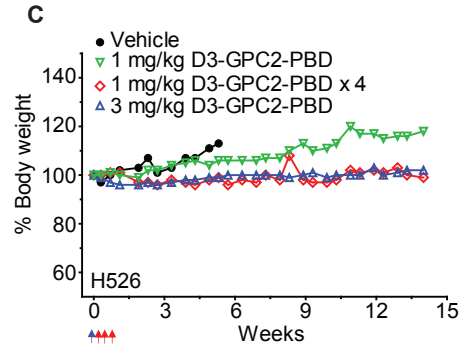
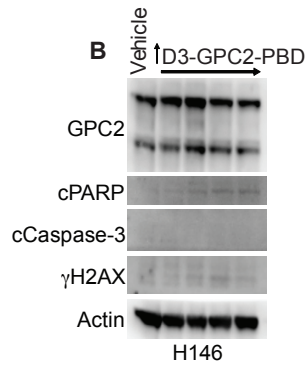
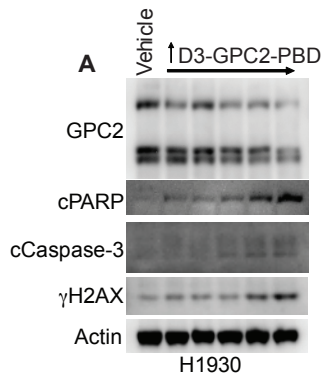


Figure S7. The D3-GPC2-PBD ADC induces diffuse intratumoral DNA damage and apoptosis without any systemic side effects in SCLC and neuroblastoma *in vivo* murine models. Related to Figures 5, 6 and 7.

(A) Western blot of H1930 SCLC cells 3 days after treatment with D3-GPC2-PBD (1.6, 8, 40, 200 and 1000 pM).

(B) Western blot of H146 SCLC cells 3 days after treatment with D3-GPC2-PBD (1.6, 8, 40 and 200 pM).

(C) Mean percent body weights from baseline of mice in the SCLC H526 xenograft treatment arms shown in **Fig. 5E**.

(D) Mean percent body weights from baseline of mice in the NB-1643 PDX treatment arms shown in **Fig. 6A**.

(E) High-power images of cCaspase-3, cPARP and γ H2AX IHC of locally advanced NB-1643 PDX tumors after treatment with 1 mg/kg D3-GPC2-PBD x 3 (ADC dosed on study days 0, 3 and 7 and tumor collected on day 9; also see **Fig. 6C-F**).

(F) Low-power images of cCaspase-3, cPARP and γ H2AX IHC of locally advanced NB-1643 PDX tumors after treatment with 1 mg/kg D3-GPC2-PBD x 3 (ADC dosed on study days 0, 3 and 7 and tumor collected on day 9).

(G) Mean percent body weights from baseline of mice of the CHLA-79 xenograft treatment arms shown in **Fig. 6G**.

(H) Mean percent body weights from baseline of mice in the CHLA-79 xenograft treatment arms shown in **Fig. 6L**.

(I) Individual tumor growth curves for SKNAS xenografts after treatment with vehicle or 1 mg/kg D3-GPC2-PBD ADC. Also shown in summary form in **Fig. 7G**.

(J) Mean percent body weights from baseline of mice in the COG-N-421x PDX treatment arms shown in **Fig. 7A**.

(K) Mean percent body weights from baseline of mice in the SK-N-AS treatment arms shown in **Fig. 7G**.

Scale bars in **E** represent 200 μ M and **F** represent 3 mM. Blue arrows in **C**, **D**, **G**, **J** and **K** represent initial ADC dose and red arrows in **C**, **D**, **G** and **K** indicate subsequent 3 ADC doses for the 1 mg/kg ADC x 4 treatment cohort.

Table S1. Summary of SCLC D3-GPC2-PBD/PBD IC₅₀ data and GPC2/MYC family expression. Related to Figures 1 and 5.

Cell line	D3-GPC2-PBD IC ₅₀ ± SEM (pM)	PBD IC ₅₀ ± SEM (pM)	TP53 mutation ^a	GPC2 RPKM ^a	MYCN RPKM ^a	MYC RPKM ^a	MYCL RPKM ^a
H82	6.9 ± 0.18	27 ± 7.9	T125T ^b	10	0.56	420	12
H209	5.6 ± 0.34	13 ± 4.8	Splice ^b	17	0.05	0.31	153
H526	7.4 ± 0.42	57 ± 9.7	Splice ^b	22	396	0.87	3.8
H146	>5000	163 ± 5.9	PKKKPLD318fs	39	0.39	66	0.60
H1930	1.2 ± 0.21	6.1 ± 2.1	G245R	88	0.67	39	1.0

^aData from the CCLE portal (<https://portals.broadinstitute.org/ccle>). ^bTP53 splice site SNP.

Table S2. Neuroblastoma PDX and cell line xenograft clinical and genomic summary. Related to Figures 2, 6 and 7.

PDX	Sex	Phase of therapy	MYCN	ALK	TP53	Other
NB-1643	Male	Diagnosis	Amp	R1275Q	WT	<i>NF1</i> splice
COG-N-440x	Female	PD-PM	Amp	WT	WT	SLX4 C1426*, <i>CIC</i> del
COG-N-421x	Male	PD-PM	Amp	WT	WT	<i>CDK4</i> amp
COG-N-453x	Male	PD-PM	Amp	F1174L	WT	
COG-N-471x	Female	PD-PM	Amp	WT	WT	
RH07-1x	n/a	n/a	n/a	n/a	n/a	
Xenograft						
SK-N-AS	Female	PD	Non-amp	WT	del ^a	NRAS Q61K
CHLA-79	Female	PD	Non-amp	WT	WT	ERCC2 E606K

^aDeletion of one *TP53* allele and expression of C-terminally truncated *TP53* variants from other *TP53* allele due to a deletion in the intron 9/exon 10 junction, although SKNAS cells may maintain some partial *TP53* functionality. PD, Progressive disease; PD-PM, progressive disease-postmortem; Amp, amplified; Non-amp, non-amplified; WT, wild-type; del, deletion; n/a, not available. Comprehensive genomic profiling data for PDXs/xenografts can be found on the PedcBioPortal for Integrated Childhood Cancer Genomics (<https://pedcbioportal.kidsfirstdrc.org>).

Table S3. Limiting dilution assay growth frequency of SH-SY5Y-Empty and SH-SY5Y-GPC2 isogenic cell dilutions. Related to Figure 2.

Dilution (cells/well) ^a	SH-SY5Y-Empty wells tested	SH-SY5Y-Empty wells with cell growth	SH-SY5Y-GPC2 wells tested	SH-SY5Y-GPC2 wells with cell growth
1000	192	185	192	192
100	192	98	192	150
10	384	192	384	202
1	768	292	768	302

^aTable represents combined data from 2 independent experiments.

Table S4. Estimation of stem cell frequency in SH-SY5Y-Empty and SH-SY5Y-GPC2 isogenic cells. Related to Figure 2.

1/stem cell frequency ^a	Lower	Estimate	Upper
SH-SY5Y-Empty	45.5	39.8	34.8
SH-SY5Y-GPC2	17.4	15.4	13.6

^aCalculated with an extreme limiting dilution analysis.⁴

Table S5. D3-GPC2-Fab/ Δ HS GPC2²⁴⁻⁴⁹³ X-ray data collection and refinement statistics. Related to Figure 3.

Data collection statistics	
Wavelength (Å)	1.033190
Resolution range (Å) ^a	28.57 - 3.30 (3.42 - 3.30)
Space group	P2 ₁
Unit cell	
a, b, c (Å)	54.94 225.65 107.25
α , β , γ (°)	90 104.754 90
Total reflections	217,876 (22,809)
Unique reflections	37,835 (3,807)
Multiplicity	5.8 (6.0)
Completeness (%)	99.8 (99.8)
Mean I/ σ (I)	7.3 (1.8)
Wilson B-factor	79.2
R _{merge} (%) ^b	19.1 (76.2)
R _{pim} (%) ^c	8.7 (33.9)
CC _{1/2} (%)	99.1 (82.6)
Refinement Statistics	
Reflections used in refinement	37,816 (3,798)
Reflections used for R _{free}	1,831 (179)
R _{work} ^d / R _{free} ^e	27.7/ 31.7
Number of non-hydrogen atoms	14,603
Macromolecules	14,603
Protein residues	1,916
RMS _{bonds} (Å)	0.003
RMS _{angles} (°)	0.62
Ramachandran favored (%)	91.9
Ramachandran allowed (%)	7.7
Ramachandran outliers (%)	0.4
Average B-factor (Å ²)	73.9

^aStatistics for the highest-resolution shell are shown in parentheses

$$^b R_{\text{merge}} = \frac{\sum_{\text{hkl}} \sum_i |I_{\text{hkl}, i} - \langle I_{\text{hkl}} \rangle|}{\sum_{\text{hkl}} \langle I_{\text{hkl}} \rangle}$$

$$^c R_{\text{pim}} = \frac{\sum_{\text{hkl}} [1/(N-1)]^{1/2} \sum_i |I_{\text{hkl}, i} - \langle I_{\text{hkl}} \rangle|}{\sum_{\text{hkl}} \langle I_{\text{hkl}} \rangle}$$

$$^d R_{\text{work}} = \frac{(\sum |F_o| - |F_c|)}{(\sum |F_o|)}$$

^e5% of the data were used for R_{free} calculation

Table S6. D3-GPC2-Fab/ Δ HS GPC2²⁴⁻⁴⁹³ interactions. Related to Figure 3.

Δ HS GPC2 ²⁴⁻⁴⁹³ amino acid residue (BSA) ^a	GPC2 AA	Murine Gpc2 AA	GPC1 AA	GPC3 AA	GPC4 AA	GPC5 AA	GPC6 AA	GPC2 mutant	GPC2 mutant K _D (nM) (fold-change K _D from WT Δ HS GPC2 ²⁴⁻⁴⁹³)	Interaction type	D3-GPC2-Fab amino acid residue ^a
Asp91 (14)	D	D	D	R	E	T	E				
<i>Asp91^O</i>										HB	H-Tyr32 ^{OH}
Thr/Ser92 (10)	S	S	S	L	Q	S	T				
<i>Thr/Ser92^O</i>										HB	H-Tyr32 ^{OH}
Phe95 (76)	F	F	V	G	H	T	F	F95V	7.1 (7.6)		
<i>Phe95</i>										VDW	H-Asp31, H-Tyr32, H-Ser96, H-Gly97
His98 (85)	H	H	A	W	A	F	T				
<i>His98^{ND1}</i>										HB	H-Ser96 ^O
<i>His98</i>										VDW	H-Ser96, H-Gly97, H-Tyr98, L-Tyr49
Thr99 (6)	T	T	M	V	V	L	T				
<i>Thr99^{OG1}</i>										HB	H-Gly97 ^O
<i>Thr99</i>										VDW	H-Gly97, H-Tyr98
Ala102 (45)	A	A	T	-	S	R	S				
<i>Ala102</i>										VDW	H-Tyr98, H-Asp99, L-Trp32
Arg103 (6)	R	R	Q	-	R	N	R	R103Q	2.8 (3.0)		
<i>Arg103^{NH1, NH2}</i>										SB	H-Asp99 ^{OD1, OD2}
<i>Arg103</i>										VDW	H-Asp99
Arg105 (50)	R	R	R	T	K	A	K				
<i>Arg105</i>										VDW	H-Tyr98, L-Trp32
Lys106 (85)	K	K	S	P	K	A	K	K106S	6.7 (7.2)		
<i>Lys106^{NZ}</i>										SB	H-Asp99 ^{OD2}
<i>Lys106^{NZ}</i>										HB	L-Leu91 ^O
<i>Lys106</i>										VDW	H-Asp103, L-Trp32, L-Asn92
Glu109 (51)	E	E	D	E	E	E	E				
<i>Glu109^{OE1, OE2}</i>										HB	L-Trp32 ^{NE1}
<i>Glu109</i>										VDW	L-Ser30, L-Trp32
Thr380 (35)	T	T	P	H	T	T	T				
<i>Thr380</i>										VDW	L-Ser28

Thr381 (40)	T	T	-	V	T	R	T					
<i>Thr381^{OG1}</i>											HB	L-Ser28 ^O
<i>Thr381</i>											VDW	L-Ser28
Ala382 (61)	A	A	-	E	A	N	A					
<i>Ala382</i>											VDW	L-Gln27, L-Ser28, L-Asn92
Arg389 (16)	R	R	K	S	R	N	R					
<i>Arg389</i>											VDW	H-Tyr58 L-Tyr94
Trp392 (26)	W	W	S	R	T	K	T					
<i>Trp392</i>											VDW	H-Thr56
Glu396 (101)	E	E	A	Q	D	N	E	E396A	5.5 (5.9)			
<i>Glu396^{OE1, OE2}</i>											HB	H-Tyr33 ^{OH} , H-Ser52 ^{OG} , H-Ser55 ^N
<i>Glu396</i>											VDW	H-Tyr33, H-Ser52, H-Ser52A, H-Ser53, H-Ser55
Arg397 (104)	R	R	Q	K	K	S	K	R397Q	1573 (1688)			
<i>Arg397^{NH1, NH2}</i>											SB	H-Glu95 ^{OE1, OE2} , H-Asp99 ^{OD1}
<i>Arg397^{NH1}</i>											HB	H-Gly97 ^O
<i>Arg397</i>											VDW	H-Tyr33, H-Glu95, H-Gly97, H-Asp99
Arg400 (88)	R	R	D	S	Q	L	L	R400D	26 (28)			
<i>Arg400^{NH2, NE}</i>											SB	H-Asp31 ^{OD1, OD2}
<i>Arg400</i>											VDW	H-Ser30, H-Asp31, H-Ser52A
Thr411 (40)	T	T	T	Y	N	Q	T					
<i>Thr411</i>											VDW	H-Thr28
Asp415 (53)	D	D	E	H	D	N	D					
<i>Asp415</i>											VDW	H-Gly26

^aSuperscript NH1, NH2, NE, OE1, OE2, OG1, OD1, OD2 and O represent the respective atom of amino acid residue involved in interaction. BSA, buried surface area; HB, hydrogen bond; SB, salt bridge; VDW, Van der Waals contact; WT, wild-type; AA, amino acid. The Clustal Omega tool⁵ (https://www.ebi.ac.uk/Tools/services/web_clustal/toolform.ebi) was used to align murine Gpc2 and human GPC1-6 protein sequences. Blue boxes indicate GPC2 identical amino acids for murine Gpc2 and human GPC1 and GPC3-6.

Supplemental References

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